
Sjögren's syndrome: from pathogenesis to novel therapeutic targets

F. Barone¹, S. Colafrancesco^{1,2}

¹Centre for Translational Inflammation Research, Institute of Inflammation and Ageing, University of Birmingham, UK; ²Department of Internal Medicine and Medical Specialties, Rheumatology Unit, Sapienza University of Rome, Italy.

Francesca Barone, MD, PhD
Serena Colafrancesco, MD

Please address correspondence to:
Francesca Barone, MD, PhD,
Centre for Translational Inflammation Research, Institute of Inflammation and Ageing,
University of Birmingham
Research Laboratories,
Queen Elizabeth Hospital,
Birmingham, B15 2WD, UK.
E-mail: f.barone@bham.ac.uk

Received and accepted on July 7, 2016.

Clin Exp Rheumatol 2016; 34 (Suppl. 98): S58-S62.

© Copyright CLINICAL AND EXPERIMENTAL RHEUMATOLOGY 2016.

Key words: primary Sjögren's syndrome, biologics, TLOs, stromal cells, chemokines, cytokines

ABSTRACT

Primary Sjögren's syndrome (pSS) is a chronic inflammatory autoimmune disease, characterised by a chronic infiltration of exocrine glands, mainly salivary glands, with the histological features of focal lymphocytic sialoadenitis. Disease spectrum is broad and the occurrence of several extra-glandular manifestations, and in rare cases lymphoma development, is well known. A specific approved treatment for pSS is still lacking and the detection of novel therapeutic biologic target is ongoing. The identification of biological fingerprints seems essential in order to stratify patients both in clinical trials and in real life. Discovery of new components of the inflammatory response will be the key in the future for the identification of novel additional therapeutic options.

Sjögren's syndrome: clinical and social relevance

Primary Sjögren's syndrome (pSS) is a relatively common chronic inflammatory autoimmune disease, with a higher incidence in female patients (9:1) and a prevalence of ~0.5% in the general population (1, 2). Autoantibody production and chronic infiltration of the exocrine glands, in particular salivary glands, with the histological features of focal lymphocytic sialoadenitis (FLS) represent pSS pathognomonic hallmarks and provide criteria for classification and diagnosis (3, 4). Inflammation results in loss of glandular function and it is responsible for the classical symptoms of dryness, increased incidence of cervical cavities and teeth loss. Ocular involvement is also typical with development of keratoconjunctivitis sicca, often complicated by infections.

The spectrum of pSS extra-glandular manifestations is broad and includes fatigue, vasculitis (leucocytoclastic vasculitis), peripheral neuropathy, joint

involvement characterised by polyarthralgia and in some cases synovitis, kidney involvement with renal tubular acidosis, interstitial lung disease, lymphoproliferative disease and immunological abnormalities (5-7). Approximately 5% of patients with pSS develop lymphoma, conferring a higher mortality risk (8, 9). Histologically, the malignancy is predominantly a non-Hodgkin's lymphoma that forms in extra-nodal sites and mainly within the acquired mucosal associated lymphoid tissue (MALT) harbouring within the affected salivary glands (9, 10). Evolution of MALT into diffuse large B cell lymphoma has been described (9). Systemic manifestations and lymphoma development are most commonly observed in immunologically active patients characterised by B cell hyperactivation, high titers of anti-SSA/Ro and anti-SSB/La autoantibodies and presence of rheumatoid factor (5, 11, 12). PSS represents a significant health and economic burden also in patients that do not develop lymphoma (13-15). Recent data highlight the increased cardiovascular risk (16) and the reduced quality of life associated to pSS (17, 18). Direct health care costs have been estimated at £1,831 to £2,546 per pSS patient per year in the UK, while indirect costs range between £7,677 and £13,502, which is approximately 80% of the costs associated with RA in the era preceding the use of biological therapy. PSS patients are significantly less likely to be in gainful employment, and are more likely to work reduced hours, be in receipt of benefits, or access health care services frequently (13, 14).

One third of pSS patients presents extraglandular manifestations, in rare cases with severe complications. In this cases the use of short courses of steroids might display limited efficacy being often not sufficient to induce and

Competing interests: none declared.

maintain remission. Disease-modifying drugs (DMARDs) can be used in severe organ involvement with variable results. For this reason, as will be further discussed, new hopes have been put in novel biological therapies that target pathways, molecules or cell types involved in disease pathogenesis.

Novel biologics in pSS: targets and challenges in clinical trial design

pSS presents a multifactorial pathogenesis. On the presence of a predisposing genetic background several external factors, mainly viruses, may act as trigger of the disease. In this context, different types of immune system cells and biological molecules provide their contribution in driving and maintaining the inflammatory response. In principle all these pathways could be targeted therapeutically. The role of IFN signature and its over expression along the development of pSS is well known. Despite the lack of clinical trials investigating the utility of anti interferon type I agents in pSS, some evidences supporting the efficacy and the rationale for using these compounds in pSS derives from studies in patients with systemic lupus erythematosus (<https://clinicaltrials.gov>).

Overexpression of several inflammatory cytokines in minor salivary glands has been demonstrated, including TNF α , IL-6, IL-1, IL-18 and IL-22 (19-26). While blocking TNF and IL-1 has been unsuccessful (27) other cytokine blocking or modulations is currently contemplated. Similarly, biological agents capable to interfere with T cells migration are currently under investigation alongside molecules able to interfere with T cell homeostasis or differentiation (<https://clinicaltrials.gov>). Targeting costimulatory molecules such as CTLA-4, ICOS and CD40L with the aim of interfering with the cross talk between T and B cells or T and dendritic cells represents another promising possibility (<https://clinicaltrials.gov>).

In pSS the use of new biological compounds has been hampered by several factors, mainly related to study design, with a key challenge represented by the variety of outcome measures to as-

sess therapeutic efficacy. Available are indexes that reflect systemic involvement, local disease (salivary flow) or a combination of the two (6, 28-31). Whilst no rationale is currently used to allocate specific tools to a population or compound, it is preferred to recruit into trials patients characterised by moderate to significant systemic involvement according to the ESSDAI, a composite score of disease activity (28). Unfortunately, currently available biological compounds, failed to demonstrate significant success in terms of ESSDAI changes in randomised clinical trials, inducing a general reflection on the ability of the clinicians to use this complex tool, the sensitivity of the index to detect changes in a short period of time and to discriminate between active arm and placebo.

An additional challenge faced when designing pSS trials is represented by the difficulties in the selection of the target population. Given the nature of the ESSDAI, used as entry criteria in a significant number of trials, the recruited population might comprise a rather heterogeneous spectrum of patients, only aligned by the common trait of B cell hyper-activation. While extremely broad in terms of clinical manifestations, this population is, however, relatively small when compared to the majority of the pSS patients, that display limited systemic involvement and are mainly characterised by dryness (32). These considerations raise ethical and practical issues when looking at the broader picture of pSS therapy.

Process driven stratification in pSS

There is a general consensus that strategies should be implemented to stratify patients and recruit into clinical trial patients identified by specific biological fingerprints. In this context, research for serum, saliva and tissue biomarkers has been implemented in several trials with the aim to stratify patients, predict and monitor response to treatment.

Baseline stratification according to biological fingerprints and correlation with clinical phenotype is also pursued. It has been recently shown that immunophenotyping of blood as well as tis-

sue isolated cells can be used to stratify patients in clusters defined by different degree of disease activity and level of glandular inflammation (33).

Changes occurring in different biological pathways in response to therapy have been only recently investigated in pSS. Using transcriptomic analysis in pre and post treatment samples from patients undergoing treatment with Rituximab differential expression of genes belonging to the IFN pathway between responders and non-responders has been demonstrated (34). Similarly, histology based stratification has been recently used in the context of clinical trials as predictor of response with contrasting results (35-38).

Several trial protocols have been recently implemented to encompass the histological analysis of salivary gland biopsies and include detailed measurements that capture changes in infiltrate size and degree of organisation, presence of germinal centres and, in selected cases transcriptomic analysis. Whilst providing a biological outcome measure of drug efficacy, the possibility to use these data in retrospective analysis to stratify responders to treatment is also considered.

Targeting stromal cells in tertiary lymphoid structures: a new therapeutic approach in pSS

The association between the degree of organisation of the salivary glands infiltrate in pSS and seric and clinical features has been clearly shown. The ectopic lymphocytic aggregates, correctly defined as tertiary lymphoid structures (TLS), have been classically associated with negative disease prognosis and lymphoma development (39, 40). Local production of autoantibodies and clonal B cell expansion (41, 42) has been also observed within fully formed TLS, thus supporting the direct pathogenic role of those structures and the rationale to target TLS formation therapeutically.

B cell targeting is, in this context, expected to modify the degree of TLS formation and interfere with the functional ability of TLS to sustain disease progression. Interestingly, this appears not to be the case. Despite the strong

rational supporting the use of B cell depleting agents in pSS, early results from randomised clinical trials using Rituximab are conflicting. Resistance to B cell depletion and loss of clinical response appears to correlate with systemic and local rebound of the levels of the B cell survival factor BAFF. This, in turn supports the homeostatic expansion of pathogenic B cell clones in the periphery and in the salivary gland during the phase of repopulation (43-45). Novel strategies aimed to overcome this problem and improve B cell targeting are under consideration and involve either combination therapies with anti BAFF neutralising agents or novel compounds aimed to broadly interfere with immune cells intracellular signals (clinicaltrials.gov).

We and others demonstrated that the activation of resident tissue stromal cells is a cardinal feature of pSS. TLS aggregates in patients with PSS contain networks of Podoplanin/gp38+ stromal cells and networks of follicular dendritic cells, whose organisation closely resemble the stroma compartment that support secondary lymphoid organs (SLOs) (46). These non-haematopoietic stromal cells are increasingly recognised as essential counterparts to leucocytes in pathogenicity.

In SLOs, the stromal cell compartment provides the scaffold that enable leucocytes migration and interaction, alongside survival and homeostatic factors required to sustain the haematopoietic cells. More recently, stromal cell have been demonstrated able to influence the size and shape of the immune cell repertoire by modulating the availability of lymphocyte survival factors and inducing deletion or expansion of auto-reactive cell clones. This central role in balancing immune stimulation *versus* peripheral tolerance is achieved by the ability of stromal cells in the lymph nodes to present a range of peripheral tissue restricted antigens and limit T cell expansion and priming through a series of mechanisms, among which the release of nitric oxide (47-56). TLS stromal cells also sustain cell migration, activation and survival of the immune compartment in persistent inflammatory conditions, likely ena-

bling disease persistence, even when lymphocytes have been depleted (46). Within TLS, persistent antigenic stimulation and presence of pro-inflammatory cytokines is responsible for the conversion of stromal cells into a lymphoid tissue-like cell phenotype. Similarly cytokines and genetic predisposition influence the epithelial compartment to contribute to disease establishment in pSS. It is well known that areas of "lymphoepithelial proliferation" or LESA represent a pathogenic histological element in the process of lymphomagenesis. The close cellular introduction between pathogenic nursing epithelial cells able to provide chemoattractive (57) and survival factors (45, 58) and the aberrant B cell clones often characterised by rheumatoid factor activity is understood to play a key role in the establishment of autoimmune associated MALT (59).

Resident stromal cells, including epithelial cells, fibroblasts, endothelium and lymphatic cells are therefore responsible for establishing the chemokines and survival factors gradients that enable migration and organisation of the pathogenic clones within the glands. CXCL13, major B cell chemoattractive factor, ligand for CXCR5, is preferentially expressed within the inner part of the aggregates and in the germinal centres by activated fibroblasts, follicular dendritic cells and few activated T cells (19, 60, 61). Its expression is regulated by lymphotoxin, TNF (62, 63) and, as recently described by our group, by proinflammatory cytokines such as IL-22 (19). The areas characterised by malignant B cell infiltration display, on the contrary, preferential expression of CXCL12 (57). Interestingly, IL-22, that is responsible for CXCL13 expression by resident activated fibroblasts, induces on epithelial cells the expression of CXCL12, thus suggesting differential regulation of the fibroblasts and epithelial compartments in the context of chronic inflammation and TLS establishment. Abrogation of the IL-22 pathway by genetic modification and therapeutic intervention leads to TLO disaggregation and loss of autoantibody production (19). These data sug-

gest the exciting prospective of targeting the pathogenic microenvironment to affect the survival and migration of the haematopoietic component.

Conclusions

To date there is no approved, specific treatment for pSS. Patients are managed with a combination of immunosuppressive drugs and, in some cases, systemic disease is treated with steroids. Efforts to identify biological fingerprints are ongoing, favoured by international initiatives and collaborative efforts, such as the EULAR endorsed Study Group for Sjögren's syndrome (www.eular.org/myUploadData/files/Investigative_Study_Group_Sjogren.pdf) with the aim to design algorithms for process driven stratification and apply precision medicine to pSS. Alongside this critical efforts are aimed to define the role of undervalued components of the inflammatory response and will provide, in the next future additional and exciting therapeutic opportunities for this orphan disease.

References

1. BRENNAN MT, FOX PC: Sex differences in primary Sjögren's syndrome. *J Rheumatol* 1999; 26: 2373-6.
2. BOWMAN SJ, PILLEMER S, JONSSON R *et al.*: Revisiting Sjögren's syndrome in the new millennium: perspectives on assessment and outcome measures. Report of a workshop held on 23 March 2000 at Oxford, UK. *Rheumatology* 2001; 40: 1180-8.
3. FOX RI, TORNWALL J, MICHELSON P: Current issues in the diagnosis and treatment of Sjögren's syndrome. *Curr Opin Rheumatol* 1999; 11: 364-71.
4. VITALI C, BOMBARDIERI S, JONSSON R *et al.*: Classification criteria for Sjögren's syndrome: a revised version of the European criteria proposed by the American-European Consensus Group. *Ann Rheum Dis* 2002; 61: 554-8.
5. ANAYA JM, DELGADO-VEGA AM, CASTIBLANCO J: Genetic basis of Sjögren's syndrome. How strong is the evidence? *Clin Dev Immunol* 2006; 13: 209-22.
6. RAMOS-CASALS M, BRITO-ZERON P, SEROR R *et al.*: Characterization of systemic disease in primary Sjögren's syndrome: EULAR-SS Task Force recommendations for articular, cutaneous, pulmonary and renal involvements. *Rheumatology* 2015; 54: 2230-8.
7. GONZALEZ S, SUNG H, SEPULVEDA D, GONZALEZ M, MOLINA C: Oral manifestations and their treatment in Sjögren's syndrome. *Oral Dis* 2014; 20: 153-61.
8. BRITO-ZERON P, RAMOS-CASALS M: [Prognosis of patients with primary Sjögren's syn-

- drome]. *Med Clin (Barc.)* 2008; 130: 109-115.
9. THEANDER E, HENRIKSSON G, LJUNGBERG O, MANDL T, MANTHORPE R, JACOBSSON LT: Lymphoma and other malignancies in primary Sjögren's syndrome: a cohort study on cancer incidence and lymphoma predictors. *Ann Rheum Dis* 2006; 65: 796-803.
 10. VOULGARELIS M, DAFNI UG, ISENBERG DA, MOUTSOPOULOS HM: Malignant lymphoma in primary Sjögren's syndrome: a multicenter, retrospective, clinical study by the European Concerted Action on Sjögren's Syndrome. *Arthritis Rheum* 1999; 42: 1765-72.
 11. ANAYA JM, TOBON GJ, VEGA P, CASTIBLANCO J: Autoimmune disease aggregation in families with primary Sjögren's syndrome. *J Rheumatol* 2006; 33: 2227-34.
 12. BRITO-ZERON P, SORIA N, MUNOZ S *et al.*: Prevalence and clinical relevance of autoimmune neutropenia in patients with primary Sjögren's syndrome. *Semin Arthritis Rheum* 2009; 38: 389-95.
 13. CALLAGHAN R, PRABU A, ALLAN RB *et al.*: Direct healthcare costs and predictors of costs in patients with primary Sjögren's syndrome. *Rheumatology* 2007; 46: 105-11.
 14. BOWMAN SJ, ST PIERRE Y, SUTCLIFFE N *et al.*: Estimating indirect costs in primary Sjögren's syndrome. *J Rheumatol* 2010; 37: 1010-5.
 15. LENDREM D, MITCHELL S, MCMEEKIN P *et al.*: Health-related utility values of patients with primary Sjögren's syndrome and its predictors. *Ann Rheum Dis* 2014; 73: 1362-8.
 16. JUAREZ M, TOMS TE, DE PABLO P *et al.*: Cardiovascular risk factors in women with primary Sjögren's syndrome: United Kingdom primary Sjögren's syndrome registry results. *Arthritis Care Res* 2014; 66: 757-64.
 17. KOTSIS K, VOULGARIS PV, TSIFETAKI N, DROSOS AA, CARVALHO AF, HYPHANTIS T: Illness perceptions and psychological distress associated with physical health-related quality of life in primary Sjögren's syndrome compared to systemic lupus erythematosus and rheumatoid arthritis. *Rheumatol Int* 2014; 34: 1671-81.
 18. LENDREM D, MITCHELL S, MCMEEKIN P *et al.*: Health-related utility values of patients with primary Sjögren's syndrome and its predictors. *Ann Rheum Dis* 2014; 73: 1362-8.
 19. BARONE F, NAYAR S, CAMPOS J *et al.*: IL-22 regulates lymphoid chemokine production and assembly of tertiary lymphoid organs. *Proc Natl Acad Sci USA* 2015; 112: 11024-9.
 20. CICCIA F, GUGGINO G, RIZZO A *et al.*: Potential involvement of IL-22 and IL-22-producing cells in the inflamed salivary glands of patients with Sjögren's syndrome. *Ann Rheum Dis* 2012; 71: 295-301.
 21. BOMBARDIERI M, MCINNES IB, PITZALIS C: Interleukin-18 as a potential therapeutic target in chronic autoimmune/inflammatory conditions. *Expert Opin Biol Ther* 2007; 7: 31-40.
 22. BOMBARDIERI M, BARONE F, PITTONI V *et al.*: Increased circulating levels and salivary gland expression of interleukin-18 in patients with Sjögren's syndrome: relationship with autoantibody production and lymphoid organization of the periductal inflammatory infiltrate. *Arthritis Res Ther* 2004; 6: R447-456.
 23. SADA PR, ISENBERG D, CIURTIN C: Biologic treatment in Sjögren's syndrome. *Rheumatology* 2015; 54: 219-230.
 24. TISHLER M, YARON I, GEYER O, SHIRAZI I, NAFTALIEV E, YARON M: Elevated tear interleukin-6 levels in patients with Sjögren syndrome. *Ophthalmology* 1998; 105: 2327-9.
 25. GARCIC-CARRASCO M, FONT J, FILELLA X *et al.*: Circulating levels of Th1/Th2 cytokines in patients with primary Sjögren's syndrome: correlation with clinical and immunological features. *Clin Exp Rheumatol* 2001; 19: 411-5.
 26. BOUMBA D, SKOPOULI FN, MOUTSOPOULOS HM: Cytokine mRNA expression in the labial salivary gland tissues from patients with primary Sjögren's syndrome. *Br J Rheumatol* 1995; 34: 326-33.
 27. MOUTSOPOULOS NM, KATSIFIS GE, ANGELOV N *et al.*: Lack of efficacy of etanercept in Sjögren's syndrome correlates with failed suppression of tumour necrosis factor alpha and systemic immune activation. *Ann Rheum Dis* 2008; 67: 1437-43.
 28. SEROR R, RAVAUD P, BOWMAN SJ *et al.*: EULAR Sjögren's syndrome disease activity index: development of a consensus systemic disease activity index for primary Sjögren's syndrome. *Ann Rheum Dis* 2010; 69: 1103-9.
 29. SEROR R, BOOTSMA H, SARAUX A *et al.*: Defining disease activity states and clinically meaningful improvement in primary Sjögren's syndrome with EULAR primary Sjögren's syndrome disease activity (ESSDAI) and patient-reported indexes (ESSPRI). *Ann Rheum Dis* 2016; 75: 382-9.
 30. SEROR R, BOWMAN SJ, BRITO-ZERON P *et al.*: EULAR Sjögren's syndrome disease activity index (ESSDAI): a user guide. *RMD Open*. 2015; 1(1):e000022.
 31. CHO HJ, YOO JJ, YUN CY, KANG EH, LEE HJ, HYON JY *et al.*: The EULAR Sjögren's syndrome patient reported index as an independent determinant of health-related quality of life in primary Sjögren's syndrome patients: in comparison with non-Sjögren's sicca patients. *Rheumatology (Oxford)*. 2013; 52: 2208-17.
 32. ONI C, MITCHELL S, JAMES K *et al.*: Eligibility for clinical trials in primary Sjögren's syndrome: lessons from the UK Primary Sjögren's Syndrome Registry. *Rheumatology* 2016; 55: 544-52.
 33. MINGUENEAU M, BOUDAUD S, HASKETT S *et al.*: Cytometry by time-of-flight immunophenotyping identifies a blood Sjögren's signature correlating with disease activity and glandular inflammation. *J Allergy Clin Immunol* 2016; 137: 1809-21.
 34. DEVAUCHELLE-PENSEC V, CAGNARD N, PERS JO, YOUINOU P, SARAUX A, CHIOCHIA G: Gene expression profile in the salivary glands of primary Sjögren's syndrome patients before and after treatment with rituximab. *Arthritis Rheum* 2010; 62: 2262-71.
 35. DELLI K, HAACKE EA, IHRLER S *et al.*: Need for consensus guidelines to standardise the assessment of germinal centres and other histopathological parameters in salivary gland tissue of patients with primary Sjögren's syndrome. *Ann Rheum Dis* 2016; 75: e32.
 36. DELLI K, HAACKE EA, KROESE FG *et al.*: In primary Sjögren's syndrome high absolute numbers and proportions of B cells in parotid glands predict responsiveness to rituximab as defined by ESSDAI, but not by SSRI. *Ann Rheum Dis* 2016; 75: e34.
 37. DELLI K, HAACKE EA, KROESE FG *et al.*: Towards personalised treatment in primary Sjögren's syndrome: baseline parotid histopathology predicts responsiveness to rituximab treatment. *Ann Rheum Dis* 2016 Jan 12. [Epub ahead of print].
 38. CORNEC D, COSTA S, DEVAUCHELLE-PENSEC V, CHICHE L, SARAUX A, PERS JO: Do high numbers of salivary gland-infiltrating B cells predict better or worse outcomes after rituximab in patients with primary Sjögren's syndrome? *Ann Rheum Dis* 2016; 75: e33.
 39. THEANDER E, VASAITIS L, BAECKLUNDE E *et al.*: Lymphoid organisation in labial salivary gland biopsies is a possible predictor for the development of malignant lymphoma in primary Sjögren's syndrome. *Ann Rheum Dis* 2011; 70: 1363-8.
 40. RISSELADA AP, LOOIJE MF, KRUIZE AA, BIJLSMA JW, VAN ROON JA: The role of ectopic germinal centers in the immunopathology of primary Sjögren's syndrome: a systematic review. *Semin Arthritis Rheum* 2013; 42: 368-76.
 41. BOMBARDIERI M, BARONE F, HUMBY F *et al.*: Activation-induced cytidine deaminase expression in follicular dendritic cell networks and interfollicular large B cells supports functionality of ectopic lymphoid neogenesis in autoimmune sialoadenitis and MALT lymphoma in Sjögren's syndrome. *J Immunol* 2007; 179: 4929-38.
 42. HUMBY F, BOMBARDIERI M, MANZO A *et al.*: Ectopic lymphoid structures support ongoing production of class-switched autoantibodies in rheumatoid synovium. *PLoS Med* 2009; 6(1): e1.
 43. DE VITA S, QUARTUCCIO L, SALVIN S *et al.*: Sequential therapy with belimumab followed by rituximab in Sjögren's syndrome associated with B-cell lymphoproliferation and overexpression of BAFF: evidence for long-term efficacy. *Clin Exp Rheumatol* 2014; 32: 490-4.
 44. QUARTUCCIO L, FABRIS M, SALVIN S, MASET M, DE MARCHI G, DE VITA S: Controversies on rituximab therapy in sjogren syndrome-associated lymphoproliferation. *Int J Rheumatol* 2009; 2009: 424935.
 45. QUARTUCCIO L, FABRIS M, MORETTI M *et al.*: Resistance to rituximab therapy and local BAFF overexpression in Sjögren's syndrome-related myoepithelial sialadenitis and low-grade parotid B-cell lymphoma. *Open Rheumatol J* 2008; 2: 38-43.
 46. BUCKLEY CD, BARONE F, NAYAR S, BEN-EZECH C, CAAMANO J: Stromal cells in chronic inflammation and tertiary lymphoid organ formation. *Annu Rev Immunol* 2015; 33: 715-45.
 47. FLETCHER AL, ACTON SE, KNOBLICH K: Lymph node fibroblastic reticular cells in health and disease. *Nat Rev Immunol* 2015; 15: 350-61.

48. DENTON AE, ROBERTS EW, LINTERMAN MA, FEARON DT: Fibroblastic reticular cells of the lymph node are required for retention of resting but not activated CD8+ T cells. *Proc Natl Acad Sci USA* 2014; 111: 12139-44.
49. CREMASCO V, WOODRUFF MC, ONDER L *et al.*: B cell homeostasis and follicle confines are governed by fibroblastic reticular cells. *Nat Immunol* 2014; 15: 973-81.
50. ACTON SE, FARRUGIA AJ, ASTARITA JL *et al.*: Dendritic cells control fibroblastic reticular network tension and lymph node expansion. *Nature* 2014; 514: 498-502.
51. SIEGERT S, LUTHER SA: Positive and negative regulation of T cell responses by fibroblastic reticular cells within paracortical regions of lymph nodes. *Front Immunol* 2012; 3: 285.
52. SIEGERT S, HUANG HY, YANG CY *et al.*: Fibroblastic reticular cells from lymph nodes attenuate T cell expansion by producing nitric oxide. *PloS One* 2011; 6: e27618.
53. LUKACS-KORNEK V, MALHOTRA D, FLETCHER AL *et al.*: Regulated release of nitric oxide by nonhematopoietic stroma controls expansion of the activated T cell pool in lymph nodes. *Nat Immunol* 2011; 12: 1096-104.
54. FLETCHER AL, LUKACS-KORNEK V, REYNOSO ED *et al.*: Lymph node fibroblastic reticular cells directly present peripheral tissue antigen under steady-state and inflammatory conditions. *J Exp Med* 2010; 207: 689-97.
55. VEGA F, COOMBES KR, THOMAZY VA, PATEL K, LANG W, JONES D: Tissue-specific function of lymph node fibroblastic reticulum cells. *Pathobiology* 2006; 73: 71-81.
56. KATAKAI T, HARA T, SUGAI M, GONDA H, SHIMIZU A: Lymph node fibroblastic reticular cells construct the stromal reticulum via contact with lymphocytes. *J Exp Med* 2004; 200: 783-95.
57. BARONE F, BOMBARDIERI M, ROSADO MM *et al.*: CXCL13, CCL21, and CXCL12 expression in salivary glands of patients with Sjögren's syndrome and MALT lymphoma: association with reactive and malignant areas of lymphoid organization. *J Immunol* 2008; 180: 5130-40.
58. PERS JO, DEVAUCHELLE V, DARIDON C *et al.*: BAFF-modulated repopulation of B lymphocytes in the blood and salivary glands of rituximab-treated patients with Sjögren's syndrome. *Arthritis Rheum* 2007; 56: 1464-77.
59. NOCTURNE G, MARIETTE X: Sjögren's Syndrome-associated lymphomas: an update on pathogenesis and management. *Br J Haematol* 2015; 168: 317-27.
60. BARONE F, BOMBARDIERI M, MANZO A *et al.*: Association of CXCL13 and CCL21 expression with the progressive organization of lymphoid-like structures in Sjögren's syndrome. *Arthritis Rheum* 2005; 52: 1773-84.
61. AMFT N, CURNOW SJ, SCHEEL-TOELLNER D *et al.*: Ectopic expression of the B cell-attracting chemokine BCA-1 (CXCL13) on endothelial cells and within lymphoid follicles contributes to the establishment of germinal center-like structures in Sjögren's syndrome. *Arthritis Rheum* 2001; 44: 2633-41.
62. NGO VN, KORNER H, GUNN MD *et al.*: Lymphotoxin alpha/beta and tumor necrosis factor are required for stromal cell expression of homing chemokines in B and T cell areas of the spleen. *J Exp Med* 1999; 189: 403-12.
63. ANSEL KM, NGO VN, HYMAN PL *et al.*: A chemokine-driven positive feedback loop organizes lymphoid follicles. *Nature* 2000; 406: 309-14.