P-1

Utility of testing for murine tissue specific autoantibodies for the diagnosis of Sjögren's syndrome

Frederick B Vivino<sup>1</sup>, Michael George<sup>1</sup>, Chadwick Johr<sup>1</sup>, Vatinee Bunya<sup>1</sup>, Giacomina Massaro-Giordano<sup>1</sup>, Brandon Eilberg<sup>1</sup>, Lakshmanan Suresh<sup>2</sup>, Long Shen<sup>2</sup>.

<sup>1</sup>University of Pennsylvania. <sup>2</sup>Trinity Biotech, Inc.

**Background.** A group of murine parotid tissue specific autoantibodies (**TSAs**) which includes anti-SP1 (salivary protein 1), anti-PSP (parotid specific protein) and anti-CA6 (carbonic anhydrase) are markers for early disease in the IL-14 $\alpha$  transgenic mouse model of SS. These TSAs are also found in NOD mice and patients (**pts**) classified with SSA+ or - Sjögren's according to the American European Consensus Group criteria.

**Methods.** We tested serum for TSAs from 6 pts groups followed in a rheumatology clinic for >1 year at a university medical center including: 1) SS who met published classification criteria (n=145), 2) non-autoimmune controls (n=32), 3) SLE (n=18), 4) RA (n=14), 5) scleroderma (n=4) and 6) chronic nonspecific sialadenitis (n=15). Saliva samples were also obtained from groups 1,2 & 6. Electronic medical records were reviewed to verify diagnoses & all pts were questioned re: the presence & duration of dry eyes/mouth & medical history. Volunteers with history of dry eyes/mouth, any autoimmune diseases or family history of autoimmune disease were excluded as controls. Serum samples were anonymously coded & assayed by ELISA (Trinity Biotech, Inc., Buffalo, NY). All laboratory personnel were blinded to pts diagnoses. Analyses was performed to determine the sensitivity, specificity & discriminative ability of the presence of  $\geq$ 1 TSAs to differentiate SS from other groups.

**Results.** Of the 145 SS pts, 133 (92%) had disease duration > 3 years. TSAs were detected in both the serum and saliva of pts with SS. Most frequently detected TSA in SS was anti-CA6 IgM (14.5%) (Table I). The presence of ≥1 TSA was not significantly different in each group: SS (43%), controls (59%), chronic sialadenitis (40%) and other connective tissues diseases (33%). No particular TSA or isotype was specific for SS. Results suggested a sensitivity and specificity of 44% and 41% respectively for the presence of ≥1 TSAs in SS. Prevalence of + ANAs/ RF IgM in each group were as follows: SS (70%/ 45%), controls (30%/41%), chronic sialoadenitis (43%/33%) and other connective tissue diseases (69%/47%) (Table II). Among controls the presence ≥1 TSAs did not significantly vary between ANA+ vs. ANA- individuals. **Conclusions.** The presence of  $\geq 1$  TSAs in the serum does not distinguish between established SS and other patient groups. The value of this assay for confirmation of early or undiagnosed SS (<3 years) remains unclear. Further studies in larger pts groups including a prospective study of TSAs in pts with early sicca symptoms are needed. Assay of TSAs in saliva or calculation of a saliva/serum TSA ratio may prove to be a more valuable diagnostic test.

### P-2

## Utility of novel autoantibodies in the diagnosis of Sjögren's syndrome among patients with dry eye

Sezen Karakus, MD<sup>1</sup>; Alan N. Baer, MD<sup>2</sup>; Devika Agrawal, BS<sup>1</sup>; Merve Gurakar, BS<sup>1,3</sup>; Robert W. Massof, PhD<sup>1</sup>; Esen K. Akpek, MD<sup>1</sup>.

<sup>1</sup>Ocular Surface Diseases and Dry eye Clinic, The Wilmer Eye Institute, Johns Hopkins University, Baltimore, Maryland, USA. <sup>2</sup>The Johns Hopkins Jerome L. Greene Sjögren's Syndrome Center, Baltimore, MD, USA. <sup>3</sup>Virginia Commonwealth University School of Medicine, Richmond, VA, USA.

**Background.** About one in 10 patients with clinically significant dry eye have underlying Sjögren's syndrome (SS). Unfortunately, SS is greatly underdiagnosed in clinical practice, not only due to its diverse symptomatic expression but also the difficulty with serological testing. Salivary protein 1 (SP1), carbonic anhydrase 6 (CA6), and parotid secretory protein (PSP) have been reported as useful markers to detect SS at an early stage. We aimed to investigate the diagnostic value of these novel autoantibodies in comparison with the traditional serological markers to detect SS among a small sample of dry eye patients.

Methods. Forty-six dry eye patients with SS (SS-dry eye), 14 dry eye patients without SS (non-SS dry eye), and 25 controls over the age of 18 were included. The 2012 American College of Rheumatology classification criteria were used for SS diagnosis. After a detailed review of systems, Ocular Surface Disease Index questionnaire, Schirmer's test without anesthesia, tear film break-up time, and ocular surface staining were performed to assess dry eye. All participants underwent serological testing using a commercially available finger prick kit.

**Results.** Thirty-seven patients with SS (80.4%) had a positive traditional antibody and 28 (60.9%) had a positive novel autoantibody. Traditional autoantibodies were absent in all non-SS dry eye patients and controls. Novel antibodies were present in 7/14 (50%) non-SS dry eye patients and 4/25 (16%) controls. Among three novel autoantibodies, anti-CA6 was significantly more prevalent in the SS and non-SS dry eye groups compared to controls (52.2% vs. 42.9% vs. 8.0%, p=0.001). Dry eye patients with positive anti-CA6 alone were significantly younger than patients with only traditional autoantibodies (43.3 vs 57.9 years, p=0.02). Anti-CA6 was associated with worse dry eye signs and symptoms.

**Conclusions.** Diagnostic criteria for SS are continually being updated as we learn more about the pathophysiology. It is still not clear whether dry eye patients with significant signs and symptoms but no positive serology or biopsy represent a separate entity or early stages of SS. We suggest that adding novel antibodies to the serologic testing, particularly anti-CA6, can improve our understanding to determine who needs close monitoring during the disease process.

## **P-3**

#### Tissue specific autoantibodies significantly improve diagnosis of primary Sjögren's syndrome and show a mild clinical course

Yuebo Jin<sup>1</sup>, Jing Li<sup>1</sup> Jiali Chen<sup>1</sup>, Ruijun Zhang<sup>1</sup>, Zhanguo, Li<sup>1</sup> and Jing He<sup>1</sup>. <sup>1</sup>Department of Rheumatology and Immunology, Peking University People's Hospital, Beijing, China.

**Background.** The presence of autoantibodies is one of several hallmarks of Sjögren's Syndrome, the detection of serum autoantibodies has a central role in the diagnosis and classification of Sjögren's syndrome. Research on correlations between serum autoantibodies and systemic damages of pSS would be beneficial to the clinical assessment and mechanism study. A group of tissue specific autoantibodies (TSAs, which includes anti-CA6, anti-SP1 and anti-PSP autoantibodies), was first identified in mouse models for SS, then confirmed in the pSS Patients. Previous studies have showed these TSAs may be linked directly to local tissue injury during the early stages of SS development. In current study, a large cohort of SS patients was recruited to address the clinical correlations of these TSAs with clinical symptoms and systemic damage during the disease progression.

**Methods.** Serum samples were collected from 316 pSS patients (28 male and 288 female) who meet the 2012 ACR Classification Criteria for Sjögren's Syndrome. SSA and TSAs autoantibodies level was examined by ELISA assays (Trinity Biotech, Buffalo, NY). All clinical and laboratory data were reviewed following protocol approved by Peking University People's Hospital IRB committee. Change of the autoantibody profiles/titers was further evaluated in 61 pSS patients between onset of the disease and 2 years follow up. Chi-square test was used to compare prevalence rate between different groups.

#### **Results.**

- In general, 70% of pSS patients were positive for SSA autoantibodies and 46% of them were positive for TSAs. When combined both SSA and TSAs, the sensitivity for detecting serum autoantibodies in pSS patients was increased from 70% to 93% (p<0.0001).</li>
- 2. The prevalence of SSA autoantibodies in pSS patients with different disease duration was 68.4% (<5 years), 80% (5-10 years, p=0.421) and 90.9% (>10 years, p=0.073) whereas the prevalence of TSAs positive (and SSA-) was 26.4% (<5 years), 10% (5-10 years, p=0.194) and 4.5% (>10 years, p=0.051).
- 3. The prevalence of SSA autoantibodies in pSS patients with different disease activities (as measured by ESSDAI score) was 55% (score 1-3), 92.6% (score 4-6, p=0.002) and 100% (score 7-10, p=0.002) whereas the prevalence of TSAs positive (and SSA-) was 30% (score 1-3), 7.4% (score 4-6, p=0.032) and 0.00% (score 7-10, p=0.019)
- Conclusions.
- In SS patients whose SSA antibody was negative, tissue specific autoantibodies can significantly improve sensitivity, which may help to recognize atypical pSS patients.
- 2. There appears to be a milder clinical course in patients presenting with tissue specific autoantibodies and SSA negative patients. TSAs might be related with the pathogenesis during the onset of the disease.

### **P-4**

Prevalence of novel candidate Sjögren's syndrome antibodies in the Dry Eye Assessment and Management (DREAM®) study

Vatinee Y. Bunya<sup>1</sup>, Gui-shuang Ying PhD<sup>1</sup>, Maureen G. Maguire PhD<sup>1</sup>, Eric Kuklinski BA<sup>2</sup>, Ellen Peskin MA CCRP<sup>1</sup>, Meng C. Lin OD PhD<sup>3</sup> and Penny A. Asbell MD<sup>2</sup>.

<sup>1</sup>Department of Ophthalmology, Scheie Eye Institute, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, Pennsylvania. <sup>2</sup>Department of Ophthalmology, Icahn School of Medicine at Mount Sinai, New York, New York. <sup>3</sup>UC Berkeley, School of Optometry, Berkeley, California.

**Background.** The novel autoantibodies salivary protein 1 (SP-1), carbonic anhydrase 6 (CA-6), and parotid secretory protein (PSP) have been described in a mouse model for Sjögren's syndrome (SS), but have not been well-characterized in dry eye disease (DED) patients, with or without SS. We evaluated the prevalence of these novel candidate SS antibodies at baseline among patients in the DRy Eye Assessment and Management (DREAM<sup>®</sup>) Study, a clinical trial designed to evaluate the effectiveness of omega-3 fatty acid supplements for the treatment of DED.

**Methods.** Baseline medical history questionnaire responses were used to categorize DREAM<sup>®</sup> patients into 3 groups: 1) no history of SS or other autoimmune disease (n=375); 2) no history of SS but with a history of other autoimmune disease (n=66); and 3) a history of SS (n=53). Ocular surface exams and serological testing for traditional and novel SS antibodies was performed. The chi-square test or Fisher exact test (count <5) was used to compare the antibody prevalence rate among the 3 groups and between Groups 1 and 3.

**Results.** Among 494 DREAM<sup>®</sup> participants, 53 (10.7%) had a history of SS and had a significantly higher prevalence of the traditional SS antibodies compared to the other two groups (Table I). The prevalence of each of the novel antibodies was similar between Group 1 (no SS) and Group 3 (SS) in that in each group approximately 20% had SP-1 antibodies (p=0.85); approximately 16% had anti-CA-6 antibodies (p=0.84), and 10% had anti-PSP antibodies (p=1.00). Participants positive for both traditional and novel antibodies (n=91) had significantly worse corneal fluorescein staining (mean=5.2) than those who were positive for the traditional antibodies alone (mean=4.7), for the novel antibodies (mean=4.1; p=0.03).

Table I. Baseline antibody testing results of DREAM<sup>®</sup> study patients by history of Sjögren's Syndrome (SS) and other autoimmune disease.

Baseline antibodies	No history of SS or autoimmune disease (n=375)	History of only non-SS autoimmune disease (n=66)	history of SS (n=53)	Overall P-value	P-value for SS vs. no autoimmune disease
Traditional SS antibodies:					
SS-A(Ro) >25 EU/ml <sup>‡</sup>	20 (5%)	6 (9%)	31 (60%)	< 0.0001	< 0.0001
SS-B(La) >25 EU/mla	2 (1%9	3 (5%)	13 (25%)	< 0.0001	< 0.0001
Positive tests for SS-A(Ro) and SS-B (La)				<0.0001§	<0.0001§
0	353 (94%)	59 (89%)	22 (42%)		
1	22 (6%)	5 (8%)	18 (34%)		
2	0 (0%)	2 (3%)	13 (25%)		
Anti-nuclear antibody ≥1:40 <sup>‡</sup>	67 (18%)	20 (30%)	32 (62%)	< 0.0001	< 0.0001
Rheumatoid factor positive	91 (24%)	20 (30%)	29 (56%)	< 0.0001	< 0.0001
Number of positive tests for				<0.0001§	<0.0001§
traditional antibodies					
0	226 (60%)	32 (48%)	14 (26%)		
1	122 (33%)	24 (36%)	6 (11%)		
2	23 (6%)	7 (11%)	11 (21%)		
3	4 (1%)	1 (2%)	11 (19%)		
4	1 (0.3%)	2 (3%)	11 (21%)		
Novel SS antibodies:					
Salivary protein 1 antibodies positive	73 (20%)	15 (23%)	11 (21%)	0.79	0.85
Carbonic anhydrase VI antibodies positive	60 (16%)	17 (25%)	9 (17%)	0.16	0.84
Parotid specific protein	39 (10%)	12 (18%)	5 (10%)	0.20	1.00
Number of positive tests for novel antibodies				$0.1^{\$}$	0.85 <sup>§</sup>
0	256 (69%)	39 (59%)	30 (58%)		
ĩ	70 (19%)	13 (20%)	19 (37%)		
2	42 (11%)	11 (17%)	3 (6%)		
3	6 (2%)	3 (5%)	0 (0%)		

<sup>8</sup>Missing data in 2 patients (one in SS group, one in group without SS and auto-immune disease. <sup>9</sup>For test of linear trend. **Conclusions.** In this large cross-sectional study, there was no difference in the prevalence rate of novel candidate SS antibodies in DED patients with or without SS. The novel antibodies may define a new class of DED patients with more severe ocular surface disease but not meeting the criteria for SS, but further studies are needed. Longitudinal changes in these antibodies in this cohort will be evaluated in the future and may yield useful insights into the patterns of prevalence in SS and non-SS DED patients.

#### P-5

## Evaluation of tissue specific autoantibodies in fibromyalgia patients with sicca and/or xerostomia

Eliana Applbaum<sup>1</sup> and Alan Lichtbroun<sup>1</sup>.

<sup>1</sup>Robert Wood Johnson Medical School, Rutgers University, New Jersey, USA.

**Background.** A significant proportion of patients with fibromyalgia complain of dry eyes and mouth. The 2010 preliminary criteria for fibromyalgia even includes dry eyes and mouth as part of the somatic symptoms severity complex used to diagnose this condition. A significant proportion of Sjögren's syndrome (SS) patients also complain of fibromyalgia symptoms, and there is literature that suggests that there is interplay between these two disorders. Recently, the presence of novel tissue specific autoantibodies (TSAs), SP-1, CA6, and PSP, has been observed in the early stages of SS. These early markers present themselves before the classic autoantibodies, such as SS-A/Ro, SS-B/La, ANA, and RF.

**Objectives.** This study aims to examine the relationship between SS and fibromyalgia by testing patients with fibromyalgia, who also complain of xerostomia and sicca symptoms, for SS related biomarkers SS-A/Ro, SS-B/La, SP-1, CA6, and PSP.

**Methods.** Over the past 5 years, we identified a cohort of 310 patients that presented with symptoms of fibromyalgia and fulfilled both the 1990 and 2010 preliminary diagnostic criteria. These patients were further questioned about xerostomia and sicca symptoms. Patients that admitted to using artificial tears at least biweekly, drinking water excessively to relieve dry mouth, or have previously experienced a blocked tear duct, but did not meet the strict diagnostic criteria for SS and did not have elevated inflammatory markers ESR or CRP, were selected for this study. Serum from study patients was sent to a tertiary lab, Immco Diagnostics, for testing of the classic autoantibodies (SS-A/Ro, SS-B/La, ANA and RF) and TSAs (SP-1, CA6, PSP). Patients testing positive for the TSA markers were provided with literature on the disease and offered appropriate treatment options, such as hydroxychloroquine.

**Results.** As of November 2017, 310 patients were selected for this study and tested for the SS markers. 91.0% of the patients were female and 8.6% of the patients were male. The average patient age was 56.5 years. Of the study patients, 271 were tested for both the TSAs and classic autoantibodies, while 39 were tested for only TSAs. Of the patients that were evaluated for both the TSAs and classic Sjögren's autoantibodies, 29.8% (81) tested positive for SS, 22.8% (71) of the patients were positive for TSAs, 9.4% (29) were positive for the classic Sjögren's autoantibodies. Further analysis of all the patients that tested positive for the TSAs (n=71), found 74.5% (53) were positive for SP-1, 8.5% (6) were positive for CA6 and 44.7% (32) were positive for PSP. Of these patients, 68.1% (48) were positive for only one of the TSA and 23 (31.9%) were positive for more than one TSA.

**Conclusion.** In this cohort of 310 fibromyalgia patients, about 1/3 of patients that were tested for both the TSAs and classic Sjögren's markers tested positive for SS, with the majority of those patients being positive for one or more of the TSAs. This suggests that autoimmunity, specifically early- stage Sjögren's syndrome, may be a confounding variable in the pathophysiology of fibromyalgia.

References

SHEN L, et al.: Clin Immunol 2012; 145: 251-55.

## **P-6**

## Clinicopathological analysis of labial salivary gland tissues from patients with IgG4-related disease

Akira Chinju<sup>1</sup>, Masafumi Moriyama<sup>1, 2</sup>, Noriko Ishiguro<sup>1</sup>, Yurie Mikami<sup>1</sup>, Akihiko Tanaka<sup>1</sup>, Takashi Maehara<sup>1, 3</sup>, Sachiko Furukawa<sup>1</sup>, Miho Ohta<sup>1</sup>, Masaki Yamauchi<sup>1</sup>, Haque A. S. M. Rafiul<sup>1</sup>, Mizuki Sakamoto<sup>1</sup>, Keita Mochizuki<sup>11</sup>, Ryusuke Munemura, Jun-Nosuke Hayashida<sup>1</sup>, and Seiji Nakamura<sup>1</sup>. <sup>1</sup>Section of Oral and Maxillofacial Oncology, Division of Maxillofacial Diagnostic and Surgical Sciences, Faculty of Dental Science, Kyushu University, Fukuoka, Japan. <sup>2</sup>OBT Research Center, Faculty of Dental Science, Kyushu University, Fukuoka, Japan. <sup>3</sup>Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts, USA.

**Objectives.** For the definitive diagnosis of IgG4-related disease (IgG4-RD), biopsies of local lesions are recommended so as to exclude other diseases, including lymphoma and cancer. However, performing biopsies of underlying organs is technically difficult and invasive. In this study, we examined the diagnostic utility of labial salivary gland (LSG) biopsy as a less invasive procedure.

**Methods.** Eighty-seven patients with suspected IgG4-RD by clinical findings or high serum IgG4 underwent LSG biopsy. We examined the relationship between the number of IgG4-positive plasma cells in LSG and clinical findings. The disease activity and damage of IgG4-RD patients were assessed by clinical findings including the site and number of affected organs, the resistance of steroid treatment, the prevalence of recurrence, and the salivary flow rate.

**Results.** The final diagnosis was 48 patients with IgG4-RD, 23 with Sjögren's syndrome (SS), 6 with suspected SS, 3 with malignant lymphoma, 6 with systemic lupus erythematosus, and 1 with Warthin's tumor. The sensitivity, specificity, and accuracy of LSG biopsy were 58.3%, 94.9%, and 74.7%, respectively. Moreover, Forty-eight IgG4-RD patients were divided into two groups: 1) 28 patients with positive of LSG biopsy (IgG4-RD B(+)) and 2) 20 patients with negative (IgG4-RD B(-)). In IgG4-RD B(+) patients, the presence of salivary gland lesions, the number of affected organs, serum IgG4 concentration, and the number and ratio of IgG4-RD B(-) patients.

**Conclusion.** These results suggest that LSG biopsy may reflect the disease activity of IgG4-RD. On the other hand, LSG biopsy alone is insufficient for the diagnosis of IgG4-RD because of its low sensitivity. However, combined with clinical findings, including serum IgG4 and number of affected organs, LSG biopsy was indicated to contribute to the diagnosis of IgG4-RD patients with affected underlying organs.

## **P-7**

## How single-cell analysis can help diagnose borderline sicca patients with chronic sialadenitis?

Cuong Q. Nguyen<sup>1,2,4</sup>, Alexandria Voigt<sup>1</sup>, Carol M. Stewart<sup>3,4</sup>, Ammon B. Peck<sup>1,4</sup>, Roland Jonnson<sup>4</sup>, Karl Brokstad<sup>5</sup>, Lida Esfandiary<sup>1</sup>, Sukesh Sukumaran<sup>6</sup>, Indraneel Bhattacharya<sup>3,4</sup>.

<sup>1</sup>Department of Infectious Diseases and Pathology, College of Veterinary Medicine, <sup>2</sup>Department of Oral Biology, <sup>3</sup>Department of Oral and Maxillofacial Diagnostic Sciences, <sup>4</sup>Center of Orphaned Autoimmune Diseases, College of Dentistry, University of Florida, Gainesville Florida, USA, <sup>5</sup>Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, Bergen, Norway, <sup>6</sup>Rheumatology Section, University of Arkansas for Medical Sciences, Arkansas Children's Hospital, Little Rock Arkansas, USA.

**Background.** Sjögren's syndrome (SS) is best characterized by chronic progressive immune attacks primarily against the salivary and lacrimal glands. Two essential diagnostic biomarkers of SS with the most weight are the formation of lymphocytic foci in the glands and the presence of serum anti-SSA/Ro autoantibodies. These two inclusive criteria preclude sicca patients with chronic sialadenitis who are borderline from fulfilling the criteria. The objective of this study was to examine labial salivary gland (LSG) biopsies of patients with chronic sialadenitis for infiltrating B cells and autoantibody production using Single-Cell Autoantibody Nanochip (SCAN).

**Methods.** LSG biopsies were obtained from non-primary SS (non-pSS) patients who did not meet the ACR/EULAR classification criteria for pSS. Paraffin-embedded sections were histologically examined for chronic sialadenitis and stained for infiltrating lymphocytes. Subsequently, single *exvivo* lymphocytes of LSG biopsies were isolated and fluorescently conjugated for cell surface markers. Cells were added onto fabricated nanowells and imaged using a high-speed epifluorescence microscope. The microengraved slides were coated with immunoglobulins to capture all secreted antibodies and hybridized with fluorescently-conjugated IgG, SSA/Ro60, SSA/Ro52, and SSB/La antigens. Microarray spots were analyzed for nanowells with single, live B cells that produced antigen-specific autoantibodies.

**Results.** Our results indicate that non-pSS patients with chronic sialadenitis exhibited significant infiltration of CD20<sup>+</sup> B cells, CD3<sup>+</sup> CD4<sup>+</sup>, and CD3<sup>+</sup> CD8<sup>+</sup> T cells. *Ex-vivo* analysis of glandular lymphocytes using SCAN demonstrated that individual B cells of these subjects produced high levels of IgG-specific anti-SSA/Ro60 in comparison to IgG-specific anti-SSA/Ro52 or IgG- specific anti-SSB/La autoantibodies.

**Conclusions.** Single-cell analysis of LSG biopsies revealed the presence of B cells producing pSS signature autoantibodies in non-pSS patients with chronic sialadenitis. This finding raises a critical issue regarding the appropriate and correct diagnosis of borderline subjects based on the latest classification criteria using focus score and serum anti-SSA/Ro.

#### **P-8**

#### A fine bioinformatical analysis of lymphocyte distribution predicts the diagnosis of primary Sjögren's syndrome among other systemic autoimmune diseases

Quentin Simon<sup>1</sup>, Bénédicte Rouvière<sup>1</sup>, Tifenn Martin<sup>1</sup>, Lucas Le Lann<sup>1</sup>, Alain Saraux<sup>1</sup>, Valérie Devauchelle-Pensec<sup>1</sup>, Concepcion Marañón<sup>2</sup>, Nieves Varela Hernández<sup>2</sup>, Aleksandra Dufour<sup>3</sup>, Carlo Chizzolini<sup>4</sup>, Ellen de Langhe<sup>5</sup>, Nuria Barbarroja<sup>6</sup>, Chary Lopez-Pedrera<sup>7</sup>, Velia Gerl<sup>8</sup>, Aurelie Degroof<sup>9</sup>, Julie Ducreux<sup>9</sup>, Elena Trombetta<sup>10</sup>, Tianlu Li<sup>11</sup>, Marta Alarcón-Riquelme<sup>12</sup>, Christophe Jamin<sup>1</sup> and Jacques-Olivier Pers<sup>1</sup>

<sup>1</sup>U1227, Université de Brest, Inserm, Labex IGO, CHU de Brest, <sup>2</sup>GENYO, Centre for Genomics and Oncological Research Pfizer, University of Granada, Andalusian Regional Government, <sup>3</sup>University Hospital and School of Medicine, <sup>4</sup>University hospital of Geneva, <sup>5</sup>University Hospital KU Leuven, <sup>6</sup>IMIBIC/Reina Sofia Hospital/University of Cordoba, <sup>7</sup>IMIBIC/Reina Sofia Hospital/ University of Cordoba, <sup>8</sup>Department of Rheumatology and Clinical Immunology, Charité University Hospital, <sup>9</sup>Institut de Recherche Expérimentale et Clinique, Université catholique de Louvain, <sup>10</sup>Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico di Milano, <sup>11</sup>Bellvitge Biomedical Research Institute (IDIBELL), <sup>12</sup>GENYO. Center for Genomics and Oncological Research.

**Background/Purpose.** We investigated 194 individuals with SADs (38 primary Sjögren's syndrome (pSS), 47 rheumatoid arthritis (RA), 46 systemic lupus erythematosus (SLE), 42 systemic sclerosis (SSc) and 21 undifferentiated connective tissue disease (UCTD) patients) and 53 healthy controls (HCs) to determine whether a fine flow cytometry analysis of T and B cell distribution in whole blood could cluster individuals according to disease diagnosis.

**Methods.** Two flow cytometry panels were designed. The first panel was dedicated to T cells and combined CD57, CD45RA, CD62L, CD27, CD38, CD3, CD4 and CD8 mAbs. The second panel was dedicated to B cells and combined IgD, TACI, CD27, CD5, CD38, CD19 and CD24 mAbs. A classical manual gating strategy and the Flow-clustering without K (FLOCK) investigation, a density-based clustering approach to algorithmically identify relevant cell populations from multiple samples in an unbiased fashion, were used.

Results. The manual gating strategy allows the identification of 17 distinct lymphocyte subsets. The prediction of the different SADs was determined by discriminant function analysis (DFA). No clustering was found. The FLOCK exploration of the merged HCs identifies 85 distinct subsets of lymphocytes used as reference when compared to SADs . The DFA analysis clearly clusters the HCs and the patients according to each SAD (see figure below). When compared to HCs, the pSS signature was discriminated by an increase in IgD<sup>hi</sup>CD24<sup>hi</sup>CD38<sup>hi</sup>CD27<sup>T</sup>ACI<sup>C</sup>D5<sup>hi</sup> transitional B cells, and an increase of CD45RA+CD27-CD62L10/-CD57hi effector CD8+ T cells. The SLE signature was discriminated by an increase in IgD CD24<sup>lo</sup>CD38 CD27 TACI+CD5 memory like B cells, an increase in CD45RA CD62L+CD38hi activated central memory CD4+ T cells. The RA signature was discriminated by an increase in IgDhiCD2410CD38-CD27-TACI+CD5- unactivated mature naïve B cells and a decrease in CD45RA+CD62L+CD38hi naïve CD8+ T cells. The SSc signature was discriminated by a decrease in CD45RA+CD62L+CD38hi naïve CD8+ T cells. Interestingly, patients with UCTD were distributed among the different clusters (28% with HC, 29% with SLE, 29% with SSc, 9% with RA and 5% with pSS clusters).



**Conclusion.** A fine bioinformatical flow cytometry analysis of T and B cell subsets clusterizes patients and HCs suggesting that each SAD can be associated with abnormal specific phenotypical distributions that could be helpful in the diagnosis.

This work has received support from the EU/EFPIA Innovative Medicines Initiative Joint Undertaking PRECISESADS grant n° 115565.www.precisesads.eu.

#### P-9

#### Withdrawn

#### P-10

## Novel radiographic features in Sjögren's syndrome related Sialo-CBCT

Keshet N.<sup>1</sup>, Aricha A<sup>2</sup>, Friedlander-Barenboim S.<sup>3</sup>, Aframian DJ<sup>1</sup>, and Nadler  $C^2$ .

<sup>1</sup>Department of Oral Medicine, Sedation, and Maxillofacial Imaging, Sjogren's Syndrome Center, Hebrew University Hadassah School of Dental Medicine, Jerusalem, Israel.<sup>2</sup>Oro-maxillofacial imaging, Department of Oral Medicine, Sedation, and Maxillofacial Imaging, Hebrew University Hadassah School of Dental Medicine, Jerusalem, Israel.<sup>3</sup>Division of Pediatric and Hospital Dentistry, The Chaim Sheba Medical Center, Ramat-Gan, Israel.

**Objective.** Sjögren's syndrome (SS) is manifested primarily by ocular and or oral dryness. Parotid Sialo-Cone-Beam Computerized Tomography (Sialo-CBCT) demonstrates the ductal architecture and gland's function. We characterized novel features in SS suspected patients and correlated them with clinical findings and known SS criteria.

**Methods.** The clinical and radiographic data of SS suspected patients; referred for Sialo-CBCT in 2011-2014, were retrospectively reviewed. Scans were evaluated using a pre-formed questionnaire by two observers for various radiographic features including duct morphology, level of branching, ductopenia and sialectasia. These features were compared between themselves, with clinical data and to 3 well established sets of SS criteria; The American-European Consensus Group (AECG) and the 2 latest American College of Rheumatology (ACR) Group.

**Results.** Sialo-CBCT scans of 67-suspected SS patients (115 parotid glands) were included. Intra-radiographic association was found between ductopenia and all other radiographic parameters. Minimal, yet important, radiographic differences were found between left and right parotid pair glands. AECG-confirmed-SS patients showed strong correlation with radiographic features, whereas ACR2012-confirmed-SS patients did not. Clinical data was insufficient for ACR2017 assessment.

**Conclusion.** Sialo-CBCT provides novel radiographic features for SS diagnosis, which may improve SS patient's diagnosis monitoring and treatment. Further studies are needed to understand their role.

## **P-11**

Patient-reported post-operative change of sensibility and pain after parotid and labial gland biopsy applied in the diagnostics of primary Sjögren's syndrome: one year follow-up study

Konstantina Delli<sup>1</sup>, Essam Dagal<sup>1</sup>, Hendrika Bootsma<sup>2</sup>, Arjan Vissink<sup>1</sup>, Fred K.L. Spijkervet<sup>1</sup>.

Departments of: <sup>1</sup>Oral and Maxillofacial Surgery, <sup>2</sup>Rheumatology and Clinical Immunology, University of Groningen, University Medical Center Groningen, the Netherlands.

**Background.** Biopsies of the parotid gland have not become common place in the diagnostics of primary Sjögren's syndrome (pSS) because of the speculated higher morbidity compared to the labial gland biopsies (1).

The aim of this study was to assess how patients perceived pain and change of sensibility of the biopsied area after having undergone both a parotid and labial gland biopsy as part of the routine diagnostic work up of pSS.

**Methods.** Adult patients suspected with pSS were included if they were willing to undergo both a parotid and labial gland biopsy in addition to the other diagnostic tests according to the ACR-EULAR criteria (2). The study protocol was approved by the ethics committee of the University Medical Center Groningen (METC approval: 2013.066).

Simultaneously, the parotid and labial salivary gland biopsy was taken under local anesthesia and loupe glass magnification (x3.5)(1,3). One week, 6 months and 12 months post-operatively, each patient was sent a postal questionnaire comprising of 4 questions in order to quantify the severity of pain and the change of sensibility in the biopsied areas with a visual analogue scale (VAS; range 0-100).



one gain on pay

Fig. 1. Patient-reported change of sensibility VAS score of patients after biopsies. Horizontal lines indicate medians.



**Results.** 110 patients were included. The median age of patients was 54 years (IQR=47-65) and 92% were female. Patient-reported changes in sensibility and pain in the biopsied area shown in Figures 1 and 2, respectively. Changes in sensibility of the biopsied area were slightly more common after a parotid biopsy than after a labial salivary gland biopsy at one week and 6 months post-operatively. Additionally, patients experienced slightly more

pain in the area of the parotid gland biopsy than in the area of the labial gland biopsy at one week and 6 months post-operatively. At 12 months post-operatively, the change in sensibility of the biopsied areas and pain level was very low in most patients and comparable for both biopsied areas

**Conclusions.** Patient-reported post-operative change of sensibility and pain in the area of the parotid and labial gland biopsy are comparable after one year. Parotid gland biopsy is a diagnostic technique well-tolerated by patients suspected with pSS.

### References

1. DELLI K, VISSINK A, SPIJKERVET FK: Oral Maxillofac Surg Clin North Am 2014; 26: 23-33

- 2. SHIBOSKI CH, SHIBOSKI SC, SEROR R et al.: Ann Rheum Dis 2017; 76: 9-16.
- 3. SPIJKERVET FK, HAACKE E, KROESE FG et al.: Rheum Dis Clin North Am 2016; 42: 485-99.

#### P-12

#### Assessment of Focus score and additional histopathological parameters for the diagnosis of primary Sjögren's syndrome in daily practice: possible bias and the need of standardisation

Valentina Donati<sup>1</sup>, Francesco Ferro<sup>2</sup>, Nicoletta Luciano<sup>2</sup>, Emanuele Calabrese<sup>2</sup>, Elena Elefante<sup>2</sup>, Chiara Baldini<sup>2</sup>.

<sup>1</sup>Unit of Anatomic Pathology II, Azienda Ospedaliero-Universitaria Pisana. <sup>2</sup>Rheumatology Unit, Department of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy.

**Background.** Recently, a standardization of labial salivary gland histopathology has been proposed for clinical trials in primary Sjögren's syndrome (pSS). Focus score (FS) remains the key element for the histological diagnosis of pSS; nonetheless additional parameters were proposed. Relatively few information are available regarding the feasibility of the assessment of these parameters in daily clinical practice, including focal lymphocytic sialoadenitis (FLS), focus score (FS) and germinal centre(GC)-like structures.

Aim of this study. Was to evaluate the impact of different bias including the areas of glandular tissue in the assessment of FS and additional histopathological parameters in daily routine pSS diagnosis.

**Methods.** Consecutive MSGBs performed in 4 different University and Hospital centers (i.e two ENT Units, one Maxillofacial Unit and one Dentistry Unit) were collected and centralized to the same Pathology Unit from January 1st 2017 to October 31st 2017. The biopsies had been performed by different operators within the same Unit. An expert pathologist evaluated the samples focusing on: glandular tissue areas, assessment of FLS versus non-specific chronic sialoadentits (NSCS), number of foci, FS and presence of GCs.

**Results.** MSGBs from 85 subjects (66F:19M) were included in the study. Out of them, 27(31.8%) were diagnosed as having a FLS with a FS≥1, 14 (16.4%) with a 0<FS<1 and 44 (51.8%) with a NSCS. The mean (SD) number of foci was 1.71 (3.30), range 0-22. Twenty of 41 (48.8%) FLS samples presented at least one GC (range 1-8). The mean (SD) FS was 1.67(1.54), range 0.31-8.48. The area of glandular tissues ranged from 1.26 to 50.42 mm<sup>2</sup> with a mean (SD) value of 7.12 (6.19). A great variability in the areas of the samples was observed inter- and intra- the 4 Units (Dentistry: 6.14(4.01), ENT1: 5.70 (2.23), ENT2:9.85 (3.58), Maxillofacial: 27.42(32.52)). In 28 out of 85 samples (32.9%) the area was lower than 4 mm<sup>2</sup>. The diagnosis of pSS remained uncertain in 5 cases presenting a MSGB area <4 mm<sup>2</sup> with a single focus and negative anti-Ro/SSA antibodies.

**Conclusions.** This study has shown that despite FS still represents the most important parameter for pSS diagnosis, in daily routine it may be biased by the variability in the area of the MSGB with clear clinical implications. This highlight the importance of standardize the MSGB procedure in daily routine practice and of considering additional histopathological parameters in assessing the severity of the inflammatory infiltrate.

### P-13

# Salivary proteomics as a promising approach to characterize differences between primary Sjögren's syndrome and IgG4-related disease

Antonella Cecchettini<sup>1</sup>, Ilaria Puxeddu<sup>2</sup>, Nadia Ucciferri<sup>1</sup>, Riccardo Capecchi<sup>2</sup>, Silvia Rocchiccioli<sup>1</sup>, Enza Polizzi<sup>3</sup>, Francesco Ferro<sup>3</sup>, Valentina Donati<sup>4</sup>, Antonio Tavoni<sup>2</sup>, Paola Migliorini<sup>2</sup>, Chiara Baldini<sup>3</sup>.

<sup>1</sup>Institute of Clinical Physiology, CNR Pisa, Italy. <sup>2</sup>Immunology Unit, Department of Clinic and Experimental Medicine, University of Pisa. <sup>3</sup>Rheumatology Unit, Department of Clinic and Experimental Medicine, University of Pisa, Pisa Italy. <sup>4</sup>Unit of Anatomic Pathology II, Azienda Ospedaliero-Universitaria Pisana.

**Background.** Primary SS (pSS) and IgG4-related sialoadenitis (IgG4-RD) share a number of clinical and serologic features, making it difficult to differentiate these two disorders in clinical practice. Nevertheless it is of primary interest to discriminate these entities either to precisely tailor the treatment or to gain insights into the mechanisms of the diseases. Recently, salivary proteomics has appeared as a valuable tool to identify novel biomarkers for pSS. Aim of this study was to explore whether pSS and IgG4-RD may have a different salivary proteomic profiling.

**Methods.** Unstimulated salivary flow was collected in standard condition from 10 subjects: 4 pSS, 2 IgG4-RD and 4 controls. High-abundance proteins were depleted using affinity and immunodepletion methodologies. A high-throughput liquid chromatography tandem mass spectrometry (LC-MS/MS) was used for the proteomic analysis. Principal component analysis (PCA) was utilized for statistical analysis.

**Results.** Over 300 proteins were identified and among these, 34 resulted as differentially expressed. Patients with IgG4-RD, but not with pSS presented a higher expression of proteins related to fibrosis and tissue remodeling including matrix metalloproteinase-9, collagen alpha-1(VI) chain, cysteinerich secretory protein 3 and cathepsin D. On the other hand, pSS patients presented a significant down-regulation of acinar proteins including cystatins C and S and prolactin-inducible protein indicating a more prominent glandular damage.

**Conclusions.** This study shows that salivary proteome may be useful to highlight different and specific pathways for pSS and IgG4-RD. Preliminary results suggest a more prominent tissue remodeling and fibrosis profile in IgG4-RD while a more explicit glandular damage in pSS syndrome.

#### **P-14**

#### Ultrasound of pleural irregularities: a promising tool in primary Sjögren's syndrome (pSS)-associated interstitial lung disease (ILD). A functional and CT correlation study

Ferro F<sup>1</sup>, Bulleri A<sup>2</sup>, Delle Sedie A<sup>1</sup>, Barsotti S<sup>1</sup>, Luciano N<sup>1</sup>, Elefante E<sup>1</sup>, Mosca M<sup>1</sup>, Baldini C<sup>1</sup>.

<sup>1</sup>Rheumatology Unit, Department of Clinical and Experimental Medicine, University of Pisa. <sup>2</sup>Radiology Unit, University of Pisa.

**Background.** Ultrasound study of pleural irregularities (PI-US) has recently been proposed as a novel approach for the assessment of interstitial lung disease (ILD) in systemic connective tissue disorders.

Aim of this work. Was to explore the correlation between PI-US, high resolution computerized tomography (HRTC) and pulmonary function tests in primary Sjögren's syndrome (pSS)-related ILD, in order to define the diagnostic value of this technique for the non-invasive assessment of lung involvement in pSS.

**Methods.** HRCT and respiratory function tests were performed in patients with a diagnosis of pSS-related ILD. Concurrently, all the patients underwent PI-US, carried out by a single operator using a MyLab-25 (Esaote), 10 MHz, 5 cm linear probe. PI was defined as the loss of the normal hyperechoic linear pleural contour (score 0-2: normal, minimal and major changes at each intercostal space). PI-US total score and partial scores were calculated, considering 6 lung fields: 2 for the anterior, 2 for postero-superior and with particular attention for the 2 postero-inferior chest surface). An expert radiologist quantified lung involvement assigning Warrick scores for each patient. **Results.** We included in this study 18 patients with pSS-related ILD (14 F:4 M, mean age =68.8±9.9 yrs). The median PI-US score was 45 (range 25.5-73.5). Both PI-US total score and partial postero-inferior PI-US score strongly correlated with the Warrick HRCT score (r= 0.813, *p*=0.000 and r= 0.914, *p*=0.000). Regarding pulmonary function tests, the Warrick HRCT score correlated with FVC (r=-0.753, *p*=0.001), TLC (r=-0.853, *p*=0.000),

and DLCO (r=-0.834, p=0.000). Similarly, total score and partial posteroinferior PI-US score correlated inversely with FVC (r=-0.849, p=0.000 and r=-0.836, p=0.000), TLC (r=-0.895, p=0.000 and r=-0.829, p=0.000), and DLCO (r=-0.953, p=0.000 and r=-0.883, p=0.001). Finally, both PI-US score and PI-US of the infero-posterior field directly correlated with FEV1/SVC (r=0.701, p=0.004 and r=0.619, p=0.01) and with FEV1/FVC (r=0.600, p=0.02 and r=0.501, p=0.05).

**Conclusions.** This study demonstrated a high correlation between PI-US, HRCT findings and pulmonary function tests, supporting the use of lung ultrasonography in clinical practice for the assessment of pSS-associated ILD. The specific correlation between PI-US scores and both FEV1/SVC and FEV1/FVC (not observed with the Warrick HRCT score) seemed to indicate a higher sensitivity of PI-US respect to HRCT in ILD assessment. Further studies are warranted to clarify the role of PI-US for the early diagnosis of ILD.

## P-15

#### Early respiratory disease activity is risk for damage in primary Sjögren's syndrome

Valéria Valim<sup>1</sup>, Wildner M Sardenberg<sup>1</sup>, Larissa Carvalho Caser<sup>1</sup>, Maria Carmen LFS Santos<sup>1</sup>.

<sup>1</sup>Hospital Universitário Cassiano Antônio de Moraes/Federal University of Espírito Santo.

**Background.** We studied clinical and epidemiological variables that could be related to greater severity and damage in primary Sjögren's syndrome (pSS).

**Methods.** Cross-Sectional study including patients with pSS (AECG 2002 or ACR/EULAR 2017). EULAR Sjögren's syndrome Disease Index (ES-SDAI) was calculated at diagnosis and currently. Patients have performed EULAR Sjögren's syndrome Patient Report Index (ESSPRI) and Sjögren's syndrome Disease Damage Index (SSDDI). Non-parametric statistics and logistic regression were applied with *p*-value of 0.05.

**Results.** 104 patients with pSS were included,  $51.49\pm12.13$  years, 95.2% (n=99/104) were women. Disease duration was  $65, 44\pm41.89$  months. Anti-Ro was positive in 68% (n=70/103), FAN in 82% (n=82/100), FR in 37.4% (n=37/99) and anti-La in 30.3%

(n=30/99), focal lymphocytic sialoadenitis in 73% (71/91). The ESSPRI was 5.62 $\pm$ 2.56 and the ESSDAI 3.63 $\pm$ 5.12. 53.8% (n=56/104) was using hydroxychloroquine and 51% (n=53/104) immunosuppressant. The higher damage score was associated with disease duration (0.270; *p*=0.007), high disease activity at the onset of the disease (initial ESSDAI) (0.226, *p*=0.021), respiratory evolvement at the onset of the disease (0.273, *p*=0.005), at some point. After multiple regression analysis, the initial involvement of the respiratory system was confirmed to be a risk factor for damage (SSDDI > 3) independent of disease duration and current disease activity (ESSDAI) (OR=8.94, 1.44-55.64CI95%; *p*<0.05). The respiratory domain was also associated with higher mortality (66.7% vs. 5%, *p*=0.011).

**Conclusion.** Early respiratory disease activity is associated with death and predicts damage in primary Sjögren's syndrome.

### P-16

#### Increased B-cell activating factor, interleukin-6, interleukin-8 and lymphocytes in induced sputum from patients with primary Sjögren's syndrome

Nilsson AM, Tufvesson E, Hesselstrand R, Olsson P, Wollmer P, Mandl T.

**Background.** Small airway disease and chronic obstructive pulmonary disease are common in primary Sjögren's syndrome (pSS). However, the underlying inflammatory mechanisms behind pSS associated airway disease have not been studied in detail. We therefore wanted to study cytokine and leucocyte levels in induced sputum in never-smoking patients with pSS.

**Methods.** Induced sputum cytokines and leucocytes were assessed in 20 never-smoking patients with pSS and 19 age and sex-matched populationbased controls. In addition, pulmonary function, disease activity, respiratory symptoms, as well as inflammatory and serological features of pSS were assessed. **Results.** B-cell activating factor (BAFF), interleukin (IL)-6 and IL-8 were significantly increased in induced sputum in pSS patients compared to population-based controls, whilst IL-1 $\beta$ , interferon- $\alpha$ , and tumor necrosis factor- $\alpha$  levels were not (Figure 1). In addition, lymphocytes were significantly increased in induced sputum from pSS patients compared to population-based controls (Figure 2). However, cytokine levels in induced sputum were not associated to pulmonary function tests, disease activity, respiratory symptoms, nor laboratory or serological features of pSS.

**Conclusions.** The increase in BAFF, IL-6, IL-8 and lymphocytes in induced sputum suggests a specific ongoing inflammatory disease process in the airways in pSS patients. Its association to pSS associated airway disease has to be further examined in future larger studies.



Fig. 1. Results of sputum supernatant levels of B-cell activating factor (BAFF), interleukin (IL)-1 $\beta$ , IL-6, IL-8, interferon- $\alpha$  (IFN- $\alpha$ ) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) in 20 primary Sjögren's syndrome patients (pSS) and 19 population based controls (Ctrl).



Fig. 2. Results of sputum differential counts of lymphocytes, eosinophils and macrophages in Sjögren's syndrome (pSS) patients and population based controls (Ctrl).

### **P-17**

A case series of neurological abnormalities associated with Sjögren syndrome – Removing inevitable confounders in complex presentations to make accurate diagnoses and provide effective treatments

Shu F.<sup>1</sup>

<sup>1</sup>University of California, Los Angeles, Medical Center, Los Angeles, California, USA.

**Background.** Sjögren Syndrome (SS) is an autoimmune disorder of exocrine glands classically resulting in xerophthalmia and xerostomia; occasionally it can be associated with a complex array of neurologic abnormalities. When the latter precedes the sicca complex or occurs in isolation, an accurate diagnosis is often impeded. We present a case series of 24 neurologic patients who fulfilled the serological and/or histopathological criteria

for SS, to 1) highlight the protean neurological manifestations, 2) discuss the association of the anti-SS-A and anti-SS-B antibodies to the neurological symptoms, or lack thereof, 3) identify mimics of SS neuropathy that could coexist with SS, and 4) discuss the effective treatments which our patients have responded to.

**Methods.** From July 2015 to December 2017, we identified 24 patients that presented to UCLA, who have fulfilled the serological and/or histopathological criteria for SS, and complained of debilitating neurological symptoms otherwise not explained by another etiology. We evaluated these patients through electromyography, skin biopsy, nerve biopsy, autonomic testing, cerebral spinal fluid analysis, and imaging. We instituted different immuno-therapies and evaluated the patients' responses.

Results. In this series, Sjögren neurological syndrome affected mostly women in their 50s. Predominantly peripheral but central nervous system (CNS) involvement was also observed. A major finding was dysautonomia characterized by tonic pupils, sudomotor dysfunction, orthostatic hypotension, esophageal dysmotility, overactive bladder, and dyssynergic defecation. Some other common features included sensory ganglionopathy, small fiber neuropathy, sensorimotor axonal polyneuropathy, sensory axonal polyneuropathy, lateral femoral cutaneous neuropathy, and trigeminal sensory neuropathy. MRI abnormalities of the brain and cervical spinal cord were also demonstrated. In addition, two patients with rapidly progressive disabling neurological symptoms demonstrated negative serology but focal lymphocytic sialoadenitis. Selected comorbidities included chronic lymphocytic leukemia, CNS vasculitis, herpes meningoencephalitis, monoclonal gammopathy, stiff person syndrome, ankylosing spondylitis, and fibromyalgia; these conditions could also exhibit neurologic symptoms. Lastly, different immunotherapies and supportive care were implemented and significant effectiveness was observed.

**Conclusions.** 1) In patients presenting with multifocal neurological symptoms without the sicca complex, Sjögren syndrome should still be considered, because neurological abnormalities could antedate the rheumatological complaints. 2) Absence of anti-SS-A and SS-B antibodies does not exclude the diagnosis, and therefore labial biopsy is encouraged if there are no contraindications. 3) Sensory ganglionopathy, autonomic neuropathy, and small fiber neuropathy are often under-recognized but actually should be indications for SS workup. 4) Neurological symptoms can be relentlessly progressive if left untreated; early disease detection and timely multidisciplinary care are the cornerstones of proper SS care.

### **P-18**

## Neuromyelitis optica spectrum disorder in primary Sjögren's syndrome: what are the relevant diagnostic aspects?

Fabiola Reis Oliveira, Paulo Louzada Junior, Antonio Carlos dos Santos, Rodolfo Dias Chiari Correia, Eduardo Melani Rocha. Ribeirao Preto Medical School, University of Sao Paulo, Ribeirao Preto, SP, Brazil.

**Purpose.** To describe the clinical, laboratory and imagining features of a series of cases of NeuromyelitisOptica spectrum disorder (NMOSD) associated with primary Sjögren's syndrome in a Brazilian referral center.

**Methods.** Patients with clinical and imagining features of NMO and primary Sjögren's syndrome (SSP) were evaluated. The SSP diagnosis was made in accordance with the American-European Criteria. NMOSD was diagnosed in the presence of optic neuritis and myelitis, magnetic resonance imaging (MRI) specific abnormalities (contiguous spinal cord lesion with 3 or more segments in length and/or MRI brain non-diagnostic for EM sclerosis) and/or serological evidence of anti-aquaporin-4 (AQP4) antibodies. All individual underwent a complete ophthalmologic and neurologic examination.

**Results.** Eleven patients were studied, 100% females. The mean age was 44+13 years old; time run from the beginning of disease was 9.2 (2-18) years. *Sicca* symptoms were present in 9/11 patients, but no one reported them before the onset of the neurological syndrome. The minor salivary gland biopsy focus score (FS) was  $\geq 1$  in 100%. Positive serum anti-SSA and anti-AQP4 were 54.5% and 50% (5/10), respectively. Optic neuropathy was the initial manifestation of 4/11. Among 11, 6 had bilateral optic neuropathy, and 5 had a single type. The clinical course of neurologic disease was chronic in 7 patients, and the average number of acute exacerbation was 4. None of the laboratory exams revealed to be predictor or biomarker of prognosis (complement, lactate dehydrogenase, beta2 microglobulin, cryoglobulins). **Conclusions.** Neurologic disorders are severe extra-glandular features of

SS and may show variable clinical courses, including acute optic neuritis, chronic optic atrophy, and myelopathy in the context of NMOSD. Longitudinally extensive myelitis and NMO, known as a rare relapsing autoimmune disease of the central nervous system (CNS), are sometimes found as the initial manifestation of primary Sjögren's syndrome. NMOSD manifestation without apparent *Sicca* symptoms makes the diagnosis more challenging due to an unknown frequency of association between both conditions. The combination of specific antibodies anti-SSA, anti-aquaporin-4, and biopsy abnormality of salivary gland are supportive for the diagnosis and treatment in atypical cases. Future studies are necessary to clarify the elements implicated in the NMOSD and SS association.

#### P-19

#### How phenotype of the small fibre neuropathy (SFN) in primary Sjögren syndrome (pSS) differs from others causes of small fibre neuropathy?

Elise Descamps<sup>1</sup> Julien Henry<sup>1</sup>, Celine Labeyrie<sup>1</sup>, David Adams<sup>1</sup>, Davide Aiello<sup>1</sup>, Xavier Mariette<sup>1</sup> and Raphaèle Seror<sup>1</sup>.

<sup>1</sup>Rheumatology and neurology departments, Hopital Bicetre, Université Paris-Sud, Le Kremlin-Bicêtre, France.

**Background.** Small fibre neuropathy (SFN) is a peripheral neuropathy characterized by neuropathic pain associated with normal routine nerve conduction study but rarefaction of intraepidermal nerve fibres (IENF). Primary Sjögren Syndrome (pSS) is one of the many aetiology of SFN.

The objective was to compare phenotype of SFN in pSS, transthyretin (TTR) familial amyloidosis and idiopathic SFN.

**Methods.** All patients referred to neurology or rheumatology departments of our hospital since October 2012 with a confirmed diagnosis of SFN associated with either pSS (ACR/EULAR 2016 criteria, TTR-amyloidosis or idiopathic were included in this monocentric retrospective study. Diagnosis of SFN was confirmed by normal nerve conduction studies and abnormal lower limb skin biopsies (defined by a reduced intraepidermal nerve fibers (IENF) density.

<7.63/mm for distal and <12.8/mm for proximal site). All patients undergo standardized diagnosis procedures within an outpatient day-clinic. Characteristics of SFN were compared between 3 groups: pSS, TTR-amyloidosis and idiopathic.

**Results.** We included 16 patients with pSS (14 (87.5 %) women, median age: 55 yrs [IQR:49.3-66.7], 7 (43.8%) anti-SSA positive, 13 (81.3%) focus score  $\geq$ 1), 17 with TTR-amyloidosis (7 (41.2%) women, median age: 47 yrs [35-56]) and 11 with Idiopathic SFN (7 (63.6%) women, median age : 47 yrs [36-56.5]).

Patients with pSS had a median ESSDAI of 5 [5-9], mainly due to the neurologic domain. One had monoclonal gammopathy, 5/14 (35.7%) rheumatoid factor, 2/14 (14.3%) hypergammaglobulinemia and none had cryoglobulin. Ten TTR amyloidosis patients had MET30 mutation and 7 other mutations. Time from first neurologic symptoms to diagnosis of SFN was significantly higher for pSS (20.5 months [7.7-48.5]) and idiopathic group (35 months [11.5-65]) than for TTR group (6 months [0-15]).

Clinical presentation was length dependant in only 3 (18.7%) patients with pSS compared to 10 (58.9%) in TTR amyloidosis (p=0.03) and 3 (27.3%) in idiopathic group (p=0.66). A "patchy" presentation (defined by asymmetrical and/or proximal symptoms involving limb, trunk and/or face), was significantly more frequent in pSS than in TTR amyloidosis (7(43.7%) vs. 1(5.9%); p=0.02).

This more frequent non-length dependant course was confirmed on skin biopsies with an IEFN at proximal site < IEFN at distal site in 8/15 (53.3%) pSS patients compared to 4 (23.5%) in TTR (p=0.14) and 1 (9.1%) in idiopathic (p=0.04) groups.

Lauria score was significantly higher in pSS than in TTR, (5 [4-8] vs. 4 [3-5], p=0.015), mainly due to sicca symptoms (n=15/16) and peripheral limb pain (n=14/16). After excluding sicca syndrome item, the score did not differ.

**Conclusion.** pSS patients with SFN had a low frequency of serum B cell activation biomarkers. Compared to other causes of SFN, in pSS SNF was characterized by a more frequent non-length dependant and patchy presentation and a higher Lauria score. This later was mainly driven by dryness and limb pain rather due to pSS than SFN.

## P-20

### Is peripheral blood lymphocyte population's distribution different in primary Sjögren's syndrome patients with lymphopenia?

Jose Loureiro Amigo<sup>1</sup> Carlos Palacio García<sup>2</sup> Roser Solans Laqué<sup>3</sup>

<sup>1</sup>PhD Programme, Medicine Department, Universidat Autónoma de Barcelona. Barcelona, Spain. <sup>2</sup>Flow Cytometry Unit, Haematology Service, Vall d'Hebron University Hospital, Universitat Autonoma, Barcelona, Spain. <sup>3</sup>Systemic Autoimmune Diseases Unit, Internal Medicine Department, Vall d'Hebron University Hospital, Barcelona, Spain

**Background.** An increasingly number of studies have given evidence for disturbances in the distribution of peripheral blood (PB) lymphocyte subsets in primary Sjögren's syndrome (pSS) patients. pSS patients exhibit a significant decrease in proportion and absolute counts of memory B cells (affecting both switched and unswitched memory B cells) and an increase in naïve B cells. Regarding T cell populations, patients with pSS have decreased T-cell counts, mostly as a result of a decrease in CD4<sup>+</sup> T cells (affecting memory, naïve and effector subsets of CD4<sup>+</sup> T cells). A significant decrease in proportion and absolute counts of Th17 cells have also been reported in pSS patients. Lymphopenia is considered to be a prognostic factor and a lymphoma predictor in pSS patients.

Our aim was to evaluate the lymphocyte population's distribution in PB from pSS patients with lymphopenia and to compare it with that in pSS patients without lymphopenia.

**Methods.** Seventy five patients with pSS were recruited at Vall d'Hebron University Hospital (Barcelona, Spain) for this study. pSS was diagnosed according to American-European Consensus Group Classification criteria for pSS. Ten patients (13.3%) had sustained lymphopenia ( $\leq 1000/mm^3$ ). Mean age of pSS without and with lymphopenia was 62 ( $\pm 14.9$ ) years and 54.1 ( $\pm 15.8$ ) years, respectively. There were three men in non-lymphopenic group.

Peripheral blood samples were collected in K3-EDTA anticoagulant. Main lymphocyte populations were analyzed by flow cytometry and defined accordingly with the Human Immune Phenotyping Consortium (HIPC) (Maecker, Nat Rev Immunol 2012). Absolute counts of lymphocyte subsets were calculated using the percentages obtained by flow cytometry and the leucocyte count obtained from an hematological analyzer.

Statistical differences were analyzed using the Mann-Whitney U test. P values less than 0.05 were considered significant.

**Results.** No significant differences were found related to proportions of T cells, CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells, B cells, monocytes and NK cells. Regarding CD4<sup>+</sup> subpopulations, we found a significant decrease in naïve CD4<sup>+</sup> proportion (38.7% vs 26.6%, p=0.048) and a significant increase in effector memory CD4<sup>+</sup> (25.6% vs 40.5%, p=0.012) in pSS patients with lymphopenia. The proportion of activated CD4<sup>+</sup> T cells and CD8<sup>+</sup> T cells were increased (3.1% vs 6.8% and 10% vs 14.9%, respectively), but this increase was only significant for CD4<sup>+</sup> (p=0.018) and did not reach statistical significance for CD8<sup>+</sup> (p=0.059). Consequently, absolute counts of activated CD4<sup>+</sup> and CD8<sup>+</sup> T cells were not significant differences were found related to proportions of CD8<sup>+</sup> subsets, CD4<sup>+</sup> T<sub>reg</sub> cells, Th1, Th2, TH17, B cells subpopulations, monocytes nor NK cells.

**Conclusions.** We found that pSS patients with lymphopenia have a lymphocyte population's distribution in PB similar to those pSS patients without lymphopenia, with decreased absolute count of all main lymphocyte subsets. However, pSS patients with lymphopenia have a lower proportion of naïve CD4<sup>+</sup> and a higher proportion of effector memory CD4<sup>+</sup> and activated CD4<sup>+</sup> than pSS patients without lymphopenia.

## **P-21**

#### Characterization of inflammatory cell infiltrate and potential survival niches of plasma cells in kidney biopsies of patients with Sjögren's syndrome

Borge H<sup>1</sup>, Marti H-P<sup>1,2</sup>, Leh S<sup>1,3</sup> and Skarstein K<sup>1,3</sup>.

<sup>1</sup>Department of Clinical Medicine, University of Bergen, Bergen, Norway. <sup>2</sup>Department of Medicine, Haukeland University Hospital, Bergen, Norway. <sup>3</sup>Department of Pathology, Haukeland University Hospital, Bergen, Norway.

**Background.** Sjögren's syndrome (SS) is an autoimmune disease affecting several organs and tissues. Renal involvement in patients with SS is known, but little is described about the histopathological findings and characteristic features of the inflammatory cell infiltrate in kidney biopsies from SS patients. Renal involvement can lead to severe kidney failure due to tubulointerstitial nephritis (TIN) or glomerulonephritis (GN).

**Objectives.** To characterize the microenvironment in the inflammatory cell infiltrate in kidney biopsies from patients with SS.

**Methods.** 83 patients who met the American-European Consensus Group Criteria for SS and with significant renal involvement (N=83), were selected from the Norwegian Kidney Biopsy Registry (NKBR). Patients with primary (pSS) where further subdivided into patients with TIN, GN or nonclassifiable glomerulonephritis (NCG). A group of patients with an undefined kidney diagnosis was also present in our patient cohort. From this cohort (n=83) we selected a group of 15 patients with TIN and pSS, 10 patients with GN and pSS, a group of 9 patients with UN together with pSS and 12 patients with a non-classifiable kidney disease and pSS. Immunohistochemistry (IHC) was used on formalin- fixed paraffin-embedded sections from patients with TIN, GN and NCG. Salivary gland biopsies from patients with pSS (n=3) was used as positive controls. Sections from patients with an undefined kidney diagnosis was not included in the IHC part of the study.

**Results.** Double staining with CXCL12 and CD138 showed a close relation between chemokines and plasma cells in the cell infiltrate and corresponds to our previous findings in salivary glands of patients with pSS. Interestingly, IL-6 was highly expressed in focal areas in biopsies from TIN and GN patients. Staining with PNAd for high endothelial venules (HEV) was negative in kidneys, but positive in salivary glands.

**Conclusion.** In the present study, we conducted a Norwegian nation-wide analysis of 83 biopsy-proven pSS-related nephropathies. Very few reports have described the histopathological findings in pSS-associated renal disease and little is known about the renal microenvironment and the inflammatory cell pattern in these patients. The detection of IL-6 and CXCL12 in close proximity to CD138\* plasma cells may indicate presence of long-lived plasma cells since these are vital factors for long-lived plasma cell survival niches.

### P-22

## Characterization of patients with association of Sjögren's syndrome and celiac disease

Antónia Szántó<sup>1</sup>, Zsófia Aradi<sup>1</sup>, Eszter Molnár<sup>1</sup>, Margit Zeher<sup>1</sup>. <sup>1</sup>University of Debrecen, Medical Faculty, Division of Clinical Immunology.

**Background.** Celiac disease (CD) is an immune-mediated enteropathy triggered by the consumption of gluten-containing cereals in genetically predisposed persons. Co-incidence of CD and Sjögren's syndrome (SS) has been observed among our patients. Therefore, we aimed to characterize the association of the two disease and to investigate how the presence of CD influences the disease course of SS.

**Methods.** Out of the patients followed-up because of SS at University of Debrecen, Medical Faculty, Division of Clinical Immunology, patients with the co-occurrence of CD were selected. Twenty-one such patients were found (SS+CD group). However, the data of 18 patients were included in the statistical analysis: one patient did not turn up during the examination period, while 2 patients exceeded 85 years of age so their data would have probably significantly influenced the results. Demographic, metabolic, endocrine and immunologic data of patients were compared to 17 age-matched patients with SS who do not suffer from CD (Control group). Statistical analysis was performed by using SPSS17.0 software. Results with p<0.05 were considered statistically significant.

Sjögren's syndrome (SS+CD: 8.83±6.81 years; Controls: 10.24±5.33 years). There was no significant difference regarding the occurrence of Hashimoto-thyroiditis, osteoporosis, glucocorticoid treatment, or in electrolyte concentrations, renal function, iron, folic acid and vitamin B12 serum levels. HbA-Ic was significantly lower in SS+CD patients ( $5.44\pm0.46\%$  vs  $5.81\pm0.39\%$ , p=0.014). SS+CD patients had significantly higher CRP ( $6.22\pm9.87$  vs  $2.67\pm4.08$  mg/l, p=0.017), prolactin ( $10.98\pm4.72$  vs  $7.85\pm3.41$ ug/L, p=0.028) and lower ANA titre values (p=0.010). In the SS+CD group, positive correlation was found between SS disease duration and prolactin levels (p=0.032). Vitamin D levels did not reach the lowest normal value (75 nmol/L) in any of the groups (SS+CD:  $59.23\pm14.28$  nmol/L; SS Controls:  $55.97\pm18.86$  nmol/L). Unfortunately, no difference was detected among patients formerly being suggested to take vitamin D and patients without such a suggestion (p=0.358).

Regarding extraglandular manifestations, polyarthritis occurred significantly more frequently in Control SS patients than in SS+CD patients (p=0.041). **Conclusions.** Diet of patients with SS+CD seems to be appropriate since their metabolism is not worse than that of the Control group. However, higher prolactin levels and lower ANA titres suggest that CD adversely influences the disease course of SS: hyperprolactinemia can accelerate autoimmune processes while lower ANA titres might indirectly indicate a higher level of autoimmunity. However, these differences are not represented in clinical symptoms, moreover, polyarthritis is more frequent in SS patients unaffected from CD.

Results of vitamin D levels refer to compliance problems: patients take insufficient dose of vitamin D and probably on an irregular basis. Accordingly, importance of vitamin D supplementation should be emphasized more effectively during the follow-up of our SS patients.

#### **P-23**

#### Subepithelial infiltrate of the vagina in primary Sjögren's syndrome: the cause of vaginal dryness?

J.F. van Nimwegen<sup>1</sup>, K. van der Tuuk<sup>2</sup>, E.R. Klinkert<sup>2</sup>, E.A. Haacke<sup>1,3</sup>, F.G.M. Kroese<sup>1</sup>, H. Hollema<sup>3</sup>, M.J. Mourits<sup>2</sup>, H. Bootsma<sup>1</sup>.

Department of <sup>1</sup>Rheumatology and Clinical Immunology, <sup>2</sup>Obstetrics and Gynaecology, <sup>3</sup>Pathology and Medical Biology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands.

**Background.** Women with primary Sjögren's syndrome (pSS) often experience vaginal dryness. Inflammation of the vaginal epithelium or endocervical mucosa might contribute to this symptom. This study evaluated whether female pSS patients have impaired vaginal health and inflammation of the vaginal epithelium or endocervical mucosa in comparison with controls.

Methods. Consecutive premenopausal women with pSS according to the AECG and ACR-EULAR criteria, with symptoms of vaginal dryness, and age-matched controls planned for a laparoscopic procedure underwent a comprehensive vaginal examination. Excluded were women with an inflammatory or infectious gynaecological disease, intra-uterine contraceptive device, or use of hormone therapy or DMARDs. Participants were screened for chlamydia, gonorrhea, and vaginal bacterial and fungal infections. The 5 domains of the vaginal health index (elasticity, fluid secretion, pH, epithelial mucosa, moisture) were scored by an experienced gynecologist on a scale of 1-5, resulting in a total score of 5-25. Low scores correspond to low vaginal health. In pSS patients, vaginal and endocervical biopsies were taken under local anesthesia. In controls, the investigation was performed under general anesthesia, prior to surgery. Formalin fixed and paraffin embedded tissue sections were stained for H&E, CD45, CD4, CD8, CD20 and CD138 and examined by a dedicated gynaecopathologist. The percentage of parenchyma stained for CD45 was calculated using the Positive Pixel Count algoritm in Aperio ImageScope v12.0.

**Results.** A total of 9 pSS patients and 8 controls was included. Median age was 36 years (IQR 33-46) in pSS patients and 41 years (IQR 36-44) in controls (p=0.61). Median vaginal health index was lower in pSS patients (19, range 14-23) compared to controls (23, range 20-25, p=0.015). The median percentage of area stained for CD45 in the vagina was higher in pSS patients (1.8%, range 1.0-5.0%) compared to controls (1.1%, range 0.4-1.4%, p=0.002). Infiltrates were mainly located in the subepithelial layer, with aggregates in dermal papillae (Figure 1) and consisted mostly of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes. Some B lymphocytes and few plasma cells were present, especially in larger infiltrates. Endocervical tissue was obtained in 6 pSS patients and in 5 controls. The median percentage of area stained for

CD45 in the endocervix did not differ significantly between groups (patients 6.4%, range 1.1-28.3%; controls 3.1%, range 0.4-22.4%, p=0.429). Endocervical infiltrates consisted of CD4<sup>+</sup> and CD8<sup>+</sup> T lymphocytes, B lymphocytes and plasma cells.

**Fig. 1.** Histopathology of vaginal biopsies from a patient with pSS and a control, stained for HE and CD45. Note the subepithelial infiltrate with aggregates in the dermal papillae of the pSS patient.



**Conclusions.** Vaginal health is impaired in premenopausal women with pSS. Women with pSS show significantly more infiltrating lymphocytes in the vagina, but not in the endocervix. A mild subepithelial infiltrate with aggregates in dermal papillae, consisting mostly of T lymphocytes, is present in the vagina of women with pSS.

#### **P-24**

#### Usefulness of "Guidance for the diagnosis of Sjögren's syndrome in pediatric patients (2015)": new criteria for the diagnosis of Sjögren's syndrome in children and adolescents

Minako Tomiita<sup>1</sup>, Ichiro Kobayashi<sup>2</sup>, Yuzaburo Inoue<sup>3</sup> Yukiko Nonaka<sup>4</sup>, Nami Okamoto<sup>5</sup>, Naomi Iwata<sup>6</sup>, Ryoki Hara<sup>7</sup>, Hiroaki Umebayashi<sup>8</sup>, Yasuhiko Itoh<sup>9</sup>, Masaaki Mori<sup>10</sup>.

Japanese Pediatric Sjögren's syndrome Study Group.

<sup>1</sup> Department of Allergy and Rheumatology, Chiba Children's Hospital, Chiba, Japan.<sup>2</sup> Department of Pediatrics, KKR Sapporo Medical Center, Sapporo, Japan. <sup>3</sup>Department of General Medical Science, Graduate School of Medicine, Chiba University, Ciba, Japan. <sup>4</sup>Department of Pediatrics, Kagoshima University Graduate School of Medicine, Cashima, Japan. <sup>5</sup>Department of Pediatrics, Graduate School of Immunology and Infectious Diseases, Aichi Children's Health and Medical Canter, Obu, Japan. <sup>7</sup>Department of Pediatrics, Miyagi Children's Hospital, Sendai, Japan. <sup>8</sup>Department of Rheumatics, Miyagi Children's Hospital, Sendai, Japan. <sup>9</sup>Department of Pediatrics, Nippon Medical School, Tokyo, Japan. <sup>10</sup>Department of Lifetime Clinical Immunology, Graduate School of Medical and Dental University, Tokyo, Japan.

**Background/Purpose.** The diagnosis of Sjögren's syndrome (SS) in pediatric patients is often difficult using currently available diagnostic or classification criteria, because patients in this age group lack sicca symptoms. Thus, new criteria for pediatric SS patients have been needed. Japanese Pediatric Sjögren's syndrome Study Group has developed new criteria for diagnosis of SS in pediatric age (Table I, II, Figure 1). Our criteria were approved by both the board of the Japanese Society for Sjögren's syndrome and the Pediatric Rheumatology Association of Japan. In this study, we analyzed the usefulness of our criteria during the follow-up period.

**Methods.** We enrolled 41 pediatric patients who were diagnosed as having SS with unanimity by 10 pediatric rheumatologists. Twenty-five were primary SS (pSS), 11 were secondary SS (sSS) and 5 were primary to secondary SS (psSS), who were diagnosed pSS first but developed other col-

## 14<sup>th</sup> International Symposium on Sjögren's Syndrome

Table I. Scoring: Evidence for autoimmunity and exocrinopathy.

Serological score Test items	Criteria	Score
Serum IgG	≥97.5 <sup>th</sup> percentile for age	1
anti-nuclear antibody	1:40-1:80	1
	1:160	2
	≥1: 320	3
Rheumatoid factor	≥15U/ml	3
anti-SSA/Ro or SSB/La antibody	positive	6
Glandular score (1) Salivary glands		
Test Items	Criteria	Score
Labial salivary gland biopsy	<1 focus / 4mm <sup>2</sup>	1
	$\geq 1$ focus / 4mm <sup>2</sup>	2
Solography (conventional or MRI)	Rubin-Holt stage ≥1	2
Salivary scintigraphy	Decreased in uptake or secretion	1
Decreased salivary flow	Saxon test ≤2.Og / 2min, or	1
(counted if at least one other test is positive)	Salivary flow rate ≤1.5ml / 15min, or Gum test ≤10ml / 10min	
(2) Lacrimal glands		
Test items	Criteria	Score
Schirmer test and Rose-Bengal test	Schirmer test <5mm/5min and	2
C C	Rose-Bengal test van Bijsterveld score ≥3	3
Schirmer test and fluorescein test	Schirmer test <5mm/5min and fluorescein test (+)	2
ACR score	≥3	2

#### Table II. Classification.

Serological score*	Glandular score**				
	≥2	1	0		
≥6	Definite	Probable	Possible		
5	Probable	Probable	Possible		
4	Probable	Probable	Possiable		
3	Probable	Possible	Need follow-up		
2	Probable	Possible	Need follow-up		
1	Possible	possible	Need follow-up		
0	Need follow-up	Need follow-up	Possibly non-SS		

\*\*For serological score is the total score of serological parameters.

\*For glandular score is the highest one: total score of score of salivary gland test items or lacrimal gland score.



Fig. 1. Algorithm for diagnosis of Sjögren's syndrome in children and adolescent.

**Results.** According to our criteria, patients were classified into 3 groups: definite, probable and possible. At the first visit, the number of patients in each group was 33, 2 and 6, and at the last visit, that was 38, 3 and 0, respectively. Four patients in the "possible" and 1 patient in the "probable" at the 1<sup>st</sup> visit were classified as "definite" at the last visit, and 2 patients in the "possible" at the 1<sup>st</sup> visit were classified as "probable" at the last visit. No patients were diagnosed as non-SS. In the other 4 criteria, the most sensitive was JPN, followed by A/E and ACR. However, even by using JPN criteria, 12% of pSS patients diagnosed as non-SS at the last visit.

**Conclusion.** Our new criteria are useful for diagnosis and follow-up of pediatric SS, and make it possible to recognize SS-associated complications at an early stage.

## P-25

Towards a better understanding of childhood Sjögren's syndrome: evaluation of the 2016 ACR/EULAR classification criteria for use in diagnosing Sjögren's syndrome in children

Scott M Lieberman<sup>1</sup>; for the International Childhood Sjögren's Syndrome Workgroup

<sup>1</sup>Stead Family Department of Pediatrics, University of Iowa, Iowa, USA.

**Background.** Clinical presentation of Sjögren's syndrome in children differs from that in adults: dryness symptoms are more common in adults, while parotitis is more common in children. Criteria developed for adult classification have demonstrated low sensitivity when applied to pediatric populations, and no child-specific criteria have been established. The latest adult classification criteria have not yet been evaluated for use in children. Our objective was to evaluate the applicability of these new criteria for use in children.

**Methods.** Retrospective chart reviews were conducted to collect individual patient level data for children diagnosed with Sjögren's syndrome (based on clinical diagnosis at age <18 years). Data including clinical features, laboratory values, imaging studies, and test items in the 2016 ACR/EULAR criteria were collected, and de-identified data were entered into a REDCap database. This study was approved by the Institutional Review Boards or equivalent regulatory bodies at individual affiliate institutions.

Results. To date, 86 children with Sjögren's syndrome were included from 11 institutions across 4 countries (data collection is ongoing). This constitutes the largest childhood Sjögren syndrome patient series to date. The majority of children (91%) were female with a mean age of 11.6 years at diagnosis (range 1-17.8 years). Twelve children (14%) also had another autoimmune disease (9 with SLE, 2 with uveitis, 1 with subacute cutaneous lupus). Frequency of clinical features were as follows: 51% with parotitis, 50% with dry eyes, 48% with dry mouth, 45% with arthralgias without arthritis, 24% with lymphadenopathy, 23% with arthritis, 14% with cytopenias, 14% with fevers, 13% with cutaneous vasculitis, 10% with weight loss, and <10% each with recurrent vaginitis, myositis, pulmonary, renal, or neurologic manifestations. Only 3 children had testing for all 5 items included in the 2016 ACR/EULAR criteria. Most children (95%) had testing for anti-SSA antibodies, but fewer underwent minor salivary gland (MSG) biopsy (50%), Schirmer testing (51%), measurement of unstimulated whole saliva flow (UWSF, 13%), or ocular surface staining (OSS, 19%). While most children studied (96.5%) were missing at least one data point, 27 of 86 children (31%) met the 2016 ACR/EULAR classification criteria for Sjögren's syndrome. Of these 27 children: 25 (93%) had positive anti-SSA antibodies; 16 (59%) had positive MSG biopsy; 21 (78%) had positive Schirmer test; 2 (7%) had positive UWSF; and 1 (4%) had positive OSS. Of the 59 children not meeting criteria: 39 (66%) had positive anti-SSA antibodies; 6 (10%) had positive MSG biopsy; 6 (10%) had positive Schirmer test; and 2 (3%) had positive UWSF.

**Conclusions.** Criteria items from the 2016 ACR/EULAR criteria are not routinely assessed in children diagnosed with Sjögren's syndrome making formal retrospective assessment of criteria difficult. Prospective study of these criteria along with defining child-specific normal values and adding child-specific criteria items (such as recurrent parotitis) are warranted. Establishing criteria for childhood Sjögren's syndrome is a key step toward better understanding and treating this condition.

P-26

Primary Sjögren's syndrome in a subset of children with recurrent salivary glands enlargement – it is high time for pediatric diagnostic criteria!

Anna Pomorska<sup>1</sup>, Wojciech Kosiak<sup>2</sup>, Dominik Świętoń<sup>3</sup>, Rafał Pęksa<sup>4</sup>, Barbara Kochańska<sup>5</sup>, Justyna Kowalska-Skabara<sup>5</sup>, Juliusz Chorążewicz<sup>6</sup>, Ninela Irga-Jaworska<sup>1</sup>.

<sup>1</sup>Department of Pediatrics, Hematology and Oncology, Medical University of Gdansk, Poland. <sup>2</sup>Laboratory of Diagnostic Ultrasound and Biopsy, Department of Pediatrics, Hematology and Oncology, Medical University of Gdansk, Poland. <sup>3</sup>Department of Radiology, Medical University of Gdansk, Poland. <sup>4</sup>Department of Pathology, Medical University of Gdansk, Poland. <sup>5</sup>Department of Conservative Dentistry, Medical University of Gdansk, Poland. <sup>6</sup>Department of Ophthalmology, Medical University of Gdansk, Poland.

**Background.** The objective was to evaluate applicability of the currently used primary Sjögren's syndrome (pSS) diagnostic criteria in pediatric population and to assess how salivary gland ultrasonography (SGUS) with elastography and salivary gland magnetic resonance with sialography (MR) might improve diagnosis in this group of patients.

**Methods.** Thirty-two patients, aged 2-17 years (15 females, 17 males) with recurrent salivary glands enlargement suggestive for diagnosis of pSS were enrolled in this prospective study. The assessement was based on the criteria included in 2002 AECG, 2012 ACR and 2016 ACR/EULAR classifications: ocular and oral dryness symptoms, serological tests (ANA, SSA, SSB, RF), ophtalmological examination (Schirmer test, ocular staining with fluoresceine and lisamine green), unstimulated whole salivary flow (UWSF) rate and labial minor salivary gland (LSG) biopsy. Moreover, parallel radiological examinations, SGUS with elastography and MR with sialography, were performed by radiology specialists. The study was approved by the Local Bioethics Committee of Medical University of Gdansk; written informed consent of the parents of all patients and consent of patients who were older than 16 years (in accordance with polish low) were obtained.

**Results.** Subjective ocular and oral dryness symptoms were reported in 5/32 (16%) and 11/32 (34%) patients, respectively. Recurrent salivary gland enlargement was observed in all patients.

Schirmer test was performed in 18/32 patients and in all of these children was normal. Ocular staining score (OSS)  $\geq$ 3 was not observed in our patients. UWSF rate  $\leq$ 0,1 was found in 7/30 patients (23%).

Serological results revealed positive ANA ( $\geq$ 1:320) in 28/32 patients (87,5%), positive SSA in 2/32 (6%) and positive SSB in 1/32 (3%). RF was elevated in 15/32 patients (47%).

LSG biopsy was performed in 19 patients: 13 patients had focus score (FS)  $\geq 1$  (68%); 5 patients had evidence of focal lymphocytic sialadenitis (FLS) with FS >0 (26%); 1 patient had FS 0.

SGUS and MR revealed SS-like changes in major salivary glands in 25/32 (78%) and 27/32 (84%) patients, respectively.

4 patients met 2002 AECG criteria, 8 patients 2012 ACR criteria and 4 patients 2016 ACR/EULAR criteria.

**Conclusions.** Currently used diagnostic criteria for adult pSS are not applicable at the onset of the disease in children. Some of the tests used in adults are imposible to perform in children because of the young age and lack of cooperation with a child. Sicca symptoms develop with delay so they are not helpful in diagnosing pSS at the early stage of the diasease. Radiological examination, especially SGUS, should be considerd to be one of the criterion in the diagnosis of childhood pSS. There is an urgent need for child-specific criteria.

## P-27

## ACR (2012) and ACR/EULAR (2016) classification criteria for primary Sjögren's syndrome on Russian cohort of patients

Khvan Y.I.<sup>1</sup>, Palshina S.G.<sup>1</sup>, Rodionova E.B.<sup>1</sup>, Safonova T.N.<sup>2</sup>, Vasiliev V.I.<sup>1</sup> Nasonova Research Institute of Rheumatology, Moscow, Russian Federation. <sup>2</sup>Research Institute of Ophthalmic Diseases, Moscow, Russian Federation.

**Objectives.** To assess ACR 2012 and ACR/EULAR 2016 criteria in 41 patients with primary Sjögren's syndrome patients fulfilling Russian criteria. **Methods.** From the middle of 2016 in the prospective study, carried out at one division of Nasonova Research Institute of Rheumatology, 41 patients (40 female, 1 male) with the mean age 52,0±13.9 years (min 28; max

82), was newly diagnosed primary Sjögren's syndrome (pSS) according to Russian criteria. There are Russian criteria for pSS: I) keratoconjunctivitis sicca (Schirmer's test <10 mm/5 min or fluorescein staining of the cornea or conjunctiva I-III degrees or tear break-up time <10 sec), II) sialadenitis (sialectasia on parotid sialography (obligatory) ± stimulated saliva flow test<2,5 ml/ 5min ± labial salivary gland biopsy with focus score (FS) of ≥2foci/4mm<sup>2</sup>), III) positive antinuclear antibody (ANA) or positive ANA with rheumatoid factor (RF) or anti-SSA (anti-Ro) or/and SSB (anti-La). pSS is proved if the first two criteria and at least one of the immunological criteria meet. We evaluated presence of ocular, oral dryness, anti-SSA/SSB (anti-Ro/La) antibodies, rheumatoid factor (RF), Schirmer's test (≤5 mm/5 minutes), stimulated saliva flow test (SFT) <2,5ml/5 min, ocular staining score (OSS) ≥5, OSS ≥3, labial salivary gland biopsy with focus score (FS) of ≥1foci/4mm<sup>2</sup>, ANA (titer ≥1/320), sialectasia on parotid sialography. Statistical analyses (Pearson Chi-square, Spearman Rank R) were performed using STATISTICA version 12.

**Results.** According to our data 24-32% patients fulfilling Russian criteria for pSS didn't have sicca symptoms and 61% patients manifested with milder KCS (OSS<3-5) and in 49% Schirmer's test was more than 5,0 mm but less than 10mm/5 min.

**Conclusion.** Patients with pSS according to ACR (2012) and/or ACR/EU-LAR (2016) criteria seem to be diagnosed on the more progressed disease stages.

### **P-28**

#### Characteristics of Korean primary Sjögren's syndrome and comparison of 3 different classification criteria for primary Sjögren's syndrome: data from Korean KISS Cohort

Jennifer Lee<sup>1</sup>, Jung Hee Koh<sup>1</sup>, Ji-Won Kim<sup>1</sup>, Yoon-Kyoung Sung<sup>2</sup>, Shin-Seok Lee<sup>3</sup>, Jung Yoon Choe<sup>4</sup>, Seung-Cheol Shim<sup>5</sup>, Hyun-Sook Kim<sup>6</sup>, Hae-Rim Kim<sup>7</sup>, Ji-Min Kim<sup>8</sup>, Sung Ryul Kwon<sup>9</sup>, Sang Il Lee<sup>10</sup>, Kichul Shin<sup>11</sup>, Chang Hoon Lee<sup>12</sup>, Seung-Ki Kwok<sup>1</sup>, Ji Hyeon Ju<sup>1</sup>, Sung-Hwan Park<sup>1</sup>. <sup>1</sup>Division of Rheumatology, Department of Internal Medicine, Seoul St Mary's Hospital, College of Medicine, The Catholic University of Korea, Seoul, South Korea.

<sup>1</sup>Division of Rheumatology, Department of Internal Medicine, Seoul St Mary's Hospital, College of Medicine, The Catholic University of Korea, Seoul, South Korea. <sup>2</sup>Department of Rheumatology, Hanyang University Hospital for Rheumatic Diseases. Seoul, South Korea. <sup>3</sup>Department of Rheumatology, Chonnam National University Medical School and Hospital, Gwangju, Republic of Korea. <sup>4</sup>Division of Rheumatology, Daegu Catholic University Medical Center, Daegu, Korea. <sup>5</sup>Division of Rheumatology, Daegu Catholic University Medical Center, Daegu, Korea. <sup>5</sup>Division of Rheumatology, Department of Internal Medicine, School of Medicine, Chungnam National University, Chungnam National University Hospital, Daejeon, Korea. <sup>6</sup>Division of Rheumatology, Department of Internal Medicine, Soonchunhyang University Seoul Hospital, Soonchunhyang University College of Medicine, Konkuk University Medical Center, Seoul, Korea. <sup>8</sup>Division of Rheumatology, Department of Internal Medicine, Keimyung University Dongsan Medical Centre, Daegu, South Korea. <sup>9</sup>Division of Rheumatology, Department of Internal Medicine, Inha University School of Medicine, Incheon, Korea. <sup>10</sup>Division of Rheumatology, Department of Internal Medicine, Gyeongsang National University School of Medicine, Snub Autonal University School of Medicine, Soul, Korea. <sup>10</sup>Division of Rheumatology, Department of Internal Medicine, Gyeongsang National University School of Medicine, Snub Autonal University School of Medicine, Norea. <sup>10</sup>Division of Rheumatology, Department of Internal Medicine, Snub Boramae Medical Center, Seoul, Korea. <sup>11</sup>Division

**Objective.** The objective of this study is to introduce the clinical and laboratory characteristics of Korean primary Sjogren's syndrome (pSS) patients enrolled in Korean Initiative of Sjögren's Syndrome (KISS). We also sought to compare the performance of newly proposed 2016 American College of Rheumatology (ACR)/European League-Against Rheumatism (EULAR) criteria to 2002 American-European Consensus Group (AECG) and 2012 ACR classification criteria for primary pSS in a well characterized registry. **Methods.** Patients with pSS from 12 university affiliated hospitals in Korea were enrolled from Oct 2013 to JAN 2017. The patients were diagnosed with pSS by either fulfilling 2002 AECG or 2012 ACR classification criteria. Data of clinical manifestations and various laboratory findings were obtained.

**Results.** Data at inclusion were available in 458 patients. The mean age was 51.9±11.8 years. Four hundred fifty patients (98.3%) were female. Mean disease duration was 33.9±46.7 months. Most common extraglandular manifestation was arthralgia (47.8%) followed by Raynaud's phenomenon (15.7%) and lymphadenopathy (13.5%). Median [interquartile range] ES-SDAI and ESSPRI was 2 [0-5], 5 [4-6.7], respectively. Among 458 patients, 328 patients had sufficient data to determine the fulfillment of each criteria. All three criteria were met by 307 patients. Among 3 patients by whom 2016 ACR/EULAR criteria were not met, one 2002 AECG- 2015 ACR+ had negative anti Ro or La while positive antinuclear antibody and rheuma-

toid factor with more than 3 ocular staining score. Two 2002 AECG+ 2015 ACR- showed no ocular sign and negative anti Ro or La while had positive focus score. Ninety-six patients had results of all items in 2016 criteria, 95 met the criteria.

Table I. Characteristics of patients with primary Sjögren's syndrome (n=41).

Parameters	Results
oral + ocular dryness	21 (51%)
ocular dryness	28 (68%)
oral dryness	31 (76%)
anti-SSA (anti-Ro) positive (>25 IU/ml)	37 (90%)
anti-SSB (anti-La) positive (>25 IU/ml)	18 (44%)
RF positive >2UNL (>30 IU/ml)	27 (66%)
ANA ≥1:320	39 (95%)
Schirmer's test (<5mm/5 minutes)	25 (61%)
stimulated SFT ≤2,5ml/5 min	32 (78%)
OSS ≥5	16 (39%)
OSS ≥3	16 (39%)
FS ≥1foci/4mm <sup>2</sup>	31 (76%)
Sialectasia on parotid sialography	39 (95%)

33/41 patients (80%) fulfilled ACR (2012) criteria for pSS, 37 patients (90%) - ACR/ EULAR (2016) criteria. In our small cohort of pSS patients we didn't find any strong correlation between parameters, even among sialectasia and FS.

**Conclusion.** We successfully established a nationwide pSS registry in Korea, which represents the characteristics of Korean pSS patients. The newly proposed 2016 ACR/EULAR criteria were met by most of the patients diagnosed with pSS according to previous criteria.

### P-29

#### Transcriptomic signatures distinguish AECG-classified primary Sjögren's from sicca control patients

Narayanan Perumal<sup>1</sup>, Angus MacDonald<sup>1</sup>, Ernst Dow<sup>1</sup>, Josh Poorbaugh<sup>1</sup>, Sean Sissons<sup>1</sup>, Karen Cox<sup>1</sup>, Kiely Grundahl<sup>2</sup>, A. Darise Farris<sup>2</sup>, Astrid Rasmussen<sup>2</sup>, Jennifer Kelly<sup>2</sup>, Robert Benschop<sup>1</sup>, Kathy Sivils<sup>2</sup>.

<sup>1</sup> Immunology – Translational Sciences, Eli Lilly & Co., Indianapolis, IN, USA. <sup>2</sup>Arthritis and Clinical Immunology Program, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA

**Background.** There is a pressing need for biomarkers to aid selection of appropriate patients and accurately measure outcomes in clinical trials for Sjögren's Syndrome (SS). We assessed therapeutic targets of interest, BAFF, IL-17A and IL-21 in plasma of patients classified with pSS by AECG criteria vs sicca controls (SC who met some of the criteria for pSS but not sufficient for definitive classification), vs healthy controls (HC). We also compared transcriptomes of pSS, SC and HC donors. The goals were to identify potential candidate biomarkers for more stringent selection of pSS subjects into clinical trials and potential new target identification in pSS.

**Methods.** Study subjects: pSS (AECG criteria), n=21, mean age 50.7 (17-71), F=21; sicca controls, n=21, mean age 49.1 (28-77), F=21; age- and sex-matched healthy controls (HC), n=41. Clinical tests were concurrent with whole blood RNA collection: Lissamine Green, Schirmer's test, Whole Unstimulated Salivary Flow (WUSF), Labial Salivary Gland (LSG) histopathology, anti-Ro (composite), anti-La, anti-Ro60, anti-Ro52 and Rheumatoid Factor. Plasma BAFF, IL-17A and IL-21 assays were measured in validated immunoassays. pSS and SC whole blood RNA samples were run on Affymetrix HTA 2.0 arrays and gene expression compared to HCs.

**Results.** There was considerable overlap between plasma concentrations of BAFF, IL-21 and IL-17A in pSS and SC compared with healthy controls. Gene expression revealed fewer than expected statistically significant transcripts (unadjusted p<1E-05) distinguishing pSS and SC (114 up and 194 down in pSS vs. SC). However, using a panel of pSS vs HC discriminatory genes as the basis for self-organizing maps, we observed that not all SC showed gene expression patterns similar to HC, with 6 of 21(28.6%) being more similar to pSS patients. At transcript level splicing, cumulative differences of SC and pSS patients vs HCs displayed a number of immune relevant genes (MYO1G, RAC2, CORO1A, LAPTM5, CD53 and LSP1) as top hits. Two clinical measures, LSG Focus Score and WUSF demonstrated positive and negative correlations respectively with patient differential expression scores, a measure of gene expression intensity derived from the discriminatory genes.

**Conclusions.** Initial plasma cytokine screening showed a substantial overlap between sicca controls and pSS compared with healthy control donors. Transcriptomic analysis, however, revealed that the pSS patients were distinct from the majority (71.4%) of sicca control donors.

## P-30

#### Addition of salivary gland ultrasound increases the feasibility of the ACR-EULAR classification criteria in primary Sjögren's syndrome

Jolien F. van Nimwegen<sup>1</sup>, Esther Mossel<sup>1</sup>, Martha S. van Ginkel<sup>1</sup>, K. Delli<sup>2</sup>, Alja J. Stel<sup>1</sup>, Frans G.M. Kroese<sup>1</sup>, Fred K.L. Spijkervet<sup>2</sup>, Arjan Vissink<sup>2</sup>, Suzanne Arends<sup>1</sup> and Hendrika Bootsma<sup>1</sup>.

Departments of <sup>1</sup>Rheumatology and Clinical Immunology and <sup>2</sup>Oral and Maxillofacial Surgery, University of Groningen and University Medical Center Groningen, Groningen, the Netherlands.

**Background.** The ACR-EULAR criteria were recently developed to reach international consensus regarding the classification of primary Sjögren's syndrome (pSS). Our objective was to assess whether addition of salivary gland ultrasound (SGUS) to the ACR-EULAR criteria influences the performance of these criteria in a large cohort of patients clinically suspected or diagnosed with pSS in daily clinical practice.

**Methods.** Included were all consecutive patients who underwent SGUS between October 2014 and July 2017, who had complete data on all ACR-EULAR items. Classification according to the criteria was determined separately in patients who underwent a labial or parotid gland biopsy. For SGUS, the average score for hypoechogenic areas in the parotid and submandibular glands on one side was applied (range 0-3)1. The optimal cut-off value for our SGUS score was determined using ROC analysis. Clinical diagnosis by the treating physician was used as gold standard. SGUS positivity was added as an item to the original ACR-EULAR criteria. The weight of the original ACR-EULAR items was kept and SGUS positivity was given a weight of 1 point. Area under the curve (AUC), absolute agreement, sensitivity and specificity of the original and adjusted ACR-EULAR criteria sets were determined.

**Results.** Of the 363 patients assessed, 254 patients had a complete data set. 156 patients were diagnosed with pSS. The accuracy of SGUS to predict clinical diagnosis was good, with an AUC of 0.873 and optimal cut-off value of  $\geq 1.5$ . The optimal cut-off value of the criteria to discriminate between pSS and non-pSS remained 4, irrespective of the type of biopsy used and whether SGUS was added to the criteria or not.

In patients who underwent a labial gland biopsy (n=130), the original ACR-EULAR criteria showed an AUC of 0.967. Absolute agreement with clinical diagnosis was 94.6%, sensitivity was 96.2% and specificity was 92.3%. After addition of SGUS, the adjusted criteria showed an AUC of 0.969, absolute agreement of 94.6%, sensitivity of 97.4% and specificity of 90.4% (Figure 1).



Fig. 1. ROC curves of the original and adjusted ACR-EULAR (addition of SGUS) classification criteria. Between brackets the type of salivary gland biopsy is shown that was used for classification.

In patients who underwent a parotid gland biopsy (n=208), the original ACR-EULAR criteria showed an AUC of 0.957. Absolute agreement with clinical diagnosis was 92.8%, sensitivity was 92.1% and specificity was 94.0%. After addition of SGUS, the adjusted criteria showed an AUC of 0.965, absolute agreement of 92.3%, sensitivity of 92.9% and specificity of 91.6% (Figure 1).

**Conclusions.** SGUS is non-invasive, cheap and easy to perform in a rheumatologist outpatient setting. Addition of SGUS to the ACR-EULAR criteria resulted in negligible changes in the performance of the criteria, irrespective of the type of biopsy performed. Thus, adding SGUS to the ACR-EULAR criteria increases its feasibility, since clinicians are offered more options that could lead to fulfillment of these criteria.

References

1. MOSSEL et al.: Ann Rheum Dis 2017: Accepted for Publication.

### P-31

#### Comparison of 2002 AECG and 2016 ACR/EULAR classification criteria and added value of salivary gland ultrasonography in a patient cohort with suspected primary Sjögren's syndrome

Divi Cornec<sup>1,2</sup>, Maëlle Le Goff<sup>1</sup>, Sandrine Jousse-Joulin<sup>1,2</sup>, Dewi Guellec<sup>1</sup>, Sebastian CostA<sup>2,3</sup>, Thierry Marhadour<sup>1</sup>, Rozenn Le Berre<sup>4</sup>, Steeve Genestet<sup>5</sup>, Béatrice Cochener<sup>6</sup>, Sylvie Boisrame-Gastrin<sup>7</sup>, Yves Renaudineau<sup>2,8</sup>, Jacques-Olivier Pers<sup>2,7</sup>, Alain Saraux<sup>1,2</sup>, Valérie Devauchelle-Pensec<sup>1,2</sup>. <sup>1</sup>Rhumatologie, CHRU Brest, Brest, France. <sup>2</sup>INSERM UMR1227, Lymphocytes B et

<sup>1</sup>Rhumatologie, CHRU Brest, Brest, France. <sup>2</sup>INSERM UMR1227, Lymphocytes B et Autoimmunité, Université de Bretagne Occidentale, Brest, France. <sup>3</sup>Anatomie et Pathologie, CHRU Brest, Brest, France. <sup>4</sup>Médecine Interne, CHRU Brest, Brest, France. <sup>5</sup>Explorations Fonctionnelles Neurologiques, CHRU Brest, Brest, France. <sup>6</sup>Dahtalmologie, CHRU Brest, Brest, France. <sup>7</sup>Odontologie, CHRU Brest, Brest, France. <sup>8</sup>Laboratoire d'Immunologie et Immunothérapie, CHRU Brest, Brest, France.

**Objective.** To evaluate concordance between 2002 AECG and 2016 ACR/ EULAR classification criteria for primary Sjögren's syndrome (pSS) and to assess how salivary gland ultrasonography (SGUS) might improve the classification of patients.

**Methods.** Patients with suspected pSS underwent a standardised evaluation, including SGUS, at inclusion into the single-centre Brittany DIApSS cohort. Agreement between the two criteria sets was assessed using Cohen's  $\kappa$  coefficient. Characteristics of discordant patients were detailed.

**Results.** We prospectively included 290 patients between 2006 and 2016, among whom 125 (43%) met ACR/EULAR criteria and 114 (39%) also met AECG criteria; thus, 11 (4%) patients fulfilled only ACR/EULAR, no patients AECG only, and 165 (57%) patients neither criteria set. Concordance was excellent ( $\kappa$ =0.92). Compared to patients fulfilling both criteria sets, the 11 patients fulfilling only ACR/EULAR criteria had similar age and symptom duration but lower frequencies of xerophthalmia and xerostomia (*p*<0.01 for each) and salivary gland dysfunction (*p*<0.01); most had systemic involvement (91%), including three (27%) with no sicca symptoms; 91% had abnormal salivary gland biopsy and 46% anti-SSA; 64% were diagnosed with pSS by the physician. SGUS was abnormal in 12% of the 165 patients fulfilling no criteria set. Including SGUS among the ACR/EULAR criteria increased sensitivity from 87.4% to 91.1% when physician diagnosis was the reference standard.

**Conclusions.** Agreement between AECG and ACR/EULAR criteria sets is excellent. ACR/EULAR criteria are slightly more sensitive and classified some patients without sicca symptoms as having pSS. Including SGUS into ACR/EULAR criteria may further improve their sensitivity.

## P-32

#### Systemic Sjögren presenting without sicca syndrome: characterization of 240 patients according to the new 2017 ACR/ EULAR Classification Criteria

Soledad Retamozo<sup>1,2,3</sup>, Nihan Acar-Denizli<sup>4</sup>, Margit Zeher<sup>5</sup>, Kathy Sivils<sup>6</sup>, Thomas Mandl<sup>7</sup>, Raphaele Seror<sup>8</sup>, Xiaomei Li<sup>9</sup>, Chiara Baldini<sup>10</sup>, Xavier Mariette<sup>8</sup>, Jacques-Eric Gottenberg<sup>11</sup>, Debashish Danda<sup>12</sup>, Roberta Prioril<sup>3</sup>, Luca Quartuccio<sup>14</sup>, Gabriela Hernandez-Molina<sup>15</sup>, Berkan Armagan<sup>16</sup>, Aike A. Kruize<sup>17</sup>, Seung-Ki Kwok<sup>18</sup>, Marie Wahren-Herlenius<sup>19</sup>, Sonja Praprotnik<sup>20</sup>, Damien Sene<sup>21</sup>, Elena Bartoloni<sup>22</sup>, Maureen Rischmueller<sup>23</sup>, Roser Solans<sup>24</sup>, Yasunori Suzuki<sup>25</sup>, David Isenberg<sup>26</sup>, Valeria Valim<sup>27</sup>, Piotr Wiland<sup>28</sup>, Gunnel Nordmark<sup>29</sup>, Guadalupe Fraile<sup>30</sup>, Hendrika Bootsma<sup>31</sup>, Takashi Nakamura<sup>32</sup>, Roberto Giacomelli<sup>33</sup>, Valerie Devauchelle-Pensec<sup>34</sup>, Benedikt Hofauer<sup>35</sup>, Michele Bombardieri<sup>36</sup>, Virginia Fernandes Moça Trevisani<sup>37</sup>, Daniel Hammenfors<sup>38</sup>, Sandra G. Pasoto<sup>39</sup>, Steven E. Carsons<sup>40</sup>, Tamer A Gheita<sup>41</sup>, Fabiola Atzeni<sup>42</sup>, Jacques Morel<sup>43</sup>, Cristina Vollenveider<sup>44</sup>, Pilar Brito-Zerón<sup>1,45</sup>, Manuel Ramos-Casals<sup>1</sup>, on behalf of the Sjogren Big Data Consortium.

Sjögren Syndrome Research Group (AGAUR), Laboratory of Autoimmune Diseases Josep Font, IDIBAPS-CELLEX, Department of Autoimmune Diseases, ICMiD, University of Barcelona, Hospital Clínic, Barcelona, Spain. <sup>2</sup>Hospital Privado Universitario de Córdoba, Instituto Universitario de Ciencias Biomédicas de Córdoba (IUCBC), Córdoba- Argentina. <sup>3</sup>Instituto De Investigaciones En Ciencias De La Salud (INICSA), Universidad Nacional de Córdoba (UNC), Consejo Nacional de Investigaciones Cientíofficas y Técnicas (CONICET) - Córdoba - Argentina. <sup>4</sup>Department of Statistics, Faculty of Science and Letters, Mimar Sinan Fine Arts University, Istanbul, Turkey. <sup>5</sup>Division of Clinical Immunology, Faculty of Medicine, University of Debrecen, Debrecen, Hungary. <sup>6</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Re-search Foundation, Oklahoma City, OK, USA. <sup>7</sup>Department of Rheumatology, Malmö University Hospital, Lund University, Lund, Sweden. <sup>8</sup>Center for Immunology of Viral Infections and Autoimmune Diseases, Assistance Publique - Hôpitaux de Paris, Hôpitaux Universitaires Paris-Sud, Le Kremlin-Bicêtre, Université Paris Sud, INSERM, Paris, France. 9Department of Rheumatology and Immunology, Anhui Provincial Hospital, Hefei, China. <sup>10</sup>Rheumatology University of Pisa, Pisa, Italy. <sup>11</sup>Department of Rheumatology, Strasbourg University Hospital, Université de Strasbourg, CNRS, Strasbourg, France. 12Department of Clinical Immunology & Rheumatology, Christian Medical College & Hospital, Vellore, India. 13Department of Internal Medicine and Medical Specialties, Rheumatology Clinic, Sapienza University of Rome, Rome, Italy. <sup>4</sup>Clinic of Rheumatology, Department of Medical and Biological Sciences, University Hospital "Santa Maria della Misericordia", Udine, Italy. 15Immunology and Rheumatology Department, Instituto Nacional de Ciencias Médicas y Nutrición Salvador Zubirán. México City, Mexico. <sup>16</sup>Department of Internal Medicine, Division of Rheumatology, Hacettepe University Faculty of Medicine, Ankara, Turkey. <sup>17</sup>Department of Rheumatology and Clinical Immunology, University Medical Center Utrecht, Utrecht, The Netherlands. <sup>18</sup>Seoul St. Mary's Hospital, The Catholic University of Korea, Seoul, South Korea. <sup>19</sup>Department of Medicine, Solna, Unit of Experimental Rheumatology, Karolinska Institutet, and Karolinska University Hospital, Stockholm, Sweden. 20Department of Rheumatology, University Medical Centre, Ljubljana, Slovenia.<sup>21</sup>Service de Médecine Interne 2, Hôpital Lariboisière, Université Paris VII, Assistance Publique-Hôpitaux de Paris, 2, Paris, France. <sup>22</sup>Rheumatology Unit, Department of Medicine, University of Perugia, Perugia, Italy. <sup>23</sup>Department of Rheumatology, School of Medicine, The University of Western Australia, Crawley, Australia. 24Department of Internal Medicine, Hospital Vall d'Hebron, Barcelona, Spain. 25 Division of Rheumatology, Kanazawa University Hospital, Kanazawa, Ishikawa, Japan. <sup>26</sup>Centre for Rheumatology, Division of Medicine, University College London, London, UK. <sup>27</sup>Department of Medicine, Federal University of Espírito Santo, Vitória, Brazil. 28 Department of Rheumatology and Internal Medicine Wroclaw Medical Hospital, Wroclaw, Poland. 29Rheumatology, Department of Medical Sciences, Uppsala Iniversity, Uppsala, Sweden. <sup>30</sup>Department of Internal Medicine, Hospital Ramón y Cajal, Madrid, Spain. <sup>31</sup>Department of Rheumatology & Clinical Immunology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands. <sup>32</sup>Department of Radiology and Cancer Biology, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan. <sup>33</sup>Clinical Unit of Rheuma-tology, University of l'Aquila, School of Medicine, L'Aquila, Italy. <sup>34</sup>Rheumatology De-partment, Brest University Hospital, Brest, France. <sup>35</sup>Hals-Nasen-Ohrenklinik und Poliklinik, Technische Universität München, München, Germany. <sup>36</sup>Centre for Experimental Medicine and Rheumatology, Queen Mary University of London, London, UK. <sup>37</sup>Federal University of São Paulo, Sao Paulo, Brazil. <sup>38</sup>Department of Clinical Science, University of Bergen; and Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>39</sup>Rheumatology Division, Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo (USP), São Paulo, Brazil. <sup>40</sup>Division of Rheumatology, Allergy and Immunology Winthrop-University Hospital, Stony Brook University School of Medicine, Mineola, NY, USA. <sup>41</sup>Rheumatology Department, Kasr Al Ainy School of Medicine, Cairo University, Cairo, Egypt. 42IRCCS Galeazzi Orthopedic Institute, Milan, Italy. <sup>43</sup>Department of Rheumatology, Teaching hospital and University of Montpellier, Montpellier, France. <sup>44</sup>German Hospital, Buenos Aires, Argentina. <sup>45</sup>Autoimmune Dis-eases Unit, Department of Medicine, Hospital CIMA- Sanitas, Barcelona, Spain.

**Background.** To analyse the epidemiological, clinical and immunological characteristics of patients presenting with systemic disease in the absence of sicca manifestations in a large international cohort of patients diagnosed with primary Sjögren syndrome (SS).

**Methods.** The new 2017 ACR/EULAR Classification Criteria for SS allows the application of the criteria to any patient with a suspicion of SS due to systemic features with at least 1 positive ESSDAI domain. We have identified how many patients from the Big Data Sjögren Registry showed this early systemic presentation at the time of the disease diagnosis. As a control group, we selected those patients who presented with the typical sicca syndrome (subjective dry mouth and dry eyes).

Results. We have identified 240 (2.5%) patients presenting with a nonsicca systemic disease among the 9545 patients included in the Registry: 211 (88%) were women and 29 (12%) were men (female: male ratio, 7:1), with a mean age at diagnosis of 46.6 years. The frequency of fulfilment of the 2017 criteria was: 63% for positive ocular staining, 76% for abnormal Schirmer test, 79% for abnormal unstimulated whole salivary flow, 84% for positive salivary gland biopsy and 88% for Ro autoantibodies. Other immunological tests included positive ANA (88%), RF (54%), low C3 levels (25%), low C4 levels (13%), and cryoglobulins (9%). In comparison with patients presenting with the typical sicca syndrome, those presenting as systemic Siggren's with no sicca syndrome were younger (46.6 vs 52.9) p < 0.001), more frequently males (12% vs 6%, p < 0.001) and less frequently classified ethnically as White (36% vs 79%, p<0.001). Immunologically, patients with systemic Sjögren's had a higher frequency of anti-Ro antibodies (88% vs 71%, p<0.001), positive antinuclear antibodies (88% vs 81%, p=0.014) and low C3 levels (25% vs 13%, p<0.001). Systemic activity at diagnosis was significantly higher in patients with systemic non-sicca Siggren's in comparison with those with sicca syndrome including higher mean ESSDAI (6.5 vs 6.0, p<0.001) and clinESSDAI (6.6 vs 6.2, p=0.003) scores. In addition, moderate systemic activity (moderate-DAS) was found in a higher frequency in patients with systemic presentation (41% vs 30%, p=0.001). With respect to the ESSDAI domains, patients with non-sicca systemic disease had a higher frequency of activity in the constitutional (13% vs 8%, p=0.01), renal (12% vs 4%, p<0.001), hematological (31% vs 22%, p=0.005) and biological (64% vs 49%, p<0.001) domains, but a lower frequency of activity in the glandular (12% vs 22%, p<0.001) and peripheral nervous system (3% vs 6%, p=0.05) domains.

**Conclusion.** Primary Sjögren's syndrome at diagnosis is presenting as a systemic disease in the absence of the typical sicca symptoms in less than 3% of cases. These patients are characterized for being younger, less frequently women and White, with a higher frequency of immunological markers (Ro, ANA and low C3), higher activity in the hematological (cytopenias) and biological (hypergammaglobulinemia) domains, and higher activity in the constitutional (fever) and renal clinical domains in comparison with patients presenting with the classical sicca syndrome.

### P-33

#### Clinical and immunological disease patterns of primary Sjögren syndrome driven by gender and age at diagnosis

Soledad\_Retamozo<sup>1,2,3</sup>, Belchin Kostov<sup>4</sup>, Margit Zeher<sup>5</sup>, Kathy Sivils<sup>6</sup>, Thomas Mandl<sup>7</sup>, Raphaele Seror<sup>8</sup>, Xiaomei Li<sup>9</sup>, Chiara Baldini<sup>10</sup>, Xavier Mariette<sup>8</sup>, Jacques-Eric Gottenberg<sup>11</sup>, Debashish Danda<sup>12</sup>, Roberta Priori<sup>13</sup>, Luca Quartuccio<sup>14</sup>, Gabriela Hernandez-Molina<sup>15</sup>, Berkan Armagan<sup>16</sup>, Aike A. Kruize<sup>17</sup>, Seung-Ki Kwok<sup>18</sup>, Marie Wahren-Herlenius<sup>19</sup>, Sonja Praprotnik<sup>20</sup>, Damien Sene<sup>21</sup>, Elena Bartoloni<sup>22</sup>, Maureen Rischmueller<sup>23</sup>, Roser Solans<sup>24</sup>, Yasunori Suzuki<sup>25</sup>, David Isenberg<sup>26</sup>, Valeria Valim<sup>27</sup>, Piotr Wiland<sup>28</sup>, Gunnel Nordmark<sup>29</sup>, Guadalupe Fraile<sup>30</sup>, Hendrika Bootsma<sup>31</sup>, Takashi Nakamura<sup>32</sup>, Roberto Giacomelli<sup>33</sup>, Valerie Devauchelle-Pensec<sup>34</sup>, Benedikt Hofauer<sup>35</sup>, Michele Bombardieri<sup>36</sup>, Virginia Fernandes Moça Trevisani<sup>37</sup>, Daniel Hammenfors<sup>38</sup>, Sandra G. Pasoto<sup>39</sup>, Steven E. Carsons<sup>40</sup>, Tamer A Gheita<sup>41</sup>, Fabiola Atzeni<sup>42</sup>, Jacques Morel<sup>43</sup>, Cristina Vollenveider<sup>44</sup>, Pilar Brito-Zerón<sup>1,45</sup>, Manuel Ramos-Casals<sup>1</sup>, on behalf of the Sjögren Big Data Consortium.

<sup>1</sup>Sjögren Syndrome Research Group (AGAUR), Laboratory of Autoimmune Diseases Josep Font, IDIBAPS-CELLEX, Department of Autoimmune Diseases, ICMiD, University of Barcelona, Hospital Clínic, Barcelona, Spain. <sup>2</sup>Hospital Privado Universitario de Córdoba, Instituto Universitario de Ciencias Biomédicas de Córdoba (IUCBC), Córdoba- Argentina. <sup>3</sup>Instituto De Investigaciones En Ciencias De La Salud (INIC-SA), Universidad Nacional de Córdoba (UNC), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) - Córdoba - Argentina. <sup>4</sup>Primary Care Research Group, IDIBAPS, Centre d'Assistència Primària ABS Les Corts, GESCLINIC, Barcelona, Spain. <sup>5</sup>Division of Clinical Immunology, Faculty of Medicine, University of Debrecen, Debrecen, Hungary. <sup>6</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA. <sup>7</sup>Department of Rheumatology, Malmö University Hospital, Lund University, Lund, Sweden. <sup>8</sup>Center for Immunology of Viral Infections and Autoimmune Diseases, Assistance Publique – Hôpitaux de Paris, Hôpitaux Universitaires Paris-Sud, Le Kremlin-Bicêtre, Université Paris Sud, INSERM, Paris, France. <sup>9</sup>Department of Rheumatology and Immunology, Anhui Provincial Hospital, Hefei, China. <sup>10</sup>Rheumatology Unit, University of Pisa, Pisa, Italy. 11Department of Rheumatology, Strasbourg University Hospital, Université de Strasbourg, CNRS, Strasbourg, France. 12Department of Clinical Immunology & Rheumatology, Christian Medical College & Hospital, Vellore, India. 13Department of Internal Medicine and Medical Specialties, Rheumatology Clinic, Sapienza University of Rome, Rome, Italy, <sup>14</sup>Clinic of Rheumatology, Department of Medical and Biological Sciences, University Hospital "Santa Maria della Misericordia", Udine, Italy. <sup>15</sup>Immunology and Rheumatology Department, Instituto Nacional de Ciencias Médicas y Nutrición Salvador Zubirán. México City, Mexico. 16Department of Internal Medicine, Division of Rheumatology, Hacettepe University Faculty of Medicine, Ankara, Turkey. <sup>17</sup>Department of Rheumatology and Clinical Immunology, University Medical Center Utrecht, Utrecht, The Netherlands. <sup>18</sup>Seoul St. Mary's Hospital, The Catholic University of Korea, Seoul, South Korea. <sup>19</sup>Department of Medicine, Solna, Unit of Experimental Rheumatology, Karolinska Institutet, and Karolinska University Hospital, Stockholm, Sweden. 20 Department of Rheumatology, University Medical Centre, Ljubljana, Slovenia. 21 Service de Médecine Interne 2, Hôpital Lariboisière, Université Paris VII, Assistance Publique-Hôpitaux de Paris, 2, Paris, France. <sup>22</sup>Rheumatology Unit, Department of Medicine, University of Perugia, Perugia, Italy. 23Department of Rheumatology, School of Medicine, The University of Western Australia, Crawley, Australia. <sup>24</sup>Department of Internal Medicine, Hospital Vall d'Hebron, Barcelona, Spain. <sup>25</sup>Division of Rheumatology, Kanazawa University Hospital, Kanazawa, Ishikawa, Japan. 26Centre for Rheumatology, Division of Medicine, University College London, London, UK. 27 Department of Medicine, Federal University of Espírito Santo, Vitória, Brazil. <sup>28</sup>Department of Rheumatology and Internal Medicine, Wroclaw Medi-cal Hospital, Wroclaw, Poland. <sup>29</sup>Rheumatology, Department of Medical Sciences, Uppsala University, Uppsala, Sweden. <sup>30</sup>Department of Internal Medicine, Hospital Ramón y Cajal, Madrid, Spain. <sup>31</sup>Department of Rheumatology & Clinical Immunology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands. <sup>32</sup>Department of Radiology and Cancer Biology, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan. <sup>33</sup>Clinical Unit of Rheumatology, University of l'Aquila, School of Medicine, L'Aquila, Italy. <sup>34</sup>Rheumatology Department, Brest University Hospital, Brest, France. 35 Hals-Nasen-Ohrenklinik und Poliklinik, Technische Universität München, München, Germany. <sup>36</sup>Centre for Experimental Medicine and Rheumatology, Queen Mary University of London, London, UK. <sup>37</sup>Federal University of São Paulo, Sao Paulo, Brazil. <sup>38</sup>Department of Clinical Science, University of Bergen; and Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>39</sup>Rheumatology Division, Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo (USP), São Paulo, Brazil. 40Division of Rheumatology, Allergy and Immunology Winthrop-University Hospital, Stony Brook University School of Medicine, Mineola, NY, USA. <sup>41</sup>Rheumatology Department, Kasr Al Ainy School of Medicine, Cairo University, Cairo, Egypt. <sup>42</sup>IRCCS Galeazzi Orthopedic Institute, Milan, Italy. 43Department of Rheumatology, Teaching hospital and University of Montpellier, Montpellier, France. <sup>44</sup>German Hospital, Buenos Aires, Argentina. <sup>45</sup>Autoimmune Diseases Unit, Department of Medicine, Hospital CIMA-Sanitas, Barcelona, Spain.

**Objective.** To analyse how the epidemiological profile modifies systemic involvement measured at the time of diagnosis in a large international cohort of patients diagnosed with primary Sjögren syndrome (SS).

**Patients.** The Big Data Sjögren Project is an international, multicentre registry formed in 2014 to take a "high-definition" picture of the main features of primary SS at diagnosis by merging international SS databases. By October 2017, the database included 9545 consecutive patients recruited from 22 countries of the five continents.

Results. Baseline ESSDAI score was available in 9118 patients (93% female, mean age at diagnosis 53 years). The median ESSDAI score at diagnosis of the entire cohort was 4 (IQR 1-8). Men showed a higher systemic activity at diagnosis than women, including a higher median ESSDAI score (5 vs 4, p<0.001), clinESSDAI score (5 vs 3, p<0.001), a higher frequency of patients presenting with a high DAS (24.1% vs 13.7%, p<0.001), a higher frequency of patients with high activity in at least one domain (14.5% vs 7.8%, p<0.001) and a lower frequency of patients having no activity (ES-SDAI = 0) (16% vs 19%, p < 0.001). With respect to the ESSDAI domains, men had a higher risk of having activity at diagnosis in the lymphadenopathy (OR 1.5, CI95% 1.20-1.88), glandular (OR 1.3, CI95% 1.14-1.49), pulmonary (OR 1.44, CI95% 1.18-1.76), peripheral nervous system (OR 2.0, CI95% 1.60-2.57) and central nervous system (OR 2.2, CI95% 1.49-3.41) domains in comparison with women. A younger onset of the disease (<35 years) was also associated with a higher systemic activity at diagnosis in comparison with patients with older onset, including a higher median ES-SDAI score (4 vs 3, p<0.001), clinESSDAI score (4 vs 3, p=0.001) and a lower frequency of patients having no activity (ESSDAI = 0) (13% vs 22%, p<0.001). With respect to the ESSDAI domains, a younger onset was associated with an enhanced risk of presenting activity at diagnosis in the constitutional (OR 1.27, CI95% 1.05-1.53), lymphadenopathy (OR 1.59, CI95% 1.34-1.89), glandular (OR 1.16, CI95% 1.04-1.29), cutaneous (OR 1.35, CI95% 1.13-1.49), renal (OR 1.4, CI95% 1.09-1.81), hematological (OR 1.18, CI95% 1.06-1.32) and biological (OR 1.33, CI95% 1.27-1.40) domains, while older onset was associated with an enhanced risk for presenting activity at articular (OR 0.89, CI95% 0.82-0.97) pulmonary (OR 0.54, CI95% 0.42-0.70), muscular (OR 0.55, CI95% 0.33-0.94) and peripheral nervous system (OR 0.59, CI95% 0.42-0.81) domains.

**Conclusion.** Gender and age at diagnosis play a key role in the severity of systemic involvement measured at the diagnosis of primary Sjögren syndrome, with men and patients diagnosed before 35 years being those presenting with the highest systemic profile. Some organs are more active when the disease is diagnosed at younger ages (constitutional, lymph nodes, glands, skin and kidneys) and others when the disease is diagnosed in the elderly (joints, lungs and neuromuscular). Clinically, the ESSDAI provides a reliable picture of systemic involvement and may help identify epidemiological subsets with high systemic activity at diagnosis and, therefore, at high risk of suffering a complicated clinical course.

#### P-34

#### How ethnicity modifies systemic activity of primary Sjögren syndrome: analysis of baseline ESSDAI scores in a multi-ethnic international cohort

Soledad Retamozo<sup>1,2,3</sup>, Belchin Kostov<sup>4</sup>, Margit Zeher<sup>5</sup>, Kathy Sivils<sup>6</sup>, Thomas Mandl<sup>7</sup>, Raphaele Seror<sup>8</sup>, Xiaomei Li<sup>9</sup>, Chiara Baldini<sup>10</sup>, Xavier Mariette<sup>8</sup>, Jacques-Eric Gottenberg<sup>11</sup>, Debashish Danda<sup>12</sup>, Roberta Priori<sup>13</sup>, Luca Quartuccio<sup>14</sup>, Gabriela Hernandez-Molina<sup>15</sup>, Berkan Armagan<sup>16</sup>, Aike A. Kruize<sup>17</sup>, Seung-Ki Kwok<sup>18</sup>, Marie Wahren-Herlenius<sup>19</sup>, Sonja Praprotnik<sup>20</sup>, Damien Sene<sup>21</sup>, Elena Bartoloni<sup>22</sup>, Maureen Rischmueller<sup>23</sup>, Roser Solans<sup>24</sup>, Yasunori Suzuki<sup>25</sup>, David Isenberg<sup>26</sup>, Valeria Valim<sup>27</sup>, Piotr Wiland<sup>28</sup>, Gunnel Nordmark<sup>29</sup>, Guadalupe Fraile<sup>30</sup>, Hendrika Bootsma<sup>31</sup>, Takashi Nakamura<sup>32</sup>, Roberto Giacomelli<sup>33</sup>, Valerie Devauchelle-Pensec<sup>34</sup>, Benedikt Hofauer<sup>35</sup>, Michele Bombardieri<sup>36</sup>, Virginia Fernandes Moça Trevisani<sup>37</sup>, Daniel Hammenfors<sup>38</sup>, Sandra G. Pasoto<sup>39</sup>, Steven E. Carsons<sup>40</sup>, Tamer A Gheita<sup>41</sup>, Fabiola Atzeni<sup>42</sup>, Jacques Morel<sup>43</sup>, Cristina Vollenveider<sup>44</sup>, Pilar Brito-Zerón<sup>1,45</sup>, Manuel Ramos-Casals<sup>1</sup>, on behalf of the Sjögren Big Data Consortium.

'sjögern Syndrome Research Group (AGAUR), Laboratory of Autoimmune Diseases Josep Font, IDIBAPS-CELLEX, Department of Autoimmune Diseases, ICMiD, University of Barcelona, Hospital Clínic, Barcelona, Spain. <sup>2</sup>Hospital Privado Universitario de Córdoba, Instituto Universitario de Ciencias Biomédicas de Córdoba (IUCBC), Córdoba- Argentina. 3Instituto De Investigaciones En Ciencias De La Salud (INIC SA), Universidad Nacional de Córdoba (UNC), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) - Córdoba - Argentina. <sup>4</sup>Primary Care Research Group, IDIBAPS, Centre d'Assistència Primària ABS Les Corts, GESCLINIC, Barcelona, Spain. 5Division of Clinical Immunology, Faculty of Medicine, University of Debrecen, Debrecen, Hungary. <sup>6</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA. <sup>7</sup>Department of Rheumatology, Malmö University Hospital, Lund University, Lund, Sweden. <sup>8</sup>Center for Immunology of Viral Infections and Autoimmune Diseases, Assistance Publique -Hôpitaux de Paris, Hôpitaux Universitaires Paris-Sud, Le Kremlin-Bicêtre, Université Paris Sud, INSERM, Paris, France. 9Department of Rheumatology and Immunology, Anhui Provincial Hospital, Hefei, China. <sup>10</sup>Rheumatology Unit, University of Pisa, Pisa, Italy. <sup>11</sup>Department of Rheumatology, Strasbourg University Hospital, Université de Strasbourg, CNRS, Strasbourg, France. <sup>12</sup>Department of Clinical Immunology & Rheumatology, Christian Medical College & Hospital, Vellore, India. 13Department of Internal Medicine and Medical Specialties, Rheumatology Clinic, Sapienza University of Rome, Rome, Italy. <sup>14</sup>Clinic of Rheumatology, Department of Medical and Biological Sciences, University Hospital "Santa Maria della Misericordia", Udine, Italy. <sup>15</sup>Immunology and Rheumatology Department, Instituto Nacional de Ciencias Médicas y Nutrición Salvador Zubirán. México City, Mexico. 16Department of Internal Medicine, Division of Rheumatology, Hacettepe University Faculty of Medicine, Ankara, Turkey. 17 Department of Rheumatology and Clinical Immunology, University Medical Center Utrecht, Utrecht, The Netherlands. <sup>18</sup>Seoul St. Mary's Hospital, The Catholic University of Korea, Seoul, South Korea. <sup>19</sup>Department of Medicine, Solna, Unit of Experimental Rheumatology, Karolinska Institutet, and Karolinska University Hospital, Stockholm, Sweden. 20 Department of Rheumatology, University Medical Centre, Ljubljana, Slovenia.<sup>21</sup>Service de Médecine Interne 2, Hôpital Lariboisière, Université Paris VII, Assistance Publique-Hôpitaux de Paris, 2, Paris, France.<sup>22</sup>Rheumatology Unit, Department of Medicine, University of Perugia, Perugia, Italy. 23Department of Rheumatology, School of Medicine, The University of Western Australia, Crawley, Australia. <sup>24</sup>Department of Internal Medicine, Hospital Vall d'Hebron, Barcelona, Spain. <sup>25</sup>Division of Rheumatology, Kanazawa University Hospital, Kanazawa, Ishikawa, Japan. 26Centre for Rheumatology, Division of Medicine, University College London, London, UK. 27Department of Medicine, Federal University of Espírito Santo, Vitória, Brazil. 28 Department of Rheumatology and Internal Medicine, Wroclaw Medical Hospital, Wroclaw, Poland. 29Rheumatology, Department of Medical Sciences, Lipsala University, Uppsala, Sweden. <sup>3D</sup>Department of Internal Medicine, Hospital Ramón y Cajal, Madrid, Spain. <sup>31</sup>Department of Rheumatology & Clinical Immunology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands. 32Department of Radiology and Cancer Biology, Nagasaki University

Graduate School of Biomedical Sciences, Nagasaki, Japan. <sup>33</sup>Clinical Unit of Rheumatology, University of l'Aquila, School of Medicine, L'Aquila, Italy. <sup>34</sup>Rheumatology Department, Brest University Hospital, Brest, France. <sup>35</sup>Hals-Nasen-Ohrenklinik und Poliklinik, Technische Universität München, München, Germany. <sup>36</sup>Centre for Experimental Medicine and Rheumatology, Queen Mary University of London, London, UK. <sup>37</sup>Federal University of São Paulo, Sao Paulo, Brazil. <sup>38</sup>Department of Clinical Science, University of Bergen; and Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>39</sup>Rheumatology Division, Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo (USP), São Paulo, Brazil. <sup>40</sup>Division of Rheumatology, Allergy and Immunology Winthrop-University Hospital, Stony Brook University School of Medicine, Cairo University, Cairo, Egypt. <sup>42</sup>IRCCS Galeazzi Orthopedic Institute, Milan, Italy. <sup>43</sup>Department of Rheumatology, Teaching hospital and University of Montpellier, Montpellier, France. <sup>44</sup>German Hospital, Buenos Aires, Argentina. <sup>45</sup>Autoimmune Diseases Unit, Department of Medicine, Hospital CIMA-Sanitas, Barcelona, Spain.

**Objectives.** To analyse the influence of ethnicity on the clinical presentation of primary Sjögren syndrome (SjS) by measuring systemic activity (ESS-DAI score) at the time of diagnosis.

Methods. The Big Data Sjögren Project is an international, multicentre registry formed in 2014 to take a "high-definition" worldwide picture of the main features of primary SjS at diagnosis by merging international SjS databases. By October 2017, the database included 9545 consecutive patients recruited from 22 countries of the five continents. Systemic involvement was defined according to the ESSDAI/clinESSDAI. Disease activity states (DAS) were categorized according to the global ESSDAI score as lowactivity (ESSDAI<5), moderate-activity (5≤ESSDAI≤13) and high-activity (ESSDAI≥14).

**Results.** Ethnicity data were available for 8746 (95%) of 9118 patients with available information on ESSDAI: 6614 (76%) patients were classified as White, 1306 (15%) as Asian, 520 (6%) as Hispanic, 130 (1%) as Black/ African American -BAA- and 176 (2%) as other ethnicities. The highest median ESSDAI score at diagnosis was found in BAA patients (6 vs 4 in W, 4 in A and 3 in H, p<0.001), as well as for median clinESSDAI score (5 vs 4 in W, 3 in A and 3 in H, p<0.001). The highest frequency of patients presenting at diagnosis with a high DAS was also found in BAA (16.2% vs 15.8% in W, 11.6% in A and 8.5% in H, p<0.001). With respect to the ES-SDAI domains, BAA patients had an enhanced risk of presenting activity at diagnosis in peripheral nervous system (OR 2, CI95% 1.27-3.15) and biological (OR 1.2, CI95% 1.03-1.4) domains in comparison with White patients. In contrast, Asian patients had a lower risk of presenting activity at diagnosis in the lymphadenopathy (OR 0.54, CI95% 0.43-0.69), glandular (OR 0.41, CI95% 0.35-0.49), articular (OR 0.57, CI95% 0.51-0.63), cutaneous (OR 0.82, CI95% 0.67-0.99), muscular (OR 0.41, CI95% 0.25-0.7), peripheral nervous system (OR 0.54, CI95% 0.4-0.73) and central nervous system (OR 0.47, CI95% 0.27-0.81) domains, but a higher risk of having activity at renal (OR 2.58, CI95% 2.11-3.16), hematological (OR 1.16, CI95% 1.05-1.29) and biological (OR 1.19, CI95% 1.13-1.26) domains in comparison with White patients. Finally, Hispanic patients had a lower risk of presenting activity at diagnosis in glandular (OR 0.63, CI95% 0.52-0.78), pulmonary (OR 0.49, CI95% 0.34-0.71), muscular (OR 0.35, CI95% 0.14-0.84) and hematological (OR 0.73, CI95% 0.6-0.89) domains in comparison with White patients.

**Conclusions.** This study provides the first evidence of a strong influence of ethnicity on the systemic phenotype of primary SjS at diagnosis. BAA and White people had the highest median ESSDAI and clinESSDAI scores and the highest frequencies of patients classified as high DAS, in contrast to Asian and Hispanic people. Organ-by-organ activity also varied significantly across ethnicities.

## P-35

#### Sjögren syndrome as the main maternal disease in mothers with babies affected with Ro-associated congenital heart block (Spanish Registry REBACC-GEAS-SEMI)

Soledad Retamozo<sup>1,2,3</sup>, Pilar Brito-Zerón<sup>1,4</sup>, Gerard Espinosa<sup>4</sup>, Ángel Robles<sup>5</sup>, Pilar Rosich<sup>6</sup>, Luis Sáez Comet<sup>7</sup>, Olga Capdevila<sup>8</sup>, José Antonio Vargas<sup>9</sup>, Lucio Pallarés<sup>10</sup>, Luis Trapiella<sup>11</sup>, José Antonio González Nieto<sup>12</sup>, Aleida Martínez Zapico<sup>13</sup>, Mónica Rodriguez<sup>14</sup>, Carles Tolosa<sup>6</sup>, Francesca Mitjavila<sup>8</sup>, Mercedes Pérez-Conesa<sup>6</sup>, José Mario Sabio<sup>9</sup>, Luis Caminal<sup>11</sup>, Joaquim Oristrell<sup>6</sup>, César Morcillo<sup>15</sup>, Alejandra Flores-Chavez<sup>1</sup>, Belchin Kostov<sup>16</sup>, Manuel Ramos-Casals<sup>1,4</sup> on behalf of the REBACC-GEAS-SEMI Registry.

<sup>1</sup>Laboratory of Autoimmune Diseases Josep Font, IDIBAPS-CELLEX, Barcelona. <sup>2</sup>Hospital Privado Universitario de Córdoba, Instituto Universitario de Ciencias Biomédicas de Córdoba (IUCBC), Córdoba- Argentina. <sup>3</sup>Instituto De Investigaciones En Ciencias De La Salud (INICSA), Universidad Nacional de Córdoba (UNC), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) - Córdoba - Argentina. <sup>4</sup>Department of Autoimmune Diseases, ICMiD, University of Barcelona, Hospital Clínic, Barcelona, Spain. <sup>5</sup>Department of Internal Medicine, Hospital La Paz, Madrid. <sup>6</sup>Department of Internal Medicine, Hospital Parc Taulf, Sabadell. <sup>7</sup>Systemic Autoimmune Diseases Unit, Hospital Miguel Servet, Zaragoza. <sup>8</sup>Department of Internal Medicine, Hospital de Bellvitge <sup>9</sup>Department of Internal Medicine, Hospital Virgen de las Nieves, Granada. <sup>10</sup>Department of Internal Medicine, Hospital Son Espases, Palma de Mallorca. <sup>11</sup>Department of Internal Medicine, Hospital Son Espases, Palma de Mallorca. <sup>11</sup>Department of Internal Medicine, Hospital Medicine, Internal Medicine, Hospital Universitario Central de Asturias, <sup>14</sup>Department of Internal Medicine, Hospital Mutua de Terrasa, Sabadell. <sup>15</sup>Department of Internal Medi Cine, Hospital Mutua de Terrasa, Sabadell. <sup>15</sup>Department of Internal Medicine, Hospital CIMA-Sanitas, <sup>16</sup>Transversal Research Group Primary Care, IDIBAPS, Barcelona.

**Objective.** To analyze both the already-diagnosed and the underlying maternal autoimmune diseases of mothers with pregnancies affected by autoimmune congenital heart block (CHB) associated with maternal anti-Ro antibodies.

**Methods.** The REBACC Spanish Multicenter Registry was created in March 2014. It is integrated by 12 centers with substantial experience in the management of systemic autoimmune diseases. Autoimmune CHB was defined as: a) CHB of any type (I, II or III), fetal endocardial fibroelastosis (EFE) and/or cardiomyopathy, b) cardiac block diagnosed in utero or in the first postpartum month, and c) mothers carrying anti-Ro52, Ro60 and/or La autoantibodies.

Results. On October 2017, the REBACC Registry included a total of 45 anti-Ro+ mothers with 50 single pregnancies with CHB. Mean maternal age at the time of first affected pregnancy with CHB was 32.97 years (range: 22-44). All mothers were anti-Ro60 (+), 17/17 anti-Ro52 (+) and 32/44 (73%) anti-La (+). The mean gestational age at diagnosis of CHB was 23 weeks (range 16-37). Information about fetal outcomes was available in 45 pregnancies: AV blocks were of type I in 2 pregnancies (4.5%), type II in 15 (33%) and type III in 27 (60%); 1 had an isolated EFE (2.5%); therapies used included dexamethasone or betamethasone (n=25), intravenous immunoglobulins (n=5), and plasma exchanges (n=3); 11 pregnancies were interrupted due to bad fetal prognosis (24%) and 34 (76%) were successfully carried to term, and pacemaker implantation was required in 18/34 babies (53%). At diagnosis of the first affected pregnancy, 31 (69%) mothers did not have any autoimmune disease and the remaining 14 (31%) had Sjögren syndrome (n=7), SLE (n=5) and undifferentiated autoimmune disease (n=2). At the last visit, of the 33 women who initially did not have any autoimmune diagnosis (none/undifferentiated autoimmune disease), 22 (67%) developed a systemic autoimmune disease (SS in 16, SLE in 5 and SS+ SLE in 1). Also, 1 mother previously diagnosed with SLE, was subsequently diagnosed with Sjögren syndrome.

**Conclusions.** At the last visit, 23/45 (51%) mothers with affected babies with Ro-associated congenital heart block have a diagnosis of Sjögren syndrome (overwhelmingly primary); in contrast, 70% of mothers have no identified systemic autoimmune disease at the first affected pregnancy, as anti-Ro antibodies can be detected several years before SS is diagnosed. Autoimmune CHB is one of the first early signs of primary SS in women of childbearing age.

### P-36

## Re-analysis of the JOQUER trial after stratifying patients into clinical phenotypes

Alexis Collins<sup>1</sup>, Dennis Lendrem<sup>1</sup>, Xavier Mariette<sup>2</sup>, Eric Hachulla<sup>3</sup>, Veronique Leguern<sup>4</sup>, Jacques-Eric Gottenberg<sup>5</sup>, Wan Fai Ng<sup>1</sup>.

<sup>1</sup>Institute of Cellular Medicine, Newcastle University & NIHR Newcastle Biomedical Research Centre, UK. <sup>2</sup>Hôpital Bicêtre, Hôpitaux Universitaires Paris-Sud,Paris, France. <sup>3</sup>Service de Médecine Interne et Immunologie Clinique, CHU Lille, Centre national de référence maladies systémiques et auto-immunes rares, Université de Lille, 59037 Lille cedex, France. <sup>4</sup>Department of Internal Medicine, Cochin Hospital, Assistance Publique-Hopitaux de Paris, Paris, France. <sup>5</sup>Rheumatology Department, National Center for Rare Systemic Autoimmune Diseases, Hôpitaux Universitaires de Strasbourg, Strasbourg, France.

**Introduction.** The JOQUER trial evaluated the efficacy of hydroxychloroquine (HCQ) for patients with Primary Sjögren's Syndrome (PSS). This randomized control trial concluded that there was no overall disease benefit from the drug. However, we have recently identified four distinct clinical phenotypes in PSS each with differences in underlying pathobiological profiles - the high symptom burden (HSB), pain dominant with fatigue (PDF), dryness dominant with fatigue (DDF) and low symptom burden (LSB) phenotypes. In this study, we re-analysed the JOQUER trial after stratifying by patient phenotype.





**Methods.** The JOQUER trial involved 120 patients with PSS from 15 different centres in France. Participants were assessed at baseline, weeks 12, 24 and 48. Patients were randomized (1:1) to receive HCQ (400 mg once daily) or placebo from baseline until 24 weeks. Between weeks 24 and 48, all participants were prescribed HCQ. Assessment of the disease activity included ESSPRI, ESSDAI and objective measures including dryness tests (Schirmer's and salivary flow) and systemic blood analysis. For the re-analysis of the trial, patients were stratified into four clinical phenotypes. The - sample sizes for the HSB, PDF, DDF and LSB groups were 32, 39, 19, and 14 respective-

ly. Multivariate analysis of variance (MANOVA) analysis permitted testing of changes over time from baseline, weeks 12 and 24. Contrasts were formed testing for changes in improvement in patient-reported outcome measures for HSB, PDF, DDF, and LSB phenotypes over time.

**Results.** By just 12 weeks, there was a statistically significant improvement with HCQ in ESSPRI of 1.27 points (95%CI: 0.33, 2.22; *p*-value=0.0032) and improvement at 24 weeks (P-value= 0.0137) compared to placebo. Patient reported dryness scores contributed most to this ESSPRI improvement. While there were no statistically significant improvements in either pain or fatigue scores, there was a significant reduction in dryness score over the 12 weeks with improvement of 1.77 points (95% CI 0.6687, 2.8697; *p*-value= 0.0006) and a significant improvement after 24 weeks (*p*-value= 0.0006) and a significant improvement was the most notable in nasal dryness after 24 weeks (*p*-value= 0.029) and oral dryness at 24 weeks (*p*-value=0.0194).

**Conclusion.** While the mechanism of action of HCQ in PSS is not fully understood, re-analysis of the JOQUER trial suggests HCQ may benefit the HSB subset of PSS patients. A stratified RCT powered to detect improvements in the four phenotypes is needed to confirm the efficacy of HCQ for HSB patients.

## **P-37**

#### Comparison of distinguishing characteristics between patients with primary Sjögren's syndrome and non-primary Sjögren's syndrome patients

\*A.M.L. Pedersen<sup>1</sup>, M.L. Sembler-Møller<sup>1</sup>, D. Belstrøm<sup>1</sup>, <sup>†</sup>N.H.H. Heegaard<sup>2.3</sup> A.L. Carlsen<sup>2</sup>, H. Locht<sup>4</sup>

<sup>1</sup>Section of Oral Medicine & Pathology, Periodontology & Oral Microbiology, Department of Odontology, Faculty of Health and Medical Sciences, University of Copenhagen, Denmark. <sup>2</sup>Department of Autoimmunology & Biomarkers, Statens Serum Institut, Copenhagen S, Denmark <sup>3</sup>Department of Clinical Biochemistry & Pharmacology, Odense University Hospital, University of Southern Denmark, Odense C, Denmark <sup>4</sup>Department of Rheumatology, Frederiksberg Hospital, University of Copenhagen, DK-2000 Copenhagen, Denmark.

**Objectives.** To evaluate potential systemic, oral and salivary distinguishing characteristics for patients with primary Sjögren's syndrome (pSS) from non-primary Sjögren's syndrome (non-pSS) patients.

**Methods.** Forty patients referred for diagnosis of pSS underwent an interview including exocrine and non-exocrine symptoms and manifestations, assessment of fatigue using the Multidimensional Fatigue Inventory MFI-20, an oral (dental and periodontal status) and ocular (Schirmer's test, tear break-up time and Lissamine green staining) examination, measurements of unstimulated (UWS) and chewing-stimulated whole saliva (SWS) flow rates, a labial salivary gland biopsy and test for serum autoantibodies.

Results. Fourteen females and one male (aged 57±12 years) fulfilled the American-European Consensus Classification Criteria, whereas 22 females and 3 males (aged 55±14 years) did not. No significant differences were found in symptoms of oral and ocular dryness, concomitant systemic diseases, no. of prescribed medication, mean score of decayed-missed-filledteeth, levels of plaque, gingival inflammation and probing depth. However, patients with pSS had lower UWS (0.04±0.06 ml/min vs. 0.13±0.12 ml/ min, p=0.026) and SWS (0.47±0.50 ml/min vs. 0.84±0.60 ml/min, p=0.038) flow rates, and higher fatigue scores (p=0.046). Lymphocytic infiltration, i.e. with focus score ≥1, was found in the salivary gland biopsies from 47% of the patients with pSS and in none of those from non-pSS patients (mean focus score 2.3±4.0 vs. 0.02±0.06, p<0.001). In the remaining 53% of pSS patients, the salivary gland tissue was characterized by atrophy, fibrosis and diffuse inflammation. All patients with pSS had elevated levels of circulating anti-Ro/SSA serum autoantibodies as compared to 28% in the non-pSS group (p<0.001).

**Conclusion.** Our preliminary findings indicate that oral, ocular and systemic symptoms and manifestations are poor distinguishing characteristics. A salivary gland biopsy with a focus score >1 had a high predictive value for the diagnosis of pSS. Presence of anti-Ro/SSA autoantibodies had a high sensitivity for pSS but a lower specificity due to several false positive serum samples. Our on-going study includes a larger cohort to substantiate our preliminary findings and discover more specific biomarkers for pSS.

#### **P-38**

## The potential continuum of risk of dry eye syndrome, Sjögren's syndrome and B-cell non-Hodgkin lymphoma

Ben Eli H.<sup>1,2</sup>, Solomon A.<sup>1</sup>, Willhauck-Fleckenstein M.<sup>3</sup>, Pawlita M.<sup>3</sup>, Kleinstern G.<sup>4</sup>, Abu Seir R.<sup>5</sup>, Kedar Tirosh A.<sup>2</sup>, Ben Chetrit E.<sup>6</sup>, Mevorach D.<sup>7</sup>, Aframian DJ<sup>7</sup> and Paltiel O.<sup>2,5</sup>

<sup>1</sup>Department of Ophthalmology, <sup>2</sup>Braun School of Public Health and Community Medicine Hadassah-Hebrew University Medical Center Jerusalem , <sup>3</sup>Infection and Cancer Program, German Cancer Research Center (DKFZ), Heidelberg, Germany. <sup>4</sup>Department of Health Sciences Research, Mayo Clinic, Rochester, MN, USA, <sup>5</sup>Department of Hematology, <sup>6</sup>Unit of Rheumatology, <sup>7</sup>Department of Internal Medicine and <sup>8</sup>Department of Oral Medicine & Sjögren's syndrome Center Hadassah-Hebrew University Medical Center, Jerusalem.

**Background.** Dry eye syndrome (DES) is a manifestation of Sjögren's syndrome (SS), an autoimmune disease (AID) with a high lifetime risk of B-cell non-Hodgkin lymphoma (B-NHL). We aimed to explore whether an etiologic continuum exists from DES through SS to B-NHL by assessing environmental and infectious exposures and cytokine levels in these disorders. **Methods.** In a clinic-based case-control study 702 participants: 91 SS, 120 DES, 211 controls (age and sex-matched), and 280 B-NHL cases were recruited and interviewed regarding exposures. Antibody titers to HCV, HBV, EBV, CMV, *H. pylori*, and *C. trachomatis* were tested by multiplex serology. Serum cytokines IL4, IL6, IL10, IL12, IL17, TNF $\alpha$ , INF $\gamma$  and IL1 $\beta$  were tested on SS and DES participants using multiplex ELISA.

Results. SS showed a female predominance (9.2:1). Factors inversely associated with B-NHL, DES and SS include alcohol consumption (OR=0.47, 95%CI: 0.32-0.71; OR=0.54, CI: 0.33, 0.88; OR=0.27, CI: 0.15, 0.49, respectively), and East European ancestry for SS (OR=0.43; CI: 0.23-0.79), compared to controls. Self-reported infection requiring hospitalization was more common in B-NHL (OR=1.91; CI: 1.22-2.98), DES (OR=3.22; CI: 1.93-5.35) and SS (OR=4.58; CI: 2.56-8.18) than in controls. B-NHL cases were more likely to report 1st degree relatives with hematologic cancer (OR=1.91; CI: 1.00-3.62), while 1st degree relatives with AID were more common among SS (OR=5.23; CI: 2.58-10.58) and DES patients (OR=3.56; CI: 1.84-6.89). IL10 and IL12 levels were higher in SS than in DES, while controls had intermediate levels (p<0.001). A higher proportion of SS patients had antibodies to HCV, EBV-EA-D and CMV (p=0.02, 0.02, 0.01, respectively) than B-NHL, DES or controls. CMV seropositivity was more common in SS patients than among controls (OR=3.56; CI: 1.14-11.04), while that of C. trachomatis was decreased in DES (OR=0.40; CI: 0.19-0.84) compared to controls.

**Conclusions.** While some factors (eg alcohol, hospitalization for infection) appear to be associated with all 3 conditions, some were specific to one or two of them. Cytokine activation does not show a continuum from controls $\rightarrow$ DES $\rightarrow$ SS. Patients with DES and SS appear distinct in terms of infectious exposures. Further work is required to understand events leading to B-NHL in autoimmune disease.

### P-39

Depicting the spectrum of Sjögren's syndrome patients: correlating clinical clustering characteristics with gene expression

Leyla Y. Teos, Mayank Tandon, Monisha Billings, Ilias Alevizos Sjögren's Syndrome and Salivary Gland Dysfunction Unit, National Institute of Dental and Craniofacial Research, NIH, Bethesda, Maryland, USA.

**Background.** Sjögren's Syndrome (SS) is a progressive, chronic autoimmune disease that is difficult to diagnose. Its trademark features include the presence of immune cells infiltrating exocrine glands, mainly the lacrimal and salivary glands, leading to symptoms of dryness. The systemic manifestations cause an array of debilitating symptoms and further confound diagnosis. The pathophysiology of SS is still unknown, yet there is evidence of both epithelial and immune dysfunction, leading to a heterogenous phenotype. To characterize the diversity within Sjögren's syndrome patients, we first identified patient clusters based on a number of clinical characteristics and then used whole RNA-Seq to identify the molecular pathways associated with each of these clusters.

**Methods.** Whole RNA-Seq was carried out on 44 primary (SS) patients and 13 healthy controls (HCs). Reconstructed reads were aligned to the hg19 reference genome and read counts were generated using the HT Seq python

## 14<sup>th</sup> International Symposium on Sjögren's Syndrome

module. Clinical data for 8 measures – salivary flow, presence of antibodies, abnormally increased levels of C3 and C4, and (FS) focus score – were encoded and input as a numerical matrix for decomposition with principal components analysis (PCA). Coordinates of the first 3 PCs were used for kmeans clustering into 4 groups. The Gene Set Enrichment Analysis (GSEA) algorithm from the Broad Institute was used to compute enrichment of pathways in the Reactome database in each patient cluster when compared to the HC cluster.

Results. The unsupervised clustering analysis defined the characteristics of 4 clusters. Cluster 1 consists mostly of healthy controls with no clinical abnormalities. Cluster 2 includes classically symptomatic patients with elevated focus scores (> 4), mildly reduced salivary flow, and autoantibodies against SSA and/or SSB. Cluster 3 captures SS patients with less clinical activity among the included criteria: low focus scores, mild salivary flow, and no autoantibodies or C3/C4 elevation. Patients in Cluster 4 also have low focus scores, but they have greatly reduced salivary flows, presence of elevated C3/C4 and tend to harbor antibodies against SSA only. Pathwaylevel enrichment analysis revealed a downregulation of metabolic pathways and upregulation of immune and signal transduction pathways across all patient clusters. Metabolic pathways related to the respiratory electron transport chain and mitochondria protein import were downregulated specifically within Cluster 4 samples. Likewise, some immune pathways like Antigen Processing Cross Presentation were upregulated only in Cluster 2, and Potassium Channels were downregulated in Cluster3.

**Conclusions.** Clustering by clinical characteristics has identified unique molecular pathways that might be important in the pathogenesis of SS. Overall, not only were previously known pathways highlighted in the analysis, but new phenotypic clusters of patients and their affected gene pathways were explored. This classification may lead to a more thorough understanding of the molecular mechanisms underlying SS as a spectrum, leading to a personalized approach based on the individual patient pathophysiology.

#### **P-40**

## Gene networks describe symptom based phenotypes in primary Sjögren's syndrome

Dennis W Lendrem<sup>a,1</sup>, Jessica Tarn<sup>a,1</sup>, Nadia Howard Tripp<sup>a,1</sup>, Peter Mc-Meekin<sup>2</sup>, Andrew Skelton<sup>1</sup>, Katherine James<sup>1</sup>, Colin Gillespie<sup>1</sup>, Shereen Al-Ali<sup>129</sup>, Kate Hackett<sup>1</sup>, B Clare Lendrem<sup>1</sup>, Ben Hargreaves<sup>6</sup>, John Casement<sup>1</sup>, Sheryl Mitchell<sup>6</sup>, Simon J Bowman<sup>7</sup>, Elizabeth Price<sup>8</sup>, Colin T Pease<sup>9</sup>, Paul Emery<sup>9</sup>, Peter Lanyon<sup>10</sup>, John Hunter<sup>11</sup>, Monica Gupta<sup>11</sup>, Michele Bombardieri<sup>12</sup>, Nurhan Sutcliffe<sup>13</sup>, Costantino Pitzalis<sup>14</sup>, John McLaren<sup>15</sup>, Annie Cooper<sup>16</sup>, Marian Regan<sup>17</sup>, Ian Giles<sup>18</sup>, David Isenberg<sup>18</sup>, Saravanan Vadivelu<sup>19</sup>, David Coady<sup>20</sup>, Bhaskar Dasgupta<sup>21</sup>, Neil McHugh<sup>22</sup>, Steven Young-Min<sup>23</sup>, Robert Moots<sup>24</sup>, Nagui Gendi<sup>25</sup>, Mohammed Akil<sup>26</sup>, Bridget Griffiths<sup>6</sup> , John D Isaacs<sup>1</sup>, Wan-Fai Ng<sup>1</sup> on behalf of the UK Primary Sjögren's Syndrome Registry\*.

**Background.** We have previously described a novel methodology for patient stratification in primary Sjögren's syndrome (PSS) using five patient reported outcome measures (ISSS2014). The stratification resulted in four phenotypic groups; Low Symptom Burden (LSB), High Symptom Burden (HSB), Dryness Dominant with Fatigue (DDF) and Pain Dominant with Fatigue (PDF). There are several clinical and haematological differences (lymphocyte count and serum IgG) between the four patient phenotypes. In this study we identify further biological differences between the phenotypes using historical whole blood transcriptomics data. Transcriptional differences separate the phenotypes provide insight into the underlying biology between the four groups.

**Methods.** Historical whole-blood transcriptomic data from two studies from the United Kingdom Primary Sjögren's Syndrome Registry (UKPSSR) were used to reconstruct gene networks and model phenotype as a function of gene expression data. We used ARACNE software to reconstruct gene networks using data from 186 patients assayed in 2015 (training data). We built a partial least squares discriminant analysis (PLS-DA) model from the training data and tested this model in an independent, time-separated cohort of 119 patients assayed in 2013 (testing data).

**Results.** The ARACNE network reconstruction revealed three main gene networks (N1, N2, N3) (Figure 1). Network N1 is a large cluster of gene interactions, including genes known to be relevant in pSS (*e.g.* TROVE2). Network N2 contains several genes involved in erythropoeisis and network N3 is comprised almost exclusively of IFN related genes. The PLS-DA

model built with the training data performed well in predicting phenotypic group membership. The PLS-DA model predicted the phenotypic groups for the testing data with greater than 90% accuracy. Four latent variables (LV1-LV4) could be extracted from the model, which represent linear combinations of the gene expression data. LV1 and LV2 are representative of gene expression differences in DDF and LSB respectively, whereas LV3 and LV4 are representative of gene expression differences between PDF and HSB. By mapping the latent variables to the ARACNE network we are able to visualize differentially activated pathways for the four phenotypes. The gene expression differences in DDF phenotype are focussed in N1 (Figure 1, a), whereas the LSB phenotype shows the biggest differences in the IFN related network (N3) (Figure 1, b). The differences in gene expression between the HSB and PDF phenotypes can be observed in all three networks (Figure 1, c,d).



 $Fig.\,1.\,\mathsf{a},\mathsf{b},\mathsf{c},\mathsf{d}.$ 

**Conclusions.** This analysis of historical whole blood transcriptomics data has granted further insight into the biological differences underpinning clinical phenotypes of pSS that our group has previously described. Further work is necessary to fully interrogate specific network interactions.

#### **P-41**

#### Sjögren syndrome profile in the Brazilian population: demographic, clinical, laboratory and imaging analysis and comparison with controls

Eduardo Melani Rocha, Carolina Maria Módulo, Amanda Pires Barbosa, Fabiola Reis Oliveira, Ana Carolina Fragoso Motta, Denny Marcos Garcia, Valdair Francisco Muglia, Paulo Louzada Junior.

Ribeirao Preto Medical School, Ribeirao Preto Dental School, University of Sao Paulo, Ribeirao Preto, SP, Brazil.

**Purpose.** Sjögren syndrome (SS) is present worldwide, with unknown cause and cure. The present work describes a large series of SS patients observed in Brazil and compare the data among SS and non-SS (NSS) patients. **Methods.** A spontaneous sample of individuals with *sicca* symptoms and controls were evaluated and were classified as SS or NSS. The demographic, clinical, laboratory and imaging data were compared among those individuals. The OSDI, PhQ-9 and neuropathic pain questionnaires were also applied. Thirty-nine controls and 19 SS individuals we submitted to MRI on the 3.0 Tesla Magnetic Resonance Scanner. Images were analyzed for signal intensity ratio of LG and vitreous (LG/V), signal intensity ratio of pisilateral parotid gland to vitreous (PG/V), apparent diffusion coefficient (ADC) of LG and PG (DWI sequence with b=1000 mm/s<sup>2</sup>) and Trigeminal ganglion (TG) volume (mm<sup>3</sup>).

**Results.** One hundred-twenty-three completed and were classified as SS or NSS (84 and 44, respectively). The mean age is  $52\pm15$  years old, and 95% are women. The groups have a similar frequency of dry eye and dry mouth. The positivity for ANF, SSa, SSb and focus score are high in SS groups compared to non SS (p<0.05, Chi-square test). Mean values of Schirmer test, tear break-up time and salivary flow were lower and corneal staining score was higher in SS groups. The OSDI and PhQ-9 questionnaires presented higher score in nonSS patients (p<0.05, Mann-Whitney U). The

signal intensity ratios of LG/V and PG/V were significantly higher in the SS group (p=0.01, Tukey's test). ADC of LG was higher in) SS patients (p=0.003, Tukey's test). LG volume was larger in young SS patients compared to age-matched controls (p=0.03). The volume of TG is 218.9±16.1 in the control and 158.3±11.6 mm<sup>3</sup> in the SS group (p=0.008). The SS TG is smaller at younger ages, but the difference is attenuated after 70 years old. There was a positive correlation between cornea fluorescein staining and signal intensity ratio of LG (r=0.63 e p=0.003) and PG (r=0.53 e p=0.01). **Conclusions.** Our data confirm the world demographic and laboratory parameters for SS. It also reveals that MRI of LG, PG, and TG identify changes in SS patients and imaging findings correlate with clinical exams. Depressive symptoms and neuropathic pain need further investigation in conjunction with the clinical signs to identify the mechanisms, potential distinctive causes and better treatment for SS and NSS *sicca* syndrome. Financial Support: FAPESP, CNPQ, FAEPA, CAPES.

### P-42

## Comorbidity and polypharmacy burden in primary Sjögren syndrome

Wan-Fai Ng<sup>1</sup>, Simon Bowman<sup>2</sup>, the UK primary Sjogren's syndrome registry, Samira Tatiyama Miyamoto<sup>3</sup>.

<sup>1</sup>Musculoskeletal Research Group, Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, United Kingdom. <sup>2</sup>University Hospital Birmingham, UK. <sup>3</sup>Department of Integrated Education in Health, Federal University of Espírito Santo, Vitória-ES, Brazil.

**Background.** The Comorbidity-Polypharmacy Score (CPS) aims to provide an accurate measurement of the epiphenomena of comorbidity and polypharmacy, where the severity of comorbidities can be objectively compared and quantified. The CPS is an indirect estimate of the "physiologic age" of the patient(1). The aim of our study was to describe comorbidities and polypharmacy in a British cohort of pSS patients.

**Methods.** A retrospective study was performed using data of the UK Primary Sjögren's Syndrome Registry (UKPSSR, <u>www.sjogrensregistry.org</u>) from 30 British centers(2). 688 patients fulfilling the American European Consensus Group classification criteria for pSS were studied. All patients were assessed using ESSDAI, EQ-5D, ESSPRI, Profile of fatigue and discomfort (PROFAD) and Hospital Anxiety & Depression Score (HADS). Clinical and laboratory data were also collected. CPS was calculated by adding all comorbid conditions and medications based on self-reported, with exception of symptoms of dryness, fatigue and arthritis, and non-prescribed medications or supplements such as multi-vitamins. Patients were subsequently categorized according to CPS into four groups: 0-7 (minor), 8-14 (moderate), 15-21 (severe), or  $\ge 22$  (morbid) (1). Data analyses included descriptive statistics, chi-square test or Fisher's exact test for categorical data, as appropriate, and Kruskal-Wallis for continuous variables. Statistical significance was set at alpha = 0.05.

**Results.** The mean age of the 688 patients of the UKPSSR cohort was of 58.47 ( $\pm$  12.51) and 94.6% were woman. Most of them having low (58.4%) or moderate (36.5%) levels of disease activity as measured by ESSDAI. However, 60% of the patients having ESSPRI (score  $\geq$ 5). Patients in the morbid CPS category tended to be older and with higher levels of disease activity (ESSDAI, ESR and CRP) and more disease/symptoms duration. These patients also tended to have worse quality of life. However, there was no statistical difference between CPS groups in all continuous variables. The frequency of patients with positive ANA in morbid CPS group was higher (73.3%) than in the other groups (p=0.032). Musculoskeletal disorders (68%), gastrointestinal disorders (65%) and cardiovascular/hematologic diseases (44%) were the most frequent group of comorbidities. 11% of patients had malignancy and 45% of all malignancy cases (n=33) have lymphoma. Oesophageal disorders (39.4%), hypertension (39%), hypothyroidism (19.3%), dyslipidemia (11.5%), chronic cystitis (11.3%), oeteoporosis (11.2%) and asthma (10%) were the most common comorbidity.

**Conclusion.** Musculoskeletal disorders gastrointestinal disorders and cardiovascular/hematologic diseases were the most frequent group of comorbidities. 11% of patients had malignancy and 45% of all malignancy cases have lymphoma. Patients in the morbid CPS category tended to be older and with higher levels of disease activity and more disease/symptoms duration. The frequency of patients with positive ANA in morbid CPS group was higher than in the other groups.

#### References

1. EVANS DC et al.: J Am Geriatr Soc 2012; 60: 1465-70.

2. NG W-F et al.: Rheumatology 2011; 50: 32-9.

## **P-43**

#### Stability of clinical phenotype membership in primary Sjögren's syndrome: longitudinal follow up data from 244 UK and 237 French patients

Nadia Howard Tripp<sup>1,2</sup>, Dennis W Lendrem<sup>1</sup>, Jessica R Tarn<sup>1</sup>, Alain Saraux<sup>3</sup>, Valérie Devauchelle-Pensec<sup>3</sup>, Raphaèle Seror<sup>4</sup>, Xavier Mariette<sup>4</sup>, Simon J Bowman<sup>5</sup>, Jacques-Eric Gottenberg<sup>6</sup> on behalf of the French ASSESS cohort and Wan-Fai Ng<sup>1, 2</sup> on behalf of the UK Primary Sjögren's Syndrome Registry.

<sup>1</sup>Institute of Cellular Medicine, Newcastle University, UK. <sup>2</sup>Newcastle upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. <sup>3</sup>CHU Brest and INSERM 1227, Université Bretagne occidentale, Brest, France. <sup>4</sup>Université Paris-Sud, Hôpital Bicêtre, Hôpitaux Universitaires Paris-Sud, Paris, France. <sup>5</sup>University Hospital Birmingham, Birmingham, UK. <sup>6</sup>Rheumatology Department, National Center for Rare Systemic Autoimmune Diseases, Hôpitaux Universitaires de Strasbourg, Strasbourg, France.

**Background.** Four distinct clinical subgroups have been demonstrated in primary Sjögren's syndrome (pSS), based on cluster analysis of pain, fatigue, dryness, anxiety and depression symptom scores. Validated in two independent cohorts, these subgroups display clinical, biological and transcriptomic differences supporting the existence of pathobiological endotypes underpinning these clinical phenotypes. This is the first study examining the longitudinal stability of these clinical phenotypes and factors affecting migration between groups over time.

**Methods.** Retrospective data for 244 patients over a mean of 4 years in the UK Primary Sjögren's Syndrome Registry (UKPSSR) and for 237 patients over 5 years in the French Assessment of Systemic Signs and Evolution in Sjögren's Syndrome (ASSESS) cohort were analysed. Clinical phenotype membership was determined at two time-points using a validated Excel Macro model based on the aforementioned symptom scores. Phenotype membership at time-point 1 was compared to membership at time-point 2 to determine phenotype stability over time in each cohort. Non-parametric analysis was used to determine differences across phenotypes. Predictors of phenotype membership were analysed using nominal logistic regression modelling.

**Results.** The majority of patients remain the same phenotype (63% in UKPSSR, 57% in ASSESS) over this time period (Table). Clinical and biological measures (IgG, ESR, salivary flow and Schirmer's test) remain significantly different and with a similar distribution across the four phenotypes at both time points within the UKPSSR. The primary predictor of phenotype membership in both the UKPSSR and ASSESS cohorts is the initial phenotype ( $p \le 0.0001$  for both). In addition, both ESR and Immuno-globulins were significantly associated with migrations from one phenotype to another (p < 0.005) in the two cohorts. Other predictors included CRP (p=0.0076) and BMI (p=0.0089) in the ASSESS cohort, and Salivary Flow (p=0.0046) in the UKPSSR cohort.

**Conclusions.** Phenotype membership is generally stable over a 4 and 5 year period, in two independent pSS cohorts. This has implications for future studies, including clinical trials, and clinical management pathways, which may benefit from using this stratification method.



The above tree maps display cluster membership at two time points for the UMPISR and ASSESS colords within each phenotype. The left hand column of each thee map shows the original tating phenotype two each group, and the right hand column shows the tables up phenotype at 4 and 5 years respectively written the UMPISR and ASSESS colords. This figure shows the phenotypes in eigeneaity shall be used on years are marking by mempiate to certain phenotype. The right hand column of each thee map shows The legand displays the 4 different phenotypes. LSB-Low Symptom Durden, HSD-High Symptom Durden, DGF-Dyness Dominant Fatgue, POF-Pain Dominant Fatgue.

#### **P-44**

## Long-term evolution to additional autoimmune diseases in patients with primary Sjögren's syndrome

Susumu Nishiyama, Tetsushi Aita, Yasuhiko Yoshinaga, and Shoji Miyawaki. Rheumatic Disease Center, Kurashiki Medical Center

**Background.** French cohort study revealed only a few percent of patients with primary Sjögren's syndrome (pSS) developed other autoimmune diseases and those patients had active immunological profile and extra-glandular manifestations (1); however, clinical features of patients developing secondary autoimmune diseases are not fully understood. The aim of this study is to investigate clinical features of patients with pSS who develop secondary autoimmune diseases.

Methods. A total of 140 patients with pSS according to the 1999 revised Japanese criteria visited our hospital between 1989 and 2006. Clinical and laboratory data were collected from their medical charts, and 80 patients (79 women) who had visited our hospital more than 10 years were selected. Results. Average age (SD) at the onset of pSS, at the first visit to our hospital, and at the final observation was 43.7 (13.1), 49.4 (12.0), and 66.7 (12.7) years old, respectively. Average observation period (range) was 17.3 (10~28) years. Number of patients with positive anti-Ro/SSA and positive anti-La/SSB was 76 (95.0%) and 45 (56.3%), respectively. Seventee patients had the other auto-antibodies (anti-RNP, anti-centromere 6, anti-DNA 3, and anti-mitochondria 1). Ten patients developed additional autoimmune diseases (rheumatoid arthritis 4, systemic lupus erythematosus 3, mixed connective tissue disease 3, systemic sclerosis 2, and dermatomyositis 1) 1 to 29 years after the onset of pSS. There was no significant difference of clinical and laboratory features except age at disease onset between patients with and without secondary autoimmune diseases. Evolution of additional autoimmune diseases was associated with younger (<35 years old) onset of pSS. Comparing 22 of younger onset patients and rest of 58 patients, significant difference of aseptic meningitis (13.6% vs. 0.0%, p<0.05), leucocytopenia (68.2% vs. 36.2%, p<0.05), and evolution of additional autoimmune diseases (31.8% vs. 5.2%, p < 0.01) were observed. Evolution of systemic lupus erythematosus was observed only in younger onset group.

**Conclusions.** Long-term evolution of additional autoimmune diseases was frequently observed in younger onset patients with pSS, who had higher complication rate of aseptic meningitis and leucocytopenia.

#### Reference

 FAUCHAIS AL *et al.*: Immunological profile in primary Sjögren syndrome Clinical significance, prognosis and long-term evolution to other auto-immune disease. *Autoimmunity Reviews* 2010; 9: 595-9.

#### **P-45**

#### Progression of glandular damage but not other disease manifestations of Sjögren's syndrome in patients evaluated years after initial classification

Astrid Rasmussen<sup>1</sup>, Lida Radfar<sup>2</sup>, Kimberly Hefner<sup>3</sup>, David M. Lewis<sup>2</sup>, C. Erick Kaufman<sup>4</sup>; Donald U. Stone<sup>5</sup>, Kerry M Leehan<sup>1</sup>, Kiely Grundahl<sup>1</sup>, Judy Harris<sup>1</sup>, Sarah Cioli<sup>1</sup>, Cherilyn Pritchett-Frazee<sup>1</sup>, Laura Battiest<sup>1</sup>, Sharon Johnson<sup>1</sup>, Wesley Daniel<sup>1</sup>, Janice Gales<sup>1</sup>, Christopher J. Lessard<sup>1</sup>, A. Darise Farris<sup>1</sup>, R. Hal Scofield<sup>1,4,6</sup>, Kathy L. Sivils<sup>1</sup>.

<sup>1</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA. <sup>2</sup>University of Oklahoma College of Dentistry, Oklahoma City, OK, USA. <sup>3</sup>Hefner Eye Care, Oklahoma City, OK, USA. <sup>4</sup>Department of Medicine, The University of Oklahoma Health Sciences Center, Oklahoma City, OK, USA. <sup>5</sup>Department of Ophthalmology, Wilmer Ophthalmological Institute, Johns Hopkins University, Baltimore, MD, USA. <sup>6</sup>US Department of Veterans Affairs Medical Center, Oklahoma City, OK, USA.

**Background.** Classical connective tissue diseases, such as SLE and RA have well documented progression of disease and damage accrual. However, the natural history of Sjögren's syndrome (SS) has been less well documented, particularly in research settings where comprehensive multidisciplinary assessments can be performed at more than one timepoint. The objective of the present study was to re-evaluate past participants in the OMRF Sjögren's research clinic (SRC) with the same protocol that was used for their initial research classification with the goal of documenting changes in their disease.

**Methods.** Questionnaires assessing health changes and willingness to be re-evaluated were mailed to 800 SRC participants. Twenty respondents (17

SS and 3 incomplete SS [iSS] both by AECG and ACR-EULAR criteria) participated in this pilot study in which all procedures performed in their initial evaluation were repeated.

Results. 356 (45%) questionnaires were answered (161 SS and 195 iSS). Subjectively, respondents reported equal or better status of ocular and oral symptoms but significantly worse fatigue and arthralgias (p<0.001) with no differences between SS and iSS. The 20 re-evaluated patients returned after an average of 5.4 years (range 2-9). Thirteen (65%) retained the initial disease classification, but 6 subjects (30%) went from SS to iSS and 1 (5.0%) from iSS to SS. It is noteworthy that this subject only met SS criteria by AECG and not by ACR-EULAR because his serology was anti- La (+) only with negative biopsy. Furthermore, only 2 subjects had a net increase in the number of criteria, while 18 had the same number (n=8) or fewer (n=10). There were no consistent patterns of change in the objective measures of lacrimal and salivary gland function: 5 subjects became Schirmer's (+) and 1 reversed to (-); the opposite was the case for the ocular staining, with 5 becoming (-) and 1 becoming (+). The only unchanged results in all the recalls were the anti-Ro/anti-La status. The most intriguing results were the changes in the minor salivary gland biopsy; 4 (20%) subjects went from positive to negative biopsy resulting in a change in classification from SS to iSS in 3 cases. Moreover, the focus score was lower in the second biopsy of 10 (50%) cases, 4 (20%) had a higher score and the remaining 6 (30%) were unchanged. The morphology of the salivary gland tissue of these reversed cases showed extensive fibrosis, fatty infiltration and atrophy of the gland, precluding the lymphocytic infiltrates from meeting the definition of being surrounded by normal tissue. Further supporting the notion of worsening gland architecture, final focus score trended to inversely correlate with WUSF (r=- 0.4; p=0.089). Complement levels and hypergammaglobulinemia did not predict stability or worsening of SS.

**Conclusions.** Re-evaluated patients showed little disease progression but steady presence of autoantibodies. Detrimental changes in salivary gland morphology were observed in later biopsies, even when classified as "negative" using current criteria. These results suggest that significant tissue destruction in long standing disease may lead to false negative biopsy results in spite of progressive glandular dysfunction.

#### **P-46**

## Diagnosis of keratoconjunctivitis Sicca: identifying the signs of dry eye disease

John Gonzales<sup>1,2</sup>, Vatinee Bunya<sup>3</sup>, Thomas Lietman<sup>1,2</sup>

<sup>1</sup>Francis I. Proctor Foundation, University of California San Francisco. <sup>2</sup>Department of Ophthalmology, University of California San Francisco. <sup>3</sup>Scheie Eye Institute, Department of Ophthalmology, University of Pennsylvania.

**Purpose.** There is no gold standard for the definitive diagnosis of keratoconjunctivitis sicca (KCS). Instead, ophthalmologists must rely on clinical signs and tests to diagnose dry eye disease. Clinical signs and tests include tear-break-up time (TBUT), Ocular Staining Score (OSS), Schirmer 1, and tear osmolarity. The Sjögren's International Collaborative Clinical Alliance (SICCA) developed new classification criteria for Sjögren's syndrome (SS), and seeks to better characterize the SS phenotype and genotype, and establish a SS data and specimen repository to support future research.

**Methods.** Levels of sensitivity and specificity for each sign (variable) were assessed using latent class analysis. Our modeling was based on four predictor variables that relate to signs of KCS. Latent class analysis allowed for estimation of sensitivity and specificity using the model-based classification as a "gold standard". We utilized R package for latent class analysis (R version 3.3.2 and RStudio 1.0.136, R, Boston, MA).

**Results.** A total of 3,514 participants from 9 international sites were enrolled into SICCA. Women made up the majority of participants (n=3,185 or 91%). SS as defined by ACR/EULAR criteria was diagnosed in 1,541 participants (52.9%) and 116 participants (3.3%) could not be classified. With latent class analysis, we found a best fit model with two groups, a gold standard-positive group and a gold standard-negative group. For the gold standard-positive group, having an abnormal TBUT, Schirmer 1, tear osmolarity, and OSS had a sensitivity of 100%, 19%, 43%, and 88%, respectively. In the gold standard-negative group having an abnormal TBUT, Schirmer 1, tear osmolarity, and OSS had a specificity of 40%, 100%, 72%, and 91%, respectively.

**Conclusions.** The Ocular Staining Score differentiated the gold standardpositive group from the gold standard-negative group better than other KCS parameters.

## **P-47**

In vivo confocal microscopy evaluation of corneal sub basal nerve and dendritic cell in Sjögren syndrome patients

Qin Zhang, Fangting Li, Jing He, Yuebo Jin, Yadan Zou

**Background.** To analyze the relationship between in vivo confocal microscopic corneal density, dendritic cell density and clinical evaluation in patients with Sjogren syndrome (SS).

**Methods.** Seventeen patients with SS and sixteen healthy age- and sexmatched Non-Sjögren dry eye disease patients control subjects were included. A clinical evaluation of dry eye(Non-invasive tear meniscus height, non-invasive break-up time, meibography of both upper and lower meibomian gland), in vivo confocal microscopy analysis of the central cornea were performed for all patients.

**Results.** Non-invasive tear meniscus height (NI-TMH), non-invasive breakup time (NI-BUT), the loss of upper meibomian gland and corneal sub basal nerve density were significantly lower in the SS group as compared with the control group (p<0.01). Dendritic cell density of central cornea increased significant in SS group (p<0.01). The loss of upper meibomian gland was correlated to age (r=0.378, p=0.033). In the SS group, corneal sub basal nerve density was correlated to NI- BUT (r=0.543, p=0.001).

**Conclusions.** The dry eye, the loss of both meibomian gland and corneal sub basal nerve was more severe in SS patient than in non-Sjogren dry eye disease patients. Confocal microscopy can be an important diagnostic tool in evaluation the ocular surface change in SS patients.

#### **P-48**

## Quantification of proteoglycan 4 (PRG4) / lubricin in normal and Sjögren syndrome human tears

SC Regmi<sup>1</sup>, S Srinivasan<sup>2</sup>, M Heynen<sup>2</sup>, GD Jay<sup>3</sup>, BD Sullivan<sup>4,5</sup>, L Subarraman<sup>2</sup>, B Caffery<sup>6</sup>, L Jones<sup>2</sup>, TA Schmidt<sup>7</sup>. <sup>1</sup>Faculty of Kinesiology, University of Calgary, Calgary, CA, Canada. <sup>2</sup>Centre for

<sup>1</sup>Faculty of Kinesiology, University of Calgary, Calgary, CA, Canada. <sup>2</sup>Centre for Ocular Research and Education, School of Optometry & Vision Science, University of Waterloo, Waterloo, ON, Canada. <sup>3</sup>Emergency Medicine, Brown University, Providence, USA. <sup>4</sup>TearLab, San Diego, USA. <sup>5</sup>Lubris BioPharma LLC, Framingham, USA. <sup>4</sup>Toronto Eye Care, Toronto, ON, Canada. <sup>7</sup>Biomedical Engineering Department, University of Connecticut Health Center, Farmington, USA.

**Background.** Sjögren's syndrome (SS) is an autoimmune disease with hallmark clinical symptoms of dry eye and dry mouth. Proteoglycan 4 (PRG4), or lubricin, is a mucin-like glycoprotein that is naturally present on the ocular surface, and in tears, where it contributes to ocular surface health. PRG4 functions as a boundary lubricant to reduce friction between ocular surfaces, and also demonstrates anti-inflammatory properties. Recently, recombinant human PRG4 was shown to be clinically effective in improving signs and symptoms of dry eye in SS patients. Interestingly, PRG4 in human tears was also shown to be susceptible to proteolytic digestion by cathepsin S, an enzyme with increased activity in SS tears, which destroyed the *in vitro* ocular surface boundary lubricating ability. However, whether levels of PRG4 are diminished in SS tears remains to be determined. The objective of this study was to quantify PRG4 levels in normal and SS human tears.

Methods. Tears were collected from 17 SS (15 F, 2 M, 56.2±16.7 years old) and 20 asymptomatic (n=20, 7 M, 13 F, 31.2±11.4 years old) participants, with approval from the Office of Research Ethics (UWaterloo). SS participants were diagnosed using the American European Consensus Criterion. Tears were collected without anaesthetic, from the inferior temporal tear meniscus of each eye, using a disposable microcapillary tube and frozen at -80C until use. The concentration of PRG4 was determined via a sensitive. competitive amplified luminescent proximity homogeneous assay using recombinant human PRG4 as the control. Total mass of PRG4 was calculated by normalizing concentration by tear volume, using 5.0 ul for normal tears and measured SS tear volume (0.1 to 2.3 ul). Data is reported as mean±SD, nonparametric statistics were employed (Mann-Whitney U & Levine tests). Results. The concentration of PRG4 in SS (28.6±44.3 ug/ml) was not significantly different than that of normal tears  $(2.6\pm2.0 \text{ ug/m1}, p=0.15)$ , but did demonstrate significantly greater variation (p<0.001). The mass of PRG4 in SS tears (10.6±4.8 ng) was significantly diminished compared to normal tears (12.8±1.4 ng, p<0.05).

**Conclusions.** PRG4 concentration is significantly more variable in SS tears, and when normalized by volume, the PRG4 mass in SS tears is diminished

compared to normal tears. These data suggest either a reduction in PRG4 production or an increase in PRG4 catabolism in SS tear relative to normal tears, which could be the cause of the variability of PRG4 concentration in SS tears. Given the role PRG4 plays in ocular surface health and its susceptibility to degradation by cathepsin S in SS tears, diminished endogenous levels of PRG4 could contribute to signs and symptoms of dry eye in SS.

### **P-49**

#### Correlations between the severity of xerophthalmia and inflammation state in primary Sjögren syndrome

Yadan Zou<sup>1,2</sup>, Yuebo Jin<sup>2</sup>, Qin Zhang<sup>3</sup>, Han Wang<sup>2</sup>, Jiali Chen<sup>2</sup>, Yifan Wang<sup>2</sup>, Huaqun Zhu<sup>2</sup>, Jing He<sup>2</sup>, Zhanguo Li<sup>2</sup>.

<sup>1</sup>Department of Rheumatology and Immunology, Peking University International Hospital, Beijing, China, <sup>2</sup>Department of Rheumatology and Immunology, Peking University People's Hospital, Beijing, China. <sup>3</sup>Department of Ophthalmology, Peking University People's Hospital, Beijing, China.

**Objective.** To assess the relationship between ocular surface parameters and inflammation state in patients with primary Sjögren syndrome (pSS).

**Methods.** 62 female patients with primary Sjögren syndrome (pSS). **Methods.** 62 female patients with pSS (age49.79±12.43 years) and 16 healthy control subjects (age  $55.67\pm13.53$  years) were recruited. Clinical Characters were documented. The immunoglobulin levels, the cytokine levels complement levels, measurements of tear break-up time (TBUT), mebomian gland (MG) evaluations, Schirmer I test, non-contact infrared meibography and lid margin morphology examination using slitlamp microscopy were performed. The MG loss, calculated as (tarsal area-MG area)/tarsal area, was evaluated in upper lids (UL). The T-test and chi-square test were used to analyze the associations between the groups. p<0.05 was considered statistically significant. Pearson and Spearman's tests were used for correlation analysis. Logistic regression analysis of meibomian gland damage was conducted for SS patients.

**Results.** (1) The ocular parameters are highly relevant and valuable for the dry eye detection. The ocular parameters of the non-invasive keratograph tear break-up time (NIKBUT), Schirmer I test , tear break-up time(BUT), the number of meibomian gland and the MG loss for the pSS group were significantly lower than those for the healthy control subjects (p<0.05). (2) The tear meniscus height(TMH) was negatively correlated with Treg and Breg level (r=-0.255, p<0.05); Schirmer I test has a negative correlation with r–globulin (r=0.29, p<0.05) and a positive correlation with transforming growth factor-beta1 (TGF-beta1) (r=0.27, p<0.05). The number of meibomian glands has a negative correlation with sCD25 and IL-17a level (respectively, r=-0.232, r=-0.201, p<0.05). The MG loss is negatively correlated with r–globulins level and Treg/Tfh+Th17 level (respectively, r=-0.256, p<0.05).

**Conclusion.** Patients with pSS has more sever xerophthalmia than controls. The severity of xerophthalmia in patients with pSS correlates with the levels of IgG, TGF-beta, IL-17a and sCD25.

#### P-50

## IL-14 as a putative biomarker for stratification of dry eyes in primary Sjögren's syndrome

Yichen Liang<sup>1</sup>, Dehua Fu<sup>2</sup>, Shuang Liu<sup>2</sup>, Yang Yao<sup>2</sup>, Chun Gao<sup>2</sup>, Guixiu Shi<sup>1</sup>and Jing He<sup>3</sup>.

<sup>1</sup>Department of Rheumatology and Immunology, First Affiliated Hospital of Xiamen University, Xiamen, China. <sup>2</sup>Department of Gastrointestinal Surgery, Tongji Hospital of Tongji Medical College of Huangzhong University of Science & Technology, Wuhan, China. <sup>3</sup>Department of Rheumatology and Immunology, Peking University People's Hospital, Beijing, China.

**Background.** The pathogenesis of primary Sjögren's syndrome (pSS) is associated with abnormal B cell activation, resulting in production of excessive autoantibodies and disorder to the cytokine network. IL-14 (also known as Taxilin) is a cytokine that was shown to enhance B cell proliferation, especially of germinal center B cells. Transgenic mice overexpressing human IL-14 alpha can develop many clinical features of pSS in the same relative time frame as seen in patients. While upregulation of IL-14 gene expression has been shown in the peripheral blood leukocytes of pSS patients, till now the measurement of IL-14 serum levels has not been possible due to lack of validated assay. Our purpose was to evaluate IL-14 as a biomarker and its

correlation to B cell activating factor (BAFF), a well-established cytokine in pSS through upregulating innate immune activation and chronic autoimmune B cell activation, in a cohort of patient with non-SS dry eye (NSDE), pSS, diseases and healthy controls (HC).

Methods. Total of 181 fresh serum samples were collected and stored in -80 degrees freezer. Among them, 65 were pSS patients (age 53.15±14.08 years) who meet the 2012 ACR Classification Criteria for Sjögren's Syndrome, 20 were dry eye patients excluding SS (age 44.85±11.39 years, NSDE), 50 were Rheumatoid Arthritis patients (age 54.95±15.35 years, RA) and 46 were healthy controls (age 43.49±14.57 years, HC). Serum level of IL-14 was evaluated by quantitative Western Blots assay and BAFF level was evaluated by ELISA assay (R&D System). All clinical and laboratory data were reviewed following protocol approved by Peking University People's Hospital IRB committee. Statistical analysis was done by software Prism 6.0 with unpaired t tests.

**Results.** 1) After normalized with internal control, the relative intensity ratio for serum IL-14 level in HC group was  $2.13\pm0.81$ , NSDE group was  $2.11\pm0.98$  (p=0.99), pSS group was  $2.92\pm0.93$  (p<0.0001) and RA group was  $2.47\pm0.95$  (p=0.15). Serum BAFF level (pg/ml) in HC group was  $323.56\pm65.85$ , DE group was  $355.21\pm87.86$  (p=0.22), pSS group was  $455.94\pm155.16$  (p<0.0001) and RA group was  $448.38\pm220.07$  (p=0.0002).

2) For age <40 years, the serum level of HC group was  $2.26\pm0.73$ , NSDE group was  $2.21\pm0.93$  (p=0.89), pSS group was  $3.48\pm0.88$  (p=0.0003) and RA group was  $3.28\pm0.87$  (p=0.08). For age 40 to 60 years, the serum level of HC group was  $2.08\pm0.93$ , NSDE group was  $1.93\pm1.11$  (p=0.69), pSS group was  $2.83\pm0.98$  (p=0.01) and RA group was  $2.23\pm0.76$  (p=0.87). For age >60 years, the serum level of HC group was  $1.93\pm0.68$ , NSDE group was  $2.56\pm0.59$  (p=0.21), pSS group was  $2.76\pm0.81$  (p=0.02) and RA group was  $2.49\pm1.04$  (p=0.12).

3) In pSS patients, the serum level of IL-14 decrease as age increase (<40 years,  $3.48\pm0.88$ , 40-60 years,  $2.83\pm0.98$ , (p=0.048) and >60 years,  $2.76\pm0.81$ , (p=.023)). Whereas the serum level of BAFF (pg/ml) increase as age increases (<40 years,  $414.22\pm119.94$ , 40-60 years,  $406.22\pm148.29$ , (p=0.87), >60 years,  $524.57\pm159.46$ , (p=0.008)).

**Conclusions.** 1) Elevation of serum IL-14 level can serve as a key cytokine biomarker for the stratification of SS vs NSDE.

2) IL-14 and BAFF may work in different fashions to maintain the abnormal B cell activation as seen is pSS patients.

#### P-51

## AIM2 is a novel interferon-inducible autoantigen targeted in Sjögren's syndrome

Brendan Antiochos<sup>1</sup>, Alan Baer<sup>1</sup>, Jungsan Sohn<sup>2</sup>, Livia Casciola-Rosen<sup>1</sup> and Antony Rosen<sup>1</sup>

<sup>1</sup>Johns Hopkins University School of Medicine, Division of Rheumatology and <sup>2</sup>Johns Hopkins University School of Medicine, Department of Biophysics and Biophysical Chemistry.

**Background.** A variety of cellular antigens are targeted by autoantibodies in Sjögren's syndrome (SS). Interferon-inducible protein 16 (IFI16) is one such autoantigen, belonging to the PYHIN family of cytoplasmic DNA sensors. Absent in melanoma 2 (AIM2) is another IFN-induced DNA sensor in this protein family. Given the similar IFN-induced expression, DNA sensing function, and structural properties of AIM2 and IFI16, we sought to determine whether AIM2 was also targeted by autoantibodies in SS.

**Methods.** Sera from 132 consecutively enrolled primary SS patients were used to immunoprecipitate <sup>35</sup>S-methionine labelled AIM2 protein. 49 healthy controls were used to establish a threshold for assay positivity. Clinical features of SS patients with and without AIM2 autoantibodies were compared using Fisher's exact test and the Mann-Whitney test.

**Results.** Anti-AIM2 antibodies were found in 46/132 (35%) of SS patients. Antibodies against AIM2 were associated with the presence of anti-Ro52, Ro60 and La. Interestingly, there was no correlation between anti-AIM2 positivity and anti-IF116 antibodies. Anti-AIM2+ patients had higher focus scores on labial salivary gland biopsies than anti-AIM2- subjects. We found a positive correlation between anti-AIM2 antibody titer and lip biopsy focus score (Spearman r=0.2164, p=0.0495). The presence of high titer anti-AIM2 antibodies was associated with leukopenia (43% vs 14%, p=0.019) and lower whole unstimulated salivary flow rates (0.45 $\pm$ 0.78 mL/5min vs 0.94 $\pm$ 0.96 mL/5min, p=0.0493). ANA, rheumatoid factor, and hypergammaglobulinemia were also strongly associated with anti-AIM2 antibodies. There was no difference in measures of eye dryness between patients with and without anti-AIM2 antibodies. Posters

Table I. Demographic and phenotypic characteristics of 132 Sjögren's syndrome patients.

Feature	Number (%)
Age (years) at blood draw, mean ± SD Female	53.2 ± 11.8 120/132 (91%)
Female Race • Caucasian • African American • Asian • Hispanic/Latino LSG Biopsy Positive Focus score, mean ± SD Ro60 Positive Ro52 Positive La Positive IF116 Positive ANA ≥ 1:320 Rheumatoid Factor Positive Huppergemendebulinemia	$\begin{array}{c} 120/132 \ (91\%) \\ 115/132 \ (87.2\%) \\ 11/132 \ (8.3\%) \\ 3/132 \ (2.3\%) \\ 3/132 \ (2.3\%) \\ 80/128 \ (63\%) \\ 2.43 \pm 1.5 \\ 82/132 \ (62\%) \\ 80/132 \ (61\%) \\ 44/132 \ (33\%) \\ 40/132 \ (53\%) \\ 44/130 \ (34\%) \\ 35/129 \ (27\%) \end{array}$
Low C4	14/132 (11%)
MGUS Leukopenia Whole Unstimulated Salivary Flow, mean ± SD Schirmer's Test, mean ± SD SICCA Ocular Staining Score, mean ± SD	$\begin{array}{c} 12/117  (11\%) \\ 25/131  (19\%) \\ 0.87 \pm 0.95 \\ 5.70 \pm 6.99 \\ 7.56 \pm 3.42 \end{array}$

Numerators correspond to number of patients with indicated feature positive and denominators to total number of patients with indicated feature recorded in the cohort, followed by percent (%) positive.

LSG: Labial salivary gland; Focus score: Lymphocytic foci / 4 mm<sup>-2</sup>; ANA: Antinuclear antibody; Hypergammaglobulinemia: IgG >1560 mg/dL or IgG >304 mg/dL; Low C4: <12 mg/dL; MGUS: Monoclonal gammopathy of undetermined significance; Leukopenia: WBC <4000/µL; Whole Unstimulated Salivary Flow: mL saliva / 5 minutes; Schirmer's Test: mm wetting / 5 minutes.

**Conclusions.** AIM2 is an IFN-induced autoantigen in SS. Anti-AIM2 antibodies are associated with increased labial salivary gland inflammation and serologic markers of severe disease. IFN signaling in the salivary gland may contribute to SS pathogenesis by inducing the expression of autoantigens.

## P-52

## Aberrant expression of the innate restriction factor bone marrow stromal antigen-2 in primary Sjögren's syndrome

Chan Chen<sup>1</sup>, Huan Shi<sup>1</sup>, Ningning Cao, Chuangqi Yu<sup>2</sup>, Lingyan Zheng<sup>2</sup>. <sup>1</sup>Department of Oral Maxillofacial Surgery, Ninth People's Hospital Shangai Jiao Tong University School of Medicine. <sup>2</sup>Shanghai Key Laboratory of Stomatology and Shanghai Research Institute of Stomatology, National Clinic Research Center of Stomatology.

**Background.** Bone marrow stromal antigen-2 (BST-2) is a transmembrane innate immune protein that was detected to play important role in some autoimmune diseases. The objective of this study was to analyze BST-2 levels in labial glands, total peripheral blood mononuclear cells (PBMCs) and PBMC subpopulations from primary Sjögren's syndrome (pSS) patients and determine the correlation between BST-2 expression and clinical characteristics.

Methods. PBMC subsets were positively separated using magnetic microbeads. BST-2 mRNA levels in labial glands, total PBMCs and PBMC subsets of 30 pSS and 16 healthy control (HC) subjects were investigated using real-time polymerase chain reaction (RT-PCR). Distribution of BST-2-positive cells in the labial glands was assessed by immunohistochemistry. **Results.** BST-2 was significantly increased in pSS labial glands and was positively correlated with the VAS value for parotid gland swelling and rheumatoid factor and  $\beta$ 2-microglobulin serum levels. BST-2 levels were statistically different between pSS patients with positive and negative expression of anti-SSA antibody. Positive glandular epithelial cells, ductal epithelial cells and adjacent infiltrating lymphocytes were observed in labial glands from pSS patients, while there were a few scattered positive ductal epithelial cells in controls. BST-2 was also up-regulated in CD19+ B cells and the remaining CD4-CD8-CD19- PBMCs.

**Conclusion.** BST-2 was aberrantly expressed in pSS patients, and expression in labial glands was positively correlated with important clinical characteristics; thus, it may be a potential biomarker of pSS activity.

P-53

## Autoantibodies to ox-LDL in Sjögren's syndrome: are they atheroprotective?

Ilir Cinoku<sup>1, 2\*</sup>, Clio P. Mavragani<sup>1, 3\*</sup>, Costantinos C. Tellis<sup>2</sup>, Adrianos Nezos<sup>3</sup>, Alexandros A. Tselepis<sup>2\*\*</sup>, Haralampos M. Moutsopoulos<sup>1,4\*\*</sup> \*equally contributed as first authors, \*\*equally contributed as last authors. <sup>1</sup>Department of Pathophysiology, School of Medicine, National and Kapodistrian University of Athens, Athens, Greece. <sup>2</sup>Department of Chemistry, Laboratory of Biochemistry, Medical School, University of Ioannina, Ioannina, Greece. <sup>3</sup>Department of Physiology, School of Medicine/Interview of Athens, Athens, Athens, Greece. <sup>4</sup>Chair Medicine/Immunology, Academy of Athens, Athens, Greece.

**Background.** The higher incidence of atherosclerosis and cardiovascular disease (CVD) in patients with systemic autoimmune diseases cannot be attributed exclusively to traditional risk factors for CVD. Antibodies to oxidized Low Density Lipoprotein (ox-LDL) seem to have a crucial role in atherogenesis.

**Methods.** Sera from 63 consecutive patients with primary Sjögren's Syndrome (SS), 121 with Systemic Lupus Erythematosus (SLE), 79 with Rheumatoid Arthritis (RA) and 26 apparently healthy individuals were evaluated for the presence of antibodies to ox-LDL by an ELISA method. The femoral and/or carotid Intima Media Thickness (IMT) and plaque formation as well as traditional CVD risk factors and disease related features were recorded for all study participants.

**Results.** Anti-ox-LDL antibody levels were significantly reduced in SS and RA patients, but not in SLE patients, compared to their healthy counterparts. Subsequently, SS patients were divided into two groups according to antibody levels to ox-LDL, using as cut off the median of each group studied. SS patients with high titers of antibodies to x-LDL displayed higher rates of autoantibodies to Ro/SSA and La/SSB antigens, purpura, low complement levels and increased SS activity index. On the other hand, the high anti-oxLDL group was characterized by reduced rates of carotid and/or femoral plaque after adjusting for potential confounders (OR [95%CI]: 0.14 [0.03-0.72]). Such associations were not shown in all other groups included in the study. **Conclusions.** These findings suggest that antibodies to ox-LDL, possibly resulting from B cell hyperactivity, might exert a protective role in the development of atherosclerosis among primary SS patients.

### **P-54**

#### Positivity for anti-RNP in primary Sjögren syndrome patients is associated with a more active disease and a higher risk of muscular and pulmonary involvement

S. Abbara<sup>1</sup>, R. Seror<sup>1</sup>, J Henry, P Chretien<sup>2</sup>, A Gleizes<sup>2</sup>, S Hacein Bey<sup>2</sup>, G. Nocturne<sup>1</sup>, X. Mariette<sup>1</sup>.

<sup>1</sup>Department of Rheumatology, Hôpitaux universitaires Paris-Sud, France. <sup>2</sup>Department of Immunology, Hôpitaux universitaires Paris-Sud, France.

Primary Sjögren Syndrome (pSS) can be associated with anti-Sm and/or anti-ribonucleoprotein (RNP) antibodies without anti-DNA antibodies. Whether these auto-antibodies are associated with a specific phenotype is unknown.

**Objectives.** To describe the clinical and paraclinical characteristics of patients displaying a pSS with anti-Sm and/or anti-Rnp antibodies, without anti-DNA antibodies.

**Methods.** Patients fulfilling ACR/EULAR 2016 criteria for pSS without other connective tissue disease diagnosis and having anti-Sm and/or anti-RNP antibodies, without anti-DNA antibodies were screened in the database from the French National Reference Center, Paris-Sud University. We compared them to all pSS patients from the Paris-Sud cohort with negative anti-Sm, anti-Rnp and anti-DNA antibodies.

**Results.** At inclusion twenty three patients (n=18 women, 3 men) were in the anti-Sm and/or anti-Rnp group (anti-Sm: n=7, anti-RNP; n=22), and 446 in the anti-Sm and anti-RNP negative group (n=426 women, 20 men). All the patients fulfilled the ACR EULAR 2016 criteria for pSS. All patients had negative anti-DNA antibodies, and none had a diagnosis of lupus according to the SLICC criteria. In the anti-Sm and/or anti-RNP positive group, 5 patients fulfilled previously described criteria of Mild Connective Tissue Disorder (Sharp; n=1, Kasukawa; n=4, Alarcon-Segovia; n=3, Kahn; n=1). Anti-Sm and/or anti-RNP positive patients had a lower mean age at onset of pSS symptoms (40 vs 48 years, p=0.02), a higher mean ESSDAI at inclusion (13 vs 4, p<0.01), more frequent objective xerostomia or xerophtalmia

(95% vs 61%, p<0.01), myositis (26% vs 2%, p<0.01), pulmonary (30% vs 6%, p<0.01), cutaneous (65% vs 36%, p<0.01) and peripheral nervous system involvement (13% vs 3%, p=0.04). Moreover, anti-Sm and/or anti-RNP positive patients had higher mean gammaglobulins (25 vs 14 g/l, n<0.01) and CPK levels (660 vs 113 U/l, n=0.02), more frequent anti-SSA antibodies (91% vs 67%, p=0.01), but less frequent lymphocytic sialadenitis with a focus score>=1 (61% vs 86%, p<0.01). Although sclerodactyly was found in 13% of patients with anti-Sm and/or anti-RNP antibodies and limited cutaneous sclerosis in 4%, no patients had a diagnosis of scleroderma.

**Conclusion.** pSS with anti-Sm and/or anti-RNP antibodies subjects seem to display a more active systemic disease, with a more frequent pulmonary and muscular involvement.

Table I. Characteristics of the 23 patients with primary Sjögren's syndrome and anti-
Rnp and/or anti-Sm antibodies at inclusion, as compared to 446 pSS patients without
anti RNP and without anti Sm.

Characteristics	pSS patients with anti-Rnp and/or anti-Sm, n=23	pSS patients without anti- RNP/anti-Sm n=446	p-value
Classification			
Objective xerostomia or xerophtalmia, n (%	b) 18/19 (94,7)	263/432 (60,9)	0.003
Lymphocytic sialadenitis	14 (60,9)	365/427 (85,5)	0.002
(focus score $\geq 1$ ), n (%)			
Mean Chisholm score, [IQ1-IQ3]	2,7 [2,0-4,0]	3,1 [3,0-4,0]	0.27
Positive anti-SSA antibodies, n (%)	21 (91,3)	298/446 (66,8)	0.014
Positive anti-SSB antibodies, n (%)	5 (21,7)	162/443 (36,6)	0.183
Age at onset of Sjögren's symptoms, mean [IO1-IO3]	40,1 [30,5-47,5]	48,3 (36,0-62,0)	0.021
ESSDAI at inclusion, mean [IQ1-IQ3]	14,0 [4,0-18,0]	4,0 [1,0-6,0]	< 0.001
Systemic manifestations			
General symptoms, n (%)	3 (13,0)	1/146 (0,7)	0.008
Parotidal involvment, n (%)	7 (30,4)	170/438 (38,8)	0.421
Joint involvment, n (%)	20 (87,0)	335/444 (75,4)	0.208
Myalgia, n (%)	7 (30,4)	132/438 (30,1)0.976	
Myositis, n (%)	6 (26,1)	10/436 (2,3)	< 0.001
Pulmonary parenchyma involvment, n (%	) 7 (30,4)	25/437 (5,7)	< 0.001
Pulmonary hypertension with cardiac	1 (4,3)	0 (0,0)	0.049
Cutaneous involvment, n (%)	15 (65.2)	161/442 (36.4)	0.006
Peripheral nervous system involvment, n	(%) 3 (13,0)	13/419 (3,1)	0.041
Central nervous system involvment, n (%	0.0)	5/433 (1.2)	1
Renal Involvment, n (%)	0 (0.0)	1/124 (0.8)	1
Lymphoma, n (%)	1 (4,3)	16/443 (3,6)	0.584
Biology			
Positive ANA Antibodies. n (%)	23 (100.0)	328/439 (74.7)	0.006
Positive RF. n (%)	11 (47.8)	205/436 (47.0)	0.940
Lymphopenia, n (%)**	5 (21.7)	57/433 (13.2)	0.222
Gammaglobulins or $\lg G > 16g/L_{\odot}$ n (%)	18 (78.3)	154/438 (35.2)	< 0.001
Cryoglobulinemia n (%)	1 (4 3)	7/423 (17)	0.348
Decreased C4. n (%)	7 (30.4)	88/406 (21.7)†	0.325
Mean gammaglobulins, g/L, [IO1-IO3]	24.6 [16.4-30.0]	14.1 [10.1-16.3]	< 0.001
Mean CK value, U/L, [IQ1-IQ3]	659,5 [85,0-512,0]#	112,6 [57,0-125,0]€	0.020

CK: Creatine Kinase; ANA: Antinuclear antibodies; RF: Rheumatoid Factor; EMG: Electromyogram. \*\*Normal CK value<170 U/L, normal lymphocytes level<1.0 G/L, normal Hb level<12g/dL, normal neutrophils level<1.0 G/L, normal platelets<100 G/L. \*Normal C4 value>=0.15g/L. \*Data available for 16 patients. \*Data available for 15 patients. \*Data available for 425 patients.

**Conclusion.** pSS with anti-Sm and/or anti-RNP antibodies subjects seem to display a more active systemic disease, with a more frequent pulmonary and muscular involvement.

#### P-55

#### Autoantibodies to Ro/SSA, La/SSB, Sm, or Sm-RNP not detectable in the serum are often present in the saliva

Kristi A. Koelsch, PhD<sup>1,2,3</sup>, Joshua Cavett, MS<sup>1,2,3</sup>, Kenneth Smith, PhD<sup>2</sup>, Jacen S. Moore, PhD<sup>1,2,3,\*</sup>, Astrid Rasmussen, MD, PhD<sup>2</sup>, C. Erick Kaufman, MD<sup>4</sup>, David M. Lewis, DDS<sup>5</sup>, Lida Radfar, DDS<sup>6</sup>, Christopher J. Lessard, PhD<sup>1,2</sup>, Biji T. Kurien, PhD<sup>1,2,3</sup>, Judith A. James, MD, PhD<sup>1,2</sup>, Kathy L. Sivils, PhD<sup>1,2</sup>, A. Darise Farris, PhD<sup>1,2</sup>, R. Hal Scofield, MD<sup>1,2,3</sup>

<sup>1</sup>University of Oklahoma Health Sciences Center, Oklahoma City Oklahoma; <sup>2</sup>Arthritis and Clinical Immunology Program, Oklahoma Medical Research Foundation, Oklahoma City, Oklahoma; <sup>3</sup>Department of Veterans Affairs Medical Center, Oklahoma City, OK; <sup>4</sup>Department of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City, OK;; <sup>5</sup>Department of Oral and Maxillofacial Pathology, University of Oklahoma College of Dentistry, Oklahoma City, OK; <sup>6</sup>Oral Diagnosis and Radiology Department, University of Oklahoma College of Dentistry, Oklahoma City, OK \*Present affiliation: Department of Clinical Laboratory Sciences, University of Texas, El Paso. TX

#### 14<sup>th</sup> International Symposium on Sjögren's Syndrome

**Background.** The involvement of B cells in the pathogenesis of Sjögren's syndrome (SS) is evidenced by the presence of autoantigen-specific B cells and the incidence of autoantibodies to Ro/SSA (Ro52 and Ro60), and La/SSB. Production of autoantibodies occurs within the salivary glands, but it is unknown if autoantibodies present in the serum originate from these cells. We hypothesize that if serum autoantibodies originate from the antibody secreting cells in the glands, they would likely be present in the saliva prior to their detectible levels in the plasma. To explore this, we tested the saliva autoantibody profiles in a group of SS patients and sicca controls and compared them to their respective IgG serum autoantibody profiles.

**Methods.** Following written informed consent 27 sequential participants all symptomatic for dry yes and mouth, were evaluated for American/European Consensus Group (AECG) primary SS inclusion/exclusion criteria in the OMRF Sjögren's Research Clinic (OSRC). Serum, stimulated parotid saliva samples and data were obtained from all subjects. 14 subjects met the American/European Consensus Group (AECG) primary SS inclusion criteria, and 1 also met the American College of Rheumatology criteria for SLE. 13 subjects did not meet the SS classification criteria and served as sicca controls. Direct enzyme-linked immunosorbent assays (ELISAs) were performed by applying saliva (1:20 dilution, in duplicate) to antigencoated (Ro/SSA, La/SSB, Sm, and Sm-RNP; ImmunoVision, Inc.) plates and detected using anti-human IgA-, or IgG-alkaline phosphatase and substrate. Four control subjects negative for all SS inclusion criteria and all other measures were used to establish positive thresholds for each ELISA (mean+5SD). To confirm stringency of this measure, we determined that the Q3+1.5\*IQR threshold was similar. We then compared the positive specificities for the saliva ELISAs to the IgG specificities (Ro/SSA, La/SSB, Sm, and Sm-RNP) measured in the serum by Ouchterlony double immunodiffusion, INNO-LIA, and Bioplex 2200.

Classification	IgG Serum				Calico Co	ocificitio				
& ID	Specificities/ANA	Saliva Specificities								
	Status									
		Ro,	SSA	La/	SSB	l s	Sm		Sm/RNP	
		lgG	IgA	lgG	lgA	lgG	lgA	lgG	IgA	
Control 1	ANA pos	NEG	NEG	NEG	NEG	NEG	NEG	NEG	NEG	
Control 2	ANA pos	POS	POS	POS	POS	NEG	POS	POS	POS	
Control 3	ANA pos	NEG	POS	POS	NEG	NEG	NEG	NEG	NEG	
Control 4	ANA neg	NEG	NEG	POS	NEG	NEG	NEG	NEG	NEG	
Control 5	ANA pos	NEG	POS	NEG	POS	NEG	NEG	NEG	NEG	
Control 6	ANA pos	NEG	POS	POS	NEG	NEG	POS	POS	POS	
Control 7	ANA pos	NEG	NEG	NEG	NEG	NEG	NEG	NEG	NEG	
Control 8	Ro, ANA neg	NEG	POS	NEG	NEG	NEG	NEG	NEG	NEG	
Control 9	Ro, ANA Pos	POS	POS	NEG	NEG	NEG	NEG	NEG	NEG	
Control 10	ANA pos	NEG	POS	NEG	NEG	NEG	NEG	NEG	NEG	
Control 11	ANA neg	POS	POS	NEG	POS	NEG	POS	POS	POS	
Control 12	ANA pos	NEG	POS	NEG	NEG	NEG	NEG	NEG	NEG	
Control 13	ANA pos	POS	POS	NEG	NEG	NEG	NEG	NEG	NEG	
	Ro, La, SmB, nRNP,									
pSS/SLE-1	RNP-A, ANA Pos	POS	NEG	POS	NEG	NEG	NEG	POS	POS	
pSS-2	Ro, ANA Pos	POS	POS	NEG	NEG	NEG	NEG	NEG	NEG	
pSS-3	Ro, ANA Pos	POS	POS	POS	POS	POS	POS	NEG	NEG	
pSS-4	Ro, La, ANA Pos	NEG	POS	NEG	NEG	NEG	NEG	NEG	NEG	
pSS-5	La, ANA Pos	POS	POS	POS	POS	NEG	POS	POS	POS	
pSS-6	La, ANA Pos	POS	POS	POS	POS	NEG	POS	NEG	NEG	
pSS-7	ANA pos	NEG	NEG	NEG	POS	NEG	NEG	POS	POS	
pSS-8	Ro, ANA Pos	POS	POS	NEG	NEG	NEG	NEG	NEG	NEG	
pSS-9	Ro, La, ANA Pos	POS	POS	NEG	POS	NEG	NEG	NEG	NEG	
pSS-10	ANA pos	NEG	NEG	NEG	NEG	NEG	NEG	NEG	NEG	
pSS-11	ANA neg	NEG	POS	NEG	NEG	NEG	NEG	NEG	NEG	
pSS-12	Ro, ANA Pos	POS	POS	POS	POS	NEG	POS	POS	POS	
pSS-13	ANA neg	NEG	POS	NEG	POS	NEG	NEG	NEG	NEG	
pSS-14	ANA pos	NEG	POS	NEG	NEG	NEG	NEG	NEG	NEG	

**Results.** 10/12 sicca control sera were ANA positive, but only two were seropositive for Ro. The comparison of sera and saliva results showed that 5/14 SS patients, and 6/12 sicca controls had IgG-, and 8/14 SS patients and 8/12 sicca controls had IgA-saliva specificities not detectable in the sera.

**Conclusions.** Saliva from some SS patients and sicca controls contain autoantibodies not present in the serum. It will be important to test these saliva positive, seronegative subjects longitudinally for future development of serum autoantibodies. If this does occur, it would be likely that certain sicca controls would then fulfill the criteria for primary SS classification, suggesting that early testing of saliva for autoantibodies may be indicated for prediction of progression to systemic disease.

## P-56

## Clinical and diagnostic significance of immunoglobulin a rheumatoid factor in primary Sjögren's syndrome

Kyung-Ann Lee<sup>1</sup>, Kyoung-Woon Kim<sup>2</sup>, Bo-Mi Kim<sup>2</sup>, Ji-Yeon Won<sup>2</sup>, Han-Ah Kim<sup>3</sup>, Hee-Won Moon<sup>3</sup>, Hae-Rim Kim<sup>1</sup>, Sang-Heon Lee<sup>1</sup>

<sup>1</sup>Division of Rheumatology, Department of Internal Medicine, Konkuk University Medical Center, Konkuk University School of Medicine, Neungdong-ro 120-1, 05030, Seoul, Korea. <sup>2</sup>Convergent Research Consortium for Immunologic Disease, The Catholic University, Banpodae-ro 222, 06591, Seoul, Korea. <sup>3</sup>Department of laboratory medicine, Konkuk University Medical Center, Konkuk University School of Medicine, Neungdong-ro 120-1, 05030, Seoul, Korea.

**Background.** To investigate the diagnostic accuracy of rheumatoid factor (RF) isotype for the detection of primary Sjögren's syndrome (pSS) and evaluating clinical and serological associations of immunoglobulin (Ig)A RF in patients with pSS.

**Methods.** RF levels were measured in 85 and 38 patients with pSS and idiopathic sicca syndrome, respectively, using the ELISA and analysed with respect to clinical and laboratory disease characteristics. ROC curves were used to determine and compare the diagnostic accuracy of IgA RF with other diagnostic tests.

**Results.** Serum levels of IgA RF were significantly higher in patients with pSS than in those with idiopathic sicca syndrome. IgA RF showed a sensitivity, specificity, positive, and negative predictive value of 90.7%, 78.9%, 89.5%, and 81.1%, respectively, for pSS diagnosis. IgA RF was associated with xerostomia; abnormal Schirmer's test; severe sialoscintigraphic grade; low unstimulated salivary flow rate (USFR); antinuclear antibody and anti Ro/SSA positivity; high IgG and IgM/G RF levels; and low C3 levels in patients with pSS. IgA RF titres had positive correlations with sialoscintigraphic grade and IgG and IgG/M RF levels and had negative correlations with USFR, Schirmer's test value, and C3 levels.

**Conclusions.** Our findings confirmed the potential of IgA RF to distinguish pSS from idiopathic sicca syndrome. The presence of IgA RF in patients with pSS was associated with significantly worse exocrine function and active serologic profile. No association between IgA RF and extra-glandular manifestations was noted.





Serum 1gA, 1gG, 1gM RF levels were measured using enzyme-linked immunosorbent assay (ELISA) in patients with pSS (n=85), idiopathic sicca syndrome (n=38), and RA (n=33). Each synbol represents the RF level in a single patient's serum. RF: rheumatoid factor; pSS: primary Sjögren's syndrome; RA: rheumatoid arthritis.



Fig. 2. Correlation between IgA RF and USFR (A), IgG (B), C3 (C), IgM RF (D), IgG RF (E), and ESR (F) in patients with pSS. Each dot reoresents individual value (correlation coefficient and p value by Spearman's rank corr3elation test). RF: rheumatoid factor; USFR: unstimulated salivary flow rate; Ig immunoglobulin; C: complement; ESR: erythrocyte sedimentation rate.

Posters

## P-57

## Correlation studies between EliA<sup>™</sup> Ro/SSA antigen Well and Ro52 / Ro60 tests

Nina Olschowka, Isabel Gehring, Ekaterina Eimer, and Peter Höpfl. Phadia GmbH, Munzinger Straße 7, 79111 Freiburg.

**Background.** Sjögrens Syndrome is a chronic inflammatory autoimmune disease of unknown cause characterized by diminished lacrimal and salivary gland secretion resulting in keratoconjunctivitis sicca and xerostomia. In half of patients, the disease occurs as a primary pathologic entity (primary Sjögren's syndrome). In the other half, it occurs in association with rheumatoid arthritis or other connective tissue disorders. The disease can occur at any age, but is more frequent in older women. Many patients develop Sjögren's syndrome as a complication of another autoimmune disease, such as rheumatoid arthritis or lupus.

Detection of the Ro/SS-A antibody aids clinicians in the diagnosis of the disease. The complete Ro antigen comprises two proteins, Ro52 and Ro60. Most of the Ro positive patients show antibodies to both Ro60 and Ro52. However, there are subsets of patients who only produce antibodies to Ro52 or to Ro60.

The objective of this study was to determine the detection capability of the Ro/SSA antibodies by the EliA Ro Well compared to individual Ro 52 and Ro 60 antibody tests from another manufacturer.

Methods. Analytical runs were performed at two different laboratories. Runs for the serum panel-1 were performed at the Thermo Fisher Scientific facility in Freiburg, Germany using EliA Connective Tissue Disease assays processed on the Phadia<sup>™</sup> Laboratory Systems by following the instructions of the manufacturer.

Laboratory-2 tested the serum panel-2 using BioPlex Ro 52 and Ro 60 assays on Bio-Rad instruments by following the instructions of the manufacturer.

**Results.** 96 clinical defined patient samples with a known diagnosis of Sjögrens Syndrome were used in a correlation study between the EliA Ro Well, the BioPlex 2200 ANA Screen, and the individual EliA and Bio-Plex Ro52 and Ro60 parameter tests. The correlation study showed an overlap of 100% between the EliA Ro Well and the single parameter tests independent from the supplier (Thermo Fisher Scientific or Bio-Rad).

Combining the test results from Ro52 and Ro60 results in a sensitivity of 84.4% which is identical with the sensitivity for the EliA Ro Well.

**Conclusion.** The EliA Ro Well offers the same sensitivity as the separate Bio-Rad Ro 52 and Ro60 assays ensuring detection Ro52 and/or Ro60 antibodies with the convenience and efficiency of processing a single test versus two.

Compared with the single marker tests for Ro 52 and Ro60, the risk of underdiagnosing a patient based on the results of the EliA Ro Well is very low. From viewing my poster the participants should be able to:

- a. Understand the use of the Ro/SS-A antibodies assay as part of the diagnosis and classification criteria of Systemic Lupus Erythematosus
- b. Demonstrate the clinical utility and efficiency of testing for Ro/SSA antibodies with the EliA Ro Well and mitigating the need of testing for separate antigens
- c. Identify how this information helps the viewer inform clinicians about the laboratory test selection to help them avoid disease misclassifications.

### P-58

#### ANTI-Ro52 antibodies in suspected primary Sjögren's syndrome (PSS)

Roser Solans-Laqué<sup>1</sup>, Jose Loureiro<sup>1</sup>, Ferran Martinez<sup>1</sup>, Javier Puig<sup>2</sup>, Ana Marin<sup>3</sup>.

Internal Medicine Department<sup>1</sup>, Ophthalmology Department<sup>2</sup> and Immunology Department<sup>3</sup>. Vall d'Hebron University Hospital. Barcelona. Spain.

**Background.** anti-Ro52 antibodies have been described in patients with pSS but there are limited data on its significance.

**Objectives.** To assess the diagnostic value of anti-Ro52 antibodies in patients with suspected pSS.

**Methods.** retrospective study of all patients diagnosed at our Department between January 2006 and January 2016 with suspected SS and negative anti-Ro/SSA antibodies, in whom anti-Ro52 abs. were determined. Patients were classified as having pSS, according to the European-American Consensus Group (EACG) criteria. Anti-Ro60 and anti-Ro52 antibodies were determined by ChemiLuminescent immune assay (CLIA) and immunoblot. Statistical analysis was done using SPSS vs.20 package.

Results. 160 patients (6 men), mean age at diagnosis 60.84±11.52 y (range 22-92), were included. Subjective xerostomia and/or xerophthalmia were reported by 82% of patients, and fatigue by 54 (33.8%). Ocular tests (Schirmer's, BUT and ocular staining) were performed in all patients, and were positive in 75.5%, 59.5% and 59.4% of cases, respectively. Salivary scintigraphy showed III/IV stage in 118 (73.6%) cases. Overall, 101 (63.1%) patients had arthralgias; 27 (16.9%) arthritis; 40 (25%) Raynaud's phenomenon; 20 (12.5%) parotitis; 14 (8.8%) lung fibrosis; 8 (5%) peripheral neuropathy, and 5 (3.1%) renal involvement. Anemia was detected in 11 (6.9%) patients, lymphopenia in 15 (9.4%), hypergammaglobulinemia in 32 (20%) and hypocomplementemia in 12 (7.5%) patients. Anti-Ro52 abs. were positive in 20 (12.5%) patients. The presence of positive anti-Ro52 antibodies was significantly related to lung fibrosis (OR 4.85, 95% CI 1.44-16.38 p=0.018), peripheral neuropathy (OR 6.70, 95%CI 1.63-27.5, p=0.015), and lymphoma (OR 7.94, 95%CI 1.48-42.82, p=0.027) development, and to the presence of anemia (OR 4.71, 95%CI 1.24-13.89), hypergammaglobulinemia (OR 8.93, 95%CI 3.24-24.56, p<0.001) and hypocomplementemia (OR 3.55, 95%CI 1.18-10.67, p=0.044) at baseline. No relationship was found between the presence of anti-Ro52 antibodies and Raynaud's phenomenon, arthritis, parotitis, renal involvement and fatigue. Salivary gland biopsy (SGB) was performed in 120 (75%) patients, and was consistent with pSS in 57 (35.6%) of cases. Eleven (55%) patients with anti-Ro52 positive antibodies had a positive SGB.

**Conclusions.** In patients with suspected pSS with negative anti-Ro60/SSA antibodies, the detection of anti-Ro52 antibodies could help in diagnosis, especially identifying patients at risk of development of lung fibrosis, poluneuropathy and lymphoma. Salivary gland biopsy is positive in 55% of cases.

#### P-59

#### Clinical and laboratory features of primary Sjögren's syndrome associated with anticentromere antibodies

B. Chaltsev<sup>1</sup>, V. Vasiliev<sup>1</sup>, S. Palshina<sup>1</sup>, E. Rodionova<sup>1</sup>, T. Safonova<sup>2</sup>. <sup>1</sup>Nasonova Research Institute of Rheumatology, <sup>2</sup>Research Institute of Ophthalmic Diseases, Moscow, Russian Federation.

**Background.** The prevalence of ACA among patients with pSS varies from 2% to 27%. This subtype differs from the "classical" one in a number of laboratory and clinical manifestations and characterized by an increased risk of developing SSc and AMA+ biliary lesions.

**Objectives.** to evaluate clinical and laboratory features of ACA-positive primary pSS; to evaluate its conformity to the ACR2012 and ACR/EU-LAR2016 classification criteria; to evaluate prevalence of SSc according to ACR2013 criteria; to evaluate prevalence of MALT-lymphoma; to evaluate prevalence of AMA+ biliary lesions.

Table I. Characteristics of pSS patients with positive ACA.

Parameters	Value	e (n, %)
Sialography of parotid gland: sialectasis	61/64	(95%)
Xerostomia (grade I-III) stimulated parotid saliva flow rate	35/56	(62.5%)
<0,5ml/5min (III)	27/43	(62.8%)
KKS with OSS ≥5	36/58	(62%)
Schirmer's test ≤5mm/5 minutes	36/61	(59%)
Focal lymphocytic sialadenitis ≥1 foci/4 mm <sup>2</sup>	37/40	(92%)
aRo (≥50 IU/ml)	14/63	(22%)
aLa (≥50 IU/ml)	3/62	(4.8%)
IgM RF (>30 IU/ml)	11/63	(17%)
Low C4 (<0.1 ng/ml)	2/41	(4.8%)
High ESR (>30 mm/h Westergren)	7/36	(19.4%)
Hypergammaglobulinemia (>20%)	8/34	(23.5%)
High IgG (>16g/l)	5/37	(13.5%)
Leucopenia (<4x10 <sup>9</sup> )	2/37	(5.4%)
Neuropathy	1/13	(7.6%)
AMA (>10 IU/ml)	17/37	(45%)
AMA+biliary lesions	12/64	(18.7%)
Histologic features of PBC	0/12	
MALT-lymphoma	9/64	(14%)
MALT-lymphoma - RF+	2/9	(22%)
MALT-lymphoma - aRo+	3/9	(33%)
MALT-lymphoma - aRo-aLa-RF -	4/9	(45%)
Raynaud's phenomenon	33/60	(55%)
Digital ulcers	1/60	(1.6%)
Abnormal nailfold capillaries	22/35	(62%)
Sclerodactyly	9/50	(18.3%)
Puffy fingers	3/50	(6%)
PAH	0/47	
Interstitial lung disease	13/45	(28%)

**Results.** Clinical and laboratory manifestations are listed in the Table I. In our study, 58.7% ACA+pts with pSS didn't fulfill ACR2012 criteria, because of the lack of aRo/aLa or RF+ANA. According to the criteria 2016 also just 60-68% pts will have  $\geq$ 4 scores for diagnosing pSS. Limited form of SSc due to 2013 criteria might be revealed in 20% cases with sclerodac-tyly, puffy fingers and digital ulcers. MALT-lymphoma occurred in 14%. AMA+ biliary lesions occurred in 18% of cases, all of which had no histological features of PBC.

**Conclusions.** ACA-positivity in pSS is associated with specific clinical and laboratory characteristics, such as combination with SSc and AMA+ biliary lesions, but only two-thirds of patients with ACA and definite pSS meet classification criteria 2012, 2016. This fact raises the question about necessity to consider ACA one of the pathogenically relevant autoantibodies for pSS.

#### P-60

Clinicopathological characteristics of anti-centromere antibody-positive Sjögren's syndrome in the presence or absence of systemic sclerosis

Yasunori Suzuki, Hiroshi Fujii, Ichiro Mizushima, Kazunori Yamada, Mitsuhiro Kawano.

Division of Rheumatology, Department of Internal Medicine, Kanazawa University Graduate School of Medicine, Kanazawa, Japan.

**Objectives.** Few studies have clarified differences in sicca symptoms, ESS-DAI score, and organ involvement in anti-centromere antibody (ACA)-positive Sjögren's syndrome (SS) with and without systemic sclerosis (SSc). We compared clinicopathological characteristics between these two groups using our cohort of patients with ACA-positive SS.

**Methods.** We studied 33 patients with ACA+ primary SS and 16 with ACA+ SS with SSc in a retrospective cohort study. All SS patients met Japanese and/or ACR criteria, and those whose results exceeded the focus score by 1 underwent labial salivary gland biopsy. All SSc patients met ACR/EULAR criteria. We analyzed SS and SSc data at diagnosis and organ involvement during follow-up.

**Results.** No significant differences were seen in age at diagnosis (mean 65.2 in ACA+ primary SS vs 66.9 in ACA+ SS with SSc), sex (97.0% vs 100%), anti-SS-A/Ro antibody (42.4% vs 25.0%), anti-SS-B/La antibody (16.7% vs 0%), laboratory data including leukocytes, lymphocytes, serum IgG, and complement levels, ESSDAI score at diagnosis (1.91±2.47 vs 1.81±4.23, respectively), organ involvement, and treatment. Focus score was 3.34±2.98 in ACA+ primary SS and 3.57±2.04 in ACA+ SS with SSc (p=0.055). Germinal center-like structures were 24.2% vs 18.8%; scores for Saxon's test, 0.59 g vs 0.54 g (p=0.57); Schirmer's test, 6.4 mm vs 5.4 mm (p=0.56); and Raynaud's phenomenon, 21.2% vs 87.5% (p<0.001), respectively. No skin sclrerosis developed during the follow-up period.

**Conclusion.** ACA+ SS with and without SSc showed severe sicca symptoms. Focus score was higher in the SS+ SSc group than in the ACA+ primary SS group.

### **P-61**

#### Anti-centromere antibody positive Sjögren's syndrome is associated with worse sicca symptoms than primary Sjögren's syndrome alone

#### Yifan Li, Arthur A. M. Bookman

Multidisciplinary Sjogren's Clinic. University Health Network, University of Toronto

**Background.** Sicca symptoms are the most prevalent symptoms in primary Sjögren's Syndrome (pSS). The presence of anti-centromere antibody has been reported in 1.4 to 10.85%1 of patients with pSS. The purpose of our study was to determine whether the presence of anti- centromere antibodies (ACA) affects the incidence or severity of sicca symptoms in patients diagnosed with pSS.

**Methods.** Patients were pre-screened for objective evidence of dry eyes or dry mouth, abnormal serology or history of salivary gland enlargement as an adult. The evaluation included a visual analogue score (VAS) for severity of xerophthalmia and xerostomia, Schirmer's-1 test (S1T), van Bijsterveld staining score and unstimulated whole salivary flow (USSF). A

minor salivary gland biopsy was performed on all patients. Assessments were performed by the same pathologist on protocol for evidence of Focal Lymphocytic Sialadenitis pathology and focus score. Serological profile included the evaluation for the presence of anti-Ro, anti-La and anti- centromere antibody (ACA). All subjects were evaluated for CREST features. pSS was classified according the American European Consensus Group (AECG) Criteria. Patients that met the pSS classification criteria were further categorized into ACA+ SS and ACA- SS. A 2- tailed student t-test with heterogeneous variance was used to compare the two groups for measures of severity of dry eyes and dry mouth.

**Results.** Within the pSS group (n=446), there were 26 patients with positive ACA serology. On a 10 cm visual analog scale (VAS), the subjective severity of ocular sicca symptoms was 7.0 (out of 10) in ACA+ SS patients and 6.4 in ACA- SS patients (p=0.197). The mean S1T in ACA+ SS patients was 3.5mm/5 mins and in ACA- SS pts was 4.1mm/5 mins. The difference was significant (p=0.038). The severity of oral sicca symptoms was 8.5 in ACA+ SS patients and 6.7 in ACA- SS patients (p<0.001). The mean value for USSF was 0.1ml/15 mins for ACA+ SS and 0.4 mL/15 mins for ACA-SS. This difference was highly significant (p<0.001). RP occurred more commonly in ACA+ SS patients (88%) than in ACA- SS patients (28%) (p<0.001). Only 35% of ACA+ SS patients (p<0.001).

### Table I. Comparison between ACA+ SS and ACA- SS.

	ACA+ SS	ACA- SS	p-value
	(n=26)	(n=429)	
Demographics			
Age, mean (years)	55.7±10	).5 53.2±13.4	Ļ
Sex Male	0	39	
Female	26	381	
Clinical Differences			
Prevalence of Raynaud's phenomenon (%	) 88	28	<i>p</i> <0.001
Serological Markers			
Prevalence of positive Ro/La antibodies (	%) 35	77	<i>p</i> <0.001
Prevalence of elevated serum IgG (%\$)	24	57	<i>p</i> <0.001
Average levels of serum IgG (gm/L)	12.4	19.4	<i>p</i> <0.001
Xerophthalmia			
Prevalence of xerophthalmia (%)	96	96	NS
Severity of xerophthalmia (on VAS, max	10) 7±2.4	6.4±2.6	NS
Average Rose Bengal score	5.7±2.	2 5.6±2.3	NS
Schirmer's-1 test (mm/5 mins)	3.2±1.	8 4.2±4.4	<i>p</i> <0.05
Duration (years)	5.4 (0-2	20) 7.5 (0–50	) NS
Xerostomia			
Prevalence of xerostomia (%)	100	98	NS
Severity of xerostomia (on VAS, max 10)	8.5±1.	4 6.7±2.4	p<0.001
USSF (mL/15 min)	0.1	0.4	p<0.001
Duration (years)	5.8 (0-2	22) 6.8 (0-45	) NS
Salivary gland biopsy			
Focus score ≥1 (%)	92	84	p<0.001
Average focus score	5.5±4.	3 4.0±3.3	NS
Average fibrosis score (out of 3)	1.0±0.8	32 1.1±0.68	NS

NS: not significant.

**Conclusions.** ACA+ SS is associated with more severe subjective xerophthalmia and more severe subjective and objective xerostomia compared to ACA- SS. Furthermore, the majority of ACA+ SS patients meet the AECG criteria for pSS despite having negative serology for anti-Ro/SSA or anti-La/SSB antibodies.

#### Reference

 BALDINI C, MOSCA M, DELLA ROSSA A et al.: Overlap of ACA-positive systemic sclerosis and Sjögren's syndrome: a distinct clinical entity with mild organ involvement but at high risk of lymphoma. Clin Exp Rheumatol 2013; 31(2): 272-80.

## **P-62**

#### Clinical and immunological features of anti-centromere antibody-positive primary Sjögren's syndrome

#### Masako Tsukamoto<sup>1,2</sup>, Katsuya Suzuki<sup>2</sup>, Tsutomu Takeuchi<sup>1</sup>.

<sup>1</sup>Division of Rheumatology, Department of Internal Medicine, Keio University School of Medicine, Tokyo, Japan. <sup>2</sup>Department of Rheumatology, National Tokyo Medical Center, Tokyo, Japan.

**Background.** Anti-centromere antibody (ACA)-positive Sjögren's syndrome (SS) is considered as a subtype in SS. Recent international collaborative large scale cohort study highlighted several clinical features such as Raynaud's phenomenon, sclerodactyly and extra glandular dysfunction (Arthritis Care Res. 2016). Assessment of ACA is potentially valuable for definitive diagnosis of this subtype and medical management in a certain number of patients uncovered by current 2016 ACR/EULAR classification criteria. However, enough information of clinical and immunological features of ACA positive SS has not been accumulated and clinical significance of ACA in SS may not be fully established. The aim of this study is to clarify clinical and immunological features of ACA positive SS.

**Methods.** All patients with primary SS who visited to our Division of Rheumatology at Keio University Hospital in Tokyo between May 1995 and July 2017 were enrolled. Clinical information and immunological tests including immunoglobulins (Ig) and serum autoantibodies were collected and statistically analyzed.

Results. Six hundred and one patients were clinically classified as primary SS (female: 94%, mean age: 56±15). They were divided into 4 groups by serum ACA and anti-SS-A antibody status. Only discrete-speckled pattern in anti-nuclear antibodies (ANA) test and/or anti-centromere antibodies positive (ACA alone) were detected in 33 patients (5.5%), while only anti-SS-A antibodies with no ACA (SS-A alone) were detected in 465 patients (77.4%). Number of patients with both ACA and anti-SS-A antibody (Double positive) was 29 (4.8%), while 74 patients had neither ANA nor anti-SS-A antibody (Seronegative). Then we statistically compared these 4 groups. The proportion of dryness was no difference among 4 groups. The proportions of Raynaud's phenomenon or sclerodactyly were higher in ACA alone and Double positive groups (p<0.01 or p<0.01). The extraglandular involvements of SS were significantly less in the ACA alone group than in the SSA alone and Double positive groups (p<0.01). The proportions of increase of serum IgG or IgA were 10% or 6% in ACA alone group, 61% or 20% in SS-A alone group, 50% or 29% in Double positive group and 20% or 4% in Seronegative group (p<0.01 or p<0.01). Existence of anti-SS-A antibody, not ACA associated to high concentration of IgG or IgA, while there was no difference between 4 groups as IgM (p=0.49). Regarding the proportion of low C3, C4 or CH50, there were no differences among 4 groups. Remarkably, the proportion of leukocytopenia in ACA alone group was significantly lower than the others (p<0.01). As compared with major organ involvements, such as pulmonary, cardiac or articular involvements, no differences were found among 4 groups.

**Conclusions.** Our large-scale study identified distinct characteristics of ACA-positive SS patients different from anti-SS-A antibody-positive or seronegative SS patients in Japanese population.

#### P-63

#### The breakthrough of the pathogenesis of anti-centromere antibody positive Sjögren's syndrome by analyzing MicroRNA expression from the minor salivary gland

Shinichiro Tsunoda<sup>1,2</sup>, Takahiro Yoshikawa<sup>1</sup>, Nozomu Moriya<sup>3</sup>, Yuichi Yokoyama<sup>1</sup>, Masahiro Sekiguchi<sup>1</sup>, Naoaki Hashimoto<sup>1</sup>, Kiyoshi Matsui<sup>1</sup>, Hajime Sano<sup>1</sup>.

<sup>1</sup>Division of Rheumatology, Department of Internal Medicine, Hyogo College of Medicine, Nishinomiya, Japan. <sup>2</sup>Division of Immunology & Rheumatology, Department of Internal Medicine, Sumitomo Hospital, Osaka, Japan. <sup>3</sup>Japan Poison Information Center, Tsukuba, Japan.

**Background.** Anti-centromere antibody (ACA) is one of the specific autoantibodies of systemic sclerosis. Recently it is also recognized in patients with Sjögren's syndrome (SS) who differ from classical SS patients with anti-Ro/SS-A and anti-La/SS-B antibodies in several clinical and laboratory parameter. MicroRNAs (miRNAs) are small conserved non-coding RNA molecules that post-transcriptionally regulate gene expression by targeting the 3' untranslated region of specific messenger RNAs for degradation or translational repression. MiRNAs play an important role in innate immunity and acquired immunity. We investigated miRNAs from the minor salivary gland (MSG)to elucidate the pathogenesis of SS due to different autoantibodies between anti- centromere antibody, and anti-Ro/SSA antibody and anti-La/SSB antibody.

**Method.** We performed lip minor salivary gland biopsy in 12 female patients with primary SS: 7 patients have ACA and 5 patients have anti-SS-A/SS-B antibody. We extracted miRNA using RNeasy Mini Kit (QIAGEN) from MSG and did the comprehensive analysis of 2565 types of miRNA expression using miRNA oligo chip (3D-Gene<sup>TM</sup> from Toray Industries). Then, we compared the expression of miRNAs with the pathological classification by the extent of cell infiltration in the lip salivary glands (Greespan classification) between ACA positive patients and anti-SS-A/SS-B antibody positive patients. Next we validated by real time PCR of these miRNAs. Finally, we used Ingenuity Pathway Analysis software to construct a molecular interaction network.

Results. We classified patients based on types of autoantibody and Greenspan's(GS) pathological classification, then we defined them as follows; Group I: ACA positive and GS grade 1 or grade 2, Group II: ACA positive and GS grade 3 or grade 4, and Group III: Anti-SS-A/SS-B antibody positive and GS grade 3 or grade 4. Group I had 4 patients and Group II had 3 patients, and Group III had 5 patients. Comparing Group II and Group III (Group III/Group II ratio) as the same degree of tissue injury, the upregulated expression of miRNA were hsa-miR-155-5p, hsa-miR150-5p, hsa-miR- 146a-5p and hsa-miR-142-5p and the downregulated expression of miRNAs were has- miR-133b, hsa-miR-1-3p, hsa-miR-203a-3p, hasmiR-144-3p and hsa-miR-744-3p. Comparing Group I + II and Group III (Group III/Group I + II ratio) as the differences in the autoantibodies, the upregulated expression of miRNA were hsa-miR-155-5p, hsa-miR-150-5p, hsa-miR-146a-5p, hsa-miR-142-5p and hsa-miR-142-3p, and the downregulated expression of miRNAs were hsa-miR-133a-3p, hsa-miR-1-3p, hsamiR-133b, hsa-miR-203a-3p, and hsa-miR-144-3p. By the validation of these miRNAs, hsa- miR-744-3p and hsa-miR-144-3p in MSG from ACA positive patients were downregulated significantly than that from anti-SS-A/SS-B antibody positive patients.

From the Ingenuity Pathway Analysis, IL-6 and TNF were associated with the different pathogenesis of SS.

**Conclusions.** From the analysis of miRNA from MSG having different autoantibodies, the different expression of microRNA could relate the regulation of inflammatory cytokines.

#### **P-64**

#### P2X7 and P2Y<sub>2</sub> receptors as the rapeutic targets in Sjögren's syndrome mouse models

Adam L. Martin, Mahmoud G. Khalafalla, Kimberly J. Jasmer-McDonald, Lucas T. Woods, Jean M. Camden, Laurie Erb and Gary A. Weisman. Department of Biochemistry and Christopher S. Bond Life Sciences Center of the University of Missouri, Columbia, MO, USA 65211-7310.

Background. Salivary gland dysfunction and other disorders related to hyposalivation affect the quality of life for millions of patients who are severely impacted by dry mouth, oral infections and poor nutrition. Loss of saliva secretion is symptomatic of Sjögren's syndrome (SS), an autoimmune exocrinopathy associated with lymphocytic infiltration of the salivary gland, autoantibody production and tissue degeneration. Clinical trials for drugs targeting P2 receptors for extracellular nucleotides have been carried out for the treatment of human diseases, including rheumatoid arthritis, cystic fibrosis and Crohn's disease. Previously, we demonstrated that activation of the extracellular adenosine 5'-triphosphate (ATP)- gated ionotropic P2X7 receptor (P2X7R) in salivary epithelium enhances salivary gland inflammation by promoting cell apoptosis, reactive oxygen species production and cytokine release. We have also shown that upregulation and activation of the G protein-coupled P2Y<sub>2</sub> receptor (P2Y<sub>2</sub>R) for extracellular ATP and uridine 5'-triphosphate (UTP) during salivary gland inflammation contributes to proliferation and migration of immune cells through transactivation of growth factor receptors and integrins, metalloprotease-mediated growth factor release and cytokine production. Thus, we are investigating the effects of selective antagonists or deletion of the P2X7R or the P2Y2R in mouse models of SS.

**Results.** Selective antagonism of the P2X7R with A-438079 or the P2Y<sub>2</sub>R with AR-C118925 reduces inflammation and improves carbachol-induced

saliva flow in SS-like mice. Knockout of the  $P2Y_2R$  in the IL-14 $\alpha$  mouse model of SS significantly reduced immune cell infiltration in salivary glands. **Conclusions.** These data suggest that targeting the P2X7 and P2Y<sub>2</sub> receptors is an effective therapeutic strategy to limit inflammation associated with salivary gland disorders.

This study was supported by National Institutes of Health (NIH) R01 grants DE007389 and DE0223342 from the National Institute of Dental & Craniofacial Research (NID-CR).

#### P-65

#### TUDCA reduces inflammatory markers in salivary gland cells decreasing MUC1 accumulation. A potential therapeutic agent to ameliorate inflammation in Sjögren's syndrome

González S<sup>1</sup>, Albornoz N<sup>2</sup>, Barrera MJ<sup>2</sup>, Aguilera S<sup>3</sup>, Castro I<sup>2</sup>, Urzúa U<sup>2</sup>, Biunno I<sup>4</sup>, Petruscu S<sup>5</sup>, and González MJ<sup>2</sup>.

<sup>1</sup>Escuela Dental, Facultad de Ciencias, Universidad Mayor. <sup>2</sup>Instituto de Ciencias Biomédicas, Universidad de Chile, Santiago, Chile. <sup>3</sup>Clínica INDISA, Santiago, Chile. <sup>4</sup>Institute of Genetic and Biomedical Research (IRGB) of the National Research Council, Milano, Italy. <sup>5</sup>Institute of Biochemistry of Romanian Academy, Bucharest, Romania

**Background.** Salivary glands (SG) of Sjögren's syndrome patients produce high levels of pro-inflammatory cytokines and show evidence of endoplasmic reticulum (ER) stress, such as ER cistern dilation, mucin (MUC1) accumulation and increased levels of ER-associated protein degradation (ERAD) machinery components (1). Tauroursodeoxycholic acid (TUDCA) is a chemical chaperone that has demonstrated cytoprotective, membranestabilizing and anti-apoptotic properties in several neurodegenerative and autoimmune diseases. TUDCA alleviates ER stress and decreases the expression of proteins implicated in inflammation (2). In this study, we determined the effect of TUDCA on MUC1 expression and secretion, expression of inflammatory markers and levels of ERAD components in acinar epithelial cells stimulated with pro-inflammatory cytokines.

**Methods.** Differentiated human salivary gland (HSG) cells were incubated with 10 ng/mL of TNF- $\alpha$  or IFN- $\gamma$  for 6 hours, and then co-incubated with TUDCA up to 24 hours. The mRNA and protein levels of SEL1L and EDEM (ERAD markers) were determined by semi-quantitative reverse transcriptase (RT)-qPCR and western blot analysis, respectively. The MUC1 mRNA and protein levels were evaluated in HSG cell cultures treated with cytokines and/or TUDCA, and/or stimulated with 10 mM carbachol for 45 minutes. The subcellular localization of SEL1L, EDEM1, MUC1 and RelA/ p65 was determined by confocal immunofluorescence. Additionally, mRNA levels of IL-1 $\beta$ , IL-6, and TNF- $\alpha$  were determined by (RT)-qPCR.

**Results.** The protein and mRNA levels of SEL1L and EDEM1 were increased in HSG cells stimulated with IFN- $\gamma$  or TNF- $\alpha$ , while the co-incubation with TUDCA inhibited this effect. IFN- or TNF- $\alpha$  stimulation increased cellular levels of MUC1 protein and co-incubation with TUDCA decreased this effect in presence or absence of carbachol. An increase of mRNA levels of MUC1 after cytokine stimulation was also observed, and this effect was inhibited by TUDCA. Similar results were obtained for RelA/p65, IL-1 $\beta$ , IL-6, and TNF- $\alpha$  mRNA levels. TUDCA also decreased RelA/p65 nuclear translocation induced by pro-inflammatory cytokines.

**Conclusions.** Our data indicates that TUDCA inhibited the action of proinflammatory cytokines in HSG cells through down-regulating protein levels of the ERAD machinery components SEL1L and EDEM1. Additionally, TUDCA reversed the effects of TNF- $\alpha$  and IFN- $\gamma$  over MUC1 mRNA and protein levels. Interestingly, TUDCA reduced NF $\kappa$ B nuclear translocation and pro-inflammatory markers expression in HSG cells treated by TNF- $\alpha$ and IFN- $\gamma$ , suggesting inactivation of the NF $\kappa$ B pathway. Based on these findings we propose that TUDCA has an anti-inflammatory effect and alleviates the ER stress of SG from Sjögren's syndrome patients.

Funding. Fondecyt 1160015 and Fondecyt Postdoctoral Grant 3170023. References

- BARRERA MJ et al.: Pro-inflammatory cytokines enhance ERAD and ATF6α pathway activity in salivary glands of Sjögren's syndrome patients. J Autoimmun 2016; 75: 68-8.
- YANGUAS-CASAS N et al.: TGFβ Contributes to the Anti-inflammatory Effects of Tauroursodeoxycholic Acid on an Animal Model of Acute Neuroinflammation. *Mol Neurobiol* 2016; 54: 6737-6749.

#### **P-66**

## Discovery of drug candidates for primary Sjögren's syndrome that target a BAFF receptor

Keiko Yoshimoto<sup>1,2</sup>, Katsuya Suzuki<sup>1</sup>, Noriyasu Seki<sup>3</sup>, Kunio Sugahara<sup>3</sup>, Kenji Chiba<sup>3</sup>, Tsutomu Takeuchi<sup>1</sup>

<sup>1</sup> Div. of Rheumatology, Dept. of Internal Medicine, Keio University School of Medicine, Tokyo, Japan. <sup>2</sup>Clinical and Translational Research Center, Keio University Hospital, Tokyo, Japan. <sup>3</sup>Research Unit/Immunology & Inflammation, Mitsubishi Tanabe Pharma Corporation.

**Background and Purpose.** We have reported that peripheral monocytes of patients with primary Sjögren's syndrome (pSS) produce higher amount of IL-6 upon stimulation with BAFF *in vitro* as compared to healthy controls. We have also found that the expression level of a BAFF receptor (BR3) is elevated in pSS monocytes and that the level is significantly and positively correlated with serum IgG level of the patients. These data collectively suggest that the elevated expression of BR3 on monocytes is involved in the pathogenesis of pSS and that BR3 is a possible therapeutic target to treat pSS. We have successfully discovered two pyrrolopyrimidine derivatives, BIK-12 and BIK-13, as BR3 antagonists using our original high-throughput screening system. In this study, we investigated the effects of these compounds *in vitro* and *in vivo* to explore the possibility if the compounds are candidates of drugs to treat pSS.

**Methods.** Peripheral monocytes and B cells were co-cultured and the amount of IgG in the culture supernatant was measured. The mixed cells were stimulated with soluble BAFF (sBAFF) in the presence or absence of BIK-12 or BIK-13 to investigate if the compounds could antagonize the stimulation. The effects of the compounds on differentiation of B cells were examined by stimulating PBMC with B cell stimuli in the presence or absence of the compounds, followed by analysis of the expression levels of CD19/CD38/IgD/CD138 and activation-induced cytidine deaminase by FACS and quantitative RT-PCR, respectively. In order to show the effects of BIK-12 and BIK-13 in vivo, these compounds were administered i.p. three times a week to MRL/Ipr and NZBWF1 mice. At the end of the treatment, the mice were sacrificed by euthanasia, and immunohistochemical analysis of lacrimal and salivary glands was carried out.

**Results.** IgG production by B cells in a co-culture with monocytes was significantly suppressed by BIK12 and BIK13 in a dose dependent manner when the cells were stimulated with sBAFF. These data, together with the results that the compounds suppressed sBAFF-induced IL-6 production by peripheral monocytes, suggest that the compounds suppress IgG production through inhibiting BAFF-binding to BR3 on not only B cells, but also monocytes. Notably, differentiation of activated B cells into plasmablasts and/or plasma cells was inhibited by these compounds in a dose dependent manner. The analysis of *in vivo* effects of the compounds revealed that the compounds suppressed the increase in an anti-dsDNA antibody in both MRL/Ipr and NZBWF1 mice after 16 weeks' of treatment. In addition Immunohistocemivcal analysis indicated that the compounds suppressed in-filtration of lymphocytes to lachrymal and salivary glands of these mice.

**Conclusions.** Our results collectively suggest that BIK-12 and BIK-13 inhibit activation of not monocytes but also B cells and cross talk between B cells and at least monocytes plays a role in pathogenesis of pSS and that BR3 is involved in the interaction. In addition, BR3 is a possible therapeutic target to treat pSS. Moreover, our findings strongly suggest that BIK12 and BIK13 are drug candidates for hyper-activated B cell-related autoimmune diseases, such as pSS.

#### **P-67**

## Sjögren's syndrome is associated with reduced sex hormone exposure: a cross-sectional study

Sara S. McCoy<sup>1</sup>, Emmanuel Sampene<sup>2</sup>, Alan N. Baer<sup>3</sup>

<sup>1</sup>Department of Medicine University of Wisconsin School of Medicine and Public Health, Madison, WI; <sup>2</sup> Department of Biostatistics, University of Wisconsin School of Medicine and Public Health Madison, Madison, Wisconsin. <sup>3</sup>Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

**Background:** Primary Sjögren's Syndrome (SS) is an autoimmune disease with female predominance and perimenopausal onset, indicating a potential role for sex hormones in its pathogenesis. The goal of this study was to evaluate whether cumulative sex hormone exposure impacts the risk of development of SS.

Methods. This is a cross-sectional study of women from the Sjögren's International Collaborative Clinical Alliance (SICCA) registry. 1320 cases of SS satisfied the ACR/EULAR 2016 criteria. 1520 sicca controls were defined as women with sicca signs who did not meet ACR/EULAR criteria for SS, lacked anti-SSA antibody, and had normal minor salivary gland biopsy. Composite estrogen score (CES) was calculated by point assignment for early menarche (≤10 years), high parity, hysterectomy, use of hormone therapy, and late menopause (≥53 years). Cumulative menstrual cycling (CMC) years for premenopausal subjects was calculated as the age of the subject minus date of onset of sicca symptoms minus time pregnant. CMC years for postmenopausal women was calculated as age of menopause or date of onset of sicca symptoms (whichever was first) minus age of menarche minus time pregnant. Covariates included age, referral source, race, education level, employment status, smoking status, and recruitment site. Multivariable logistic regression was used for outcomes against the predictors of interest and all results are interpreted in terms of odds ratios.

Table I. Odds ratios for association of sex	k hormone exposure and Sjögren's syndrome
---	---

		OR	95% CI	р
CES	0	ref		
	1	0,85	0.70-1.02	0.08
	2	0.78	0.60-1.01	0.06
	3	0.56	0.33-0.96	0.03
	4	1.79	0.11-29.78	0.68
CMC (yrs) premenopausal	≤13	ref		
	>13-≤16	0.77	0.45-1.31	0.33
	>16-≤18	0.79	0.46-1.38	0.41
	>18	0.54	0.35-0.85	0.01
CMC (yrs) postmenopausal	≤13	ref		
	>13-≤16	0.6	0.25-1.43	0.25
	>16-≤18	0.6	0.23-1.58	0.3
	>18	0.61	0.37-0.99	0.047

OR: odds ratio; CAS: composite estrogen score; CMC: cumulative menstrual cycles.

**Results.** SS subjects had a statistically significant reduced risk of cumulative sex hormone exposure, when compared to controls, as measured by CES of 1-3 (Table). This risk reduction increased progressively from the CES1 to CES3 strata. This association was not significant for the highest stratum of CES, but the number of subjects in this stratum was small, leading to wide confidence intervals. This finding was corroborated by the analysis of CMC. At the highest level of premenopausal CMC (>18 years), there was a 46% reduced risk of cumulative sex hormone exposure among premenopausal SS subjects relative to the control group, after adjusting for other covariates. This reduction was statistically significant (p=0.01). However, this finding was not observed at lower CMC levels. Similarly, among postmenopausal SS subjects, the risk was also significantly reduced at CMC>18 years (p=0.047)

**Conclusions.** Women with SS have lower estrogen exposure and cumulative menstrual cycling compared to a non-autoimmune sicca control group. As estrogen exposure and cumulative menstrual cycling increased, there was a trend toward decreased risk of SS. Further longitudinal studies of sex hormone exposure in SS are needed to confirm these findings.

#### **P-68**

#### Sex hormone exposure may influence clinical characteristics of Sjögren's syndrome: a cross-sectional study

Sara S. McCoy<sup>1</sup>, Emmanuel Sampene<sup>2</sup>, Alan N. Baer<sup>3</sup>

<sup>1</sup>Department of Medicine, University of Wisconsin School of Medicine and Public Health, Madison, WI; <sup>2</sup> Department of Biostatistics, University of Wisconsin School of Medicine and Public Health Madison, Madison, Wisconsin. <sup>3</sup>Department of Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

**Background.** Primary Sjögren's Syndrome (SS) is an autoimmune disease with female predominance and perimenopausal onset, suggesting a role for sex hormones in its pathogenesis. The goal of this study was to evaluate whether sex hormone exposure may impact disease phenotype of women with SS.

**Methods.** This is a cross-sectional study of women from the Sjögren's International Collaborative Clinical Alliance (SICCA) registry. 1320 women with SS satisfied the ACR/EULAR 2016 criteria. Composite estrogen score (0-5) was calculated by points for early menarche ( $\leq$ 10 years), high parity, hysterectomy, use of hormone therapy, and late menopause ( $\geq$ 53 years). Cu-

mulative menstrual cycling (CMC) for premenopausal subjects was calculated as the age of the subject minus date of onset of sicca symptoms minus time pregnant. CMC for postmenopausal women was calculated as age of menopause or date of onset of sicca symptoms (whichever was first) minus age of menarche minus time pregnant. CES and CMC were compared in SS subjects with and without the following phenotypic features: abnormal ocular staining score (OSS), unstimulated whole salivary flow (UWS), arthritis (joint stiffness>1 hour, joint pain or swelling, or synovitis on examination), rheumatoid factor (RF) positive, anti-SSA antibody positive, focus score  $\geq 1$ , and germinal center (GC) presence. Covariates included age, referral source, race, education level, employment status, smoking status, and recruitment site. Multivariable logistic regression was used for outcomes against the predictors of interest and all results are interpreted in terms of odds ratios.

**Results:** Statistical analyses are reported for CES levels of 1-3, given the paucity of subjects with CES levels of 4 or higher (Table). Among SS subjects with CES levels of 3 compared to those with CES levels of 0, there was a statistically significant reduced risk of abnormal OSS, rheumatoid factor, and SSA antibodies and increased risk of arthritis. These findings were not corroborated by an analysis of CMC.

**Conclusions:** Among women diagnosed with SS, estrogen exposure was associated with reduced risk of abnormal OSS, RF, and anti-SSA antibody and increased risk of arthritis. CMC did not parallel these findings, potentially indicating estrogen exposure and other sex hormones involved in menstruation may differentially influence features of SS. Further longitudinal studies are required.

Table Odds ratios for association of sex hormone exposure with features of Sjögren's syndrome.

	UWS abro	rmal 05	S abnormal	Arthritis		RF positive		A positive	Focus score ≥1		GC present		t.
	OR 95%CI	P OR	95%CI P	OR 95%CI P	OR	95%CI P	OR	95%CI P	OR 95%	1 P	OR	95%CI	Ρ
CES													
0	ref	ref		ref	ref		ref		ref		ref		
1	125 104 150	0.02 0.76	0.63-0.92 0.01	1.20 0.99-1.45 0	05 0.90	0.74-1.09 0.2	7 0.84	0.70-1.02 0.08	0.84 0.65-1	.08 0.18	1.05	0.78-1.45	0.70
2	115 0.89-1.48	0.29 0.75	0.57-0.97 0.03	129 0.98-1.69 0	07 0.82	0.62-1.08 0.1	5 0.71	0.54-0.54 0.02	0.71 0.50-1	.01 0.05	1.05	0.67-1.68	0.81
3	0.58 0.59-1.63	0.93 0.37	0.21-0.66 0.00	263 133-5.19 0	01 0.49	0.25-0.94 0.0	3 0.49	0.25-0.91 0.02	0.84 0.38-1	.84 0.66	0.67	0.19-2.31	0.52
4		157	0.1-37.67 0.78				3.5	0.21-58.0 0.39					
CMC Premenopausal													
\$13	ref	ref		ref	ref		ref		ref		ref		
>13-515	152 0.90-2.55	0.12 0.94	0.53-1.65 0.82	0.75 0.44-1.31 0	32 0.70	0.39-1.23 0.2	1 0.87	0.50-1.51 0.62	0.68 0.31-1	45 0.32	1.08	0.47-2.47	0.85
>16-518	201 117-3.45	0.01 0.83	0.45-1.48 0.52	0.75 0.44-1.32 0	33 0.88	0.49-1.56 0.6	6 0.74	0.41-1.31 0.30	0.90 0.40-1	.99 0.79	0.75	0.31-1.81	0.53
>18	227 148-350	⊲.0001 0.68	0.43-1.07 0.10	0.74 0.47-1.17 0	20 0.80	0.50-1.27 0.3	4 0.66	0.42-1.05 0.08	0.67 0.35-1	27 0.22	1.01	0.51-2.01	0.98
Post Menopausal													
\$13	ref	ref		ref	ref		ref		ref		ref		
>13-516	114 0.43-2.75	0.75 1.25	0.52-3.07 0.61	2.27 0.67-7.62 0	19 0.75	0.31-1.84 0.5	3 102	0.42-2.45 0.96	1.13 0.33-3	90 0.85	1.07	0.23-4.91	0.93
>15-518	140 0.53-3.67	0.50 2.30	0.81-6.50 0.12	0.93 0.30-2.87 0	90 1.22	0.47-3.17 0.6	9 0.81	0.30-2.13 0.66	499 0.55-4	53 0.15	169	0.33-8.55	0.53
>18	141 0.85-2.33	0.17 0.92	055-152 0.74	0.65 0.37-1.17 0	16 0.53	0.32-0.87 0.0	1 0.73	0.44-1.19 0.21	0.85 0.44-1	.64 0.64	0.77	0.35-1.71	0.53
CES: Composite Estrogen S	icore; CMC:cumu	lative menstrual	cycles; UWS: uns	timulated whole sa	alivary flo	w; OSS: ocular	staining	score; RF: rheur	matoid factor; GC	germinal ce	nter		

#### **P-69**

#### Gene expression profiles in primary Sjögren's syndrome with and without active systemic manifestations

Claudio Vitali<sup>1</sup>, Marzia Dolcino<sup>2</sup>, Romina Andracco<sup>3</sup>, Andrea Pelosi<sup>4</sup>, Piera Fiore<sup>4</sup>, Wanda Maglione<sup>3</sup>, Eleonora Zaccara<sup>3</sup>, Nicoletta Del Papa<sup>3</sup>, Antonio Puccetti<sup>4</sup>.

<sup>1</sup>Section of Rheumatology, San Giuseppe Institute, Como; <sup>2</sup>Dept. of Medicine, University of Verona, Verona; <sup>3</sup>DH Reumatologia, Gaetano Pini Hospital, Milan; <sup>4</sup>Immunology Area, Bambino Gesù Hospital, Rome, Italy.

**Background.** Different phenotypes characterize the clinical spectrum of primary Sjögren's syndrome (SjS). Patients with a clinical expression limited to glandular features (GFs) are classically distinguished from patients with extra-glandular manifestations (EGMs). The former patients often complain higher level of fatigue and widespread pain (WP) (Segal *et al.* 2013). This suggests that gene expression pattern may be different in the two subgroups. The present study is aimed at investigating the differences of gene expression in SjS patients with and without EGMs.

**Methods.** All of the enrolled patients met the 2016 ACR-EULAR classification criteria for SjS. Gene expression of peripheral blood mononuclear cells (PBMCs) from 2 patients with SjS and EGMs and 2 patients with GFs alone was preliminarily analyzed using Clariom D human Affymetrix gene chip (Affymetrix, Santa Clara, CA, USA), and compared to that found in healthy controls. Differences in gene expression were evaluated by the analysis of variance (ANOVA) and Step-Up FDR-controlling procedure, being FDR corrected *p* value  $\leq 0.01$  and fold change >1.5 considered as statistically significant.

Validation of the gene overexpression was performed by quantitative Real Time(qRT)-PCR in PBMCs from 9 SjS patients with EGMs and 6 with GFs alone, using the  $\Delta\Delta$ Ct method for comparing relative fold expression differences.

**Results.** The total group of SjS patients was composed by 18 females and 1 male. All of the patients had a positive lip biopsy, while anti-SSA/Ro antibodies were detected in 10/11 and 6/8 of the SjS patients with and without EGMs, respectively. ESSDAI value ranged from 7 to 55 in SjS patients with EGMs (median 17), and from 0 to 2 in patients with GFs alone (median 1). In both types of patients, the functional analysis of the two transcriptomes showed a large number (>1000) of modulated genes that are involved in the biological processes (*i.e.*, apoptosis, inflammatory response, immune response, type I and type II interferons, and Toll-like receptors signaling) strictly connected to the known pathological processes of SjS. Genes involved in sensory perception and in nociceptive signaling (*i.e.*, ANPEP, TNRF1, P2RY1, 5HT1) were modulated only in patients with GFs alone. The significantly different expression of these selected genes in the two SjS subgroups was confirmed by the qRT-PCR analysis.

**Conclusions.** These data indicate that in SjS patients with GFs alone a dysregulation of pain signaling pathways (namely beta-adrenergic receptor signaling and Notch signaling) may play a role in the development of WP that is common in this subset of patients. The biological mechanisms underlying the activation of these genes remain to be clarified.

#### P-70

## IL10, TNF-α and TNFAIP3 genes polymorphism in patients with dry eye syndrome and Sjögren's syndrome

Ben Eli H.<sup>1,2</sup>, Solomon A.<sup>1</sup>, , Gomel N.<sup>3</sup>, Abu Seir R.<sup>4</sup>, Perlman R.<sup>4</sup>, Ben Chetrit E.<sup>5</sup>, Mevorach D.<sup>6</sup>, Kleinstern G.<sup>7</sup>, Paltiel O.<sup>2,4</sup> and Aframian DJ.<sup>8</sup>

<sup>1</sup>Department of Ophthalmology, <sup>2</sup>Braun School of Public Health and Community Medicine, <sup>3</sup>School of Medicine, <sup>4</sup>Department of Hematology, <sup>5</sup>Unit of Rheumatology, <sup>9</sup>Department of Internal Medicine <sup>7</sup>Department of Health Sciences Research, Mayo Clinic, Rochester, MN, USA. <sup>8</sup>Department of Oral Medicine & Sjögren's syndrome Center Hadassah-Hebrew University Medical Center, Jerusalem.

**Background.** Cytokine-related genes are assumed to be key players in dry eye syndrome (DES) and Sjögren's syndrome (SS) pathogenesis. However the association between specific genes variants and both DES and SS are unclear, and comparisons between these two diseases has not yet been performed. In this study we compared single nucleotide polymorphism (SNP) variation in genes encoding cytokine levels among SS and DES patients in Israel.

**Methods.** A total of 180 subjects were recruited, 82 with SS and 98 with DES. Using a candidate gene approach and allele-specific PCR technique for genotyping, the proportions of risk alleles in TNF- $\alpha$  (rs1800629), IL10 (rs1800896) and TNFAIP3 (rs2230926) SNPs were compared between study groups.

**Results.** The allelic distribution of the study groups was found to be very similar and match to Caucasians (CEU – Northern Europeans from Utah) population distributions in these SNPs. While none of the SNPs variants were found to be statistically significant associated to SS or DES in a recessive model, in an additive model the TNF $\alpha$  (rs1800629)-G risk allele was found among a higher proportion of SS patients compared to DES (Homozygote-G: 70.8% vs. 64.7%; Heterozygote: 26.9% vs. 11.2%, respectively, *p*=0.02). After adjusting for possible confounders, none of the tested SNPs were associated with SS compared to DES.

**Conclusions.** The frequency of IL10 (rs1800896-A) and TNFAIP3 (rs2230926-G) alleles was not found significantly differ between SS and DES patients. These findings may be due to limited power of the sample size of 180 participants. The TNF- $\alpha$  (rs1800629-G) SNP seems to be associated with SS in an additive model. TNF- $\alpha$  protein levels are known to be associated with inflammation, outcome of infection, and susceptibility to autoimmune diseases such as SS. The gene has also been associated with non-Hodgkin lymphoma, a serious complication of SS. Further comparison to healthy controls is required, as well as exploring other SNPs variants relating to the immune pathway in order to understand the genetic basis of DES and SS etiology.

## **P-71**

Identification of dysregulated immune-related gene networks in primary Sjögren's syndrome

Jeremy Kiripolsky<sup>1</sup>, Akinsola Oyelakin<sup>1</sup>, Rose-Anne Romano<sup>1</sup>, Guan Yu<sup>2</sup>, and Jill M. Kramer<sup>1</sup>.

<sup>1</sup>Dept. of Oral Biology, School of Dental Medicine, State University of New York at Buffalo, NY, USA. <sup>2</sup>Dept. of Biostatistics, School of Public Health and Health Professions, State University of New York at Buffalo, Buffalo, NY, USA.

Primary Sjögren's Syndrome (pSS) is an autoimmune disease with both oral and systemic disease manifestations. Similar to pSS patients, NOD.B10Sn-H2b/J (NOD.B10) mice develop lymphocytic infiltration in exocrine tissues, lose salivary flow, and display anti-nuclear autoantibodies. While pSS is clearly mediated by immune dysfunction, the pathways and networks that mediate disease are incompletely understood. Our objective was to characterize the genetic landscape in splenic tissue from NOD.B10 female mice with clinical disease in order to identify novel immune-related pathways that may be targeted therapeutically in disease. Spleens were harvested from female NOD.B10 mice with clinical disease and age and gender-matched C57BL/10SnJ controls (n=3 each). RNA was isolated and RNA-sequencing (RNA-seq) performed. Raw sequence reads were aligned and mapped to the current reference Mus musculus genome sequence using RNA-seq algorithms. Gene expression levels were calculated as fragments per kilobase of transcript per million mapped reads. We employed Cufflinks and Cuffdiff to determine gene transcript levels. Differential gene expression (DEG) analysis was carried out using DESq2. The DEGs were analyzed by Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis to determine the gene networks that were altered between pSS and control animals. We identified numerous pathways that were differently expressed between splenic tissue derived from NOD. B10 females and that from healthy controls. Many of these were related to innate and adaptive immune dysfunction, including B and T cell receptor pathways, cell adhesion molecule signaling, and networks involved in viral responses. In conclusion, our RNA-seq results identified numerous signaling pathways that are dysregulated in splenic tissue derived from a pSS mouse model. These data provide a foundation for future studies to establish the therapeutic relevance of these networks to human disease.

#### **P-72**

## Genetic differentiators of Sjögren's Syndrome subtypes in an international cohort

Kimberly E. Taylor<sup>1</sup>, Quenna Wong<sup>2</sup>, David M. Levine<sup>2</sup>, Kimberly Doheny<sup>3</sup>, Mi Y. Lam<sup>4</sup>, Caroline Shiboski<sup>4</sup>, Lindsey A. Criswell<sup>1</sup>, for the Sjögren's Syndrome Collaborative Clinical Alliance (SICCA).

<sup>1</sup>University of California, San Francisco, Russell / Engleman Rheumatology Research Center, San Francisco, CA; <sup>2</sup>University of Washington, <sup>3</sup>Johns Hopkins University, <sup>4</sup>University of California, San Francisco.

**Background.** While much has been learned in recent years about the genetics of Sjögren's Syndrome, little has been studied at the level of the major disease subphenotypes, which are important for understanding disease etiology and potential treatments. Our goal is to understand the genetic contributions to differences in the clinical manifestations of Sjögren's Syndrome, particularly with respect to ocular versus oral involvement.

**Methods.** We studied 3,355 participants in the Sjögren's International Collaborative Clinical Alliance (SICCA; contract # HHSN 268201300057C) registry genotyped on the Illumina HumanOmni 2.5-Quad marker set. Participants were enrolled at nine international sites, including Argentina, China, Denmark, India, Japan, the UK and three in the US. We also utilized 4,023 external controls from dbGap. Principal components analysis was used to characterize each participant for both continental and intra-continental genetic ancestry. We analyzed the entire group and the two largest strata by ethnicity, European and Asian.

We analyzed associations between patients with a high focus score (FS  $\geq 2.6$ , the median in our FS-positive subjects) and control subjects who were healthy or negative for the FS criteria (FS <1). Similarly, we analyzed associations between patients with a high ocular staining score (OSS  $\geq 7$ ) and control subjects who were healthy or had OSS <3. Genome-wide analyzes were performed for SNPs, genes, and pathways. We also examined the distribution of risk SNPs from our recent case-control GWAS in patients posi-

tive for the FS criteria but not the OSS criteria, compared to those who were OSS positive and FS negative.

**Results.** In our all-subjects and European analyses, numerous genes in the MHC region were significantly associated with both high FS (top all-subjects p=2e-25, OR=2.7; top European p=2e-22, OR=2.8) and high OSS (top all-subjects p=1e-19, OR=2.1; top European p=4e-17, OR=2.1). MHC genes were involved in the top pathways of both subphenotypes, including interferon gamma signaling and the MHC class I protein complex. As seen previously for SS, MHC variation appears to have a much lesser role in Asian patients for these subphenotypes. SNPs in *IRF5* were significantly associated with high FS (top SNP kgp2820799; all-subjects OR=1.5, p=9e-10; European OR=1.7, p=2e-8) whereas SNPs in *STAT4* (all-subjects top SNP kgp9676217, OR=1.4, p=1e-8; European top SNP rs7574865, OR=1.6, p=1e-9) were significantly associated with high OSS.

In a multivariate model of risk SNPs in European patients, FS-positive/ OSS-negative patients were more likely to have risk alleles of *GTF21* (kgp10686875, OR=1.9, p=0.021), *HLA-DRA* (rs6903608, OR=1.7, p=0.004), *NFAT5* (rs244418, OR=1.7, p=0.003), *BLK* (rs2735340, OR=1.7, p=0.009), *RELN* (rs7341475, OR=1.6, p=0.042), and *IRF5* (rs4728142, OR=1.5, p=0.026). In multivariate analysis of risk SNPs in Asian patients, FS-positive/OSS-negative patients were more likely to have risk alleles of *HLA-DRA* (rs6903608, OR=3.7, p=0.001) and *TNIP1* (rs3792785, OR=3.2, p=0.025).

**Conclusions.** The FS and OSS subphenotypes have a combination of shared and distinct genetic contributors, which also vary according to ancestry.

### **P-73**

#### The role of GPR78 in Sjögren's syndrome

Maria C. Guimaro<sup>1</sup>, Paola P. Riveros<sup>1</sup>, Melodie L. Weller<sup>2</sup>, Drew G. Michael<sup>1</sup>, Sandra A. Afione<sup>1</sup>, Giovanni Di Pasquale<sup>1</sup>, William Swaim<sup>1</sup>, Hongen Yin<sup>1</sup>, Malini Ahuja<sup>1</sup>, and John A. Chiorini<sup>1</sup>.

<sup>1</sup>National Institute of Dental and Craniofacial Research (NIDCR), National Institutes of Health, Bethesda, MD. <sup>2</sup>University of Utah, Salt Lake City, Utah.

**Background.** The molecular mechanism underlying the gender disparity in prevalence of several autoimmune diseases is poorly understood. Primary Sjögren's syndrome (pSS) is much more common in women than men, having a 9:1 female/male ratio. Large differences in the transcriptomes of minor salivary glands from male and female healthy volunteers were observed in previous data from our lab. We hypothesize that salivary glands (SG) from male and female patients with pSS will have different gene expression profiles.

**Methods.** Minor salivary glands were surgically excised from individuals and clinical parameters and disease status for primary Sjögren's syndrome were recorded. Microarray analysis was performed per the manufacturers standard protocol (Agilent, Santa Clara, CA). Following feature extraction, data were normalized via 75<sup>th</sup> percentile shift and probes subject to high levels of noise (<20<sup>th</sup> percentile expression) removed from downstream analyses. The measurement of cAMP was made using a cAMP response element on a luciferase reporter assay. Caspase 3 experiments were measured using an activated caspase 3/7 green detection reagent and by Western blot. Apoptosis experiments were performed by flow cytometry using the Annexin V assay. Markers of early and late stages of apoptosis were analyzed by Western blot.

**Results.** Our transcriptome analysis showed a significant increase in Gprotein coupled receptor 78 (GPR78) expression in the minor SG RNA isolated from male SS patients compared with male healthy volunteers. This increase was not observed in female patients when compared with female healthy volunteers. GPR78 is an orphan receptor that triggers intracellular cAMP signaling. No mouse ortholog of this protein is reported and the function of GPR78 is unknown with only a few studies published about this protein. Cells transfected with GPR78 showed an increase in cAMP activity as previously reported but also had less viability compared to control cells. Further analysis indicted these transfected cells have an increase in caspase 3 activation, decrease in Bid, and an increase in markers of early and late stage apoptosis. Additional studies showed that expression of GPR78 also interfered with muscarinic receptor activity and expression.

**Conclusions.** This onset of apoptosis aligns with previous literature suggesting an increase in apoptosis in the salivary glands of pSS patients. Our results suggest that GPR78 is likely involved in the pathogenesis associated with pSS in male patients.

#### **P-74**

## Transcription factors ETS1 and LEF1 as potential pathogenic biomarkers of Sjögren's syndrome

Nirav R. Shah, PhD, Braxton D. Noll, BS, Jenene Noll, RN, Mike T. Brennan, DDS, MHS, Ricardo Padilla, DDS, Farah K.B. Mougeot, PhD, Jean-Luc C. Mougeot, PhD.

Department of Oral Medicine, Carolinas Medical Center, Charlotte, NC, USA.

Background. Sjögren's syndrome (SS), is a chronic systemic autoimmune disease mainly characterized by severe dry eyes and dry mouth and mostly affecting women. Despite extensive research, the etiology of this disease is not fully understood. Currently, the American college of rheumatology/European league against rheumatism (ACR-EULAR) criteria are being used for the classification of primary SS. Under the ACR-EULAR classification, lymphocytic infiltration in the minor salivary gland (MSG) corresponding to a focal score (FS)  $\geq 1$  can be designated as SS positive. Using a computational meta-analysis, we previously identified ETS1 and LEF1 as candidate biomarkers of SS. In this study, for the first time, dual staining immunofluorescence was used to analyze the protein expression and localization of ETS1, LEF1, and MMP9 in SS patients compared to non-SS sicca control patients. Methods. The paraffin embedded MSGs were cut (5 µm) from 11 SS and 13 non-SS sicca patient control samples. These sections were deparaffinized and processed for antigen retrieval. Next, tissue sections were blocked and further incubated with primary antibodies (ETS1-CD4, LEF1-CD4 or MMP9-TIMP1) for 60 minutes, followed by incubation with Cy3-Cy5 (green-red) conjugated secondary antibody for dual staining. Sections were mounted and images were captured using confocal microscope. Total fluorescence intensity and H-score for individual staining were calculated.

**Results.** Our immunofluorescence analysis revealed that all MSG tissue sections from SS patients with FS  $\geq$ 1 had most areas with moderate to high intensity of LEF1-ETS1-MMP9 colocalization, whereas MSG tissue sections from SS patients with FS <1 had mostly very low to moderate intensity areas of LEF1-ETS1-MMP9 colocalization. Since ETS1 and LEF1 are transcription factors known to increase MMP9 expression in cancer cells, it can be hypothesized that MMP9 upregulation is due to increased ETS1 and/ or LEF1 expression in the MSGs of SS patients.

**Conclusions.** Our analysis showed that both ETS1 and LEF1 are significantly upregulated and co-localized with MMP9 expression in the MSGs of SS patients, but not in non-SS sicca control patients. These results suggest the potential importance of ETS1 and/or LEF1 upregulation as candidate biomarker(s) for the diagnosis of SS.

#### **P-75**

#### Functional characterization of the Sjögren's syndrome-associated locus DDX6-CXCR5

Sharmily Khanam<sup>1</sup>, Michelle L Joachims<sup>1</sup>, Nicholas Means<sup>1,2</sup>, Indra Adrianto<sup>3</sup>, Astrid Rasmussen<sup>1</sup>, Simon J Bowman<sup>4</sup>, David M Lewis<sup>5</sup>, Lida Radfar<sup>1,6</sup>, Rolad Omdal<sup>7</sup>, Marie Wahren-Herlenius<sup>8</sup>, Ilias Alevizos<sup>9</sup>, Torsten Witte<sup>10</sup>, Roland Jonsson<sup>11,12</sup>, Maureen Rischmueller<sup>13</sup>, Patrick M Gaffney<sup>1</sup>, Judith A James<sup>1,2,14</sup>, Lars Rönnblom<sup>15</sup>, Elke Theander<sup>16</sup>, Nelson L Rhodus<sup>17</sup>, Barbara M Segal<sup>18</sup>, R Hal Scofield<sup>1,14,19</sup>, Courtney G Montgomery<sup>1</sup>, Xavier Mariette<sup>20</sup>, Wan-Fai Ng<sup>21</sup>, for UK Primary Sjögren's Syndrome Registry, Gunnel Nordmark<sup>15</sup>, Kathy L Sivils<sup>1,2</sup>, and Christopher J Lessard<sup>1,2</sup>.

<sup>1</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, Oklahoma, USA. <sup>2</sup>Department of Pathology, University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma, USA.<sup>3</sup> Department of Public Health Sciences, Henry Ford Health System, Detroit, MI, USA. <sup>4</sup>Rheumatology Department, University Hospital Birmingham, Birmingham, UK. <sup>5</sup>Department of Oral and Maxillofacial Pathology, University of Oklahoma College of Dentistry, Oklahoma City, Oklahoma, USA. <sup>6</sup>Oral Diagnosis and Radiology Department, University of Oklahoma College of Dentistry, Oklahoma City, Oklahoma, USA. <sup>7</sup>Clinical Immunology Unit, Department of Internal Medicine, Stavanger University Hospital, Stavanger, Norway. <sup>8</sup>Department of Medicine, Karolinska Institute and Karolinska University Hospital, Stockholm, Sweden. <sup>9</sup>Sjögren's Syndrome Clinic, National Institute of Dental and Craniofacial Research, Bethesda, MD, USA. <sup>10</sup>Department of Clinical al Immunology and Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>11</sup>Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>12</sup>Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, Norway. <sup>13</sup>Department of Rheumatology, The Queen Elizabeth Hospital and Discipline of Medicine, University of Adelaide, South Australia. <sup>14</sup>Department of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City,

Oklahoma, USA. <sup>15</sup>Department of Medical Sciences, Rheumatology and Science for Life Laboratory, Uppsala University, Uppsala, Sweden. <sup>16</sup>Department of Rheumatology, Skane University Hospital, Lund University, Malmø, Sweden. <sup>17</sup>Department of Oral Surgery, University of Minnesota School of Dentistry, Minneapolis, Minnesota, USA. <sup>18</sup>Division of Rheumatology, University of Minnesota Medical School, Minneapolis, Minnesota, USA. <sup>19</sup>Department of Veterans Affairs Medical Center, Oklahoma City, Oklahoma, USA. <sup>20</sup>Université Paris-Sud, Assistance Publique–Hôpitaux de Paris (AP-HP), Hôpitaux Universitaires Paris-Sud, Institut National de la Santé et de la Recherche Médicale (INSERM) U1012, Paris, France. <sup>21</sup>Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK.

**Background.** Sjögren's syndrome (SS) is a chronic, heterogeneous disease with hallmark features of auto-inflammation and autoantibody production. We previously identified association between the *DDX6-CXCR5* locus and SS surpassing genome-wide significance. The goal of this study is to determine the mechanism by which this association contributes to disease.

Methods. Fine mapping and imputation approaches were used to enrich existing genetic datasets using 1916 SS cases and 3684 controls to include ~2x the testable variants as previous, 971, in the DDX6-CXCR5 interval. Statistical approaches and bioinformatics data were used to prioritize candidate variants. Electromobility shift assays (EMSAs) and pull-downs (PDs) followed by mass spectrometry were used to determine allelic-specific differences in binding using lysates from HSB-2 (T), Jurkat (T), Reh (B), Ramos (B), Daudi (B), THP-1 (monocyte), and HEK 293T (epithelial) cells. Results. In the DDX6-CXCR5 region, imputation showed a pattern of association spanning beyond the length of the DDX6 coding sequence to the proximal promoter of CXCR5. Bioinformatic analysis of the top associated variants (rs7125066 and rs7119038) did not yield evidence of regional functionality. However, 46 other candidates that span the region of association were identified. Chromatin methylation pattern data from the Roadmap database showed several variants in this region were within transcription start sites or enhancer elements depending on the cell type and state. Using RegulomeDB, Haploreg, and other databases, 4 variants showed strong evidence they affect binding and/or expression of one or more target genes in the region. Thus, rs4938572, rs12365699, rs57494551, and rs10892294 were selected for further study. Using EMSAs, the risk allele resulted in a statistically significant decrease in binding when compared to the non-risk allele for rs4938572 (p<0.01) using cell lysates from HSB-2, Reh, THP-1, Ramos, Jurkat, and Daudi but not HEK 293T cells. However, the differences were more restricted for rs10892294, rs12365699, and rs57494551, which decreased binding with the risk allele only uisng Ramos cells. Moreover, the risk allele decreased binding at rs10892294 and rs57494551 increased binding at rs12365699. The rs57494551 risk allele also decreased binding within the THP-1 cell lysate. Preliminary mass spectrometry analysis showed several immune related transcription factors likely binding this region, including: IRF8, GTFTI, RFX5, IKZF3, PAX5, and IKZF1. Cell type-specific expression of the genes in the region shows the expression of DDX6 spans many immune cell subsets while the expression of CXCR5 is more restricted.

**Conclusions.** Genetic variants in the *DDX6-CXCR5* locus likely alter both protein coding genes in a cell/context specific manor. Ongoing studies will assess the role of these sequences as promoters and/or enhancers using luciferase assays and CRISPR/Cas9 deletions of the elements of interest in various cell types.

#### **P-76**

#### What's old is new again: ancient aryuvedic oil-pulling technique combats the microbiome associated with the oral maladies of Sjögren's syndrome patients

Leslie P. Laing<sup>1</sup>, Yona Vandersluis<sup>2</sup>, and Javid Al-Noor Karim<sup>3</sup> <sup>12</sup>University of Toronto, Faculty of Dentistry, <sup>1</sup>Discipline of Prosthodontics, <sup>2</sup>Discipline of Orthodontics, Toronto, Ontario, M5G 1G6; <sup>3</sup>Western University, London, Ontario, N6A 3K7, CANADA

**Background.** Sjögren's Syndrome (SS) is an auto-immune disorder with a plethora of oral manifestations including dry mouth, rampant dental decay, gingivitis, candidiasis, and hypogeusia/ dysgeusia, and subsequently wreaking havoc on quality of life (QoL). Since the natural oral buffer capacity of saliva is reduced in these patients, it is of vital importance that agents be identified that might protect the tooth surface from demineralization. Lipids may play such a role through the inhibition of carious demineralization by providing a diffusion barrier within the organic protein-lipid-water matrix of enamel which may decelerate caries demineralization (Featherstone & Rosenberg 1984). During a literature search to find agents that could either rehydrate or at least prevent further oral dehydration of xerostomic patients, a description of the ancient Ayurvedic oil-pulling technique was discovered claiming several benefits to oral health (Asokan 2008). The purpose of the study was to determine 1) the *in vivo* effects of oil-pulling in patients with xerostomia due to SS and 2) the *in vitro* effects of the edible oils in protection against erosion.

**Methods.** 1) *in vivo*: participants previously diagnosed with primary SS (pSS) or otherwise healthy salivators (HS) were recruited into the study. QoL questionnaires were completed concurrently with the collection of saliva, supra- and sub-gingival plaque, and tongue scraping samples were collected pre- and post-testing and. Samples were analysed for levels of *Strep. mutans*, Lactobacilli *sp.*, and Candida *sp.* Participants were randomly assigned to a rinsing agent and followed the traditional oil-pulling technique of 15 min/day for 3 weeks first thing in the morning using virgin coconut oil (VCO), olive oil (OO), sesame oil (SEO), sunflower (SUO), chlorhex-idine (CHX, 1 min/day, 2 weeks only), or distilled water. 2) *in vitro:* Human extracted teeth were subjected to 10 cycles of 5 min pre-treatment with the above products, 30 min artificial saliva, 3 min 1% citric acid (pH 2.3), 60 min artificial saliva then subjected to optical profilometer analysis.

**Results.** In SS participants, levels of *Strep. mutans* were reduced 10-fold by OO and SEO, 100-fold by VCO and completely by CHX; of Lactobacilli *sp.* 1000-fold by CHX, otherwise not at all; and of Candida *sp.* up to 100-fold by VCO, 10-fold by SUO and CHX. After usage of VCO, SS participants stated "my guns don't bleed after flossing", "my teeth look brighter", "I can taste again", and "I don't have that sour dough smell". Protection against enamel and dentin loss was greater with VCO than SEO or SUO and not all with OO.

**Conclusion.** Oil-pulling with VCO offers the xerostomic SS patient a more pleasant oral condition, an improvement in QoL, a decrease in micro-organisms associated with various oral maladies, and a somewhat protective effect against xerostomia-related tooth demineralization.

### **P-77**

## The bacterial community of the periparotid epithelium in patients with Sjögren's syndrome

Mayank Tandon, Robert J. Palmer Jr., Loreto Abusleme Ramos, Eileen Pelayo, Pamela J. Gardner, Niki Moutsopolous, Ilias Alevizos. National Institute of Dental and Craniofacial Research, NIH, Bethesda, Maryland, USA.

**Background.** Sjögren's Syndrome (SS) is a common autoimmune exocrinopathy that affects primarily the salivary and lacrimal glands, causing hyposalivation and chronic dryness of the mouth and eyes. Because salivary function is critical for oral health, SS patients have an increased risk of dental caries and oral candidiasis, which is partly attributable to the fundamental role of saliva in the metabolism of oral microorganisms. 16S rRNA has previously been used to study the oral microbiome of SS patients using DNA extracted from whole saliva, but, at best, only mild disease-specific alterations were reported. Here, we examine phylogenetic diversity in bacterial communities present on the oral mucosa adjacent to the parotid gland of SS patients and healthy controls (HCs).

**Methods.** DNA was extracted from buccal swabs of the parotid periglandular region of SS patients (n=12) and controls (n=13). 16S rRNA sequencing was performed using primers directed to the hypervariable V1V2 region on the Illumina MiSeq platform. Sequenced reads were processed according to standard procedures for mothur analyses. Reads were filtered for length and quality, then clustered into Operational Taxonomic Units (OTU: species defined by rRNA gene sequence) according to the recreated SEED database from SILVA version 123, provided with mothur. The phyloseq package in R was used to perform statistical analyses.

**Results.** On average, 134 species were observed in each sample. While no significant difference was found between patients and controls (p=0.19), SS patients tended to have fewer species. Further, no statistically significant differences were found by the alpha diversity measures Chao1, Shannon, Simpson, Fisher, and ACE indices. However, principal coordinates analysis (PCoA) of the Jensen-Shannon (JS) distance – a measure of beta diversity – showed differences between patients and healthy controls. Statistical testing on the Jaccard and JS distances between SS and HC samples proved the differences to be highly significant (p=0.001). Linear discriminant analysis Effect Size (LEfSe) procedures demonstrated SS patients to have a greater

presence of OTUs belonging to taxa in the Escherichia/Shigella group and in Enterobacteriales than did the HCs, while OTUs within phylum Firmicutes and family Erysipelotrichaceae were more important in HCs.

**Conclusions.** Statistically significant differences were found in the oral flora of the glandular epithelium between SS patients and HCs. Differences in beta diversity (the relative distribution of organisms), but not in alpha diversity (number and types of organisms), between health and disease suggests that the communities differ significantly. The taxa enriched in SS patients are representative of non-resident bacteria, such as enterococci and coliform bacteria, whereas typical oral taxa (some being minor components of the oral flora) were more abundant in healthy subjects. Given the proximity of the sample collection site to the parotid gland, a major target organ in SS, a potential exists that the SS-associated organisms have a role in the severity or progression of the disease through their intimate relationship with the glandular region.

#### **P-78**

#### Dysbiosis of the Oral Microbiome in Patients with Primary Sjögren's Syndrome

T.A. van der MeuleN<sup>1</sup>, F.G.M. Kroese<sup>2</sup>, A. Vich Vila<sup>3</sup>, S.C. Liefers<sup>2</sup>, H. Bootsma<sup>2</sup>, F.K.L. Spijkervet<sup>1</sup>, H.J.M. Harmsen<sup>4</sup> and A. Vissink<sup>1</sup>.

<sup>1</sup>Dept. of Oral and Maxillofacial Surgery, <sup>2</sup>Dept. of Rheumatology and Clinical Immunology, <sup>3</sup>Dept. of Gastroenterology, <sup>4</sup>Dept. of Medical Microbiology; UMC Groningen, University of Groningen, the Netherlands.

**Background.** Early differentiation of primary Sjögren's syndrome (pSS) patients from patients with oral dryness due to other causes is a diagnostic challenge. If the oral microbiome is specific for pSS, then the oral microbiome might serve as an early biomarker to distinguish pSS from non-SS dry mouth patients (non-SS). Therefore, the aim of our study was to assess the oral microbiome of pSS and non-SS patients in relation to healthy controls and disease controls.

**Methods.** The oral microbiome of 72 pSS patients, 90 non-SS sicca patients, 33 systemic lupus erythematosus (SLE) patients and 14 healthy controls (HC) was assessed with 16S rRNA sequencing of oral washing samples. All pSS patients fulfilled the 2016 ACR/EULAR classification criteria. Patients referred for a diagnostic workup for SS not fulfilling the classification criteria were grouped as non-SS patients. All SLE patients fulfilled the 2012 SLICC criteria. Sequencing data and diversity analyses were processed in QIIME 1.9.1. Samples contained at least 4000 reads/sample. Bacterial richness, diversity, overall composition and relative abundance of bacterial taxa were compared between the four groups.

**Results.** Oral washings from pSS patients showed a lower bacterial richness (number of different bacterial taxa/sample) than those of non-SS patients, SLE patients and HCs (p=0.001, p=0.03, p=0.01 respectively, Wilcoxon test, Figure 1). Also bacterial diversity (Shannon index) was lower in pSS patients compared to non-SS patients, SLE patients and HCs, although only statistically significant compared to non-SS patients (p=0.004, Wilcoxon test, Figure 1). Furthermore, the overall bacterial composition of oral washings from pSS patients significantly differed from non-SS, SLE and HCs (p<0.001 for all groups). However, disease status explained only 6% of the variation in distances between all samples and 4% of the variation between pSS and non-SS samples (statistics: adonis from vegan package in R, Figure 2).



Fig. 1. Boxplots representing the median with interquartile range of bacterial richness and bacterial diversity, measured by respectively the number of different taxa per sample and the Shannon index.



Fig. 2. Principal coordinate analysis (PCoA) based on the Unweighted Unifrac distance of all oral washing samples. 34.1% of the variation between the samples is explained by the first and second principal component (PC1, PC2). Samples from pSS and non-SS patients shift towards a higher PC1. The four groups differ significantly from each other. Disease status explained 6% of the total variation in samples and 4% of the variation between pSS and non-SS patients. This is apparent in by the large spread and overlap of samples from the different groups.

The relative abundance of 12 bacterial genera was significantly lower in pSS patients compared to non-SS patients. Only genus *Lactobacillus* was higher in pSS patients (statistics: MaAsLin in R, false discovery corrected *p*-value<0.1). Thus, the oral microbiome of pSS patients is characterized by a lower bacterial richness, lower diversity and an increased relative abundance of *Lactobacillus*. Therefore the oral microbiome of pSS patients can be classified as dysbiosis.

**Conclusions.** The oral microbiome of pSS patients is characterized by dysbiosis and differs from the oral microbiome of non-SS patients, SLE patients and healthy controls.

#### P-79

#### Oxidative damage in Sjögren's syndrome

Biji T. Kurien<sup>1,2,3</sup>, Sona Nuguri<sup>4</sup>, Bre'ana Byrd<sup>1,2,5</sup>, Joey Maher<sup>1,2,6</sup>, Rohit Thomas<sup>1,4</sup>, Huyen Tran<sup>1,2,5</sup>, Stephanie L. Cummings<sup>1,2</sup> R. Hal Scofield<sup>1,2,3</sup>. <sup>1</sup>Arthritis & Clinical Immunology Program, Oklahoma Medical Research Foundation; <sup>2</sup>Department of Medicine, University of Oklahoma Health Sciences Center; <sup>3</sup>Veterans Affairs Medical Center, Oklahoma City; <sup>4</sup>Oklahoma School of Science and Mathematics, <sup>5</sup>Oklahoma City; University of Central Oklahoma, <sup>6</sup>Edmond; University of Oklahoma, Norman, Oklahoma.

Sjögren's syndrome (SS) is a lingering inflammatory, autoimmune condition with reduced lacrimal and salivary gland secretion resulting in keratoconjunctivitis sicca and xerostomia. Autoantibodies targeting the 60 kD Ro autoantigen occurs in up to 90% of patients with SS. Oxidative damage mediated by free radicals has not been well characterized previously in SS. Therefore, we studied oxidative damage (conjugate diene formation) and modification of serum proteins by the lipid peroxidation by-product 4-hydroxy-2-nonenal (HNE) or malondialdehyde (MDA) in SS, and age-and sex matched controls.

**Materials.** We studied 69 primary SS subjects, 25 age and sex matched subjects that did not meet criteria (incomplete SS), and 18 normal controls. Indices of oxidative damage, namely conjugate diene formation, and HNE or MDA-protein adducts were investigated in the sera of SS, incomplete SS and normal controls. Diluted sera from SS subjects or normal controls were coated on ELISA plates as antigen source. HNE or MDA adducts in serum proteins bound to the plates was determined with rabbit anti-HNE or anti-MDA antibodies purchased commercially. To identify specific proteins modified by HNE, sera from SS or normal controls were electrophoresed, transferred by electroblotting and subjected to immunoblotting with rabbit anti-HNE antibody. For determination of conjugate diene, 25 µl of SS sera or incomplete SS were extracted with chloroform:methanol (2:1) and the samples were centrifuged. Two ml of the clear supernatam was evaporated to dryness at 45°C and reconstituted in one ml methanol. The spectra ranging from 200 to 360 nm was read using a spectrophotometer.

**Results.** Significantly increased oxidative damage was found in the sera of primary SS subjects compared to normal controls by ELISA and immunoblotting. Serum proteins from SS subjects were found to contain HNE adducts. There was significantly more HNE-modified proteins in SS sera (n=10) compared to controls (n=10) by ELISA ( $0.074\pm0.017$  versus  $0.046\pm0.007$ ; p=0.00015; average OD±SD). However, there was no sig-

nificant difference in MDA-modified proteins between SS and controls by ELISA. When SS sera (n=34) were analyzed by immunoblotting, we found HNE adducts in several serum proteins, and significantly in a protein migrating at 18 kD. Normal control sera did not show significant HNE-modification (n=8). Our preliminary results for conjugate diene formation show that there is no significant difference between conjugate diene levels in the Sjögrens's syndrome patients (n=25) and incomplete SS subjects (n=25). We are pursuing HNE modification in the sera of incomplete SS subjects and also identifying the protein bands in SS subjects with HNE adducts by matrix assisted time of flight mass spectrometry.

**Conclusion.** Oxidative damage occurs in SS, as evidenced by the significantly elevated HNE protein adducts—but not MDA-protein adducts—in the sera of SS subjects compared to normal controls.

#### **P-80**

#### Transcriptomic profiling of pDCs from patients with pSS indicates an activated phenotype, enhanced anti-viral state, and increased susceptibility to apoptosis

Maarten R Hillen<sup>\*12</sup>, Aridaman Pandit<sup>\*12</sup>, Sofie LM Blokland<sup>12</sup>, Sarita AY Hartgring<sup>12</sup>, Kim van der Wurff-Jacobs<sup>1</sup>, Aike A Kruize<sup>1</sup>, Marzia Rossato<sup>12</sup>, Joel AG van Roon<sup>\*12</sup>, Timothy RDJ Radstake<sup>\*12</sup>.

<sup>1</sup>Laboratory of Translational Immunology, University Medical Centre Utrecht, Utrecht University, Utrecht, the Netherlands. <sup>2</sup>Department of Rheumatology & Clinical Immunology, University Medical Centre Utrecht, Utrecht University, Utrecht, the Netherlands.

\*Equal contribution

**Background.** Type-I interferons (IFN) are thought to play an important role in pSS pathogenesis. Plasmacytoid dendritic cells (pDCs) produce large amounts of type-I IFN upon activation via Toll-like receptors. pDCs infiltrate the salivary glands of patients with pSS and their numbers correlate with local IFN-production. Furthermore, pDCs are decreased in number in the peripheral blood of patients with pSS. To understand the molecular mechanisms behind systemic dysregulation of pDCs, we performed RNA sequencing on pDCs isolated from peripheral blood of patients with pSS, incomplete SS (iSS) and healthy controls.

**Methods.** We established two independent cohorts (each n=31), of patients and controls. pSS patients (n=25) were classified according to the 2002 AECG criteria. iSS patients (n=20) were included in both cohorts. These individuals presented with dryness complaints without a known cause, did not have any generalized autoimmune disease including pSS, and did not fulfil the classification criteria for pSS. Healthy donors (n=17) were included as control group. Peripheral blood pDCs were isolated using magnetic bead-associated cell sorting.  $\pm 20$  million paired-end sequencing reads per sample were obtained using using Illumina HiSeq 2500 platform.

**Results.** 8556 genes were differentially expressed (*p*-value <0.05) between all three groups in the discovery cohort. Of these, 3144 genes were also differential in the replication cohort. We generated gene modules from both discovery and replication cohorts, and found 5 gene clusters comprising 1259 genes that were consistently dysregulated in both cohorts. Interestingly, this dysregulation was regulated by a few key transcription (co)factors. Pathway analysis showed that the 5 modules contain genes, as well as genes involved in regulation of apoptosis and intracellular transport. Generally, pDCs from patients with iSS displayed an intermediate phenotype but a few iSS patients exhibited similar degree of dysregulation in their pDCs transcriptome compared to the pSS patients.

**Conclusions.** Using blood pDC transcriptomics, we identified 5 gene clusters that show that pDCs from pSS patients exhibit an activated phenotype. These gene clusters are robustly replicated in two independent cohorts. Taken together, our data indicate that pDCs from the blood of patients with pSS, and to a lesser extent those with iSS, have an activated phenotype and are more susceptible to induction of migration and apoptosis.

## Clinical and Experimental Rheumatology 2018

#### **P-81**

## Immune activation following seasonal influenza vaccination in patients with primary Sjögren's syndrome

Albin Björk<sup>1</sup>, Gudny Ella Thorlacius<sup>1</sup>, Johannes Mofors<sup>1</sup>, Marika Kvarnström<sup>1</sup>, Marie Wahren-Herlenius<sup>1</sup>.

<sup>1</sup>Department of Medicine, Karolinska Institutet, Stockholm, Sweden.

**Background.** Immune triggering by environmental factors in genetically predisposed individuals has been suggested as an underlying pathogenic mechanism in development of autoimmune diseases. Although highly warranted, studies of primary infections in humans pose great logistic and temporal challenges. However, monitoring responses following vaccination enables systematic studies of immune activation to microbial agents *in vivo* under standardized and safe conditions. In the present study, we therefore longitudinally observed untreated and hydroxychloroquine (HCQ) treated patients with primary Sjögren's syndrome (pSS) and healthy controls (HC) vaccinated against seasonal influenza to identify differences in immune activation and resulting cellular and serological responses.

**Methods.** The study included pSS patients without treatment (n=17), pSS patients receiving HCQ treatment (n=8), and HC (n=16). All participants were female and all pSS patients were positive for Ro/SSA autoantibodies. All individuals received vaccination with a non-adjuvanted trivalent seasonal influenza vaccine as part of the standard vaccination program. Blood samples were collected before vaccination, 24 hours after, on day 28, and on day 90. Clinical parameters were registered using a questionnaire based on ESSPRI. RNA expression analysis was performed in CD14<sup>+</sup> monocytes using nanostring. Antibody titers were analysed by ELISA.

Results. Differences in the immediate response to immune activation was observed in terms of a significant decrease in the total leukocyte count 24 hours after vaccination in untreated patients with pSS (p<0.02), which was not observed in the HCQ treated or HC group. Investigation of mRNA expression in CD14+ monocytes before and 24 hours after vaccination as expected demonstrated an IFN-signature in patients before vaccination, and revealed a differential transcriptional response to vaccination in untreated patients compared to HC. In particular, higher activation of genes in the NFkB signaling pathway was observed in untreated patients following vaccination, with the top hit being the NF-kappa-B essential modulator (NEMO) (p<0.001). Further, untreated patients with pSS, but not HCQ treated patients, responded with significantly higher vaccine specific IgG titers than HC (p<0.03), and there was a positive, significant correlation of the early, higher expression of genes in the NFkB signaling pathway and the higher serological response to the vaccine one month after vaccination (Spearman  $R_{NEMO} = 0.44$ ,  $p_{NEMO} < 0.02$ ). Notably, levels of anti-Ro52 autoantibodies increased in untreated patients, but not in HCQ-treated patients after vaccination. No significant changes in self-reported clinical parameters were registered.

**Conclusion.** We observed augmented innate and adaptive immune responses to vaccination in untreated patients with pSS, suggesting an underlying hyper-responsiveness to immune challenges. The higher serological response was associated with a higher early activation of the NFkB pathway in monocytes, indicating the importance of NFkB signaling in enhanced immune responses of autoimmune individuals.

#### **P-82**

## Optimized footprinting pipeline for profiling diseased cell state changes

Thomas JF Pranzatelli, Drew Michael, Ida Shinder, John A. Chiorini.

**Background.** Sjögrens' syndrome is a complex multifactorial disease with an associated loss of salivary tissue function. Salivary gland tissues of individuals with Sjögren's syndrome have altered transcription compared to tissues of healthy volunteers. To profile the regulatory state changes concomitant with Sjögren's syndrome, we have optimized a pipeline for analyzing footprinting information against known protein binding information from ChIP-seq.

**Methods.** To optimize the pipeline, two replicates each of DNase1- and ATAC-seq data for the lymphocytic cell line GM12878 were tested in pipelines varying across eight parameters for a total workload of 106 computational years. Pipelines with different combinations of parameters were tested for the correlation between intermediate files of the two replicates,

representing the reproducibility of the result, as well as the correspondence between footprinting information and known protein binding via ChIP-seq. Pipelines with high ChIP-seq values did not necessarily have high correlations between replicates in intermediate files, so pipelines optimized for reproducibility are reported alongside pipelines optimized for recapitulation of biological information.



**Results.** Reproducibility-optimized pipelines showed a nearly twofold increase in correlation between alignments, called regions of open chromatin, and footprint placement between replicates. ChIP-seq-optimized pipelines showed a roughly 16% increase in AUC against ChIP-seq relative to default parameters when using the footprinting algorithm HINT-BC and recovered non-random prediction when using the footprinting algorithm Wellington. **Conclusions.** These pipelines can be used similarly to ensemble machine learning, where multiple models vote on predictions. The results from well-validated pipelines of different stringencies and strengths adds information about footprints that are gained or lost between diseased and healthy individuals, and this paves the way for understanding how changes in gene regulatory network subgraphs define disease.

### **P-83**

## Kynurenines pathway biomarkers for primary Sjögren's syndrome.

Valéria Valim<sup>1</sup>, Eliana Zandonade<sup>1</sup>, Johan G Brun<sup>2</sup>, Roland Jonsson<sup>2</sup>, Per Ueland<sup>3</sup>, Piotr M Mydel<sup>2</sup>.

<sup>1</sup>Hospital Universitário Cassiano Antônio de Moraes/Federal University of Espírito Santo. <sup>2</sup>Bröegelman Research Laboratory, Haukeland Hospital, University of Bergen. <sup>3</sup>Bevital Laboratory, Haukeland Hospital, University of Bergen.

**Objective.** To investigate kynurinines pathway metabolites and their association with clinical manifestation in primary Sjögren's syndrome (pSS).

**Methods.** Cross-sectional study including 97 patients with pSS (AECG 2002 or ACR/EULAR 2017) and 63 age matched healthy controls. Tryptophan, kynurenines, neopterin, and vitamin B6 forms were measured in plasma by liquid chromatography-tandem mass spectrometry. Non-parametric statistics, Spearman correlation, Roc curves and logistic regression were applied with a p-value of 0.05.

**Results.**  $50\pm11$  anos, 95% women, anti-SSA-Ro 63%. Kynurine/tryptophan rate (KTR), kynurinine (KYN), quinolinic acid (QA), 3-Hydroxykynurenine (HK) xanthurenic acid (XA), anthranilic acid (AA) were higher and tryptophan was lower in pSS compared to controls (p<0.05).

KTR was associated with disease duration, higher creatinine, hypergammaglobulinemia, CRP, and low hemoglobin, C3/C4 (r=0,211-0,588,  $p \le 0.05$ ). Higher KTR was found in those with biological (0,033±0,016 vs. 0,029±0,013; p=0,003) and glandular ESSDAI evolvement (0,037±0,014 vs. 0,029±0,013; p=0.007). Neopterine was associated with glandular manifestation, ESSDAI, CPR, hypergammaglobulinemia, hyper IgG, and low unstimulated whole salivary flow, hemoglobin, neutrophils, hemathuria, and complement (r=0,233-0,455,  $p \le 0.05$ ). Higher levels of Kyn (OR=2,51, 1.44-55.64 C195%; 0.005), Kyn/Trp (OR=3.45, 1.76-6.74 C195%; p=0.001), Neopterine (OR=4,28, 1.95-9.42; p=0.001), Par index (Pyridoxic acid/(Pyridoxal 5' phosphate + Pyridoxal)) (OR=3.34, 1.19-9.39) were risk for pSS. **Conclusion.** KTR, neopterine and Par index are biomarkers for pSS and they are associated with most clinical manifestation.

## **P-84**

## Unexplored functions of indoleamine 2,3 dioxygenase-1 and -2 (IDO1 and IDO2) in Sjögren's syndrome

S. Nayar<sup>1</sup>, C. Vitali<sup>2</sup>, J. Campos<sup>1</sup>, C.D. Buckley<sup>1</sup>, B. Fisher<sup>1</sup>, S. Bowman<sup>1</sup>, N.D. Papa<sup>2</sup>, F. Barone<sup>1</sup>

<sup>1</sup>Rheumatology Research Group, University of Birmingham, UK, <sup>2</sup>Sezione di Reumatologia, Istituto San Giuseppe, Italy.

Background/Objective. Autoimmunity is not only overt activation of inflammatory pathways but also disruption in immune-modulatory pathways. The question which is recently being considered while developing biologic treatment for various autoimmune diseases is will understanding the mechanism of immune-modulation and its disruption in autoimmunity help in restoring THE balance? Indoleamine 2,3 dioxygenase (IDO) is believed to promote immune tolerance via tryptophan catabolism that suppresses antigen-driven T-cell responses, either directly or by activating regulatory T cells. There are two isoforms identified IDO1 and IDO2. Serum tryptophan/ kynurenine ratios are increased in patients with chronic diseases such as Siögren's Syndrome (SS) and rheumatoid arthritis which implicates IDO in autoimmune disease pathogenesis. However, the relative roles played by IDO1 and IDO2 in modulating immune responses remains controversial. In CIA, IDO1 inhibition induces worsening of disease whilst IDO2-null mice display less aggressive joint inflammation in K/BxN arthritis. In this work we have investigated the implications of IDO1 and IDO2 activation during salivary gland (SG) inflammation in SS.

**Methods.** Immunofluorescence (IF) for IDO1 and IDO2 was done on SG biopsies from patients with SS and non-specific chronic sialoadenitis (NSCS). Inducible and resolving mouse model of SS was used to explore relative roles of IDO1 and IDO2 *in vivo*. Murine SGs were studied by IF, flow cytometry/CyTOF and qPCR.

**Results.** Significant expression of IDO was detected in SS biopsies, as compared to NSCS controls. Interestingly, IDO2 expression significantly correlated with the presence of lymphocytic aggregates. IDO1 was primarily detected in non-hematopoietic cells and plasmacytoid dendritic cells whereas

Gene expression profiling in inflamed murine SGs taken at different stages of the inflammatory process demonstrated IDO2 upregulation during the early and peak phases of inflammation. In contrast, IDO1 showed significant increase towards the resolution of inflammation. Differential inhibition of IDO1 versus IDO2 with Methyl-DL-Tryptophan (1-MT) given at different stages of inflammation clearly showed that IDO2 had a pro-inflammatory role wherein it was important for induction of Th17 cells and aggravation of the inflammatory process. In contrast, 1-MT treatment at later time-points (coinciding with increased IDO1 expression) resulted in persistence/retention of lymphocytic aggregates. This anti-inflammatory IDO1 function was mediated by impairing the survival/proliferation of lymphocytes *in situ*.

**Conclusions.** These data demonstrates engagement of IDO1 and IDO2 in different cell populations and moreover very distinct functions in SS pathogenesis.

#### P-85

## RANK/RANK-ligand interaction regulates pathogenic T cell recruitment in Sjögren's syndrome

S. Nayar<sup>1</sup>, B.A. Fisher<sup>1</sup>, D. Gardner<sup>1</sup>, J. Campos<sup>1</sup>, A. Dumusc<sup>1</sup>, C. Smith<sup>1</sup>, V. Iannizzotto, C.D. Buckley<sup>1</sup>, S. Bowman, C. G. Mueller, F. Barone<sup>1</sup>. <sup>1</sup>Rheumatology Research Group, University of Birmingham, UK, <sup>2</sup>Institut de Biologie Moléculaire et Cellulaire, Strasbourg, France.

**Background.** The RANKL (ligand)-RANK-OPG triad, members of the TNF(R) superfamily, is implicated in lymphoid organ development and bone homeostasis. It has recently been demonstrated that RANK-activated astrocytes release CCL20 and attract T cells to the central nervous system in a model of Multiple Sclerosis and that transgenic RANK expression in the skin promotes aberrant epithelial cell proliferation and is sufficient to induce ectopic formation of tertiary lymphoid structures (TLS). Ductal epithelial cells (SGEs) have been implicated in Sjögren's Syndrome (SS) pathogenesis where they mediate immune recruitment by expression of pro-inflammatory chemokines and support the formation of pre-malignant myoepithelial lesions.

**Objective/Methods.** A combination of human and mouse studies were used to address the role of RANK-RANKL interaction in primary (p) SS. Salivary glands (SGs) and saliva samples from patients recruited in the OASIS cohort (University of Birmingham) were studied to evaluate this pathway in human disease. Consecutive stimulated saliva samples (n=69) were analysed using Proseek Multiplex INF<sup>96x96</sup>, covering 92 unique inflammation-related protein biomarkers. Taking advantage of a viral induced model of pSS we studied the effect of this pathway with a RANKL blocking antibody and by inducing gain of function with direct cannulation in the salivary glands of recombinant RANKL. Murine SGs were studied by immunofluo-rescence, flow cytometry and qPCR on total tissue and sorted cells.

**Results.** Fourteen proteins in saliva were significantly separated between pSS and sicca controls, and elevated levels of just two proteins, RANKL and TNF $\beta$ , could classify pSS or sicca with 75% accuracy. Levels of salivary RANKL and CCL20 were strongly correlated (r=0.6; *p*<0.01). We demonstrated that both human and murine inflamed SGEs upregulate both RANK and CCL20, a chemokine known to recruit pathogenic T cells. Upregulation of RANKL was found in human Th2 cells, classically associated with humoral responses and germinal centre (GC) formation. SGs from mice treated with anti-RANKL antibody showed decreased epithelial proliferation, reduced T cell infiltration and defective TLS establishment. On the contrary, viral infected SGs treated with recombinant RANKL showed increased T cells infiltration, CCL20 expression and enhanced differentiation of GC B cells.

**Conclusions.** *In vivo* RANK/RANKL interaction mediates recruitment of activated T cells that are skewed toward a Th2 phenotype. These, in turn, will favour the establishment of TLS in the SG. Those data were confirmed in human pSS, where expression of RANK is found in inflamed epithelium and RANKL detection in saliva is able to differentiate patients with pSS from sicca controls, thus candidates this pathway both for drug targeting and patient stratification.

**P-86** 

#### T follicular-helper cells (TFH) enrichment and follicular T regulatory cells (TFR) exclusion from ectopic germinal centers in salivary glands of Sjögren's Syndrome patients.

E. Pontarini<sup>\*1</sup>, W. Murray-Brown<sup>1</sup>, C. Croia<sup>1</sup>, E. Corsiero<sup>1</sup>, Liliane Fossati-Jimack<sup>1</sup>, E. Astorri<sup>1</sup>, D. Lucchesi<sup>1</sup>, N. Lepse<sup>1</sup>, N. Sutcliffe<sup>2</sup>, A. Tappuni<sup>3</sup>, C. Pitzalis<sup>1</sup>, M. Bombardieri<sup>1</sup>

<sup>1</sup>Centre of Experimental Medicine and Rheumatology, William Harvey Research Institute, London, United Kingdom <sup>2</sup>Barts Health, <sup>3</sup>Institute of Dentistry, Queen Mary University of London.

**Background.** Lymphocytic aggregates in the salivary glands (SG) of Sjögren's syndrome (SS) can organize in ectopic lymphoid structures (ELS) forming functional germinal centers (GCs), which are linked to the development of MALT lymphoma (MALT-L). T follicular-helper cells (Tfh) and follicular T regulatory cells (Tfr) are specialized CD4<sup>+</sup> T helper cells that positively and negatively regulate, respectively, the magnitude of the GC response and possibly the development of autoimmunity.

**Objectives.** To characterize the infiltration of Tfh and Tfr in the SG of patients with SS in the presence/absence of ectopic GCs and MALT-L.

**Methods.** SG biopsies with matching histology and RNA from 37 SS and 38 non-specific chronic sialadenitis (NSCS) patients were stratified as ELS-/ELS+ based on CD3/CD20/CD21/CD138 immunostaining (IHC). Histological samples and mRNA from 12 parotid MALT-L were also studied. Gene expression was performed with Taqman rt -PCR.

Multicolor immunofluorescence/confocal microscopy for CD3, CD4, CD45RO, ICOS, PD1, BCL6 and FoxP3 was used to identify Tfh and Tfr. Results. Tfh cells (CD4+CD45RO+PD1+ICOS+Bcl6<sup>+</sup>) and Tfr cells (CD4+CD45RO+PD1+ ICOS+FoxP3+) are significantly increased in the ELS+ SG tissues from SS patients vs ELS- and NSCS. Tfh cells densely infiltrate the B cell rich areas and preferentially localize within ectopic GCs in the SG tissues. Furthermore, Tfh infiltration correlated with SG IL-21 mRNA expression, which in turn was strongly correlated with CD3, CD20 and CD138 IHC scores and with CXCL13, LTb, BAFF, AID and Pax5 gene expression. Finally, MALT-L samples displayed 10-fold higher IL-21 mRNA and twice as much PD1+ICOS+BCL6+ Tfh-cells/field compared to ELS+ SS samples. The Tfh:Tfr ratio in ELS+ SG was approximately 2:1. Interestingly, while in tonsils Tfr were routinely detected within GC in ELS+ SG Tfr were predominantly excluded from the B cell follicles and accumulated in the T cell rich areas at the periphery of the B-cell aggregates. Conclusions. Within the SG of SS patients Tfh cells closely segregate with lesional IL-21 expression, localize within ELS and are strongly enriched during MALT-L development. Conversely, although Tfr cells are also present in ELS+ SG in SS patients, they are excluded from ectopic GCs. This suggests that Tfr in SS SG fail to exert their physiological immunoregulatory function in controlling the magnitude of the GC response and B cell autoreactivity, as observed in secondary lymphoid organs.

Acknowledgements. This work was supported by project grants from the Medical Research Council (MR/N003063/1 to MB) and Arthritis Research UK (grant 20089 to MB). Disclosure of Interest. E. Pontarini: None declared, W. Murray-Brown: None declared, C. Croia: None declared, E. Astorri: None declared, D. Lucchesi: None declared, N. Lepse: None declared, N. Sutcliffe : None declared, A. Tappuni: None declared, C. Pitzalis: None declared, M. Bombardieri Consultant for: Amgen/Medimmune, GSK and UCB.

### **P-87**

## A preliminary exploration of the role of follicular T helper cell subsets in primary Sjögren's syndrome

Han Wang, Yuebo Jin, Qingwen Wang, Jing He.

<sup>1</sup>Department of Rheumatology and Immunology, Peking University Shenzhen Hospital, Shenzhen, China. <sup>2</sup>Department of Rheumatology and Immunology, Peking University People's Hospital, Beijing, China.

**Objective.** Primary Sjögren's syndrome(pSS) is an autoimmune disease characterized by the production of autoantibodies due to abnormal humoral immunity. Follicular T helper cells (Tfh) play an important role in the pathogenesis of autoimmune diseases by directing the differentiation of B cells into memory B and plasma cells in secondary lymphoid follicles. To examine abnormalities of Tfh cells in SS patients, the percentage of Tfh cells were compared to particular clinical manifestations in patients with pSS.
**Methods.** Peripheral blood was collected from 57 SS patients and 16 healthy controls. Tfh cell subsets were analyzed via flow cytometry. The clinical data and laboratory parameters were collected. Serum levels of cytokines were detected by ELISA.

**Results.** The proportion of Tfhem cells in peripheral blood of patients with SS was significantly higher than that of health controls  $(24.08\%\pm3.57\% \text{ vs.} 8.7\%\pm2.0\%, p<0.05)$ . The frequency of Tfhem cells was correlated with sCD25, Cre, eGFR, Platelet count,  $\gamma$ -globulin (rscd25=0.337; rCre=0.338; reGFR=-0.365; rPlt=-0.266;r  $\gamma$ -globulin =0.280; p<0.05). There was a significantly positive correlation between Tfh17 and IgA (rIgA=0.334; p<0.05); Tfh1 and IgG (rIgG=0.325; p<0.05).

**Conclusion.** Tfh cells are elevated along with increases in sCD25, IgA, IgG and decreases in platelet counts and GFR in patients with SS. Tfh cells may play an essential role in the development of SS and sustaining antibody production, which ultimately results in the activation of disease and organ damage, esp. kidney.

#### **P-88**

# A potential role of NR4A2 overexpression in CD4<sup>+</sup> T cells in the pathogenesis of Sjögren's syndrome

Hiroyuki Takahashi, Hiroto Tsuboi, Hiromitsu Asashima, Hanae Kudo, Yuko Ono, Saori Abe, Yuya Kondo, Isao Matsumoto and Takayuki Sumida. Department of Internal Medicine, Faculty of Medicine, University of Tsukuba

**Backgound.** In Sjögren's syndrome (SS), emerging roles of Th17 cells in the pathogenesis were recently suggested in some studies. Among Th17-related molecules, nuclear receptor subfamily 4, group A, and member 2 (NR4A2), was reported to promote Th17 differentiation via IL-21 production in an animal model of experimental autoimmune encephalomyelitis. The purpose of this study is to clarify a potential role of NR4A2, which was up-regulated in labial salivary glands (LSGs) of patients with SS, in the pathogenesis of SS.

#### Methods.

- Gene expression was analyzed by DNA microarray in LSGs of SS patients, IgG4-related disease (IgG4-RD) patients and healthy controls (HC). Validation analysis of a differentially expressed gene (DEG) upregulated in SS was performed by quantitative PCR (qPCR).
- Protein production of the DEG in LSGs of SS patients and IgG4-RD patients was examined by immunofluorescence staining (IF).
- Functional analysis of the DEG was performed using peripheral CD4<sup>+</sup>T cells of SS patients.
- NR4A2 expression in CD4<sup>+</sup>T cells in lymphoid tissue of a T cell specific RORγt transgenic (Tg) mouse, exhibiting SS-like spontaneous sialadenitis, was analyzed.

#### **Results**.

- Among 1320 DEGs up-regulated in SS, NR4A2 up-regulation in LSGs of SS patients was validated in comparison with those of IgG4-RD patients by qPCR.
- IF of LSGs revealed higher expression of NR4A2 in CD4<sup>+</sup> T cells in SS patients than in IgG4-RD patients and localization of NR4A2 in nuclei of IL-17-producing cells.
- 3) NR4A2 mRNA expression in peripheral CD4<sup>+</sup> T cells was significantly higher in SS patients than in HC. Population of IL-17-producing peripheral CD4<sup>+</sup> T cells after Th17 polarization in vitro was significantly increased in SS patients than in HC and significantly correlated with NR4A2 mRNA expression at baseline. Protein expression of NR4A2 analyzed by cellular IF was localized in nuclei of CD4<sup>+</sup> T cells specifically under Th17-polarizing conditions in comparison with Th1 and Th0 conditions. Nuclear NR4A2 expression in Th17-polarized CD4<sup>+</sup> T cells analyzed by densitometry for cellular IF was significantly increased in SS patients compared with HC. In addition, an inhibitor of importin-β, importazole, inhibited nuclear transport of NR4A2 and Th17 polarization along with IL-21 mRNA expression in CD4<sup>+</sup> T cells under Th17-polarizing conditions.
- 4) NR4A2 mRNA expression in CD4<sup>+</sup> T cells in spleen and thymus was up-regulated in a RORγt Tg mouse compared with a wild-type mouse. Conclusion. NR4A2 seems to promote Th17 polarization via increased expression and intranuclear localization in CD4<sup>+</sup> T cells of SS patients, which could play a critical role in the pathogenesis of SS.

#### **P-89**

#### Sjögren's syndrome minor salivary gland CD4<sup>+</sup> T cells associate with oral disease features and have a T follicular helper-like transcriptional profile

Michelle L. Joachims<sup>1</sup>, Kerry M. Leehan<sup>1</sup>, Mikhail G. Dozmorov<sup>1,\*</sup>, Zijian Pan<sup>1</sup>, Astrid Rasmussen<sup>1</sup>, Lida Radfar<sup>2</sup>, David M. Lewis<sup>2</sup>, Donald U. Stone<sup>3,4</sup>, Kiely Grundahl<sup>1</sup>, R. Hal Scofield<sup>1,5,6</sup>, Christopher J. Lessard<sup>1</sup>, Jonathan Wren<sup>1</sup>, Kathy L. Sivils<sup>1</sup>, Jacen Maier-Moore<sup>1,†</sup> and A. Darise Farris<sup>1</sup>.

<sup>1</sup>Arthritis and Clinical Immunology Program, Oklahoma Medical Research Foundation (OMRF), Oklahoma City, OK. <sup>2</sup>College of Dentistry, University of Oklahoma Health Sciences Center (OUHSC), Oklahoma City OK. <sup>3</sup>Department of Ophthalmology, Johns Hopkins University, Baltimore, MD. <sup>4</sup>King Khaled Eye Specialist Hospital, Riyadh, KSA. <sup>5</sup>Department of Medicine, OUHSC, Oklahoma City, OK. <sup>6</sup>Department of Veteran's Affairs Medical Center, Oklahoma City, OK, USA.

\*Current affiliation: Department of Biostatistics, Virginia Commonwealth University, Richmond, VA, USA

<sup>†</sup>Current affiliation: Clinical Laboratory Science Program, University of Texas at El Paso, El Paso, TX, USA

**Background.** The predominant salivary gland (SG) T cell types contributing to disease in Sjögren's syndrome (SS) are unclear. This study assessed the frequency and number of SG CD3<sup>+</sup> T cell subtypes for association with SS disease features and compared the SG CD4<sup>+</sup> T cell transcriptome of primary SS subjects to that of sicca controls not meeting criteria for SS.

**Methods.** CD3<sup>+</sup> T cells from SG biopsy tissue of subjects with primary SS and non-SS sicca were evaluated for proportion (n=45 SS, n=66 non-SS) and number/mg biopsy tissue (n=34 SS, n=56 non-SS) of T cell subsets defined by CD3, CD4, CD8 and CD45RA. Memory CD4<sup>+</sup>/CD8<sup>+</sup> T cell ratios were evaluated for correlation with clinical and oral disease parameters. Sorted memory CD4<sup>+</sup> T cells from a subset of focus score positive SS cases (n=17) and focus score negative non-SS controls (n=15) were evaluated for global gene expression by microarray.

**Results.** Proportions of CD4+CD45RA<sup>-</sup>T cells (mean±SEM SS: 31.6%±2.0, non-SS: 20.0%±1.1, p<0.0001) but not that of other CD3<sup>+</sup> T cell subsets were increased in SS cases compared to non-SS sicca controls. Ratios of SG CD4+/CD8<sup>+</sup> memory T cells positively correlated with SG focus score (r=0.49, p<0.0001), morphologic area of SG fibrosis (r=0.35, p=0.006), and vanBjsterveld corneal damage score (r=0.37, p<0.0001), with relationships remaining after age correction. Differentially expressed (DE) genes in SS cases versus non-SS sicca controls were enriched for T folicular helper (Tfh), interferon, T cell homeostasis, resistance to apoptosis, atypical lymphoid trafficking and elevated inflammatory response pathways but not Th17 profile. Predicted upstream drivers of the DE genes included CXCL13, CD40/CD40 ligand and Bcl6, while predicted decreased effects included FoxP3, Fas, STAT6 and mTOR.

**Conclusions.** Proportion and number of SG memory CD4<sup>+</sup> T cells selectively associate with key SS disease features, and SG memory CD4<sup>+</sup> T cells are enriched for a predominant Tfh-like cell profile.

#### **P-90**

# Single cell TCR analysis of Sjögren's syndrome salivary gland CD8<sup>+</sup> T cells reveals extensive clonal expansions and the presence of viral-specific cells

Michelle L. Joachims<sup>1</sup>, Christina Lawrence<sup>1</sup>, Richard Pelikan<sup>1</sup>, Kerry M. Leehan<sup>1,2</sup>, Lida Radfar<sup>3</sup>, David Lewis<sup>4</sup>, Astrid Rasmussen<sup>1</sup>, R. Hal Scofield<sup>1,2,5</sup>, Kiely Grundahl<sup>1</sup>, Kathy L. Sivils<sup>1,2</sup>, Linda F. Thompson<sup>1</sup>, A. Darise Farris<sup>1,2</sup> <sup>1</sup>Arthritis and Clinical Immunology Program, Oklahoma Medical Research Foundation, Oklahoma City, OK; <sup>2</sup>Department of Pathology, College of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City, OK, <sup>3</sup>Department of Oral Diagnosis and Radiology, College of Dentistry, University of Oklahoma Health Sciences Center, Oklahoma City, OK, <sup>4</sup>Department of Oral Pathology, College of Dentistry, University of Oklahoma Health Sciences Center and <sup>5</sup>Section of Endocrinology and Diabetes, College of Medicine, University of Oklahoma Health Sciences Center, Oklahoma Oklahoma Health Sciences Center and <sup>5</sup>Section of Endocrinology and Diabetes, College of Medicine, University of Oklahoma Health Sciences Center, Oklahoma

**Background.** Sjögren's syndrome (SS) is an autoimmune disease characterized by lymphocytic infiltration of the salivary and lacrimal glands, severe dry eyes and mouth, fatigue and musculoskeletal pain. How the T cell-dominated salivary gland (SG) inflammation is connected to pathologic and clinical features of SS is poorly understood. We reported that SG CD4<sup>+</sup> T cell clonal expansion is antigen-driven and correlated with reduced sali-

vary flow and increased SG fibrosis. To determine whether the extent of SG CD8<sup>+</sup> T cell clonal expansion is related to pathologic features of disease, we evaluated the SG CD8<sup>+</sup> TCR repertoire of the same subjects.

**Approach.** Multiplex single cell RT-PCR was used to retrieve paired TCR  $\alpha$  and  $\beta$  sequences from SG and peripheral blood (PB) memory CD8<sup>+</sup> T cells from 11 subjects meeting the 2016 ACR/EULAR classification criteria for primary Sjögren's syndrome. The extent of SG and PB CD8<sup>+</sup> T cell clonal expansion was compared between SG and PB and evaluated for relationships with pathologic and clinical disease features using Mann-Whitney and Spearman rank correlation tests.

Results. Our cohort repertoire consisted of over 2,800 TCR sequences isolated from a median of 81 (range 15-119) SG and 100 (range 40-146) PB cells evaluated per subject. Clonal expansions of SG and PB CD8+ T cells were detected in all subjects. Expansions were extensive in both tissue compartments, ranging from 8.3-66.3% in SG (mean 34.9) and 5-48.5% in PB (mean 23.1). Although the percentage of clonally expanded CD8+ T cells did not differ significantly between SG and PB, there were significantly higher levels of clonally expanded SG CD8+ T cells compared to SG CD4+ T cells from the same individuals. Further, many subjects exhibited large expansions, with 60% of individuals having at least one clonal expansion of 4 or more cells. In contrast to our prior observations in CD4+ T cells, the degree of SG CD8+ T cell clonal expansion did not significantly correlate with measures of oral disease. However, CD8+T cell expansions were more extensive in patients with ESSDAIs ≥2. Identical clonal expansions were detected in SG and PB from 5/11 patients (45%), indicating significant trafficking of CD8+ T cells in both tissues. Further, several clonotypes of expanded cells in SG and PB were identified as EBV viral-specific, indicating the presence of viral-specific CD8 T cells in the SG of SS patients

**Conclusion.** Clonally expanded CD8<sup>+</sup> T cells were abundant in both the SG and PB of SS patients, but did not correlate with measures of oral disease. Known viral-specific T cells were found in SG and PB from SS patients, adding evidence for viral-initiated or -driven CD8<sup>+</sup> T cell proliferation in SS.

### P-91

#### Transcriptomics of CCR9-expressing pathogenic T helper cells from primary Sjögren's syndrome patients reveal dysregulated pathways involved in effector T cell function

SLM Blokland<sup>1,2</sup>, CGK Wichers<sup>1,2</sup>, A Pandit<sup>1,2</sup>, AA Kruize<sup>1</sup>, JAG van Roon<sup>1,2</sup>,TRDJ Radstake<sup>1,2</sup>.

<sup>1</sup>Rheumatology & Clinical Immunology, University Medical Center Utrecht, The Netherlands. <sup>2</sup>Laboratory of Translational Immunology, Department of Immunology, University Medical Center Utrecht, Utrecht University, The Netherlands.

**Background.** CCR9+ Th cells produce large amounts of IFN- $\gamma$  and IL-10 (Papadakis JI 2003), lack CXCR5 expression but have features similar to T follicular helper cells and the recently described pathogenic PD1+CXCR5-cells, including expression of ICOS, PD-1, IL-21 and Bcl6, but no CXCR5 expression (McGuire Immunity 2011, Rao Nature 2017). CCR9 T helper (Th) cells and their ligand CCL25 are elevated in salivary glands of primary Sjögren's syndrome (pSS) patients (Blokland A&R 2017). Since CCR9 Th cells strongly induce antibody production and robustly respond to IL-7 (Blokland A&R 2017), which is indicated to play an essential role in pSS pathogenesis (Van Roon A&R 2013) and in formation of ectopic lymphoid structures (Seo J Virol 2014), these cells may play an important role in pSS regulation of CCR9 Th cells in pSS patients.

Iminunopaniously. Inc. goal of the regulation of CCR9 Th cells in pSS patients. **Methods.** CCR9<sup>+</sup>, CXCR5<sup>+</sup> and CCR9-CXCR5<sup>-</sup> Th cells were sorted from peripheral blood of 7 healthy individuals and 7 pSS patients and RNA sequencing of these sorted cell subsets was performed. Computational analysis was used to investigate significantly differentially expressed genes (DEG) and to identify gene expression networks. Pathway enrichment analysis was performed in order to assess differentially regulated pathways. Target genes identified using these analyses are being validated on protein level by flow cytometry. Knockdown experiments will assess the functional role of identified targets.

**Results.** The sorted Th subsets could robustly be distinguished based on their transcriptomes. In the CCR9<sup>+</sup> Th cell subset 2777 DEG (1249 up and 1528 down) were identified between healthy controls and pSS patients, and 1416 and 1077 in the CXCR5<sup>+</sup> and CCR9-CXCR5<sup>-</sup> subsets, respectively. Using network analysis 15 modules were constructed, consisting of genes showing coherent expression patterns. Four modules of interest were selective dbased on pathway enrichment analysis, revealing pathways involved in *e.g.* cytokine and chemokine production, proliferation and migration. DEG of interest within these networks were selected, including upregulated ex-

pression of integrin  $\alpha E$ , integrin  $\alpha 1$  and downregulation of regulatory T cell associated genes FoxP3 and IL2RA. Expression of these markers is being validated using flow cytometry. In addition, knockdown of predicted key transcription factors is studied to reveal their role in the pathogenic potential of CCR9 Th cells.

**Conclusion.** Transcriptomic analysis of CCR9 Th cells from pSS patients revealed multiple dysregulated pathways previously shown to be involved in increased effector T cell function. Upregulation of genes associated with pathogenicity and downregulation of regulatory T cell associated genes were found in pSS patients. Targeting predicted key molecules might reveal (novel) therapeutic targets to halt pathogenic processes induced by CCR9 Th cells.



**A.** Multidimensional scaling (MDS) of transcriptomes distinguished CCR9+, CCR5+ and CXCR5-CCR9- (DN) T helper subsets.

B. Network analysis reveals modules consisting of coherent differentially expressed genes.

C. Examples of up (ITGA1) and downregulated (FOXP3) genes in CCR9+ Th cells from pSS patients.

(HC: healthy control; pSS: primary Sjögren's syndrome; DN: double negative, gene expression is presented in normalized read counts (Iog2)). Blokland *et al.*, ISSS 2017 abstract.

# P-92

#### Single cell based phosphorylation profiling identifies alterations in Toll-like receptor 7 and 9 signaling in patients with primary Sjögren's syndrome

Richard Davies<sup>1</sup>, Irene Sarkar<sup>1</sup>, Daniel Hammenfors<sup>1,2</sup>, Brith Bergum<sup>1</sup>, Petra Vogelsang<sup>1</sup>, Sonia Gavasso<sup>3</sup>, Johan G. Brun<sup>2,4</sup>, Roland Jonsson<sup>1,2</sup>, Silke Appel<sup>1</sup>. <sup>1</sup>Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, Nergen, Norway. <sup>2</sup>Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>3</sup>Department of Clinical Science, University Hospital, Bergen, Norway. <sup>4</sup>Department of Clinical Science, University of Bergen, Norway.

**Background.** Primary Sjögren's syndrome (pSS) is associated with polymorphisms and mRNA expression profiles that are indicative of an exaggerated innate and type I IFN immune response. Excessive activation potential of signaling pathways may play a role in this profile, but the intracellular signaling profile of the disease is not well characterized.

**Methods.** To gain insights into potential dysfunctional intracellular signaling profiles of pSS patients, we conducted an exploratory analysis of MAPK/ERK and JAK/STAT signaling networks in peripheral blood mon-

onuclear cells (PBMC) from 25 female pSS patients and 25 female agematched healthy donors using phospho-specific flow cytometry. We analysed unstimulated samples, and samples during a 4-hour time period following activation of Toll-like receptor (TLR) 7 and 9. Expression levels of MxA, IFI44, OAS1, GBP1 and GBP2 in PBMC were analysed by real-time PCR. We will confirm the flow cytometry results using mass cytometry.

**Results.** Principal component analysis (PCA) showed that basal phosphorylation profiles could be used to differentiate pSS patients from healthy donor samples through stronger phosphorylation in NK and T cells relative to B cells. Stimulation of PBMC with TLR7 and -9 ligands showed significant differences in the phosphorylation profiles between samples from pSS patients and healthy donors. Including clinical parameters such as extraglandular manifestations (EGM), PCA revealed stronger responses through NF- $\kappa$ B and Stat3 S727 in EGM-negative patients than in patients with EGM and healthy donors. In addition, 70% of the patients had a positive IFN score. These patients differed from the IFN score-negative patients regarding their phosphorylation profiles. Mass cytometry data will be shown to confirm this.

**Conclusion.** We here report increased signaling potential in PBMC from pSS patients after TLR7 and -9 stimulation, mainly through STAT3 S727 and NF- $\kappa$ B in B cells, which correlates with a type I IFN signature.

### P-93

#### DNA microarray analysis of labial salivary gland in Sjögren's syndrome indicates a role for innate immune responses in its pathogenesis via Toll like receptor 8

Mizuki Sakamoto<sup>1</sup>, Masafumi Moriyama<sup>1,2</sup>, Keiko Oyama<sup>1</sup>, Akihiko Tanaka<sup>1</sup>, Takashi Maehara<sup>3</sup>, Sachiko Furukawa<sup>1</sup>, Miho Ohta<sup>1</sup>, Masaki Yamauchi<sup>1</sup>, Noriko Ishiguro<sup>1</sup>, Haque A. S. M. Rafiul<sup>1</sup>, Akira Chinju<sup>1</sup>, Keita Mochizuki<sup>1</sup>, Ryusuke Munemura<sup>1</sup> Jun-Nosuke Hayashida<sup>1</sup>, and Seiji Nakamura<sup>1</sup>.

<sup>1</sup>Section of Oral and Maxillofacial Oncology, Division of Maxillofacial Diagnostic and Surgical Sciences, Faculty of Dental Science, Kyushu University, Fukuoka, Japan. <sup>2</sup>OBT Research Center, Faculty of Dental Science, Kyushu University, Fukuoka, Japan. <sup>3</sup>Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts, USA.

**Background.** Sjögren's syndrome (SS) is an autoimmune disease characterized by lymphocytic infiltration into the salivary and lacrimal glands with concomitant autoantibody production and destruction of the glandular tissue. Recent studies suggest that Toll like receptors (TLRs) plays a important rule in innate immune responses and associate with pathogenesis of autoimmune diseases including Systemic lupus erythematosus and Psoriasis. However, the involvement of TLRs in the pathogenesis of SS is still not clear. In this study, we thus addressed to identify the disease-associated genes, especially TLR related molecules.

**Methods.** Gene expression was analyzed by using DNA microarray in labial salivary gland (LSG) from patients with typical SS (n=3) and healthy controls (n=3). DNA microarray analysis was performed in three groups to screen for TLR family (TLR1-TLR10). Up-regulated TLRs were validated by real-time PCR and immunohistochemical staining in SS (n=19), and healthy controls (n=10).

Results. Gene expression patterns in the 2 groups were quite different from each other by the pvclust method and principal components analysis. In SS, 2361 up-regulated genes and 1388 down-regulated genes were identified (adjust *p*-value<0.01, and ratio  $\geq 2$  or ratio  $\leq 0.5$ ). Functional analysis indicated that the up-regulated genes in SS encoded proteins involved in T/B cell activation and chemotaxis. Regarding to TLR family, the gene expression of TLR1, TLR7, TLR8, TLR9, MyD88, IRF1, IRF7, and IRF8 in SS were up-regulated compered with that of healthy control. PCR validated that the expression of TLR8 in SS was significantly higher than that of healthy controls (p<0.01). TLR8 is considered to play an important role in the innate immune response by recognizing viral RNA, self- RNA, and several classes of small molecule agonists. Furthermore, immunofluorescence double staining and flow cytometry analysis confirmed that the expression of TLR8 almost merged with that of the monocyte/macrophage marker CD68. Conclusions. TLR8 was identified as a disease-associated molecule in SS by DNA microarray. Moreover, TLR8+ monocytes/macrophages might be involved in the pathogenesis of SS

# **P-94**

#### Activated M2 macrophage via its Toll-like receptor 7 contributes to the pathogenesis of IgG4-related disease

Noriko Ishiguro<sup>1</sup>, Masafumi Moriyama<sup>1,2</sup>, Katsuhiro Furusho<sup>1,3</sup>, Sachiko Furukawa<sup>1</sup>, Miho Ohta<sup>1</sup>, Takashi Maehara<sup>1</sup>, Akihiko Tanaka<sup>1</sup>, Masaki Yamauchi<sup>1</sup>, Mizuki Sakamoto<sup>1</sup>, Haque A.S.M. Rafiul<sup>1</sup>, Akira Chinju<sup>1</sup>, Keita Mochizuki<sup>1</sup>, Yuko Ono<sup>1</sup>, Ryusuke Munemura<sup>1</sup>, Jun-Nosuke Hayashida<sup>1</sup>, and Seiji Nakamura<sup>1</sup>

<sup>1</sup>Section of Oral and Maxillofacial Oncology, Division of Maxillofacial Diagnostic and Surgical Sciences, Faculty of Dental Science, Kyushu University, Fukuoka, Japan. <sup>2</sup>OBT Research Center, Faculty of Dental Science, Kyushu University, Japan. <sup>3</sup>Division of Innate Immunity, Department of Microbiology and Immunology, Institute of Medical Science, University of Tokyo, Tokyo, Japan

**Background.** IgG4-related dacryoadenitis and sialoadenitis (IgG4-DS) is a unique inflammatory disorder characterized by the elevation of serum IgG4 and infiltration of IgG4-positive plasma cells in lacrimal and salivary glands (SGs). Regarding the immunological aspects of this disease, it is well known that IgG4 is induced by T helper type 2 (Th2) cytokines such as IL-4 and IL-13. We previously reported that these Th2 cytokines contributed to IgG4 production in IgG4-DS. In addition, recent studies indicated that the activation of innate immunity also plays a key role in the IgG4 production upon stimulation with toll-like receptor (TLR) ligands. In this study, we thus examined the expression of innate immune molecules, especially TLRs in SGs from patients with IgG4-DS.

**Methods.** Gene expression was analyzed by DNA microarray in submandibular glands (SMGs) from patients with IgG4-DS (n=6), chronic sialoadenitis (CS) (n=3), and controls (n=3). TLR family (TLR1-TLR10) was validated by real-time PCR and immunohistochemical staining in SGs from patient with IgG4-DS (n=15), Sjögren's syndrome (SS) (n=15), CS (n=9), and controls (n=9). Up-regulated TLR as TLR7 is considered to express on macrophages and dendritic cells (DCs) in secondary lymphoid organs (SLOs), we examined the distribution of macrophages (CD68, CD163), DCs (CD11c, CD123), and TLR7 in the SMGs from patients with IgG4-RD and normal SLOs such as lymph nodes and tonsils. Finally, the phenotype of human TLR7 (huTlr7) transgenic (Tg) C57BL/6 mice before/after stimulation with TLR agonist (R848) was assessed compared with that of wild-type C57BL-6 mice.

**Results.** In patients with IgG4-RD, TLR4, TLR7, TLR8, and TLR9 were overexpressed. PCR validated the up-regulation of TLR7 in IgG4-RD compared with the other groups. Immunohistochemical analysis confirmed strong infiltration of TLR7-positive cells in the SGs of patients with IgG4-RD. Double immunofluorescence staining showed that TLR7-positive cells mainly co-localized with CD123-positive cells in SLOs. In contract, in IgG4-RD tissues, TLR7-positive cells mainly co-localized with CD163-positive cells.

The mRNA expression of TLR7 in IgG4-RD was positively correlated with that of IL-33, which is a Th2 activating cytokine. In huTlr7 Tg mice, the focus and fibrosis score in SMGs, pancreas, and lungs were significantly higher than those in wild-type mice. Moreover, the concentration of serum IgG and IgG1 in huTlr7 Tg mice was significantly higher than that in wild-type mice and distinctly increased upon stimulation with TLR7 agonist.

**Conclusions.** TLR7-expressing M2 macrophages might promote the activation of Th2 immune responses via the local inflammation with IL-33 secretion in IgG4-DS. At more thorough understanding of the role of TLR7+CD163+ M2 macrophages in IgG4-RD could lead to the establishment of a mouse model of IgG4-RD and to the eventual development of novel pharmacological strategies to interrupt TLR7 or TLR7-downstream signals as a further means of inhibiting disease initiation or progression.

#### P-95

#### Withdrawn

# P-96

#### Th17 cytokines as a potential salivary biomarker for morphological changes in Sjögren's Syndrome

Giovanna P. Florezi<sup>1</sup>; Sheyla B. Bologna<sup>1</sup>; Milena M. de Souza<sup>2</sup>, Sandra G. Pasoto<sup>3</sup>, Silvia V. Lourenço<sup>1</sup>

<sup>1</sup>Department of Stomatology, Dental School, University of Sao Paulo, Sao Paulo-SP, Brazil. <sup>2</sup>Department of Dermatology, Medical School, University of Sao Paulo, Sao Paulo-SP, Brazil. <sup>3</sup>Department of Rheumatology, Medical School, University of Sao Paulo, Sao Paulo-SP, Brazil.

DISCLOSURE: FAPESP 2014 / 11020-5; FAPESP 2017/11806-7

**Background.** The pathophysiology of Sjögren's Syndrome is mainly described as a focal lymphocyte infiltration in exocrine glands. One of the most creditable mechanisms to the development of the disease is the involvement of epithelial cells as a target called "autoimmune epithelitis". The epithelium activation leads to the secretion of pro-inflammatory cytokines which can attract lymphocytes and induce inflammatory responses. These cytokines are mediated by T helper (Th) lymphocytes, being the Th17 subtype abundantly found in minor salivary glands with a severe destruction<sup>1</sup>. These histological changes are often associated with the decrease in salivary flow and xerostomia<sup>2</sup>, however, there's still the necessity of understanding the mechanisms that leads to the salivary gland secretion impairment and the development of a specific biomarker to differentiate SS from other autoimmune diseases.

**Methods.** Twenty-seven (27) patients diagnosed with SS were selected using the American-European consensus criteria. All patients underwent a minor salivary gland biopsy and the morphological analysis considered acinar and ductal alterations and inflammatory infiltrate characterization. A saliva sample from these patients and from 15 healthy volunteers was collected and analyzed by multiplex for the concentration of cytokines mediated by Th17 lymphocytes: CD40L, IFN- $\gamma$ , IL-1 $\beta$ , IL-10, IL-17A, IL-17E/IL-25, IL-17F, IL-21, IL-22, IL-23, IL-31, IL-4, IL-6, TNF- $\alpha$ . These concentrations were then correlated and associated through statistical analysis to the observed histological changes.

**Results.** The main finding in these specimens was the presence of an inflammatory infiltrate with a moderate (55%) to severe (41%) intensity around ductal structures. There was a statically significant difference between the concentrations of IL-17E (p=.0458); IL-17A (p=.0006); IL-21 (p<.0001); CD40L (p<0.0001) in SS saliva and healthy controls. It was possible to observe a significant positive model when correlated the concentration of IL1- $\beta$  and the focus score (p<.000; r=.748) and a decrease in saliva severe acinar atrophy showed a higher concentration of IL-21 (p=.0427), IL-10 (p=.0321), IL-17F (p=.0286) and CD40L (p=.0008). Acinar fibrosis was also found generalized in patients with a higher concentration of IFN- $\gamma$  (p=.0183) and IL-17F (p=.0039).

**Conclusion.** These findings show that IL-21, CD40L, and IL-17 are involved in the salivary gland impairment, and could be potential biomarkers in SS. Other cytokines such as IL-1 $\beta$ , IL-10 and IFN- $\gamma$  may participate in the atrophic, sclerotic and inflammatory development in this disease. These cytokines may be associated with the disease severity, as they were found in a higher concentration in specimens with severe alterations.

#### References

- TZIOUFAS AG, KAPSOGEORGOU EK, MOUTSOPOULOS HM: Pathogenesis of Sjögren's syndrome: What we know and what we should learn. *J Autoimmun* 2012; 39(1–2): 4–8.
- JENSEN SB, VISSINK A: Salivary Gland Dysfunction and Xerostomia in Sjögren's Syndrome. Oral Maxillofac Surg Clin North Am 2014; 26(1): 35-53.

# P-97

#### Clinical significance of chemokine expression in the minor salivary glands of patients with Sjögren's syndrome

Kyung-Eun Lee<sup>1</sup>, Ji-Hyoun Kang<sup>1</sup>, Dong-Jin Park<sup>1</sup>, Kyung Chul Yoon<sup>2</sup>, Ji Shin Lee<sup>3</sup>, Shin-Seok Lee<sup>1</sup>.

<sup>1</sup>Department of Rheumatology, Chonnam National University Medical School & Hospital, Gwangju, Republic of Korea, <sup>2</sup>Department of Ophthalmology, Chonnam National University Medical School & Hospital, Gwangju, Republic of Korea, <sup>3</sup>Department of Pathology, Chonnam National University Medical School & Hospital, Gwangju, Republic of Korea. **Methods.** Sociodemographic data on 106 SS patients were obtained and the glandular and extraglandular manifestations of the disease were documented. In addition, minor salivary gland biopsies were performed and the patients' laboratory findings were analyzed. European League Against Rheumatism SS disease activity index (ESSDAI) values of SS disease activity ity at the time of biopsy and the SS disease damage index (SSDDI) values were also recorded. An immunohistochemical approach was used to semiquantitatively measure the CCL21 and CXCL13 expression in the minor salivary glands.

**Results.** The minor salivary glands of SS patients stained positively for CCL21 and CXCL13 in 46.2% (49/106) and 70.7% (75/106) of all cases, respectively. Higher-level expression of CCL21 and CXCL13 was associated with increases in ESR, IgG and rheumatoid factor levels, as well as anti-SS-A and -SS-B titers. A higher focus score and ESSDAI value at the time of biopsy were also associated with these chemokines. In patients with extraglandular manifestations of SS, the prevalence of lymphadenopathy increased with increasing CCL21 levels.

**Conclusion.** The expression levels of CCL21 and CXCL13 within the lymphocytic infiltrates of SS patients were associated with several laboratory features of the disease as well as lymphadenopathy and the extent of clinical disease activity. CCL21 and CXCL13 levels can therefore serve as useful markers to predict the disease activity and prognosis of patients with SS.

#### **P-98**

#### Distinct and persistent cytokine profiles associated with histopathological features in primary Sjögren's syndrome

Tove Ragna Reksten<sup>1</sup>, Daniel Hammenfors<sup>2</sup>, Johan G. Brun<sup>1,3</sup>, Roland Jonsson<sup>1</sup>, Malin V. Jonsson<sup>4</sup>.

<sup>1</sup>Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, Norway. <sup>3</sup>Section for Rheumatology, Department of Clinical Science, University of Bergen, Norway. <sup>3</sup>Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>4</sup>Section for Oral and Maxillofacial Radiology, Department of Clinical Dentistry, University of Bergen, Norway.

**Background.** The cytokine network in Sjögren's syndrome (SS) is highly dysregulated, with local and systemic overexpression of pro-inflammatory cytokines. We have previously found that serum cytokine levels vary with morphological features in minor salivary glands, particularly with the presence of ectopic germinal centre (GC)-like structures (Reksten *et al.*, 2009). In this longitudinal study we aimed to assess if and which cytokines change over time in SS patients.

**Methods.** Seventy SS patients were recruited in a follow-up study, 43 of whom were classified as GC-, 15 as GC+ and 12 with negative focus score at the time of study inclusion. Serum samples obtained at time of study inclusion and at 8-year follow-up were analysed for cytokines using a Luminex assay (Human Cytokine 25-Plex Panel, Invitrogen, #LHC0009M).

**Results.** Of 25 measured cytokines, overall levels of IL-5, IL-8, IL-10, IL-13, IL-17a, GM-CSF and MIG were significantly increased at follow-up, whereas IL-4, MCP-1 and TNF- $\alpha$  levels were decreased (Wilcoxon p-values ranging between 0.3 and <0.0001. Fig. 1). To assess cytokine level variations in GC-, GC+ and focus score (FS)- patients, we conducted two-way repeated measure ANOVA analyses and identified IL-1RA, IL-13, IL-17, MIP-1 $\alpha$  and TNF- $\alpha$  as significantly associated with GC-status at both time-points (Fig. 2). We also found that GM-CSF, IL-13, IL-8 and MCP-1 levels significantly changed with time across all groups. Further, using mixed linear model analyses we identified between-group differences in Eotaxin (*p*=0.009), IL-12 (*p*=0.04) and MIG (*p*=0.01) level changes with time (Fig. 3). We confirmed GC-status associated with IL-1RA (*p*=0.02), IL-17a (*p*=0.007) and MIP-1 $\alpha$  (*p*=0.04) levels, as well as time effect on GM-CSF (*p*<0.0001), IL-8 (*p*=0.06), IL-13 (*p*<0.0001) and MCP-1 (*p*<0.0001) levels. No differences were observed between patients with normal and hyposalivation.

**Conclusions.** Our study confirms that serum cytokine levels are abnormal in SS, and only vaguely fluctuate with time. GC+ patients had overall higher

serum cytokine levels. With the exception of pro-inflammatory, autoimmunity associated cytokines IL-17a and the IL-23 subunit IL-12p40, chemokines rather than inflammatory cytokines were the discriminating factors between GC-status subgroups both at individual time-points and over time.



Fig. 1. Serum cytokine and chemokine levels were over all relatively constant across all three subgroups between the two time points, though levels of IL-4, MCP-1, and TNF- $\alpha$  dropped slightly, whereas IL-5, IL-8, IL-10, IL-13, IL-17a, GM-CSF, and MIG increased.







Fig. 3. Changes in serum levels of Eotaxin (CCL 11), Monokine induced by gamma interferon (MIG/CXCL9), and interleukin 12 (IL-12) at time of inclusion (S2A) and time of follow-up (S2B) was significantly different between pSS patient with and without ectopic GC-like structures in minor salivary glands (GC+ and GC-) and patients with negative focus score (FS).

# P-99

#### Role of CXCL13 and CXCL12 in Sjögren's syndrome: association with histological, clinical and laboratory features.

Colafrancesco S<sup>1</sup>, Priori R<sup>1</sup>, Pipi E<sup>2</sup>, Campos J<sup>2</sup>, Nayar S<sup>2</sup>, Cerbelli B<sup>3</sup>, Arienzo F<sup>1</sup>, Giordano C<sup>3</sup>, Bombardieri M<sup>4</sup>, Valesini G<sup>1</sup>, Barone F<sup>2</sup>. <sup>1</sup>Department of internal medicine and medical specialties, Rheumatology Unit, Sapienza University of Rome, Rome, Italy. <sup>2</sup>Centre for Translational Inflammation Research, University of Birmingham, Birmingham, UK. <sup>3</sup>Department of Radiological, Oncological and Pathological Sciences, Sapienza, University of Rome, Rome, Italy. <sup>4</sup>Department of Experimental Medicine and Rheumatology, Barts and the London NHS Trust and Barts and the London School of Medicine and Dentistry, London, UK.

**Background.** Ectopic production of the lymphoid chemokines CXCL13 and CXCL12 has been described in tertiary lymphoid structures (TLS) that harbour in the salivary glands of patients with Sjögren's Syndrome (pSS). Whilst CXCL13 expression was found to correlate with clinical features, its potential role as a biomarker to monitor the organization and severity of the salivary gland infiltrate has been hampered by the lack of sensitive tools to describe TLS extent and features.

**Methods.** We studied histological features of the minor salivary glands (MSG) and sera of respectively fifty and seventy (Table) unselected consecutive patients with pSS (AECG criteria) enrolled in the Sapienza University of Rome, Italy. Concentration of CXCL13 and CXCL12 were evaluated by ELISA in patient sera and eleven healthy age-matched controls (HC). Paraffin embedded MSG were studied by haematoxylin/eosin and anti-CD3, anti-CD20, anti-CD21 staining. Images analysis was used to calculate focus score (FS), mean foci area, percentage of infiltration (%i), segregated foci (%SF), %GCs and lymphoepitelial lesions (%LEL). GCs from MSG and tonsils were microdissected and quantitative PCR was used to test CXCL12 and CXCL13 transcripts.

#### Table.

Patients	Serum analysis	Histological analysis
Demographic features	Mean ± SD	Mean ± SD
Mean age	54.6 ±12.5	54.8 ±12.5
Mean age at diagnosis	51.7 ±12.2	52.1 ±12.2
Laboratory/clinical features	Number/70(%)	Number/50(%)
ANA	50/70 (71.4)	37/50 (74)
Anti-Ro/SSA antibodies	28/70 (40)	16/50 (32)
Anti-La(SSB antibodies	20/70 (28.5)	13/50 (26)
Rheumatoid Factor	20/70 (28.5)	12/50 (24)
Hypergammaglobulinaemia	23/70 (32.8)	15/50 (30)
Cryoglobulins	2/70 (2.8)	2/50 (4)
Monoclonal component	8/70 (11.4)	8/50 (16)
Hypocomplementaemia	9/70 (12.8)	5/50 (10)
Leucopenia	12/70 (17.1)	5/50 (10)
Xerostomia	61/70 (87.1)	46/50 (92)
Xerophtalmia	60/70 (85.7)	46/50 (92)
Gland swelling	24/70 /34.2)	15/50 (30)
Lymphoma	8/70 (11.4)	0 (0)



#### Image 2.

Results. Histological analysis unveiled strong correlations between the mean foci area with the %i and the presence of SF; positive correlations were also observed between the %i and both the FS and %SF. This was significantly higher in patients exhibiting SF. The % of SF positively correlated with FS, presence of %GC and %LEL that also correlated with the %i and the %SF (image 1). Mean CXCL13 and CXCL12 serum levels were significantly higher in pSS compared to HC [(124.12±119.73 pg/ml vs 8.9±15.4 pg/ml (p=0.001) and 34.6±54.2 pg/ml vs 2.5±8.3 pg/ml (p=0.05), respectively]. CXCL13 was significantly higher in patients with SF, with GCs and LEL and correlated with the mean foci area, the %i, the FS and the percentage of LEL. Higher CXCL13 levels were associated with the presence of antibodies and other biological findings including hyperglobulinemia. Higher CXCL13 levels were also able to discriminate patients with lymphoma (p=0.009). CXCL12 levels correlated with the FS, %i and % of LEL. Transcript analysis showed no difference in the expression of CXCL13 between MSG and tonsil GC, whilst CXCL12 was found significantly higher in MSG (p<0.0001) (image 2).

**Conclusion.** Our results suggest the utility to expand the parameters of histologic evaluation of MSG, whilst reinforcing the role of the FS as reliable instrument to reflect the severity of inflammation. Accurate analysis of MSG infiltration and foci segregation was able to identify subjects

with increased proliferative risk (as demonstrated by presence of GCs and LELs). We demonstrated that serum CXCL13, but not CXCL12 is a robust biomarker of histological severity of the disease, and is able to stratify patients with lymphoma. The high levels of CXCL12 in MSG GC suggest a differential biology of TLS in the SG, probably implicated in aberrant B cell clone survival that should be further investigated.

# **P-100**

#### Germ-free environment worsens lacrimokeratoconjunctivitis in a mouse model of Sjögren syndrome

De Paiva,  $CS^1$ ; Zaheer,  $M^1$ ; Yu,  $Z^1$ ; Bian<sup>1</sup>, F; Swennes,  $AG^2$ , Britton,  $RA^3$ , Pflugfelder,  $SC^1$ .

<sup>1</sup>Department of Ophthalmology, Baylor College of Medicine, Houston, Texas. <sup>2</sup>Center for Comparative Medicine and Department of Molecular Virology and Microbiology, Baylor College of Medicine, Houston, Texas. <sup>3</sup>Center for Metagenomics and Microbiome Research, Department of Molecular Virology and Microbiology, Baylor College of Medicine, Houston, Texas.

**Background.** To investigate the role of the gut microbiota in the appearance and development of dacryoadenitis in the IL-2 receptor alpha knock-out (CD25KO) model of Sjögren Syndrome.

Methods. Eight-week old germ-free (GF) KO mice were compared to KO mice raised in specific pathogen free (SPF) conditions. Corneal barrier function was assessed by dye staining. Eyes and adnexae were excised and prepared for histology. Conjunctival goblet cell density was counted in PAS stained sections. Total cell infiltrates in lacrimal gland (LG) were visualized in histologic sections. T helper (Th) phenotype in LG and cervical lymph nodes (CLN) of recipients was investigated by flow cytometry. CD4<sup>+</sup>T cells were isolated from CLN and spleens and adoptively transferred into RAG1KO recipients. Four-week-old GF KO mice were gavaged with fecal slurry from C57BL/6 mice, and dry eye phenotype was evaluated 4 weeks later. LG lysates were used to investigate the expression of IFN- $\gamma$ , MHC II, IL-12, TNF- $\alpha$  and IL-1 $\beta$  mRNA transcripts by real-time PCR.

**Results.** GF KO mice have significantly lower number of goblet cells and significantly greater corneal barrier disruption and LG infiltration than KO SPF mice. There was a greater percentage (%) of CD4<sup>+</sup> IFN- $\gamma^+$  in GF KO LG and CLN compared to SPF KO group (7.65±3.26 vs. 5.23±3.13 and 4.90±1.07 vs.3.49±1.07%, *p*<0.05 for both, respectively). Adoptive transfer of GF KO CD4<sup>+</sup>T cells recapitulated the most severe phenotype in RAG-IKO recipients compared to SPF CD4<sup>+</sup>T donors 5 weeks post-transfer. GF KO animals gavaged with normal mouse fecal slurry had a significantly higher number of goblet cells and significantly lower corneal barrier disruption than GF KO mice while displaying decreased LG infiltration score. This was accompanied by a 50% or greater decrease in IFN- $\gamma$ , MHC II, IL-12, TNF- $\alpha$  and IL-1 $\beta$  mRNA levels in the LG of recolonized animals.

**Conclusions.** Lack of commensal bacteria accelerates the onset and severity of dacryoadenitis and generates autoreactive CD4<sup>+</sup>T cells with greater pathogenicity. These results indicate the commensal bacteria or products secreted by them have immunoregulatory properties that protect exocrine glands in the CD25KO SS model.

Support. This work was supported by the NIH/NEY EY026893 (CSDP), Alkek Center for Metagenomics and Microbiome Research (CSDP), Biology of Inflammation Center (SCP and CDP), NIH Training Grant T32-AI053831 (FB), RPB Stein Innovation Award (RAB), RPB Research to Prevent Blindness (SCP), The Oshman Foundation (SCP), William Stamps Farish Fund (SCP), The Hamill Foundation (SCP), and by the Cytometry and Cell Sorting Core at Baylor College of Medicine, which is funded by the NIH NIAID P30AI036211, NCI P30CA125123, and NCRR S10RR024574.

# P-101

#### Analysis of suppressive ability and its mechanisms of rice seeds expressing altered peptide ligands against M3 muscarinic acetylcholine receptor (M3R) induced sialadenitis

Hanae Kudo<sup>1</sup>, Hiroto Tsuboi<sup>1</sup>, Hiromitsu Asashima<sup>1</sup>, Hiroyuki Takahashi<sup>1</sup>, Yuko Ono<sup>1</sup>, Saori Abe<sup>1</sup>, Yuya Kondo<sup>1</sup>, Yuya Wakasa<sup>2</sup>, Fumio Takaiwa<sup>2</sup>, Isao Matsumoto<sup>1</sup> and Takayuki Sumida<sup>1</sup>.

<sup>1</sup>Department of Internal Medicine, Faculty of Medicine, University of Tsukuba. <sup>2</sup>Plant Molecular Farming Unit, Division of Biotechnology, Institute of Agrobaiological Sciences, National Agriculture and Food Research Organization.

Purpose. We previously reported that Rag1-/- mice inoculated with splenocytes from M3 muscarinic acetylcholine receptor (M3R) knockout mice immunized with M3R peptides mixture (N-terminal regions; N1, N2, N3, and three extracellular loops; 1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup>) developed sialadenitis like Siögren's syndrome (M3R induced sialadenitis; MIS). We also found that intravenous administration on day 7 and 10 after the cell transfer of altered peptide ligands (APLs) of N1 and 1st which were T cell epitopes of MIS suppressed MIS. In this study, we aimed to evaluate the suppressive ability and its mechanisms of rice seeds expressing APLs against MIS. Methods.

1) M3R peptides (N1 and 1st), N1-APL, 1st-APL, and fluorescent substance (mCherry) were constructed as the fusion structures with glutelin which expresses in protein bodies of rice seeds. Transgenic rice seeds expressing N1, 1st, N1-APL, 1st-APL, and mCherry were generated.

- 2) Rice seeds expressing mCherry were orally administrated to mice. After 0, 3, 6, 12, 24, and 48 hours, fluorescence in the ileum, mesenteric lymph node (MLN), and spleen were investigated by the fluorescence microscope. Simultaneously, co-localization of mCherry with CD3+T cells and CD11c<sup>+</sup>dendritic cells was also analyzed by immunofluorescent staining.
- 3) 150 mg of rice seeds expressing 30-40 µg of N1, 1st, N1-APL, and 1st-APL, or non-transgenic rice seeds were orally administrated to each of the MIS mice every day from day 7 to day 21 after the cell transfer. The change of saliva volume and pathological findings of salivary glands were compared between treatment groups.

#### Results.

- 1) Expression of M3R peptides (N1 and 1st), N1-APL, 1st-APL, and mCherry was detected in the protein bodies of rice seeds.
- 2) mCherry was detected in the ileum, MLN, and spleen at 3, 6, 12, 24, and 48 hours after orally administration of rice seeds expressing mCherry. mCherry was co-localized with CD11c+dendritic cells, while not with CD3+T cells.
- 3) We are now analyzing the histological findings of MIS treated by rice seeds expressing N1, 1st, N1-APL, and 1st-APL.

Conclusion. After oral administration, mCherry expressed in rice seeds could be transported into MLN and spleen, and then entrapped by CD11c+dendritic cells, with keeping the fluorescence.

#### **P-102**

#### Bone morphogenetic protein 6 receptor inhibition restores salivary gland function in a mouse model of primary Sjögren's syndrome

Hongen Yin<sup>1</sup>, Lovika Kalra<sup>1</sup>, Arif Karim<sup>1</sup>, Zhennan Lai<sup>1</sup>, Maria C, Guimaro1, Lauren Aber1, Blake Warner1, Bill Swaim1, Sandra Afione1, Alexandria Voigt<sup>2</sup>, Cuong Q. Nguyen<sup>2</sup>, Paul Yu<sup>3</sup>, Donald B. Bloch<sup>4</sup> and John A. Chiorini<sup>1</sup> <sup>1</sup>Molecular Physiology and Therapeutics Branch, National Institute of Dental and Craniofacial Research, National Institutes of Health, Bethesda, MD, USA. <sup>2</sup>Department of Pathology and Infectious Diseases, University of Florida, Gainesville, FL, USA. 3Cardiovascular Division, Brigham and Women's Hospital, Harvard Medical School, Boston, MA. 4Center for Immunology and Inflammatory Diseases and the Division of Rheumatology, Allergy, and Immunology of the Department of Medicine, Massachusetts General Hospital, Harvard Medical School, Boston, MA, Boston, MA.

Background. Bone morphogenetic protein 6 (BMP6) plays a critical mechanistic role in decreasing salivary gland dysfunction in primary Sjögren's syndrome (pSS) patients. BMP6 is reported to be over expressed in the salivary glands of over 50% of pSS patients and has been linked to a decrease in salivary gland function in patients. BMP6 signals through type 1 receptors, which results in phosphorylation of SMAD1/5/8 transcription factors that ultimately alter gene expression within the nucleus. Two inhibitors, LDN-212854 and LDN-193189, have been developed to selectively target the ALK2 and ALK3 BMP type 1 receptors. This study examined the ability of these inhibitors to block BMP6 signaling, and their effect on expression of key proteins involved in salivary gland function and inflammation

Methods. BMP6 expression was detected in minor salivary glands (MSG) from female pSS patients by immunofluorescent staining. In vitro water permeability was tested by regulated volume decrease (RVD) assay in BMP6 treated HSG cells with or without LDNs treatment, followed by observation of change of phospho-SMAD (pSMAD)1 expression as detected by Western blotting. BMP6 overexpression mice were generated by retroductal cannulation of AAV5-BMP6 in submandibular salivary glands (SMG) of 6-8wks old female C57/B6 mice. In vivo activity of the ALK inhibitors was tested in C57BL/6.NOD-Aec1Aec2 mice and BMP6 overexpressing mice, which have established SS like disease, by daily i.p. injection for 24 days or 3 days

respectively. Subsequent pilocarpine stimulated saliva flow were assessed during and at the end of the study. Local and systemic immune response was investigated by flow cytometric assay. The levels of pSMAD1, ID3 and aquaporin-5 (AQP-5) expression in SMG from LDN treated C57BL/6. NOD-Aec1Aec2 mice were measured by immunofluorescent staining.

Results. Elevated BMP6 was found in N=43/79 (54.4%) of pSS patients examined in this study. In humans, ALK2 and ALK3 receptors were found on both ductal and acinar cells. In vitro treatment of HSG cells with ALK2/3 inhibitors resulted in decreased BMP6 signaling and SMAD 1/5/8 phosphorylation and led to a recovery of fluid movement. Furthermore, daily treatment of BMP6 overexpressing mice or C57BL/6.NOD-Aec1Aec2 mice with either inhibitor restored salivary flow rate. Associated with this increase in salivary flow was an increased expression of AQP5, a protein critical for membrane water permeability in salivary glands, and decreased pSMAD1 and ID3, which are downstream signaling pathways for ALK2/3. LDN treatment also decreased infiltrating IFN-gamma producing CD4+ T cells in submandibular glands from C57BL/6.NOD-Aec1Aec2 mice.

Conclusions. BMP6 expression is increased in a majority of pSS patients. Treatment with BMP6 inhibitors can reverse the loss of function within the salivary gland as well as decrease inflammation. These findings suggest that inhibition of BMP6 is a promising approach to the treatment of primary Sjögren's syndrome.

#### P-103

#### Targeting B-cell activating factor (BAFF) impairs ectopic lymphoneogenesis in murine models of Sjögren's syndrome

Campos J<sup>1</sup>, Slocombe T<sup>2</sup>, Nayar S<sup>1</sup>, Iannizzotto V<sup>1</sup>, Gardner DH<sup>1</sup>, Buckley CD<sup>1</sup>, Havnes A<sup>2</sup>, Henderson R<sup>2</sup>, Barone F<sup>1</sup>.

<sup>1</sup>Centre for Translational Inflammation Research, Institute of Inflammation and Ageing, College of Medical and Dental Sciences, University of Birmingham, UK. 2Immunoinflammation TAU, GSK, Stevenage, London, UK.

Background. Tertiary lymphoid structures (TLS) characterised by germinal centre formation and B cell proliferation represent the histological hallmark of primary Sjögren's syndrome (pSS). However, the events preceding the formation of such ectopic structures and factors driving their persistence are unknown. Overexpression of BAFF, also known as B cell lymphocyte stimulator (BLyS), in pSS patients has been linked with the presence of autoreactive B cells and autoantibody production (1). Furthermore, in pSS salivary glands BAFF is associated with the expansion of specific B cell subsets, and with B cell repopulation post rituximab treatment (2). In this work we aimed to dissect the dynamics of B cell subsets within tertiary lymphoid structures following BAFF-targeted treatment in both inducible and chronic animal models that mimic the histological features of pSS

Methods. Submandibular salivary glands of C57BL/6 mice were intra-ductally cannulated with luciferase-encoding replication-deficient adenovirus to induce TLS formation as previously described (3). Prior to salivary gland cannulation, mice were treated with two doses (i.p.) of either anti-BLyS mAb or isotype control. Salivary glands were dissected at day 15 postcannulation and TLS formation in both groups was assessed. NOD.B10. H2b mice were similarly treated with anti-BLyS mAb at 26 weeks old and salivary gland infiltrates assessed 21 days later.

Results. Histological analysis of salivary glands from anti-BLyS treated C57BL/6 animals unveiled severely compromised TLS formation. Post anti-BLyS treatment, salivary glands were infiltrated by T cell clusters but only few, and scattered, B cells were present, contrasting with fully developed and organised TLS in the salivary glands of mice treated with isotype control. Significantly lower numbers of B cells, particularly from the B2 subset, as well as plasmablasts, infiltrated salivary glands of anti-BLyS treated mice. However, treatment with anti-BLyS did not affect numbers of infiltrating T cells (both CD4 and CD8), proliferative T cells, or plasma cells in inflamed salivary glands. In a chronic setting, salivary glands from NOD. B10.H2b mice were also infiltrated by significantly lower numbers of B2 B cells following anti-BLvS treatment.

Conclusions. Our data highlights BAFF as a key player in ectopic lymphoneogenesis during inflammation as well as a subset-specific role for BAFF in B cell maturation. Furthermore, these results support future studies of BAFF-targeted therapeutics in pSS. References

1. PERS et al.: Ann NY Acad Sci 2005. 2. PERS et al .: Arthritis Rheum 2007

3. BOMBARDIERI, BARONE et al.: JI 2012.

Clinical and Experimental Rheumatology 2018

#### 14<sup>th</sup> International Symposium on Sjögren's Syndrome

# P-104

Effect of cenerimod, a sphingosine-1-phosphate receptor 1 (S1P1) modulator, on the formation of tertiary lymphoid structures in a mouse model of sSögren's syndrome

S. Nayar<sup>1</sup>, J. Campos<sup>1</sup>, C. Smith<sup>1</sup>, C. D. Buckley<sup>1</sup>, S. Froidevaux<sup>2</sup>, K. Wartha<sup>2</sup>, C. Seemayer<sup>2</sup> and F. Barone<sup>1</sup>.

<sup>1</sup>Rheumatology Research Group, University of Birmingham, UK, <sup>2</sup>Idorsia Pharmaceuticals Ltd, Allschwil, Switzerland.

**Background/Objective.** Tertiary lymphoid structures (TLS) often develop in target tissues of autoimmune diseases (AID) such as systemic lupus erythematosus (SLE), multiple sclerosis (MS), rheumatoid arthritis, and Sjögren's syndrome (SS). These structures consist of aggregates of B and T cells with varying degree of organization and are proposed to promote the generation of autoreactive effector cells and autoantibody production. Modulation of the S1P<sub>1</sub> receptor inhibits egress of pathogenic lymphocytes from lymphoid organs and reduces their availability in circulation. This has proven to be an effective target for the treatment of AID including MS and is currently being considered for early phase clinical trials in SLE and SS. Here we investigated the functional targeting of the S1P<sub>1</sub> receptor in a murine model of SS.

**Method.** Cenerimod, an orally active, selective  $S1P_1$  receptor modulator, was used either preventively (early in inflammation) or therapeutically (established inflammation) in an inducible model of SS to evaluate the efficacy of cenerimod *in vivo*.

Results. Cenerimod induced disaggregation of the lymphocytic structures and resolution of salivary gland (SG) inflammation with a concomitant decrease in focus score, lymphoid structure size and T/B-cell follicular organization. Mice treated with cenerimod displayed significantly decreased T (naïve, central memory and effector) and B (including CD138+ plasma cells) lymphocyte infiltration in cannulated SG, relative to vehicle treated mice. Interestingly, the lymphocytes from cenerimod treated mice exhibited significantly reduced proliferation as well as reduction in pro-inflammatory cytokine RNAs such as IL-17, IL-21, and IL-6. Furthermore, the gene expression profile associated with TLS formation (LT $\alpha$ , LT $\beta$ , TNF- $\alpha$ , CXCL13, CCL19) was less pronounced in cenerimod treated samples. The cervical lymph nodes draining the salivary glands showed a slight reduction in T lymphocytes, but no significant defects were observed in structure, organization and production of lymphoid chemokine/cytokines, suggesting that homeostatic regulation of tertiary and physiological lymphoid organs differentially relies on lymphocyte/stromal cell cross-talk during inflammation.

**Conclusion.** Together, these data demonstrate that the  $S1P_1$  receptor modulator cenerimod regulates TLS in mice and might therefore be a potential treatment option in AID with TLS formation such as SS and SLE. The data also unveil differential requirements for the establishment and maintenance of secondary versus tertiary lymphoid structures.

Acknowledgement. This research is funded by Idorsia Pharmaceuticals Ltd.

### P-105

#### Butyrate suppresses salivary flow rate decrease and lymphocytic infiltration in salivary and lacrimal glands of non obese diabetic mice

Jennifer Lee<sup>1</sup>, Da Som Kim<sup>2</sup>, Ji-Won Kim<sup>1</sup>, Seung-Ki Kwok<sup>1</sup>, Ji Hyun Ju<sup>1</sup>, Sung-Hwan Park<sup>1</sup>

<sup>1</sup>Division of Rheumatology, Department of Internal Medicine, School of Medicine, The Catholic University of Korea, Seoul St. Mary's Hospital, Seoul, Republic of Korea. <sup>2</sup>Rheumatism Research Center, Catholic Research Institute of Medical Science, The Catholic University of Korea, Seoul, Republic of Korea.

**Background.** Gut microbiota has been introduced as an important environmental factor in the pathogenesis of autoimmune inflammatory diseases, such as rheumatoid arthritis and inflammatory bowel disease. Gut microbiota participates in the development of the immune system and produces various metabolites. Metabolites secreted by gut microbiota maintain homeostasis in the gut through various mechanisms, of which butyrate is known to have an anti-inflammatory effect. The purpose of our study was to investigate the effect of butyrate on clinical and histopathologic aspects of Sjögren's syndrome (SS) in non-obese diabetic (NOD) mice, an animal model of SS. **Methods.** Sodium butyrate (1g/kg) or vehicle (saline) was intraperitoneally injected three times per week from 11 weeks after birth. Salivary flow rate (SFR) was addressed on every 2 weeks between 11 and 23 weeks. Histologic analysis was performed on 23 weeks.

**Results.** The SFR of NOD mice in both groups decreased over time. However, SFRs of sodium butyrate-treated mice were greater than those treated with vehicle. Histopathologic evaluation of salivary gland and lacrimal gland showed markedly reduced lymphocytic infiltration in the mice treated with sodium butyrate.

**Conclusion.** Butyrate has suppressive effect on SFR decrease and lymphocytic infiltration in salivary gland and lacrimal gland of NOD mice. Butyrate may be a novel therapeutic approach in SS.

### P-106

# Low-dose IL-2 administration expands regulatory T cells and protects against experimental Sjögren's syndrome

Yang Yao<sup>1,2</sup>, Yichen Liang<sup>1,2</sup>, Yuebo Jin<sup>1</sup>, Jing He<sup>1</sup>.

<sup>1</sup>Department of Rheumatology and Immunology, Peking University People's Hospital, Beijing, 100044, China. <sup>2</sup>Department of Gastrointestinal Surgery, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology.

**Objective.** Treg dysfunction or deficiency results in unopposed immune activation and contributes to the development of autoimmune diseases such as Sjögren's syndrome. Interleukin-2 (IL-2) is critical for the expansion, suppressive function, and maintenance of Tregs. This study is to evaluate the effect of low dose IL-2 treatment in SS.

**Methods.** 8-week old female NOD mice were administered daily subcutaneous injections of humanized recumbent Interleukin-2 (30,000 IU) every day or PBS as a control. Immunized mice were analyzed at week 16. Solenocytes were incubated with fluorophore-conjugated monoclonal antibodies and Treg (Foxp3+CD25+CD4+) cells was analyzed by flow cytometry. The animals were analyzed for the presence of anti-SSA, anti-SSB, RF, ANA by immunofluorescence or ELISA. The salivary flow rate was measured. Salivary glands were examined by H&E staining.

**Results.** The number of  $Foxp3^+$  CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells was higher in the IL-2 groups compared to the PBS control groups (p<0.05). SS related antibodies titers and lymphocytic infiltration in the salivary glands were decreased in the IL-2-treated group. Salivary flow rate increased in the IL-2 treatment group.

**Conclusions.** Our data demonstrate that low dose IL-2 effectively inhibited the progression of experimental Sjögren's syndrome autoimmunity in NOD mice and expanded Tregs *in vivo*. Low dose IL-2 may be an appropriate treatment for SS in patients.

#### P-107

#### Contributions of CXCL12 and its receptor to the T cell autoimmune response in a Sjögren's syndrome murine model

Naozumi Ishimaru, Mie Kurosawa, Rieko Arakaki, Aya Ushio, Kunihiro Otsuka, Yasusei Kudo.

Department of Oral Molecular Pathology, Tokushima University Graduate School of Biomedical Sciences, Tokushima, Japan.

Dysregulated chemokine signaling contributes to autoimmune diseases by facilitating aberrant T cell infiltration into target tissues, but the specific cytokines, receptors, and T cell populations still remain largely unidentified. Contributions of the potent chemokine CXCL12 and its receptor CXCR4 to the T cell autoimmune response were evaluated in alymphoplasia (aly)/ aly mice, a Sjögren's syndrome (SS) model bearing a point mutation of the nuclear factor (NF)-kB-inducing kinase gene. Salivary gland (SG), a major target of SS pathology in aly/aly mice exhibited higher concentrations of effector memory T (TEM) cells than aly/+ mouse SG. TEM cells from aly/aly mice demonstrated higher in vitro migratory activity toward CXCL12 than aly/+ TEM cells. Moreover, CXCL12 expression was specifically upregulated in SS target organs of aly/aly mice, and aly/aly TEM cells showed greater CXCR4 surface expression. TEM cells from RelB-/- mice but not NF- $\kappa B1^{-/-}$  mice also demonstrated greater migratory activity toward CXCL12, implicating a non-classical NF-κB2/RelB pathway in regulation of TEM cell migration. TEM cells from aly/aly mice also overexpressed

transforming growth factor (TGF) $\beta$  receptors I and II, and TGF $\beta$  enhanced both CXCR4 expression and migratory activity to a greater degree in *aly/aly* TEM cells than *aly/+* TEM cells. The CXCR4 antagonist AMD3100 suppressed autoimmune lesions in *aly/aly* mice by reducing TEM cell infiltration. Collectively, these results suggest that NF- $\kappa$ B2/RelB regulates T cell migration to autoimmune targets through TGF $\beta$ R-dependent regulation of CXCL12-CXCR4 signaling, and highlights these signaling pathways as potential therapeutic targets for autoimmune diseases.

### P-108

# Reduced corneal innervation in the CD25KO model of Sjögren syndrome

Mary Ann Stepp<sup>\*1,2</sup>, Sonali Pal-Ghosh<sup>1</sup>, Gauri Tadvalkar<sup>1</sup>, Alexa Williams<sup>1</sup>, Stephen C Pflugfelder<sup>3</sup>, and Cintia S. de Paiva<sup>3</sup>.

<sup>1</sup>Department of Anatomy and Regenerative Biology, The George Washington University School of Medicine and Health Sciences, Washington DC USA. <sup>2</sup>Department of Ophthalmology, The George Washington University School of Medicine and Health Sciences, Washington DC USA. <sup>3</sup>Department of Ophthalmology, Ocular Surface Center, Cullen Eye Institute, Baylor College of Medicine, Houston, TX, USA.

**Purpose.** Decreased corneal innervation is a frequent finding in patients with Sjögren Syndrome (SS). The purpose of this study was to investigate the density of intraepithelial corneal nerves (ICNs) and corneal sensitivity using the well-characterized IL-2 receptor alpha chain (CD25KO) model of SS.

**Methods.** Corneal barrier function was examined by uptake of a fluorescent dye and graded by two masked observers in CD25KO and wild-type (WT) mice. Whole-mount corneas were used to quantify ICN density and thickness using bIII tubulin staining. Mechanical corneal sensitivity was measured using a modified Cochet-Bonnet esthesiometer. Quantitative PCR was performed to quantify expression of beclin 1, LC3, Lamp-1, Lamp-2, CXCL-1, BDNF, NTN1, DCC, Unc5b1, Efna4, Efna5, Rgma, and p21 in corneal epithelial mRNA.

**Results.** CD25KO mice had significant greater corneal barrier disruption than WT mice. This was accompanied by a significant reduction in axon density and mechanical corneal sensitivity. Real-time PCR results indicated that CD25KO mice have increased expression of genes regulating phagocytosis and autophagy (beclin-1, LC3, LAMP-1, LAMP-2, CXCL1 and BDNF) while no change was observed in genes related to axonal targeting and extension (NTN1, DCC1, Unc5b1, Efna4, Efna5, Rgma and p21).

**Conclusions.** At 8 weeks of age, mice lacking CD25 show decreased corneal innervation accompanied by reduced corneal sensitivity and increased expression of genes regulating phagocytosis and autophagy.

#### P-109

# Pathogenic role of interleukin 27 in the nonobese diabetic mouse model of Sjögren syndrome

Scott M Lieberman<sup>1</sup>, Jennifer Y Barr<sup>1</sup>, Xiaofang Wang<sup>1</sup>, Yi-Guang Chen<sup>2</sup> <sup>1</sup>Stead Family Department of Pediatrics, University of Iowa, Iowa, USA. <sup>2</sup>Department of Pediatrics, Medical College of Wisconsin, Wisconsin, USA.

**Background.** Interleukin 27 (IL27) is a heterodimeric cytokine with immunostimulatory and immunomodulatory properties depending on the context. Sjögren syndrome patients have elevated levels of IL27 in serum, yet whether IL27 contributes to the pathogenesis of Sjögren syndrome is not known. Similar to Sjögren syndrome in humans, nonobese diabetic (NOD) mice develop spontaneous autoimmune dacryoadenitis and sialadenitis. The role of IL27 in lacrimal and salivary gland autoimmunity in NOD mice has not been reported. Our objective was to evaluate the role of IL27 signaling in the development of dacryoadenitis and sialadenitis in the NOD mouse model of Sjögren syndrome.

**Methods.** NOD mice with deletion mutations disrupting expression of genes encoding the p28 component of IL27 (*II27*) or the alpha chain of the IL27 receptor (*II27ra*) were developed through Zn-finger nuclease or CRISPR/Cas9 mediated gene editing, respectively. Development of dacry-oadenitis and sialadenitis were determined by histological analyses, and T cell phenotypes were characterized by flow cytometry. *In vivo* regulatory T cell (Treg) depletion with anti-CD25 monoclonal antibody (PC61) and

adoptive transfers were performed to determine the effects of disrupted IL27 signaling on development of dacryoadenitis. Studies were approved by the Institutional Animal Care and Use Committee of the University of Iowa. **Results.** NOD mice lacking IL27 or IL27Ra failed to develop spontaneous autoimmune dacryoadenitis or sialadenitis. Phenotypically, T cells from IL27-deficient or IL27Ra-deficient NOD mice showed no evidence of defective T cell activation based on expression of T cell activation markers analyzed by flow cytometry *ex vivo*. Depletion of Treg cells in IL27-deficient NOD mice failed to drive dacryoadenitis in these mice. In our adoptive transfer model, wild-type T cells transferred dacryoadenitis to NOD-SCID recipient mice, but IL27Ra-deficient T cells failed to transfer dacryoadenitis. When co-transferred with wild-type cells, IL27Ra-deficient CD4 effector T cells demonstrated a significant competitive disadvantage in their ability to infiltrate lacrimal glands.

**Conclusions.** IL27 is required for development of dacryoadenitis and sialadenitis in NOD mice. T cell-intrinsic IL27 signaling is required to transfer disease. Defective infiltration of lacrimal glands by IL27Ra-deficient effector CD4 T cells suggests IL27 signaling may drive upregulation of homing receptors required for lacrimal gland inflammation.

**Disclosures.** This work was supported by grants from the NIH (EY022344 to SML, DK097605 to YC), Children's Miracle Network (to SML), American Association of Immunologists (to SML), and institutional research funds from the University of Iowa (to SML). The authors declare no conflicts of interest.

#### **P-110**

# Activation of the STING pathway is involved in the induction of Sjögren's syndrome-like disease in mice

Joanna Papinska<sup>1</sup>, Harini Bagavant<sup>1</sup>, Grzegorz B. Gmyrek<sup>1</sup>, Magdalena Sroka<sup>1</sup>, Saigiridhar Tummala<sup>2</sup>, Kate Fitzgerald<sup>3</sup>, and Umesh S. Deshmukh<sup>1</sup>. <sup>1</sup>Arthritis and Clinical Immunology Program, <sup>2</sup>Comparative Medicine, Oklahoma Medical Research Foundation, Oklahoma City, OK 73104, USA. <sup>3</sup>Division of Infectious Disease and Immunology, University of Massachusetts Medical School, Worcester, MA 01655, USA.

**Background/Purpose.** Patients with Sjögren's syndrome (SS) often present with a heightened type I IFN response. Recognition of DNA within the cytosol by a multitude of cytosolic DNA sensors and downstream activation of the stimulator of interferon gene (STING) protein is a key pathway for the induction type I IFN. The major objective of this study was to investigate the role of STING activation in the etiopathogenesis of SS.

**Methods.** Female C57BL/6 mice were injected with a STING agonist dimethylxanthenone-4-acetic acid (DMXAA) and control mice were treated similarly with the vehicle. Salivary glands were monitored for gene expression by real time PCR and for inflammatory cell infiltration by immunohistochemistry and flow cytometry. Salivary gland function was evaluated by measuring pilocarpine-induced salivation. Sera were analyzed for cytokines and autoantibodies. Cultured primary salivary gland cells were used to study the expression and activation of STING.

**Results.** DMXAA treatment rapidly upregulated the expression of  $Ifn\beta$  and pro-inflammatory cytokines, both systemically and locally in the salivary glands. In the murine submandibular glands, STING expression was detected mainly in interstitial cells. In vitro activation of STING in cultured primary salivary gland cells, rapidly phosphorylated TBK1, IRF3 and induced the expression of  $Ifn\beta$  and  $TNF-\alpha$ . Within 4 weeks of treatment, in comparison with the vehicle group, DMXAA treated mice developed significantly higher incidence of sialoadenitis (1/17 versus 10/21, p=0.009). At early stages of the disease, significantly increased numbers of NK1.1+: NK cells (CD49b+CD49a-), tissue type I innate lymphoid cells (ILC1) (CD49a+CD49b-) and salivary gland ILC1 (CD49a+CD49b+) were observed in the salivary glands. The mean saliva amount in DMXAA treated group (56 $\pm$ 14 mg) was significantly lower (p=0.001) than the untreated (80±20 mg) and vehicle treated groups (78±25). The incidence of high titer ANA (>400) was significantly higher (p=0.02) in the DMXAA group (8/16) than in the vehicle treated group (0/8).

**Conclusion.** This study demonstrates that activation of STING protein induces certain features of SS in mice. Our study also suggests that following activation of innate immunity, type I innate lymphoid cells might be involved in the initial stages of salivary gland disease in SS. We would like to propose that apart from viral infections, conditions that cause cellular perturbations and accumulation of host DNA within the cytosol should be considered as possible endogenous triggers for SS.

#### Posters

# P-111

# Metabolic changes in the evolution of Sjogren's syndrome in a mouse model.

Julian L. Ambrus Jr.<sup>1</sup>, Alexander Jacob<sup>1</sup>, Gary A. Weisman<sup>2</sup> and Jing He<sup>1,3</sup>. <sup>1</sup>SUNY at Buffalo School of Medicine, Buffalo, NY USA. <sup>2</sup>Department of Biochemistry, University of Missouri, Columbia MO USA. <sup>3</sup>Peking University People's Hospital, Beijing, China.

**Background.** Previous studies have identified metabolic abnormalities in patients with Sjogren's syndrome (SS) and fatigue (*Clin. Immunol.* <u>182</u>:1, 2017). Deletion of the P2Y<sub>2</sub> nucleotide receptor (P2Y<sub>2</sub>R) alleviates symptoms and signs of SS in IL-14 alpha transgenic mice (IL14aTG). Stimulation of P2Y<sub>2</sub>R requires cellular release from the cytoplasm of ATP that is derived from mitochondria. The current studies were designed to evaluate changes in metabolism that occur in IL14aTG mice during the evolution of SS.

**Methods.** Microarray studies were performed on the spleens and submandibular glands of IL14aTG mice and control mice at 6 months of age. Abnormally expressed genes were confirmed by qPCR studies during both early disease (6 months) and late disease (12 months). Serum metabolites were evaluated by gas chromatography at 6 and 12 months of age.

**Results.** At 6 months of age, IL14aTg mice expressed two mitochondrial respiratory chain enzymes in their spleens at high levels compared to normal controls, NADH dehydrogenase and ubiquinol cytochrome C reductase. Their salivary glands express PI-3K at high levels, which can induce mTOR1 and p70S6K. Expression of ryanodine receptor 1 (RYR1) was decreased. Serum studies revealed enhanced activity of the citric acid cycle, mitochondrial respiratory chain and aerobic glycolysis, consistent with the activation of mTOR1. In contrast, serum studies at 1 year of age, when IL14aTG mice are at an advanced stage of disease, revealed less activity of the mitochondrial respiratory chain and increased amino acid metabolism. Elevated C5 carnitine, and hydroxyglutarate were consistent with acquired glutaryl CoA dehydrogenase deficiency.

**Conclusions.** The nature of metabolic abnormalities differs in the early and late stages of SS in IL14aTG mice. At 6 months of age, high energy requirements are met with increased mitochondrial respiration and aerobic glycolysis. At 1 year of age, energy requirements are altered and are met more with increased amino acid metabolism and fatty acid oxidation. A potential defect in fatty acid metabolism is noted at 1 year of age. Because of the potential central importance of mTOR1 in early metabolic events, current studies are evaluating the effects of the mTOR1 inhibitor, rapamycin, on early events in SS.

Funded by National Institutes of Health grant DE007389.

# P-112

#### RORyt antagonist suppressed spontaneous sialadenitis in RORyt transgenic mice via inhibition of IL-17 production with increase of regulatory T cells

Yuko Ono<sup>1,2</sup>, Hiroto Tsuboi<sup>1</sup>, Masafumi Moriyama<sup>2</sup>, Hiromitsu Asashima<sup>1</sup>, Hanae Kudo<sup>1</sup>, Hiroyuki Takahashi<sup>1</sup>, Yuya Kondo<sup>1</sup>, Isao Matsumoto<sup>1</sup>, Seiji Nakamura<sup>2</sup>, and Takayuki Sumida<sup>1</sup>

<sup>1</sup>Department of Internal Medicine, Faculty of Medicine, University of Tsukuba. <sup>2</sup>Section of Oral and Maxillofacial Oncology, Division of Maxillofacial Diagnostic and Surgical Sciences, Faculty of Dental Science, Kyushu University.

**Objective.** Our previous studies demonstrated that T cells specific ROR $\gamma$ t-transgenic-mice under human CD2 promoter (ROR $\gamma$ t-Tg mice) developed spontaneous sialadenitis like Sjögren's syndrome (SS) in which reduced regulatory T cells (Tregs) and ROR $\gamma$ t-overexpressed CD4<sup>+</sup>T cells contributed to the pathogenesis. The purpose of this study is to clarify suppressive ability and its mechanisms of ROR $\gamma$ t antagonist (A213) for sialadenitis in ROR $\gamma$ t-Tg mice.

**Methods.** 6-weeks aged ROR $\gamma$ t-Tg mice orally received 300 mg/kg of A213 (10  $\mu$ L/g body weight) or vehicle (PBS, 10  $\mu$ L/g body weight) every three days for 2 weeks. We analyzed 1) saliva volume, 2) histopathology of salivary glands (Hematoxylin and Eosin-staining and focus score), 3) populations of T cells subsets (naïve, central memory, and effector memory T cells) in splenocytes, 4) percentages of Tregs(Foxp3<sup>+</sup>/CD4<sup>+</sup>CD25<sup>+</sup>) in splenocytes, and 5) mRNA expression of IL-17A and IFN $\gamma$  in splenocytes.

#### **Results.**

- The ratio of saliva volume at 2 weeks to that at baseline was significantly increased in A213-treated group (1.4±0.1) compared with PBS-treated group (0.9±0.1) (p<0.05).</li>
- 2) Infiltration of mononuclear cells in salivary glands in Hematoxylin and Eosin-staining were dramatically improved in A213-treated group compared with in PBS-treated group. The focus score of sialadenitis at 2 weeks was significantly lower in A213-treated group (0.2±0.2) than in PBS-treated group (2.3±0.6) (p<0.05) (Fig. 1).</p>
- 3) The population of effector memory T cells (CD44<sup>hi</sup>CD62L<sup>ho</sup>) was 96.9%, central memory T cells (CD44<sup>hi</sup>CD62L<sup>hi</sup>) was 1.99%, and naïve T cells (CD44<sup>hi</sup>CD62L<sup>hi</sup>) was 0% in RORγt-Tg mice before administration of A213. After administration of A213, the population of effector memory T cells tended to be decreased (73.2%) and central memory and naïve T cells tended to be increased (26.0% and 0.51%, respectively), whereas PBS did not alter the population of these T cell subsets.
- 4) The percentage of Treg (Foxp3<sup>+</sup>/CD4<sup>+</sup>CD25<sup>+</sup>) in RORγt-Tg mice tended to be increased from 47.4% at baseline to 75.5% at 2 weeks after administration of A213, while PBS did not alter the percentage of Treg (40.9%).
- 5) In splenocytes at 2 weeks after treatment, the mRNA expression of IL-17A was significantly decreased in A213-treated group compared with PBS-treated group (p<0.05), while that of IFNγ was comparable between groups.</p>

**Conclusion.** A213 could suppress the sialadenitis in ROR $\gamma$ t-Tg mice via inhibition of IL-17 production with increase of Tregs.



Fig. 1. Comparison of histological focus score of inflammatory lesions in salivary glands between A213- and PBS-treated RORytTg mice.

### P-113

#### B cell receptor repertoire of signature Sjögren's syndrome antigen-specific autoantibodies in labial salivary glands

Alexandria Voigt<sup>1</sup>, Carol M. Stewart<sup>2,3</sup>, Indraneel Bhattacharya<sup>2,3</sup>, Cuong Q. Nguyen<sup>1,3,4</sup>.

<sup>1</sup>Department of Infectious Diseases and Pathology, College of Veterinary Medicine. <sup>2</sup>Department of Oral and Maxillofacial Diagnostic Sciences. <sup>3</sup>Center of Orphaned Autoimmune Diseases, <sup>4</sup>Department of Oral Biology, College of Dentistry, University of Florida, Gainesville Florida, USA.

**Objectives.** Sjögren's syndrome (SS) is characterized by massive lymphocytic infiltrations in the salivary and lacrimal glands. Additionally, anti-SSA/Ro and anti-SSB/La autoantibodies are the signature diagnostic biomarkers for the disease; Ro antigens have been implicated in the pathogenesis of SS. As an essential etiological feature of the autoimmune process, it remains unclear what B cell repertoires are expressed by these antigenspecific and autoreactive B cells. Here we examine the infiltrating B cells of SS patients *ex-vivo* using single-cell microengraving analysis and high throughput sequencing to determine the B cell receptor (BCR) repertoires of the individual antigen-specific B cells.

**Methods.** Single-cell microengraving analysis is a soft lithographic technology that is used here to isolate single cells and identify specific subsets of B cells that secrete reactive antibodies against Ro-52, Ro-60, and La antigens, as well as identify the antibody isotype, in the labial salivary glands (LSGs) of primary SS (pSS) patients and non-pSS controls. The technology uses an array of 84,672 nanowells to capture an individual cell per well. Single-cell suspensions of LSG cells were prepared from pSS and nonpSS samples labeled with CD19, CD27, and CD138, then dispersed into nanowells. Individual B cells in the nanowells were identified using automated epifluorescent microscopy to identify the presence of plasma cells, plasmablasts, and memory B cells in each well. Individual B cells were microengraved for secreting IgG isotypic anti-SSA/Ro52, anti-SSA/Ro60, and anti-SSB/La. Individual secreting B cells were isolated and subjected to high-throughput sequencing for BCR repertoire analysis.

**Results.** The data indicate that there was a disproportionately high frequency of plasmablasts present in the LSGs of pSS patients compared to non-pSS subjects. The frequency of plasmablasts is significantly higher than plasma cells and memory B cells in pSS. Interestingly, both pSS patients and non-pSS controls produced IgG isotypic anti-SSA/Ro52, anti-SSA/ Ro60, and anti-SSB/La with higher levels of these autoantibodies found in pSS patients. Lastly, high- throughput sequencing of individual BCRs of these antigen-specific B cells revealed a more restricted repertoire in the pSS patients when compared to non-pSS controls.

**Conclusion.** The results indicate an antigen-driven B cell response in the LSGs of pSS patients. Further work will be needed to understand the underlying mechanism that governs the clonal expansion during the autoimmune process of SS.

# **P-114**

#### Somatically introduced N-glycosylation of immunoglobulin – a potential non-specific mechanism for B cell activation in Sjögren's syndrome

Kristi A. Koelsch, PhD<sup>1,2,3</sup>, Joshua Cavett, MS<sup>1,2,3</sup>, Kenneth Smith, PhD<sup>2</sup>, Jacen S. Moore, PhD<sup>1,2,3,\*</sup>, Sylvain D. Lehoux, PhD<sup>4</sup>, Nan Jia, PhD<sup>4</sup>, Tim Mather, PhD<sup>2</sup>, Syed M. S. Quadri, MD<sup>1,2</sup>, Astrid Rasmussen, MD, PhD<sup>2</sup>, C. Erick Kaufman, MD<sup>5</sup>, David M. Lewis, DDS<sup>6</sup>, Lida Radfar, DDS<sup>7</sup>, Christopher J. Lessard, PhD<sup>1,2</sup>, Biji T. Kurien, PhD<sup>1,2,3</sup>, Richard D. Cummings, PhD<sup>4</sup>, Judith A. James, MD, PhD<sup>1,2</sup>, Kathy L. Sivils, PhD<sup>1,2</sup>, A. Darise Farris, PhD<sup>1,2</sup>, R. Hal Scofield, MD<sup>1,2,3</sup>.

<sup>1</sup>University of Oklahoma Health Sciences Center, Oklahoma City Oklahoma.<sup>2</sup>Arthritis and Clinical Immunology Program, Oklahoma Medical Research Foundation, Oklahoma City, Oklahoma. <sup>3</sup>Department of Veterans Affairs Medical Center, Oklahoma City, OK. <sup>4</sup>National Center for Functional Glycomics, Department of Surgery, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA. <sup>5</sup>Department of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City, OK. <sup>6</sup>Department of Oral and Maxillofacial Pathology, University of Oklahoma College of Dentistry, Oklahoma City, OK. <sup>7</sup>Oral Diagnosis and Radiology Department, University of Oklahoma College of Dentistry, Oklahoma City, OK.

\*Present affiliation: Dept. of Clinical Laboratory Sciences, University of Texas, El Paso, TX.

Background. Sjögren's syndrome (SS) is a systemic autoimmune disease characterized by lymphocytic infiltration of the salivary and lacrimal glands resulting in tissue destruction, pathological dry mouth and dry eyes and in increased risk for lymphoma. The presence of ectopic germinal centers, clonally related B cells and autoantibody production from antibody-secreting cells (ASCs) isolated from the salivary glands indicate that at least some of the ASCs arise from an antigen-driven immune response. Analyses of immunoglobulin sequences can be instrumental for determining somatic mutational patterns shaped by selective pressures, where positive selection in the Complementary Determining Regions (CDRs) and negative selection in the framework regions (FWRs) would indicate antigen-driven antibody production. An alternative would be a non-specific mode of B cell activation. Immunoglobulin variable region Nlinked glycosylation acquired by somatic hypermutation (AcN-glycs) has been strongly correlated to follicular lymphoma. Bacterial lectins can bind and activate B cells via AcN-glycs - a possible non-specific mechanism for antibody production and proliferation of B cells.

Methods. To explore immunoglobulin selective pressures in our cohort, we single cell-sorted IgG+ ASCs isolated from the minor salivary glands of 4 SS patients meeting the American-European combined and ACR criteria, and 5 sicca controls. The immunoglobulin variable regions were sequenced by RT-PCR. AcN-glyc motiffs were identified using the online NetNGlyc tool. An IMGT/V-QUEST sequence analysis was performed to confirm that all identified AcN-glyc motifs were introduced by somatic mutation and not germline-encoded. To analyze for antigen-driven selection we utilized BASELINe Version 1.3, an online tool for predicting positive or negative selective pressures.

**Results.** We sequenced 56 variable regions from 3 SS and 21 sicca control immunoglobulins. Analysis of heavy and light chain sequences revealed SS patients had an increased frequency of AcN-glycs motifs in the FWRs and a decreased frequency in the CDRs as compared to controls (20% vs 5%; 4%

vs 10%). The BASELINe analysis showed positive selection in the CDRs and negative selection in the FWRs of sequences from both SS patients and controls. When the selection strengths for immunoglobulins based on glycosylation status were compared (regardless of classification), we found that heavy chains with CDR AcN-glycs and light chains with FWR AcN-glycs had significantly less positive selection than those without AcN-glycs (p=0.05 and p=0.04, respectively).

**Conclusions.** Overall, immunoglobulins from ASCs infiltrating the minor salivary glands of SS patients and sicca controls in our cohort undergo positive selection in the CDRs and negative selection in the FWRs, indicating antigen-driven immune responses. Immunoglobulins with AcN-glycs have lower positive and negative selection pressures in the heavy chain CDRs and FWRs as compared to those that are non-glycosylated, indicating a potential non-specific mechanism for activation in these ASCs that could potentially give rise to autoreactive proliferations or lymphoproliferative neoplasms.

#### P-115

#### RNA Sequencing detection of gene dysregulation in epithelial sorted cells from salivary gland tissue reveals interesting pathways involved in Sjögren's syndrome pathophysiology

Elodie Rivière<sup>1,2</sup>, Nicolas Tchitchek<sup>1,3</sup>, Juliette Pascaud<sup>1</sup>, Alexandre Virone<sup>1</sup>, Bineta Ly<sup>1</sup>, Saida Boudaoud<sup>1</sup>, Michael Mingueneau<sup>4</sup>, Bernd Jagla<sup>5</sup>, Audrey Paoletti<sup>1</sup>, Samuel Bitoun<sup>1</sup>, Gaetane Nocturne<sup>1,6</sup>, Xavier Mariette<sup>1,6</sup>.

<sup>1</sup>Centre for Immunology of Viral Infections and Autoimmune Diseases (IMVA), Institut National de la Santé et de la Recherche Médicale (INSERM) U1184, Paris, France. <sup>2</sup>Arthritis Fondation Courtin, France. <sup>3</sup>CEA - Université Paris Sud 11 - INSERM U1184, Immunology of Viral Infections and Autoimmune Diseases, IDMIT Infrastructure, 92265 Fontenay-aux-Roses, France. <sup>4</sup>Immunology Research, Biogen, Cambridge, Mass. <sup>3</sup>Transcriptome and EpiGenome Platform, Institut Pasteur, Paris, France. <sup>6</sup>AP-HP, Hôpitaux Universitaires Paris-Sud, Université Paris-Sud, Paris, France.

**Background.** Primary Sjögren's syndrome (pSS) is a chronic autoimmune disorder characterized by lymphocytic infiltrates and destruction of the salivary glands. Several lines of evidence support the hypothesis that salivary gland epithelial cells (SGECs) are not only the target of autoimmunity in pSS patients but may also play a role for its initiation and maintenance. **Objective.** To establish high-resolution molecular maps of SGEC from pSS patients compared to controls using RNASeq analysis.

**Methods.** SGEC, B, T CD4 and CD8 lymphocytes were sorted from salivary gland biopsies from 9 pSS patients and 4 controls, using a FACS ARIA cell sorter. Total RNASeq profiling was performed using MiSeq (Illumina). For SGEC subset, 4 samples were excluded due to a contamination by B lymphocytes, thus analysis was performed on 5 pSS and 4 controls using R software, to identify transcriptional differences between pSS and control SGEC. Functional Enrichment analysis was performed using Ingenuity Pathway Analysis software.



Fig. 1. Volcano Plot representation of differential expression analysis of genes in the pSS SGEC versus controls. Each dot corresponds to a gene, and dots are represented in a 2D chart, where the x-axis shows log2 fold-changes expressions and the y-axis shows the p-values –(represented as -log10).

 Table I. Pathways identified a statistically over-represented with Ingenuity Pathways Analysis in epithelial cells from pSS patients compared to controls.

Pathways	log p-value	Genes
Primary Immunodeficiency Signaling	4,08	PTPRC, BTH, IGHG1, CD8A, TAP1, TNFRSF13C
Interferon Signaling	3,52	IFIT3, OAS1, IFI6, STAT1, TAP1
8 Cell Development	2,89	PTPRC, HLA-DRA, CD86, IL7
Role of JAK2 in Hormone-lil Cytokine Signaling	ke 2,73	STAT5A, IRS1, SH2B3, STAT1
IL-7 Signaling Pathway	2,51	STAT5A, SLC2A1, IRS1, IGHG1, STAT1, IL7

Table II. Selection of genes differentially expressed between pSS and controls in SGEC.

Gene Symbol	Gene ENSG ID	log2 fold-change	<i>p</i> -value
BST2	ENSG00000130303	4.08	0.0002
HLA-DRA	ENSG00000204287	1.84	0.0372
IL-23 A	ENSG00000110944	3.59	0.0155
BAFF-R	ENSG00000159958	4.94	0.0097
CD86	ENSG00000114013	-5.92	0.0096

**Results.** In SGEC, 495 genes were differentially expressed between pSS and controls. 280 genes were up-regulated, and 215 genes were down-regulated (Figure 1). Enrichment analysis (Table I) highlighted IL7 signaling pathways (including IL7, STAT5A, STAT1 genes) and interferon signaling (including OAS1, IFIT3, IFI6, TAP1 genes). Other genes potentially involved in immune responses and interactions between SGEC and lymphocytes were significantly up-regulated, including bone marrow stromal cell antigen 2, HLA-DRA, BAFF-R and IL23 A (Table II). CD86, a costimulatory molecule, was found to be significantly down-regulated. These results need to be confirmed by RT qPCR. However, congruent results have already been obtained in our laboratory, showing that IL7 serum level is increased in pSS patients compared to controls and that SGECs produce IL7 after interferon stimulation. The analysis of the non-coding RNA part and the other sorted cells subtypes is ongoing.

**Conclusions.** Immune interactions between SGEC and B or T lymphocytes could represent a key in the understanding of the initiation and/or maintenance of autoimmunity in pSS. Our study highlights the key role of epithelial cells in activation of immune cells. In vitro experiments are needed to confirm these results and elucidate the molecular mechanisms.

## P-116

# IL-7 in primary Sjögren syndrome (pSS) is secreted by salivary gland epithelial cells after IFN stimulation and is associated with B-cell activation

A. Virone<sup>1</sup>, J. Pascaud1, E. Rivière<sup>1</sup>, J.E. Gottenberg<sup>2</sup>, V. Le Guern<sup>3</sup>, X. Mariette<sup>1,4</sup>, G. Nocturne<sup>1,4</sup>

<sup>1</sup>UMR1184, Le Kremlin Bicêtre, France. <sup>2</sup>Dept of rheum. CHU Strasbourg, France. <sup>3</sup>Dept of int medicine, Cochin hospital, Paris, France. <sup>4</sup>Dept of rheum, Hop universitaires Paris-Sud, France.

**Background.** pSS is characterized by a strong IFN signature, ectopic germinal centers formation and a chronic blood lymphopenia. IL-7 plays a central part in T cells homeostasis and in lymphoid structures organization. We aimed to assess the role of IL-7 in pSS pathogenesis.

**Methods.** IL-7 serum level was assessed in 372 pSS patients and 73 paired controls. Primary cultures of salivary gland epithelial cells (SGEC) from patients and controls were stimulated by Poly I: C30 ng/ml, IFN- $\alpha$  600UI/ml, IFN- $\gamma$  5ng/ml and IFN- $\lambda$  (IL-28) 25ng/ml for 72 hours. IL-7 secretion was tested in culture supernatant by ELISA. IL-7 expression after 24 hours stimulation was assessed by quantitative RT-PCR. IL-7 and its receptor's expressions were evaluated in RNA-Seq analysis from cells form salivary glands biopsies (SGB) and PBMC.

**Results.** pSS patients had higher serum IL-7 levels than controls: 7.56 ng/ ml  $\pm$  8.52 (mean  $\pm$  SD) versus 4.86 ng/ml  $\pm$  5.59; p<0.0001. A positive correlation with B cells activation markers, IFN-induced chemokines and disease activity markers was observed. In multivariate analysis, serum IL-7 level was associated with CXCL13, anti-SSA, RF,  $\varkappa$  light-chain and low C4. SGEC stimulation with Poly I:C, IFN- $\alpha$ , - $\gamma$  and - $\lambda$  induced IL-7 protein secretion in the supernatant (p=0.002, p=0.004, p=0.007, p=0.004 respectively). A trend for a greater IL-7 production in pSS patients compared to controls was observed. IL-7 expression was confirmed by quantitative RT-PCR. Among cell subsets purified ex vivo from SGB and PBMC, RNA-Seq analysis showed a greater expression of IL-7 by epithelial cells of patients compared to controls (p=0.03). No difference was observed regarding T and B cells either from biopsies or PBMC. IL-7 receptor expression was equivalent between patients and controls. Analysis of T cells exhaustion profile and IL-7 intracellular signaling are on-going.

**Conclusion.** Our data demonstrate that IL-7 is secreted within the target tissue of pSS by SGEC after stimulation by IFNs. This IFN/IL-7 pathway can be involved in the organization of ectopic germinal centers found in pSS. But more importantly, since II-7 is one of the major controller of homeostasis of T cells, this IFN/IL-7 pathway could be involved in the persistent lymphopenia which is a hallmark of the disease, either by favoring exhaustion of T cells or by an impaired function of the IL-7 intracellular signaling. Both mechanisms are currently explored and will be presented.



Fig. 1. IL-7 secretion after a 72 hour-stimulation of SGEC.

## P-117

# Immune activation of epithelial cells causes primary Sjögren's syndrome-like symptoms

Wang, X.<sup>1</sup>, Saalan, A.<sup>2</sup>, Liefers, S.<sup>1</sup>, Proctor, G.<sup>2</sup>, Bootsma, H.<sup>1</sup>, Kroese, F.G.M.<sup>1</sup> & Pringle, S.<sup>1</sup>.

<sup>1</sup>Department of Rheumatology and Clinical Immunology, University Medical Centre Groningen, University of Groningen, The Netherlands. <sup>2</sup> Division of Mucosal and Salivary Biology, King's College London Dental Institute, King's College London.

**Background.** Hyposalivation and lymphocytic infiltration in salivary glands are common manifestations of primary Sjögren's syndrome (pSS). NF-kappa B (NF- $\kappa$ B) signaling is one of the most important proinflammatory pathways, and is inhibited by A20 (also known as TNFAIP3). Although mounting studies are pointing to the central role of epithelial cells in initiation of pSS, what exact function they perform the early stages remains poorly understood. In the current study we employ a mouse model using cytokeratin 14 (CK14) promoter-driven knockout of the NF- $\kappa$ B inhibitor A20 gene, to promote an inflammatory environment by epithelial cells and investigate this.

**Methods.** To generate knockout mice F2 generations of A20FL/FL mice crossed with CK14Cre/WT mice were used. A20FL/FL mice were kindly provided by Dr. Geert van Loo (Ghent University, Belgium). In the F2 offspring, the A20 gene was conditionally knocked out in CK14- expressing cells, which reside in the case of the salivary gland in the basal epithelial layer of the striated ducts, and cells of the intercalated ducts. In order to study salivary gland function, pilocarpine induced saliva production was measured and production of saliva mucins 10 and 19 were assessed by western blot and Coomassie brilliant blue/PAS stains. The degree of inflammatory infiltration of submandibular salivary glands was assessed using immunohistological staining with antibodies directed against the CD45, CD3 and B220 cell surface markers, for detection of leukocytes, T cells and B cells respectively. All data were analyzed at 10, 20 and 30 week time points, and in comparison to wild type littermate controls.

Results. Saliva production of A20FL/FL mice was significantly less

than A20WT/WT mice after correction for pilocarpine dose (Figure 1A). The level of mucin 10 in saliva of A20FL/FL mice decreased compared to A20WT/WT mice at 30 weeks of age, whereas mucin 19 appeared to increase. CD45+ and CD3+ periductal lymphocytic infiltrations started to emerge in A20FL/FL mice from 20 weeks of age, and were significantly higher than those in the control mice at 30 weeks of age. Characterizing of B cell content of A20FL/FL mice is in progress. Lymphocytes also infiltrated the striated ducts of A20FL/FL mice by 30 weeks of age, (Figure 1B). Characterization of the dominant cell type in these structures is underway. Conclusions. We present a mouse model for early epithelial cell involvement in pSS pathology development, whereby immune activation via the NF-[?]B pathway of epithelial cells is enough to generate the main characteristics of pSS, namely saliva reduction, altered composition of saliva, periductal lymphocytic infiltration and lymphoepithelial lesions. In our study, we emphasize the critical role of epithelial cells in the early stage of this autoimmune disease and open the door for further studies into the initiation of salivary gland pathology development in pSS.





#### P-118

#### Sjögren's syndrome: concerted triggers of sicca conditions

Diana Mieliauskaitė<sup>1</sup>, Ieva Narkevičiūtė<sup>2</sup> Justyna Mażul<sup>3</sup>, Irena Dumalakienė<sup>2,4</sup>, Irena Butrimienė<sup>3,5</sup>, Rita Vilienė<sup>2</sup>, Zygmunt Mackiewicz<sup>3</sup>. <sup>1</sup>Department of Innovative Medical Technologies and Health Resort Science, State Research Institute Centre for Innovative Medicine, Santariskiu St. 5, LT-08406 Vilnius, Lithuania. <sup>2</sup>Department of Immunology, State Research Institute Centre for Innovative Medicine, Santariskiu St. 5, LT-08406 Vilnius, Lithuania. <sup>3</sup>Department of Regenerative Medicine, State Research Institute Centre for Innovative Medicine, Santariskiu St. 5, LT-08406 Vilnius, Lithuania. <sup>4</sup>Department of Chemistry and Bioengineering, Faculty of Fundamental Sciences, Vilnius Gediminas Technical University, Saulėtekio al. 11, LT-10223 Vilnius, Lithuania. <sup>5</sup>Center of Rheumatology, Vilnius University, Santariskiu St. 2, LT-08406 Vilnius, Lithuania. **Background.** The aim of this study was to analyze whether Sjögren's syndrome is triggered by patient's intrinsic pathomechanisms or in cooperation with coincident hidden viral infection.

**Methods.** In 126 patients with sicca conditions immunohistochemical analysis of immune response to suggested silent persistence of mumps virus in the minor salivary glands biopsies and cytometric analysis of blood cells was done.

**Results.** Marks of mumps virus together with protein IFI16, interferons gamma and beta, dendritic cells, interleukin-3 receptor, receptor for natural killers and autophagy products were detected in the minor salivary gland biopsies from the patients with Sjögren's syndrome, rheumatoid artrhritis, but also in the non autoimmune sicca patients. Cytometric analysis of the blood cells from these patients revealed dropping amount of circulating natural killers and dendritic cells as supposed result of their central exhaustion due to massive intraglandular entrapping and degradation.

**Conclusions.** Occurence of abundant intraglandular immunohistochemical marks of mumps virus protein known by strong tropism to some epithelial cells accompanied by depletion of circulating immune cells make firm background for thought of presumable mumps or/and other viruses participation in specific epithelial damage in predisposed patients sicca syndrome.

This work was supported by a grant no. MIP-013/2014 from the Lithuanian Research Council.

# P-119

# Local glandular interferon activation in primary Sjögren's syndrome is associated with systemic interferon activation

Iris L. A. Bodewes<sup>1</sup>, Cornelia G. van Helden-Meeuwsen<sup>1</sup>, Alan N. Baer<sup>2</sup>, Marjan A. Versnel<sup>1</sup>

<sup>1</sup>Department of Immunology, Erasmus University Medical Centre, Rotterdam, The Netherlands. <sup>2</sup>Department of Medicine / Division of Rheumatology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA.

**Background.** Interferon (IFN) is considered a pathogenic factor in subsets of primary Sjögren's syndrome (pSS) patients. Upregulation of both IFN type I and IFN type II are described locally in the glands and systemically in the blood. However, there are limited data on how local and systemic IFN activation are correlated. It is critical to define the correlation between peripheral blood and minor salivary gland IFN activation patterns in the same individual as this can prevent repeated collection of salivary gland biopsies. In this pilot study we analyzed systemic IFN activation in the peripheral blood monocytes of pSS patients and correlated this with the local glandular IFN activation which was previously characterized in simultaneously collected salivary gland biopsies of patients.

**Methods.** Frozen PBMC samples were obtained from 11 pSS patients and 2 healthy controls without pSS from the Sjögren's International Collaborative Clinical Alliance (SICCA) registry. Local IFN type I and type II activation was previously determined in frozen labial salivary gland lysates by Western blotting by us (Hall *et al.* 2015). IFIT3 protein expression was used to indicate IFN type I activation (alpha) and GBP1 was used to indicate IFN type I activation (gamma). Peripheral blood monocytes were isolated from PBMC samples followed by RQ-PCR as described previously (Brkic *et al.* 2013). Systemic modular IFN expression was analyzed (Chiche *et al.* 2014, Bodewes *et al.* in press). M1.2 indicated systemic activation of IFN type I and IFN type II.

**Results.** Patients with local upregulation of IFN showed systemically higher M1.2 (IFN type I activation) and M5.12 scores (IFN type I/II activation). Patients with local gamma/alpha upregulation showed higher systemic M5.12 scores compared to patients with local gamma only. M1.2 scores were comparable between gamma/alpha and gamma only patients. Patients without local IFN activation had lower systemic M1.2 and M5.12 IFN scores than patients with local IFN activation (gamma or gamma/alpha activation). Furthermore, M1.2 and M5.12 scores of patients without local IFN activation were comparable to scores of healthy controls.

**Conclusions.** This small sample study indicates that pSS patients with local glandular IFN activation also have systemically higher IFN scores. Systemic IFN activation might therefore be a useful biomarker to monitor local inflammation.

Acknowledgements. The SICCA registry and participating patients.

**Funding.** The study was supported by a grant of the Dutch Arthritis Foundation (RF14-3-404) and the Jerome L. Greene foundation.

# P-120

TBK1 inhibition downregulates expression of interferon type I and the upregulated expression of RIG-like receptors and DNA-sensing receptors in interferon positive primary Sjögren's syndrome patients

Iris L. A. Bodewes<sup>1\*</sup>, Erika Huijser<sup>1\*</sup>, Cornelia G. van Helden-Meeuwsen<sup>1</sup>, Liselotte Tas<sup>1</sup>, Ruth Huizinga<sup>1</sup>, Paul L. A. van Daele<sup>1,2</sup>, Virgil A.S.H. Dalm<sup>1,2</sup>, P. Martin van Hagen<sup>1,2</sup>, Marjan A. Versnel<sup>1</sup>.

<sup>1</sup>Department of Immunology, <sup>2</sup>Department of Internal Medicine, Division of Clinical Immunology, Erasmus University Medical Centre, Rotterdam, The Netherlands. \*Shared first author.

Background. Upregulation of type I interferons (IFN-I) is a hallmark of systemic autoimmune diseases like primary Sjögren's syndrome (pSS). Expression of IFN-I is induced by three different receptor families: Toll-like receptors (TLRs), RIG-like receptors (RLRs) and DNA-sensing receptors (DSRs). Previously we have shown increased mRNA levels of TLRs and RLRs in plasmacytoid dendritic cells (pDC) and CD14+ monocytes of IFN-I positive (IFNpos) pSS patients (1). TANK-binding kinase (TBK1), is an important signaling hub downstream of RLRs and DSRs and leads to production of IFN-I and subsequent induction of interferon-stimulated genes (ISGs). The objective of this study was to study RLRs and DSRs in pSS and explore the potential of a TBK1 inhibitor to downregulate IFN-I activation. Methods. Expression of RLRs and DSRs was assessed by RQ-PCR and flowcytometry in CD14<sup>+</sup> monocytes, BDCA4<sup>+</sup>CD123<sup>+</sup> pDC and CD19<sup>+</sup> B cells from IFNpos pSS patients. pDCs from IFNpos pSS patients were analyzed by flowcytometry for phosphorylated-TBK1 (pTBK1). PBMCs of pSS patients were cultured with a TBK1 inhibitor, BX795, followed by analysis of IFN-I production and expression of ISGs.

Results. In addition to upregulated mRNA levels of RLRs IFIH1 (encoding for MDA5) and DDX58 (encoding for RIG-I), which we previously observed in pDC and monocytes of IFNpos pSS patients, gene expression of IFIH1 and DDX58 was also upregulated in B cells. Upregulation of mRNA levels of the DSRs IFI16 and ZBP1 was observed in monocytes and B cells from IFNpos patients. In pDC protein expression of MDA5, ZBP-1, IFI16 was increased in IFNpos pSS, while there were no differences in RIG-I. In monocytes protein expression of MDA5 was increased and a trend was visible for RIG-I and IFI16. B cells showed increased protein expression of MDA5 and a trend was observed for RIG-I, ZBP-1 and IFI16. These data indicate upregulation of RLRs and DSRs particularly in pDC of IFNpos pSS patients. To further look into the signaling of RLRs and DSRs, phosphorylation of TBK was studied in pDC, the main IFN-I producers, of pSS patients. Increased expression of pTBK1 was observed in pDCs from IFNpos pSS. Similar upregulation of pTBK1 was observed in IFNpos systemic lupus erythematosus (SLE) and systemic sclerosis (SSc) patients. Upon treatment with BX795, PBMCs from IFNpos pSS (and SLE and SSc) downregulated the production of IFN-I and mRNA expression of the ISGs MxA, IFI44, IFI44L, IFIT1 and IFIT3.

**Conclusions.** RLRs and DSRs are upregulated in IFNpos pSS. Signaling of these receptors could be blocked using a TBK1 inhibitor, which reduced IFN-I protein production and expression of ISGs in PBMCs of IFNpos pSS patients. As patented pharmacological inhibitors, amongst others a small molecule inhibitor, are available TBK1 inhibition is indicated as a potential future treatment target for IFNpos pSS.

Funding. The study was supported by a grant of the Dutch Arthritis Foundation (RF14-3-404).

#### Reference

 MARIA NI, STEENWIJK EC, IJPMA AS, VAN HELDEN-MEEUWSEN CG, VOGELSANG P, BEUMER W et al.: Contrasting expression pattern of RNA-sensing receptors TLR7, RIG-I and MDA5 in interferon-positive and interferon-negative patients with primary Sjögren's syndrome. Ann Rheum Dis 2016.

# P-121

Sjögren's-like syndrome in APECED patients is associated with down-regulated IFN pathways and impaired salivary fluid secretion genes: Calcium signaling attenuation in patients and NOD Aire -/- mice

Leyla Y. Teos<sup>1</sup>, Timothy J. Break<sup>2</sup>, Elise M.N. Ferre<sup>2</sup>, Anamaria Bondici<sup>2</sup>, Mayank Tandon<sup>1</sup>, Michail S. Lionakis<sup>2</sup>, Ilias Alevizos<sup>1</sup>.

<sup>1</sup>Sjögren's Syndrome and Salivary Gland Dysfunction Unit, National Institute of Dental and Craniofacial Research, NIH, Bethesda, Maryland, USA. <sup>2</sup>Fungal Pathogenesis Section, Laboratory of Clinical Immunology & Microbiology, National Institute of Allergy and Infectious Diseases (NIAID), NIH, Bethesda, Maryland, USA.

Background. Recognized as a rare autoimmune disorder; Autoimmune polyendocrinopathy- candidiasis-ectodermal dystrophy (APECED) is caused by mutations in the AIRE (autoimmune regulator) gene. Mutations in AIRE cause disruption of negative selection in the thymus, creating impaired autoreactive T cells leading to endocrine and non-endocrine autoimmune dysregulation. Diagnosis of APECED requires the presence of two of the three traditional hallmarks which include chronic mucocutaneous candidiasis, hypoparathyroidism, and Addison's disease. 42.9% of our American APECED cohort developed Sjögren's-like syndrome (JCI Insight. 2016;1(13) e88782) based on established American-European consensus criteria; manifesting with chronic sicca symptoms (eyes and mouth) and positive focus score on minor salivary gland biopsies. In contrast to Siögren's syndrome: Siögren'slike syndrome in APECED patients lack ENA autoantibodies, suggesting the pathogenesis of the syndrome in APECED is distinct from primary Sjögren's syndrome. The objectives were to identify key targets altered in the pathophysiology of Sjögren's-like APECED patients. We performed RNA-Seq of patient salivary glands and we completed functional imaging studies on salivary glands from patients and the APECED mouse model NOD Aire-/-.

**Methods.** Total RNA was isolated from 4 APECED patients with Sjögren'slike syndrome and 6 Healthy Controls. Sequenced data were preprocessed on the Ion Torrent server with Torrent Suite. Genomic features were counted using the HTSeq-count tool, and raw counts were analyzed for differential expression using the DESeq2 package in R. Ingenuity Pathway Analysis (IPA) was used to analyze pathway enrichment. Functional data was acquired from, patient salivary gland biopsies and submandibular glands from NOD Aire-/- mice, and minced to obtain cell clusters. Loaded clusters with the cytosolic calcium indicator (Fluo-2AM) were used for live-cell imaging using confocal microscopy to quantitate cytosolic calcium fluxes induced by parasympathetic stimulation.

**Results.** IPA analysis displayed enrichment of immune-related pathways including downregulation of IFN pathways and genes associated with saliva secretion in the patients. The most significant down-regulated gene was KCNMA1; a potassium calcium-activated channel critical for maintaining fluid secretion. In support of this finding, the functional data displayed at tenuation of calcium signaling at the level of calcium release in two of three patients and in the NOD Aire-/- mice. Aquaporin 3 (AQP3), a highly AIRE–dependent gene in medullary thymic epithelial cells, was also downregulated; AQP3 localized on the basolateral side of acinar cells is known to participate in transcellular osmotic water flow.

**Conclusion.** Our study has identified specific target genes that may be implicated in the pathogenesis of Sjögren's-like syndrome in APECED patients. Ongoing experiments are aimed at validating these target genes, at examining the transcriptome profile of Aire-/- salivary glands, and at determining the presence of autoantibodies against KCNMA1 or AQP3 in this population.

#### P-122

#### A Novel Graph-Approach Applied to Modular-Analysis Identifies Shared Gradual Whole Blood IFN Signatures in Primary Sjögren's Syndrome and Systemic Lupus Erythematosus, and Reveals New IFN-related Modules in Disease Progression

Naomi I. Maria<sup>1</sup>,Noémie Jourde-Chiche<sup>2,3</sup>, Peter K. Gregersen<sup>4</sup>, Damien Chaussabel<sup>5</sup>, Laurent Chiche<sup>6</sup>, Ilya Korsunsky<sup>4</sup>.

<sup>1</sup>Center for Autoimmunity, Musculoskeletal & Hematopoietic Diseases, Feinstein Institute for Medical Research, Manhasset, New York, USA. <sup>2</sup>Aix-Marseille Univ, Department of Nephrology, AP-HM, Hôpital Conception, Marseille, France. <sup>3</sup>Aix-Marseille Univ, UMR\_S 1076, Vascular Research Center of Marseille, Marseille, France. <sup>4</sup>Robert S. Boas Center for Genomics & Human Genetics, Feinstein Institute for Medical Research, NY. <sup>5</sup>Systems Biology Department, Sidra Medical and Research Center, Doha, Qatar. <sup>9</sup>Department of Internal Medicine, Hôpital Européen, Marseille, France. **Background.** There is significant clinical and molecular heterogeneity among patients suffering from systemic autoimmune diseases, such as primary Sjögren's Syndrome (pSS). Deciphering this heterogeneity could allow the molecular stratification of patients in terms of prognosis and therapeutic targets. Our previous work using a Modular Repertoire Analysis (MRA) has demonstrated that the IFN signature observed in systemic lupus erythematosus (SLE) patients is not restricted to a mere type I IFN signature, but involves the gradual activation of 3 distinct IFN modules driven by various IFN types including IFNg. Although type I and type II IFN signatures have been described in patients with pSS, a detailed MRA in pSS is still lacking. Here we aimed to refine MRA and discover new transcriptional signatures in pSS, by applying a novel Graph-Theoretic-Approach (GTA) to reveal the progression of module activation patterns.

**Methods.** Blood transcriptomic microarray datasets, including that of pSS patients (n=133; UKPSSR) fulfilling American European Consensus Group (AECG)-criteria and SLE patients (n=157 samples; LUPUCE cohort) fulfilling ACR-criteria, were analyzed using MRA followed by GTA. MRA was performed using a blood modular framework comprising 260 modules. A novel GTA, based on the Extended Suppes Bayes Causal Network (ESBCN), was used to generate an ordered, branching progression model of modular activation. Disease-specific causal graphs in selected datasets were built in order to generate hypotheses regarding disease progression for a particular disease. Significance to clinical characteristics was evaluated using Fisher's exact test and ANOVA, for categorical and continuous characteristics, respectively.

Results. The GTA-to-MRA analysis confirmed the previously described pattern of gradual activation of IFN modules in SLE patients: first M1.2 (81.5%), then M3.4 (67.5%) and finally M5.12 (22.3%). Interestingly, this gradual modular IFN signature was similarly observed in pSS patients who exhibited activation of 1 (64%), 2 (37%) or all 3 (8%) IFN modules. Additionally, GTA-to-MRA identified a dual mode of disease progression in SLE after the activation of the IFN modules M1.2 and M3.4: either completion of the IFN signature, to include the more IFNg- related module M5.12 or with completion of a newly identified 4th IFN-related module M8.59, or the activation of a neutrophil module M5.15 associated with renal involvement. In pSS, a dual mode of progression identified comparable completion of IFN signature to include M5.12, ending with IFN module M8.59, or activation of a 5th IFN-related module M8.95. Solely 7% of pSS patients portrayed a neutrophil signature. Conclusion. With The application of GTA to blood MRA, we are the first to show a detailed modular IFN signature in pSS. Here we show the sharing of gradual activation of IFN modules between pSS and SLE and the identification of new IFN-related modules through the observation of progression patterns, with both shared and distinct downstream progression patterns in pSS and SLE. Defining distinct molecular subgroups in pSS will aid in development of more tailored therapeutic regimens.

### P-123

#### Comprehensive proteomic profiling of Sjögren's syndrome revealed dysregulation of interferon and other immunologic pathways

Jingya Wang<sup>1</sup>, Shu Wang<sup>1</sup>, Saifur Rahman<sup>1</sup>, Varsha Kumar<sup>1</sup>, Jodi Karnell<sup>1</sup>, Molecular Medicine Team<sup>1</sup>, Roland Kolbeck<sup>1</sup>, Ilias Alevizos<sup>2</sup>, Rachel Ettinger<sup>1</sup> <sup>1</sup>MedImmune, LLC; Gaithersburg, MD, <sup>2</sup>NIH, NIDCR; Bethesda, MD

Siögren's syndrome (SS) is the second most common Rheumatic autoimmune disease with a broad spectrum of clinical and serological manifestations and no approved biologics. To better understand the pathogenesis of SS and explore potential soluble biomarkers and therapeutic targets, sera from 109 SS and 50 healthy controls (HC) were examined. Comprehensive profiling was performed using autoantigen and proteomic arrays (UTSW, 95 autoAb's; RBM's, 165 proteins; and SOMAscan, 1129 protein). Data showed that autoAb's against Ro, La, U1sn-RNP-BB', U1sn-RNP-C and Ribo phaspho protein P1 significantly increased in SS compared to HC (Fold change >1.5 and FDR <0.05) that positively correlate with FOCUS disease score (FDR<0.05). RBM and SOMAscan were highly consistent and detected 123 unique proteins up- or down- regulated in SS compared to HC. Strikingly, 71 were interferon-regulated proteins, with IP10 correlating best with FOCUS score (r=0.65). Pathway analysis also confirmed strong activation of type I interferon signaling. Type I interferon signature genes were also observed in salivary gland of the NOD.H-2h4 mice, a model that closely resembles human SS. Additional proteins dysregulated in SS patent sera include the TNF signaling, antimicrobial proteins A2M and B2M, and multiple SLAMF family members, which may shed light on disease mechanisms and novel therapeutic strategies.

# **P-124**

Aberrant cell signaling in peripheral blood mononuclear cells upon interferon alpha stimulation in patients with primary Sjögren's syndrome associates with type I interferon signature

Richard Davies<sup>1</sup>, Daniel Hammenfors<sup>1,2</sup>, Brith Bergum<sup>1</sup>, Petra Vogelsang<sup>1</sup>, Sonia Gavasso<sup>3</sup>, Johan G. Brun<sup>2,3</sup>, Roland Jonsson<sup>1,2</sup>, Silke Appel<sup>1</sup>.

Broggelmann Research Laboratory, Department of Clinical Science, University of Bergen, Bergen, Norway. <sup>2</sup>Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>3</sup>Department of Neurology, Haukeland University Hospital, Bergen, Norway.

**Background.** Primary Sjögren's syndrome (pSS) is a complex heterogeneous systemic autoimmune disease. Biomarkers for patient stratification are scarce. Several single nucleotide polymorphisms within type I interferon (IFN) signalling pathways are associated with pSS. To define novel biomarkers for pSS patient stratification, we analysed the temporal profile of MAPK/ ERK and JAK/STAT signalling networks in peripheral blood mononuclear cells (PBMC) upon stimulation with IFNα by flow cytometry.

**Methods.** PBMC from pSS patients and healthy matched donors were stimulated for 15, 30, 60, 120, 180, and 240 min with IFN $\alpha$  at 100 ng/ml. Nine different phosphor-epitopes were analyzed: STAT4(pY693), ERK1/2(pT202/pY204), NF- $\kappa$ B p65(pS529), STAT1(pS727), STAT1(pY701), p38 MAPK(pT180/pY182), STAT3(pS727), STAT3(pY705) and STAT5(pY694). To define leukocyte populations, scatter properties and cell surface markers CD3, CD20 and CD56 were utilized.

**Results.** Cells from pSS patients displayed significant differences in basal and IFN $\alpha$  induced phosphorylation levels of numerous signalling proteins compared to cells from healthy donors. Principal component analysis (PCA) using IFN $\alpha$  induced phosphorylation levels after 15 minutes showed clustering of pSS patients and pSS patient subgroups. PCA visualization showed a positive shift for pSS samples away from healthy donor samples with positive movement influenced by changes in phosphorylation of STAT1 Y701 in T, NK and B cells in PC1, and PC2 positive movement influenced by STAT1 Y701 in NK and B cells, and negative movement by STAT3 S727 in T cells. Medicated and autoantibody negative patients grouped closer to healthy donors than non-medicated or autoantibody positive patients.

**Conclusion.** pSS patients show increased responses to IFN $\alpha$  through STAT1. Increased responses to IFN $\alpha$  may in part drive an up-regulation of interferon induced genes.

# P-125

#### Imbalance in subsets of circulating innate lymphoid cells is associated with disease activity and type i interferon signature in primary Sjögren's syndrome

SLM Blokland<sup>\*1,2</sup>, LL van den Hoogen<sup>\*1,2</sup>, EFA Leijten<sup>1,2</sup>, AA Kruize<sup>1</sup>, TRDJ Radstake<sup>1,2</sup>, JAG van Roon<sup>1,2</sup>.

<sup>1</sup>Department of Rheumatology & Clinical Immunology, University Medical Center Utrecht, The Netherlands. <sup>2</sup>Laboratory of Translational Immunology, Department of Immunology, University Medical Center Utrecht, Utrecht University, The Netherlands.

Background. Recent studies indicate an important role for innate lymphoid cells (ILCs) in the pathophysiology of rheumatic diseases. In rheumatoid arthritis and spondyloarthropathies elevated numbers of subsets of ILCs have been found at the site of inflammation producing cytokines including IFN-y and IL-22 and in addition, group 3 ILC have been suggested to be involved in formation of ectopic lymphoid structures in rheumatic diseases (Shikhagaie Nat Rev Rheumatol 2017, Wenink A&R 2017). ILC3-like cells producing IL-22 have been found in the salivary glands of pSS patients (Ciccia ARD 2012). However, circulating ILC have not yet been studied in primary Sjögren's syndrome (pSS) and systemic lupus erythematosus (SLE). SLE and pSS are characterized by presence of a type I interferon (IFN) signature in a large proportion of the patients. Animal studies in HIV and asthma implicate type I IFN, produced by plasmacytoid dendritic cells (pDCs), to regulate the survival of group 2 and group 3 ILCs (ILC2 and ILC3) via increase of Fas (CD95) expression, rendering the ILC more susceptible to apoptosis (Maazi JACI 2017, Zhang JCI 2015, Duerr Nat Immunol 2016). In this study, we explored for the first time the frequency and phenotype of circulating ILC in pSS and SLE and their relation to the IFN signature.

**Methods.** Frequencies and phenotypes of ILC subsets and pDCs were assessed by flow cytometry in peripheral blood of patients with pSS (n=20), SLE (n=20) and healthy controls (n=17). Patients were stratified by the

presence or absence of an IFN signature as assessed by RT-qPCR on peripheral blood mononuclear cells as previously described (Brkic ARD 2013). **Results.** ILC1 numbers were increased in peripheral blood of patients with SLE as compared to healthy controls and in pSS patients ILC1 numbers correlated with disease activity (ESSDAI score), serum IgG levels and anti-SSB autoantibodies (all p<0.05). Numbers of ILC1, ILC2 or ILC3 did not significantly differ between patients with SLE and pSS. However, patients with a high expression of the type I IFN signature had significantly decreased numbers of ILC2 and ILC3 (p=0.04 and p=0.02, respectively). The decrease of ILC2 and ILC3 was related to increased expression of Fas (CD95) on these cells in patients with a high type I IFN signature (both p=0.01).

**Conclusion.** Both in SLE and pSS, the presence of a type I IFN signature is related to reduced numbers of circulating ILC2 and ILC3 in association with increased Fas expression on these cells possibly rendering them more susceptible to Fas/FasL-dependent apoptosis at peripheral sites.

# P-126

#### Y RNA and other circulating RNAs in Sjögren's syndrome

Gilad Wasserman DMD/MSc<sup>1,2,3,4</sup>, Devora Gur-Wahnon PhD<sup>3,5</sup>, Hadas Ben-Eli PhD<sup>6,8</sup>, Ora Paltiel MDCM/MSc/FRCPC<sup>6,7</sup>, Avraham Solomon MD8, Iddo Z Ben-Dov MD/PhD<sup>\*3,5</sup>, Doron J Aframian DMD/PhD<sup>\*1,2,4</sup>.

All authors are affiliated with the Hadassah-Hebrew University Medical Center <sup>1</sup>Department of Oral Medicine, Sedation, and Maxillofacial Radiology. <sup>2</sup>Sjögren's Syndrome Center. <sup>3</sup>Laboratory of Medical Transcriptomics. <sup>4</sup>Laboratory of Salivary Diagnostics. <sup>5</sup>Department of Nephrology. <sup>6</sup>Braun School of Public Health and Community Medicine. <sup>7</sup>Department of Hematology. <sup>8</sup>Department of Ophthalmology. \*These authors contributed equally to this work.

**Background.** There are no clear diagnostic criteria for SS and the final diagnosis is essentially clinical. Biomarkers for disease diagnosis, progression assessment, and patient stratification would undoubtedly be "game changers". Various types of non-coding RNA (ncRNA) such as miRNA, tRNAs and fragments, and Y RNA have been studied as potential biomarkers for specific diseases.

In Next Generation Sequencing (NGS) vast amounts of sequencing reads are produced in a fast and accurate manner, but is still rather costly and at times the depth of NGS may be excessive. Modern methods have been devised in order to sequence more samples in a single NGS flow cell thus reducing the cost while homing in on a more specific subset of sequences.

The objective of this work is to discover novel ncRNA SS biomarkers in serum via a modern adaptation to NGS.

**Methods.** This is an ancillary study to a previous case-control on risk factors for SS (initiated in 2011 with AECG inclusion criteria). Forty-four female patient sera from previous collection were selected: 22 Lymphoma free patients that met the 2016 classification criteria for SS and 22 age matched DES patients served as control. Patients were grouped according to subjective reporting of their highest burden: ocular dryness, oral dryness, or if they are primarily diagnosed with Rheumatoid Arthritis.

Total RNA was extracted from sera using an inhouse protocol. Barcoded pooled RNA libraries were produced as described previously by Hafner et al (Methods 58 (2012) 164–170). The libraries underwent simultaneous NGS using Illumina NextSeq.

Then samples were extracted via their barcode and analyzed insilico using "RNA WORLD", edgeR in R-Studio, and GOTTCHA.

Table I. SS vs DES edgeR analysis.

	logFC	logCPM	<i>p</i> -value
trnaV12A67G	2.8	8.5	1.00E-04
trnaV2T20CC34G	2.7	8.7	5.40E-05
trnaV2T20CG26TC34G	2.7	8.5	1.10E-04
TRNAV1	2.7	8.6	1.10E-04
trnaV2C34G	2.7	8.6	1.10E-04
trnaV2T19CT20CC34G	2.7	8.5	1.10E-04
TRNAV2	2.7	8.6	1.10E-04
TRNAV10	2.8	8.7	4.80E-05
trnaV2G26T	2.8	8.5	1.00E-04
TRNAP17	1.9	6.8	2.10E-03
RNU5B-1_ENST00000363286	-2.2	5.9	2.09E-03
mu5b-1T25A	-2.2	5.9	2.02E-03
hsa-mir-31	-4.2	5.7	1.90E-03
trnaN15A59G	-4.3	6.2	4.77E-05
SNORD38B_ENST00000384690	-5.6	6.0	5.57E-04

logFC: log, Fold change; logCPM: log, Counts per million.





**Fig. 1.**  $Log_2$  fold change of the 4 types of Y RNA in SS patients relative to DES. Subgroup E had mainly ocular (Eye) symptoms, Subgroup M had mainly oral (Mouth) symptoms, and subgroup RA were secondary to Rheumatoid Arthritis.

**Results.** Nine variants of Valine tRNA were upregulated by an average of 6.6 fold (logFC=2.7) in SS vs DES, several other RNAs were down regulated (Table I). Y RNA was depleted in SS secondary to RA vs DES while it was slightly upregulated in the other SS groups. Only the R4 variant change was statistically significant, but there is a general trend (Figure 1).

**Conclusions.** Circulating ncRNA has the potential to uncover SS biomarkers. Further large scale multinational clinical trials are needed to validate this observation. The depletion of Y RNA in serum of SS patients is particularly appealing since this highly conserved sequence is part of the Ro60 RNP and this depletion could be explained by autoimmune mediated degradation. The relevance of Y RNA in SS was recently hypothesized by Kabeerdoss *et al.*: "Y RNA derived small RNAs in Sjögren's syndrome: Candidate biomarkers?" *Int J Rheum Dis.* 2017 Nov 19.

#### **P-127**

#### Decreased expression of microRNA 130a indicates dysregulation of classical dendritic cells in patients with primary Sjögren's syndrome

Lopes AP<sup>1,2</sup>, van Roon JAG<sup>1,2</sup>, Blokland SLM<sup>1,2</sup>, Wang M<sup>4</sup>, Chouri E<sup>1,2</sup>, Kruize AA<sup>1</sup>, Burgering BMT<sup>4</sup>, Rossato M<sup>2,3</sup>, Radstake TRDJ<sup>1,2</sup>, Hillen MR<sup>1,2</sup>. <sup>1</sup>Department of Rheumatology & Clinical Immunology, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands. <sup>2</sup>Laboratory of Translational Immunology, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands. <sup>3</sup>Functional Genomics Center, University of Verona, Italy. <sup>4</sup>Center for Molecular Medicine, University Medical Center Utrecht, Utrecht University, Utrecht, the Netherlands.

**Background.** Primary Sjögren's syndrome (pSS) is a systemic autoimmune disease characterized by lymphocytic infiltration of the exocrine glands and dryness of mouth and eyes. Classical dendritic cells (cDCs) are very potent antigen presenting cells known to induce strong T-cell proliferation and cy-tokine production. Considering the critical role of microRNAs (miRNAs) in regulation of gene expression, we investigated miRNA expression in isolated CD1c-expressing cDCs of patients with pSS.

**Methods.** Two independent cohorts consisting of pSS patients and healthy controls were established: a discovery cohort (15 pSS, 6 HC) was used to screen the expression of a large panel of 758 miRNAs, while a validation cohort (14 pSS, 11 HC) tested the reproducibility of the results. CD1c-expressing cDCs were isolated from peripheral blood using MACS and miRNA profiling of 758 targets was performed using the OpenArray platform in the donors included in the discovery cohort. A selection of 16 miRNAs found to be differentially expressed in the pSS group compared to the control group (p<0.05, with a difference between the groups of >log2) was measured in the independent validation cohort using a custom-made array. Isolated cDCs from HC were stimulated with a panel of Toll-like receptor (TLR) ligands and the expression of miR-130a on protein synthesis was analysed by using the pulsed stable

isotope labelling by amino acids in cell culture (pSILAC) method (quantitative mass spectrometry-based technique) in a HEK-293T cell-line.

**Results.** A total of 24 miRNAs was downregulated in pSS patients versus HC in the discovery cohort. Of the 16 selected targets for replication, decreased miR-130a and miR-708 were validated. cDC activation through TLR3 and TLR7/8 downregulates the expression of both miRNA-130a and miRNA-708. Transfection with miR-130a resulted in downregulation of proteins involved in NF-kB signalling.

**Conclusions.** miR-130a and miR-708 are significantly downregulated in cDCs of patients with pSS. We show that the expression of these miRNAs is decreased upon cDC activation and that transfection with miR-130a downregulates the expression of proteins involved in the NF-kB pathway. As such, these miRNAs seem to be involved in cDC activation and reflect enhanced activation of cDCs from peripheral blood of pSS patients.

#### **P-128**

# Identification of dysregulated Interferon-inducible non-coding RNAs in Sjögren's syndrome

Nicolas Means<sup>1,2</sup>, John A Ice<sup>1</sup>, Indra Adrianto<sup>3</sup>, Anna M. Stolarczyk<sup>1</sup>, Michelle Joachims<sup>1</sup>, Astrid Rasmussen<sup>1</sup>, Joel M. Guthridge<sup>1</sup>, Judith A. James<sup>1,2,4</sup>, R. Hal Scofield<sup>1,4,5</sup> Kathy L. Sivils<sup>1,2</sup>, Christopher J. Lessard<sup>1,2</sup>. <sup>1</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, Oklahoma, USA. <sup>2</sup>Department of Pathology, University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma, USA. <sup>3</sup>Department of Public Health Sciences, Henry Ford Health System, Detroit, MI, USA. <sup>4</sup>Department of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma, USA. <sup>5</sup>Department of Veterans Affairs Medical Center, Oklahoma City, Oklahoma, USA.

**Background.** Sjögren's syndrome (SS) is a chronic, heterogeneous disease with hallmark features of auto-inflammation and autoantibody production. Upregulation of type I and II interferon-stimulated genes (ISGs), known as the "Interferon (IFN) Signature" is correlated with anti-Ro titers and has been observed both in the salivary glands and peripheral blood of SS patients. Within the 2p25.2 genomic interval, the long non-coding RNA (IncR-NA) negative regulator of the interferon response (*NRIR*) has been identified as inducible by type I IFN and is responsible for the downregulation of the ISGs *CMPK2* and *RSAD2* (Kambara *et al.*, *Nucleic Acids* 2014). We sought to identify additional unannotated ISGs IncRNAs that are differentially expressed (DE) in SS patients utilizing correlated expression of *RSAD2*.

**Methods.** In this study, we evaluated and compared the transcriptome of anti-Ro(+) patients (n=27) and healthy controls (n=27) using RNA-seq with DE defined as q < 0.05 and a fold change (FC)  $\geq 2$  or  $\pm 0.5$ . qRT-PCR was used to validate DE with primers specific for each RNA of interest. To understand the biological relevance of these transcripts, we performed *in vitro* time-course experiments to compare the transcriptional changes of unstimulated cells and cells stimulated with either PMA/ionomycin (PMA/I) or universal type I IFN for 36 hours measuring 7 time points.

Results. Given the importance of the IFN signature among the SS Ro(+) patients we sought to identify lncRNAs with a potential role in this pathway. One of the most overexpressed type I ISGs in the SS Ro(+) is RSAD2 (FC=8.05,  $p=3.29 \times 10E-07$ ). Since RSAD2 plays a role in the type I IFN pathway, pairwise correlation coefficients between all the DE transcripts and RSAD2 for SS Ro(+) patients were calculated. In total, we found 223 transcripts exhibiting correlation with RSAD2 expression (Pearson's r >+0.70 or <-0.60), including NRIR (FC=2.72, p=5.87x10E-03) and CMPK2 (FC=2.53, p=3.58x10E-03). Of these 223 transcripts, 14 DE expressed lncRNAs correlated with RSAD2 expression. Several antisense (AS) ncRNAs situated in close proximity to other type I ISGs correlated with RSAD2, including: AC099063.1 (FC=2.51), AC004551.1 (FC=3.35), and AP001610.1 (FC=4.12). We confirmed upregulation of these lncRNAs by qRT-PCR from the independent replication (14 Ro(+) and 36 controls) cohort (p=2.49X10E-02, 8.63X10E-06, 1.17X10E-04, respectively). Based on the locations of the lncRNAs to type I ISGs, HSB-2 cells were stimulated with PMA/I or universal type I IFN. Using qRT-PCR to measure the protein coding genes MX1, OAS1, and GBP5 along with the lncRNAs, AC099063.1 (GBP5-AS1), AP001610.1 (MX1-AS1), and AC004551.1 (OAS123-AS1) showed coordinated regulation with their protein coding counterparts with both stimuli.

**Conclusions.** Given the importance of the IFN signature to disease pathogenesis in the autoantibody positive patients, it is critical that we better understand how this complex pathway is regulated so coordinately. Since one critical function of lncRNAs is to regulate the genome, characterizing dysregulation of these mechanisms in SS could result in new therapeutic.

### P-129

### Overexpression of miR-146a correlates with the down-regulation of FADD in primary Sjögren's syndrome

Huan Shi<sup>1</sup>, Ling-yan Zheng<sup>1</sup>, Zhijun Wang<sup>1</sup>, Li-song Xie<sup>1</sup>, Ping Zhang<sup>1,2</sup>, and Chuang-qi Yu\*<sup>1</sup>.

<sup>1</sup>Department of Oral Surgery, Affiliated Shanghai 9<sup>th</sup> People's Hospital, Shanghai Jiao Tong University, School of Medicine, Shanghai Key Laboratory of Stomatology, 639 Zhi-Zao-Ju Road, Shanghai, China. <sup>2</sup>Department of Oral and Maxillofacial-Head Neck Oncology, 9<sup>th</sup> People's Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai Key Laboratory of Stomatology, 639 Zhi-Zao-Ju Road, Shanghai, China.

**Objective.** Increasing data indicate a key role for miR-146a in the maintenance of immune homeostasis. This study was undertaken to investigate the contribution of miR-146a to the pathogenesis of primary Sjögren's syndrome (pSS).

**Method.** The expression levels of miR-146a and its potential target genes IRAK-1, TRAF-6, and FADD were assessed by real-time PCR in peripheral blood mononuclear cells (PBMCs) obtained from 22 pSS patients and 22 healthy controls. The relationship between these levels and those of the cy-tokines IL-1 and TNF- $\alpha$  were also investigated.

**Result.** Compared with healthy controls, the expression level of miR-146a was significantly increased in PBMCs from pSS patients and was positively correlated with the visual analog scale of parotid swelling, the grading of labial salivary gland biopsies, and the expression level of IL-1 and TNF- $\alpha$ . The target genes FADD and IRAK-1 were down-regulated in PBMCs from pSS patients, while TRAF-6 remained unchanged. The increasing expression of miR-146a was significantly correlated with the down-regulation of FADD, but not IRAK-1 or TRAF-6, in PBMCs from pSS patients.

**Conclusions.** The overexpression of miR-146a in PBMCs from pSS patients significantly correlated with the down-regulation of FADD, but not TRAF-6 and IRAK-1, indicating the complex functions of miR-146a in pSS. On the one hand, miR-146a-mediated down-regulation of FADD leads to resistance among self-reactive lymphocytes to Fas-mediated apoptosis; on the other hand, increasing levels of miR-146a fail to regulate TRAF-6 and IRAK-1 and result in prolonged activation of the NF-κB pathway. However, both of these pathways may play key roles in the pathogenesis of pSS. **Keywords.** Sjögren's syndrome, miR-146a, FADD, IRAK-1, TRAF-6

#### **P-130**

#### Involvement of gap junctional comunication in salivary gland dysfunction related to primary Sjögren's syndrome

Letizia Mattii<sup>1</sup>, Enza Polizzi<sup>2</sup>, Francesco Ferro<sup>2</sup>, Valentina Donati<sup>3</sup>, Antonella Cecchettini<sup>4</sup>, Chiara Baldini<sup>2</sup>.

<sup>1</sup>Section Histology Department of Clinic and Experimental Medicine, University of Pisa. <sup>2</sup>Rheumatology Unit, Department of Clinic and Experimental Medicine, University of Pisa. <sup>3</sup>Unit of Anatomic Pathology II, Azienda Ospedaliero-Universitaria Pisana. <sup>4</sup>Institute of Clinical Physiology, CNR Pisa, Italy.

**Background.** Gap junctions and distinct connexin (Cx) isoforms play a crucial regulatory role in various aspects of glandular function. The acinar cells of salivary glands are coupled by Cx26 and Cx32 channels and it has been suggested that these gap junction channels may control salivary secretion via the inhibition of capacitative Ca2+ entry. To date few data are available regarding the role of connexins in salivary dysfunction related to primary Sjögren's syndrome (pSS)

**Aim of this study.** to characterize the expression of Cx26 in minor salivary gland biopsies (MSGBs) of patients with pSS in order to investigate their potential role in salivary gland inflammation and hypofunction.

**Methods.** We analyzed the expression of Cx26 in MSGBs from pSS patients and from non-immune mediated sicca subjects, (no-SS). MSGB were obtained as part of the routine diagnostic procedures when pSS was suspected. In all cases, part of the MSG specimen was fixed in neutral-buffered formalin for the assessment of the focus score (FS), and the remaining part was quickly frozen and stored at -80°C for immunoistochemical experiments. Unstimulated salivary flow was measured as well in all the subjects enrolled. Cx26 expression was evaluated by indirect immunofluorescence, performed using rabbit anti-human Cx26 and mouse anti-human ASMA primary antibodies, was observed under confocal laser scanning microscopy by two blinded, independent researchers. We defined according to a semiquantitative scale 4

levels of Cx26 expression in both acinar and myoepithelial cells (1=severely decreased, 2=moderately decreased, 3=slightly decreased, 4=no decreased). **Results.** We analyzed the expression of Cx26 in 29 MSGBs:18 from pSS patients and 11 no-SS subjects. Immunolabeling for Cx26 was observed at level of luminal, lateral and basal borders of secretory and myoepithelial cells. The expression of Cx26 in both acinar (p=0.02) and myoepithelial cells (p=0.03) from pSS patients was significantly decreased when compared to no-SS subjects. In pSS MSG with a FS≥3, the acinar (p=0.01) and myoepithelial expression (p=0.05) of Cx26 was significantly lower than in pSS samples with a FS<3. Finally, a significant association (p=0.01) was observed between Cx26 expression in MSGB and salivary flow impairment in pSS. **Conclusions.** This pilot study has shown that gap junctional communica-

tion may be involved in pSS-salivary gland dysfunction. Further studies are necessary to clarify the pathogenetic role of Cx proteins in salivary gland hypofunction and whether these proteins may represent a novel target for future therapies.

# P-131

#### Role of Nuclear factor of activated T-cells 5 in hyperosmolar stress-induced osmoadaptative and inflammatory responses in human salivary glands cells

Clara Chivasso, Angélic Bryla, Françoise Gregoire, Nargis Bolaky, Valérie Delforge, Jason Perret and Christine Delporte.

Laboratory of Pathophysiological and Nutritional Biochemistry, Université Libre de Bruxelles, Brussels, Belgium.

Background. Cells can be subjected to hyperosmotic stress (HOS) under physiological and pathological conditions. Indeed, during the steps leading to saliva secretion, acinar cells are exposed to a transepithelial osmotic gradient, resulting from NaCl secretion and accumulation in the acini lumen, which drives an important transcellular water flux through apically-localized aquaporin-5. Within the ductal lumen, the composition of the primary secreted isotonic fluid is modified, leading to the secretion of a final hypotonic saliva. In salivary glands of patients suffering from Sjögren's syndrome, characterized by gland destruction and in some cases altered aquaporin-5 subcellular localization, acinar cells are hypothesized to be subjected to chronic HOS. In response to HOS, cells rapidly initiate an osmoadaptive program that includes the activation of the transcription factor Tonicity Enhancer Binding Protein (TonEBP), also called Nuclear factor of activated T-cells 5 (NFAT5), and subsequent transactivation of osmoprotective genes (such as aldose reductase (AkR1B1) and sodium- and chloride-dependent taurine transporter (TAUT)). Under pathological conditions, HOS may significantly contribute to disease progression by triggering proinflammatory cytokines release. The aim of the present study was to investigate the role of NFAT5 in osmoadaptative and inflammatory responses triggered by HOS in HSG cells. Methods. Human salivary gland HSG cells were stably transfected by electroporation with either a plasmid coding for a scrambled RNA sequence (shCTN) or a plasmid coding for NFAT5 gene silencing short hairpin RNAs (shNFAT5). shCTN- and shNFAT5-stably transfected HSG cells were subjected to iso-osmolar condition (iso-osmolar medium; ISO) or HOS (isoosmolar medium supplemented with various concentrations of NaCl or sucrose). NFAT5 transactivation activity was measured in shCTN- and shN-FAT5-stably transfected HSG cells that were subsequently co- transfected with the reporter plasmid pTonE-SEAP (two NFAT5 binding sites (TonE) upstream of a modified secreted alkaline phosphatase (SEAP) gene) and finally exposed to ISO or HOS. mRNA levels were quantified by real time quantitative PCR (RT-qPCR). Statistical analysis was performed to compare group means using t-test for unique sample and repeated measure ANOVA with post- hoc Bonferroni t-tests.

**Results.** As compared to ISO, HOS significantly increased NFAT5 transactivation activity; NFAT5, AKR1B1 and TAUT mRNA levels; as well as CCL2, IL6, IL8, TGFß mRNA levels in HSG cells. As compared to shCTN, shNFAT5 significantly decreased NFAT5 transactivation activity; NFAT5, AKR1B1, CCL2, IL6, IL8, TGFß (but not TAUT) mRNA levels in HSG cells.

**Conclusions.** Our data suggest that NFAT5 is involved in HOS-triggered osmoadaptative and inflammatory responses in HSG cells. In patients suffering from Sjögren's syndrome, HOS may therefore significantly contribute to disease progression by triggering proinflammatory cytokines release.

# **P-132**

# RNA profiling and genome wide mapping of salivary gland gene regulation reveals core regulators of salivary flow and function

Drew G. Michael, Thomas Pranzatelli, Ida Shinder, Blake Warner, John A. Chiorini.

Extensive efforts have been directed towards understanding the differential gene expression patterns associated with primary Sjögren's syndrome induced salivary dysfunction. Although these studies have extensively characterized the changes associated with pathogenesis, relatively little is known regarding the transcriptional regulators that drive normal salivary gland gene expression.

We have utilized systems levels genomic approaches to comprehensively identify the transcription factors highly expressed and enriched within the salivary gland. Integration of RNA sequencing data with DNase1 digital genomic foot printing was utilized to construct the first whole genome gene regulatory network of salivary gland gene regulation. These techniques utilize genomic foot printing to identify open chromatin regions across the genome and characterize protein bound regions associated with transcription factor affinity and enhancer profiling, a unified map of the salivary gene regulatory network was constructed at a genome wide level. Following network construction, bivariate genomic foot printing techniques were used to compare salivary gland foot printing states with DNase data acquired from lung and heart tissues to identify the salivary specific factors associated with driving normal salivary function.

Collectively, our work serves as a foundation for the expansion of salivary gland biology into non-coding DNA, enabling further investigation into regulatory changes associated with Sjögren's syndrome and identifying the drivers of normal salivary gland differentiation and function.

# P-133

#### The levels, subcellular distribution and posttranslational processing of salivary mucins are affected by pro-inflammatory cytokines in Sjögren's syndrome patients

Isabel Castro<sup>1, 2</sup>, Inka Brockhausen<sup>3</sup>, Sergio Aguilera<sup>4</sup>, María-José Barrera<sup>1</sup>, Sergio Gonzalez<sup>5</sup>, Claudio Molina<sup>5</sup>, Ulises Urzúa<sup>1</sup>, Cecilia Leyton<sup>1, 2</sup>, María-Julieta Gonzalez<sup>1</sup>.

<sup>1</sup>Instituto de Ciencias Biomédicas, Universidad de Chile, Santiago, Chile. <sup>2</sup>Departamento de Tecnología Médica, Universidad de Chile, Santiago, Chile. <sup>3</sup>Queen's University, Kingston, Ontario, Canadá. <sup>4</sup>Clínica INDISA, Santiago, Chile. <sup>5</sup>Universidad Mayor, Santiago, Chile.

Background. Sjögren's syndrome (SS) is a chronic autoimmune disease characterized by inflammation of the exocrine glands and symptoms of oral and ocular dryness. The persistent sensation of oral dryness in SS-patients may be attributable to changes in the quality of mucins, the major component of the viscoelastic layer covering all mucosal surfaces in the body. Mucins are large glycoproteins synthesized in the rough endoplasmic reticulum (RER) and post-translationally processed in the RER and Golgi apparatus where they are N and O-glycosylated, respectively. O-glycosylation consists of the attachment of sialylated and sulfated oligosaccharides conferring mucins the property of retain water. It has been described that pro-inflammatory cytokines induce the expression of certain mucins and modulate post-translational processing of these and other glycoproteins in several cell types. Salivary glands (SG) from SS-patients show high levels of pro-inflammatory cytokines, accumulation of MUC1 and decreased sulfation of MUC5B. We hypothesized that pro-inflammatory cytokines alter the secretory pathway of SG from SS-patients, inducing changes in levels, localization and post-translational processing of salivary mucins

**Methods.** We evaluated the expression and subcellular localization of MUC1, the expression and activity of Golgi glycosyltransferases and Gal3-O-sulfotransferases (Gal3ST), and the effect of pro-inflammatory cytokines on levels and localization of mucins. Relative mRNA and protein levels of mucins and Golgi enzymes were determined by real-time RT-PCR and Western blotting in SG from SS-patients and controls and in HSG-3D acini. Localization analyses were performed by immunofluorescence. Enzymatic activities were quantified in SG using *in vitro* assays with radioactively labeled donor substrates and specific acceptor substrates.

**Results.** MUC1 was overexpressed and accumulated in the endoplasmic reticulum where unexpectedly it was also observed the Golgi tethering protein Giantin. A significant decrease of Gal3ST activity without expression changes was observed in SG from SS-patients. Gal3ST activity directly correlated with MUC5B sulfation and inversely correlated with inflammation. Stimulation of HSG-3D acini with TNF- $\alpha$  or IFN- $\gamma$  induced MUC1 expression and accumulation.

**Conclusions.** Our results suggest that pro-inflammatory cytokines induced the expression of MUC1, which accumulated in the RER of SG from SS-patients, likely by the stress condition of this compartment. As Gal3ST activity was decreased without expression changes, we speculate that altered localization of Giantin could affect the correct targeting of Gal3ST thereby explaining the hyposulfation of mucins observed in SG from SS-patients. The decrease of glandular inflammation could help to restore the secretory pathway of salivary mucins, improving the hydration of the oral mucosa in SS-patients.

Funding: FONDECYT's initiation into research-11170049, FONDECYT-1160015.

#### P-134

# Acinar atrophy and adipose tissue infiltration in salivary gland biopsy are associated with IFN- $\gamma$ pathway inflammation biomarkers

Valéria Valim<sup>1</sup>, Widner M Sardenberg<sup>1</sup>, Maria Carmen LFS Santos<sup>1</sup>, Larissa Carvalho Caser<sup>1</sup>, Johan G Brun<sup>2</sup>, Roland Jonsson<sup>2</sup>, Per Ueland<sup>3</sup>, Piotr M Mydek<sup>2</sup>.

<sup>1</sup>Hospital Universitário Cassiano Antônio de Moraes/Federal University of Espírito Santo. <sup>2</sup>Bröegelman Research Laboratory, Haukeland Hospital, University of Bergen. <sup>3</sup>Bevital Laboratory, Haukeland Hospital, University of Bergen.

**Background.** Acinar atrophy and adipose infiltration are late findings in the salivary glands histology of primary Sjögren's syndrome (pSS). We hypothesized that they could be an evidence of severe disease and associated with disease activity. We assessed inflammatory biomarkers of the IFN- $\gamma$  pathway and its association with acinar atrophy and/or adipose infiltration in the minor salivary gland biopsy in patients with pSS.

**Methods.** Cross-sectional study including patients with pSS (AECG 2002 or ACR/EULAR 2017) submitted to minor salivary gland biopsy and histological analysis. Kynurenines, and neopterin were measured in plasma by liquid chromatography- tandem mass spectrometry. Non-parametric statistics was applied with p-value of 0.05.

Results. 99 patients with pSS performed labial salivary gland biopsy (LSGB), 95.9% were women, 51.49±12.13 years, Anti-Ro positive in 68% (n=70/103) and Eular Sjögren's Syndrome Disease Activity Index (ESSDAI) 3.63±5.12. LSGB histopathology showed 73.7% (n=73/99) focal lymphocytic sialadenitis with focus score  $\geq 1$ , non-specific chronic sialadenitis in 15.2% (n=15/99), hypotrophic chronic sialadenitis in 10.1% (n=10/99) and within normal limits (no lymphocytes) in 1% (n=1/99). No case of sclerosing chronic sialadenitis, granulomatous inflammation or marginal zone (MALT) lymphoma. The majority of patients had some grade of acinar atrophy (70.4%, n=57/81), ductal dilatation (86.4%, n=70/81), adipose infiltration (51.2%, n=39/80). Acinar atrophy was present in older individuals (53.46±10.09 vs. 42.83±13.3 years, p=0.001), associated with a higher frequency of menopause (57.9% vs. 33.3%; p=0.043), dryness (ES-SPRI >5) (78.2% vs. 50%, p=0.018), more active disease (ESSDAI  $\geq$ 5) (33.3% vs. 8.3%, p=0.017) and higher levels of neopterin  $(25.54\pm1.125 \text{ vs.})$ 20.67±12.58 nmol/L, p=0.040). There was no association with disease duration, anti-Ro, glandular dysfunction (unstimulated whole salivary flow) and smoking. Adipose infiltration was also present in older individuals (53.49±12.33 vs. 47.51±11.29, p=0.016), associated with lacrimal dys-function (Schirmer ≤5 mm) (69.2% vs. 41%, p=0.012) and higher quinolinic acid (503.35±193.30 vs. 427.35±285.76 nmol/L, p=0.029), kynurenine (1.99±0.6, 54 vs. 1.61±0.46 µmol/L, p=0.006), kynurenine/tryptophan ratio (KTR) (0.030±0.09 vs. 0.025±0.01, p=0.031) and anthranilic acid (03±4.96 vs. 16.46±5.24 nmol/L, p=0.022)

**Conclusions.** Acinar atrophy and adipose infiltration are associated with disease activity, symptoms, and activation of the INF  $\gamma$  pathway. Kynurenine pathway metabolites are associated with adipose infiltration and neopterin is associated with acinar atrophy.

# P-135

Global transcriptome analysis of salivary gland epithelia from Sjögren's and non-Sjögren's patients reveals a novel subpopulation with mixed clinical and histopathological features

Rongjuan Mi<sup>1,2</sup>, Vinay Kartha<sup>3</sup>, Ariana Dela Cruz<sup>1</sup>, Taylor Reynolds<sup>4</sup>, Michael Mingueneau<sup>4</sup>, Stefano Monti<sup>3</sup>, Janicke Jensen<sup>5</sup>, Kathrine Skarstein<sup>6</sup>, \*Xaralabos Varelas<sup>2</sup> and \*Maria Kukuruzinska<sup>1</sup>.

<sup>1</sup>Department of Molecular and Cell Biology, Boston University School of Dental Medicine; <sup>2</sup>Department of Biochemistry, Boston University School of Medicine; <sup>3</sup>Department of Medicine, Boston University School of Medicine; <sup>4</sup>Biogen Idec; <sup>5</sup>Faculty of Dentistry, University of Oslo, Norway; <sup>6</sup>Department of Clinical Medicine, University of Bergen, Norway. \*co-senior authors.

**Background.** Sjögren's syndrome (SS) is a complex autoimmune disease associated with lymphocytic infiltration and secretory dysfunction of salivary and lacrimal glands that are frequently accompanied by pro-fibrotic changes in the underlying stromal compartment. Although the current model postulates that primary SS is a secondary effect of lymphocytic infiltrates, increasing evidence suggests that structural defects in the salivary gland epithelia precede and contribute to pathological immune and fibrotic responses in this disease. To gain insights into the molecular changes underlying SS epithelia that may impact immune and fibrotic responses, we carried out global gene expression analyses of labial salivary gland epithelia from SS and non-SS patients using laser capture microdissection (LCM) followed by computational genomic analyses.

**Methods.** Labial salivary glands were obtained from biopsies from 16 patients with sicca symptoms. Based on the American-European Consensus Criteria, these patients were sub-divided into 8 SS (focus score (FS) 1-2) and 8 non-SS sicca controls (FS<1). RNA was isolated from laser capture microdissected epithelia and analyzed by RNAseq followed by computational interrogation of gene expression signatures using genome-wide differential expression testing (DEseq2) for multiple group comparisons.

**Results.** Analyses of RNAseq-based gene expression profiles derived from SS and non-SS samples revealed three distinct subclusters. These included a SS subcluster (n=3), a non-SS subcluster (n=3) and a subcluster that consists of SS and non-SS samples (n=10). Differential expression analysis between the different subclusters revealed signaling events that may be associated with differential stages of SS epithelial pathology. Notable signals included the enrichment of INFgamma and JAK/STAT-regulated genes, and the induction of genes encoding secreted factors implicated in immune responses. **Conclusion.** Our study has identified potential molecular subtypes of SS pathology, defined by gene expression signatures of salivary epithelia that are associated with mixed clinical and histopathological characteristics. Our observations also suggest that gene expression alterations arising in the salivary epithelia contribute to the etiology of SS. We postulate that these molecular sub-groups reflect varying stages of disease predisposition and may offer novel insight into the signals contributing to the progression of SS.

### P-136

#### Are unfolded protein response pathways modulated by inflammation in salivary glands of Sjögren's syndrome patients?

Barrera MJ<sup>1</sup>, Aguilera S<sup>2</sup>, Bahamondes V<sup>1</sup>, Sepúlveda D<sup>1</sup>, Castro I<sup>1</sup>, Matus S<sup>1</sup>, Carvajal P<sup>1</sup>, González S<sup>3</sup>, Molina C<sup>3</sup>, Urzúa U<sup>1</sup> and González MJ<sup>1</sup>. <sup>1</sup>Instituto de Ciencias Biomédicas, Universidad de Chile, Santiago, Chile. <sup>2</sup>Clínica INDISA, Santiago, Chile. <sup>3</sup>Universidad Mayor, Santiago, Chile.

**Background.** Sjögren's syndrome (SS) is an autoimmune epitheliitis that mainly affects the salivary and lacrimal glands. The glandular inflammation has been associated with endoplasmic reticulum (ER) stress. The unfolded protein response (UPR) helps to recover and preserve ER homeostasis and is mediated by the activation of IRE1 $\alpha$ , PERK and ATF6 $\alpha$  sensors. Salivary gland (SG) acinar cells of SS-patients show an altered secretory route indicative of ER stress. The aim of this study was to evaluate the expression, promoter DNA methylation and localization of several UPR pathway components in labial SG (LSG) from SS-patients. Further, we analyzed the expression of these components induced by pro-inflammatory cytokines *in vitro*. **Methods.** mRNA and protein levels were measured by qPCR and western blot in LSG of SS-patients (n=20) and control-subjects (n=22). Promoter DNA methylation was evaluated by MS-HRM. Localization was analyzed

by immunofluorescence. Correlation analysis was performed by Pearson's test. The effect of IFN- $\gamma$  on UPR pathways was evaluated in 3D-acini.

**Results.** A significant decrease of IRE1 $\alpha$ , XBP-1u, XBP-1s, total XBP-1, and GRP78 mRNAs (IRE1 $\alpha$  pathway) was observed in LSG of SS-patients, which was correlated with increased methylation levels of their respective promoters. Consistently, the protein levels were decreased. In 3D acini IFN- $\gamma$  decreased the mRNA and protein levels of XBP-1s, IRE1 $\alpha$ , and GRP78 and increased the methylation of their promoters.

PERK pathway was activated and ATF4 protein was overexpressed in LSG from SS-patients. The ATF4 mRNA levels were decreased, which was correlated with increased promoter DNA methylation. SS-patients showed a significant increase of Xc-system (antioxidant response) expression. ATF4 protein levels correlated with scintigraphy data and serological markers while Xc-system protein levels correlated with ATF4 protein levels, sero-logical markers, and mouth and eye dryness. In 3D acini IFN- $\gamma$  decreased the ATF4 mRNA and increased its promoter's methylation.

A significant increase of ATF6 $\alpha$  mRNA (ATF6 $\alpha$  pathway) was observed in LSG of SS-patients, correlating with decreased DNA methylation of its promoter. The protein levels of ATF6f and ERAD (ER-associated degradation) machinery components were increased. A balanced expression of cIAP2 and cleaved-caspase-3 was observed in LSG of SS-patients. IFN- $\gamma$  decreased ATF6 DNA promoter methylation increased ATF6 $\alpha$  mRNA levels, ATF6 $\alpha$  protein levels, ATF6 $\alpha$  nuclear translocation and ERAD machinery components, without increasing apoptosis.

**Conclusions.** The attenuation of IRE1 $\alpha$ /XBP-1 signaling pathway could explain the glandular dysfunction in SS-patients, because this pathway is involved in biogenesis of the secretory machinery. Considering that ATF4 and ATF6 regulate the expression of genes involved in adaptive responses to cellular stress and ERAD, respectively, their increased expression and correlation with clinical data could be partial response aimed to relieve the stress arising in LSGs of SS-patients, thereby promoting cell survival. Glandular stress signals, including IFN- $\gamma$ , could modulate the expression of the UPR pathways, likely by promoter DNA methylation.

Funding: FONDECYT 1160015, Ph.D. CONICYT-fellowship, FONDECYT Postdoctoral Grant 3170023.

### P-137

#### Cytosolic accumulations of damaged DNA trigger inflammasome activation in the ductal salivary gland epithelia of Sjögren's syndrome patients

Aigli G. Vakrakou<sup>1,2</sup>, Evangelia Xingi<sup>3</sup> and Menelaos N. Manoussakis<sup>1,2</sup>. <sup>1</sup>Department of Pathophysiology, School of Medicine, National and Kapodistrian University of Athens, Athens, Greece. <sup>2</sup>Laboratory of Molecular Immunology, Hellenic Pasteur Institute, Athens, Greece. <sup>3</sup>Light Microscopy Unit, Hellenic Pasteur Institute, Athens, Greece.

**Background.** Sjögren's syndrome (SS) is characterized by chronic periductal inflammatory lesions in the salivary glands (SG) of patients. In addition, several previous studies from this laboratory had indicated that the ductal SG epithelial cell lines (SGEC) of SS patients manifest signs of intrinsic (cell-autonomous) activation, including the constitutive activation of inflammasome signaling pathways in transcriptome expression analyses. Herein, we have investigated the expression of various features related to the activation of inflammasomes in the SG epithelial cells of SS patients.

**Methods.** Confocal microscopy and several quantitative mRNA and protein analyses were used to evaluate the expression of various inflammasome activation-related molecules in SGEC lines and SG tissue specimens from 15 SS patients and 10 non-SS controls.

**Results.** The SGEC lines of SS patients (SS-SGEC), but not those of controls, were found to manifest significantly high constitutive activation of the AIM2 and NLRP3 inflammasomes, including the formation of characteristic intracellular ASC specks, the activation of caspace-1, as well as high intracellular and secreted IL-1 $\beta$  protein production. In addition, SS-SGEC lines and the ductal salivary epithelia of patients' tissues were found to display marked cytoplasmic accumulations of chromatin fragments and micronuclei (by DNA staining). These aberrant chromatin structures were found to contain significant amounts of damaged DNA, as illustrated by increased co-localization with phosphorylated histone  $\gamma$ -2AX staining (a DNA damage marker), as well as to be actively involved in the activation of inflammasome in SS-SGEC, as suggested by strong co-localization with the inflammasome-inducing DNA sensor AIM2. The cytoplasmic DNA de-

posits also co-localized with the lysosomal marker LAMP1, likely reflecting their insufficient clearance. Among various nucleases studied, the SS-SGEC and the ductal salivary tissues of SS patients manifested impaired DNase1 expression and activity, which may have a role in the aberrant cytoplasmic DNA build-up in these epithelia. In fact, the silencing of DNase1 in healthy SGEC (by DNaseI siRNA transfection) was found to lead to the induction of various inflammasome-related genes and to high IL-1 $\beta$  secretion in culture supernatants.

**Conclusion.** Our findings provide first evidence that the ductal salivary epithelia of SS patients manifest aberrant cytoplasmic accumulations of damaged DNA, which is associated with impaired cellular DNase1 activity and apparently drives the chronic constitutive activation of inflammasomes in these cells. These results appear to provide a mechanistic explanation for the intrinsic activation status that characterizes the ductal epithelia of SS patients and may indicate a key pathogenetic process in the disease.

### P-138

#### RANK-L is expressed by salivary gland stromal and epithelial cells in primary Sjögren's syndrome and could represent a key player in ectopic lymphoid structures neogenesis

Pierre-Marie Duret<sup>1,2</sup>, R. Veber<sup>2</sup>, R. Felten<sup>1</sup>, F. Monneaux<sup>2</sup>, J. Sibilia<sup>1</sup>, C. Mueller<sup>2</sup>, H. Dumortier<sup>2</sup> and Jacques-Eric Gottenberg<sup>1,2</sup>.

<sup>1</sup>Hôpitaux universitaires de Strasbourg; Service de rhumatologie; Centre national de référence des maladies auto-immunes systémiques rares. <sup>2</sup>Laboratoire «Immuno-pathologie et chimie thérapeutique», CNRS UPR3572.

Introduction. Tertiary Lymphoid Organs (TLOs) can be observed in target tissues of various auto- immune diseases such as salivary glands in primary Sjögren's syndrome. TLOs are mimicking secondary lymphoid organs (SLOs) structure and strikingly share common features with lymph nodes. SLOs organogenesis is coordonated by a complex stromal network and RANK-L (Receptor Activator of NF-kB Ligand) has been recently involved as a pivotal cytokine in precoce steps of their formation. Nethertheless, the contribution of stromal cells in TLOs neogenesis remains unclear. Thus, we hypothesized that RANK-L could be expressed by salivary gland stroma and could therefore play a critical role in TLOs establishment.

Materials and Methods. Stromal cells and RANK-L expression were analysed in salivary glands' TLOs by immuno-fluorescence on frozen sections in the NZB/NZW F1 mouse model and in minor salivary gland biopsies of patients fullfiling 2016 ACR-EULAR Sjögren's syndrome criterias and by flow- cytometry after enzymatic digestion of NZB/NZW F1 salivary glands. RANK-L expression has also been assessed by Real Time quantitative Polymerase Chain Reaction (RT-qPCR) and immunofluorescence on primary cultures of salivary gland epithelial cells (SGECs) with or without IL-1 $\beta$ or Interferon alpha (INF- $\alpha$  stimulation.

Results. Most of SLOs stromal cells populations: FRCs (Fibroblastic Reticular Cells), FDCs (Follicular Dendritic Cells), LECs (Lymphatic Endothelial Cells), BECs (Blood Endotholial Cells) and HEVs (High Endothelial Veinules) were identified in salivary TLOs of both NZB/NZW F1 mice and patients. FRCs were dominant in salivary TLOs and their number correlated with the degree of lymphocytic infiltration (r=0,7; p=0,007). The main difference between TLOs and SLOs was the lack, in salivary gland, of MRCs (Marginal Reticular Cells) which are a major source of RANK-L in lymph nodes. Despite the absence of MRCs in TLOs, we have observed an expression of RANK-L by a few T-cells within the infiltrates and strikingly by blood vessels and epithelial cells. Moreover, RANK-L expression by SGECs in primary cultures was increased after INF- $\alpha$  or IL-1 $\beta$  stimulation. Conclusion. To our knowledge, this is the first report of a RANK-L expression in Sjögren's syndrome. These results suggest that RANK-L could be an important actor of ectopic lymphoid structures neogenesis and its inhibition might represent, in the future, an alternative immuno-modulatory strategy in primary Sjögren's syndrome.

# 14th International Symposium on Sjögren's Syndrome

# P-139

Salivary gland damage in Sjogren's syndrome: are endothelial and vascular alterations associated with the secretion of IL-17?

Silvia V. Lourenço<sup>1</sup>; Sheyla B. Bologna<sup>1</sup>; Wanessa S. Cavalcante<sup>2</sup>, Sandra G. Pasoto<sup>3</sup>, Giovanna P. Florezi<sup>1</sup>.

<sup>1</sup>Department of Stomatology, Dental School, University of Sao Paulo, Sao Paulo-SP, Brazil. <sup>2</sup>Department of Dermatology, Medical School, University of Sao Paulo, Sao Paulo-SP, Brazil. <sup>3</sup>Department of Rheumatology, Medical School, University of Sao Paulo, Sao Paulo-SP, Brazil.

Disclosure. FAPESP 2014 / 11020-5; FAPESP 2017/11806-7

Background. Sjögren's syndrome (SS) is a systemic, inflammatory autoimmune disease that primarily affects the exocrine glands. The pathophysiology of the disease is mainly described as an autoimmune epithelitis, that, mediated by cytokines, growth factors and metalloproteinases leads to the destruction of the glandular parenchyma, causing a sclerotic process, which ends up impairing the salivary secretion. Recent studies have shown that systemic vascular changes are also part of the SS; these alterations involve endothelial dysfunction and vasculitis and are described in multiple organs, however the description of these changes in salivary glands are scarce. Saliva production is complex; it involves water and ionic changes throughout salivary gland ducts and the vascular flow is an important factor for saliva formation. Interleukin-17 (IL-17), a pro-inflammatory cytokine, which has been associated with a pro-thrombotic effect in multiple inflammatory and autoimmune diseases<sup>1</sup>, is also frequently associated with SS, for this reason, this study aimed to correlate the concentrations of IL-17 family in the saliva and the vascular alterations found in minor salivary gland specimens.

**Methods.** The morphological characteristics of biopsied specimens from 27 patients previously diagnosed with SS, were analyzed for the presence of vascular alterations such as vasculitis, thrombosis, congestion and hemorrhage. These patients also underwent a saliva collection for a multiplex analysis of the concentrations of IL-17A, IL-17E and IL-17F. Statistical analysis were performed to observe the concentration differences among the morphological alterations.

**Results.** Morphological analysis showed vessels congested in every specimen analyzed, and the presence of thrombi in 11 (40%) of the cases. Vasculitis was also found in 16 (59%) of the specimens. Binary logistic regression showed an increase of 1.012 in IL-17E when associated with vasculitis (p=0.034; 95%CI 1.001-1.022) with statistically significant higher concentration in patients with signs of vasculitis (p=0.0058). However, there was no statistically significant differences or associations between the IL-17A and IL-17F with the vascular changes. There was a higher concentration of IL-17E when compared to IL-17A (p<0.0001).

**Conclusions.** The IL-17A and its isoform IL-17F are often associated with large vessels inflammation such as takayasu artheritis (1), IL-17E was recently renamed to IL-25, for its antagonist function to IL-17. The functions of this cytokine are still being described, however, it is known that IL-17E is capable of mediating Th2 responses, and was found present in a higher concentration in patients with Churg-Strauss syndrome, associated with vascular changes (2), however, its role in mediating the autoimmune response in SS and the association with vascular alterations in the disease must be further investigated.

#### References

 VON VIETINGHOFF S LEY K: Interleukin 17 in vascular inflammation. Cytokine Growth Factor Rev 2010 Dec 14; 21(6): 463-9.

 MONTELEONE G et al.: Interleukin-25: A two-edged sword in the control of immune-inflammatory responses. Cytokine Growth Factor Rev 2010; 21(6): 471-5.

# P-140

# Functional analysis of saliva secretion in a mouse model of X-linked hypohidrotic ectodermal dysplasia

Takashi Munemasa<sup>1,2</sup>, Taro Mukaibo<sup>1,2</sup> and James E. Melvin<sup>1</sup>. <sup>1</sup>Secretory Mechanisms and Dysfunction Section, National Institute of Dental and Craniofacial Research, NIH, Bethesda, MD, U.S.A. <sup>2</sup>Division of Oral Reconstruction and Rehabilitation, Kyushu Dental University, Kitakyushu, Fukuoka, Japan.

**Background.** X-linked hypohidrotic ectodermal dysplasia (X-LHED) is the most common form of more than 150 types of ectodermal dysplasias. Over 200 mutations in *Ectodysplasin A (EDA)*, a gene inherited on the X-chromosome, have been associated with abnormal development of the hair, teeth, and

sweat glands. Like Sjögren's syndrome, X-LHED patients often report salivary gland dysfunction and nearly 30% of them perceive xerostomia. Thus, the purpose of this study is to further our understanding of the fluid secretion process and to investigate the mechanism of hyposalivation in X-LHED.

**Methods.** Tabby 6J (Ta/Y) male mice, an X-LHED disease model that has a single nucleotide deletion mutation in the *Eda* gene, were used. Their male unaffected littermates (X/Y) served as controls. The amount of secreted saliva from submandibular and sublingual glands (SMGs and SLGs) in response to  $0.3\mu$ M carbachol and  $1.0\mu$ M isoproterenol was determined using the *ex vivo* salivary gland perfusion technique, while *in vivo* experiments measured the amount of secreted saliva from parotid glands (PGs), SMGs and SLGs in response to 10 mg pilocarpine/kg body weight. The concentration of ions (Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup>) in the secreted saliva were determined. The cross-sectional areas of acini and ducts in the three major salivary glands were calculated from HE-stained images using ImageJ (NIH) software. Epithelial sodium channel (ENaC) subunits,  $\alpha$ -,  $\beta$ - and  $\delta$ -, mRNA expression was measured by qPCR.

**Results.** The body and gland weights of both SMG and SLG in Ta/Y mice were significantly less than in X/Y mice, whereas PG weight was comparable between X/Y and Ta/Y mice. The amount of secreted SMG saliva in both *ex vivo* and *in vivo* experiments was significantly reduced in Ta/Y mice, however, when normalized to gland weight, it was significantly increased in Ta/Y mice. In contrast, the amount of secreted saliva from SLGs and PGs was comparable between X/Y and Ta/Y mice, both for total saliva volume and when normalized to gland weight. The concentrations of Na<sup>+</sup> and Cl<sup>-</sup> in the saliva of Ta/Y mice were significantly higher than in X/Y mice, and this observation was most remarkable in SMGs. The cross-sectional area of SMG ducts in Ta/Y mice was less than in X/Y mice. mRNA expression of all ENaC subunits in Ta/Y mice was less than in X/Y mice.

**Conclusions.** In this study, we found that the amount of secreted saliva was diminished in Ta/Y mice. However, when saliva secretion was normalized to gland weight the total amount was not reduced compared to X/Y mice, suggesting that the fluid secretion pathways in acinar cells remain intact. In contrast, the Na<sup>+</sup> and Cl concentrations in the secreted saliva were increased, while both the cross-sectional area of ducts and transcript levels for ENAC subunits were reduced in Ta/Y mice. Together, these results are consistent with a compromised NaCl reabsorption mechanism in X-LHED Ta/Y mice.

# P-141

#### Increased expression of lysosomal associated membrane protein (LAMP) 3 in the salivary glands of Sjögren's syndrome patients can stimulate stalled autophagy and apoptosis

Tsutomu Tanaka<sup>1</sup>, Blake Warner<sup>1</sup>, Toshio Odani<sup>1</sup>, Hongen Yin<sup>1</sup>, Tatsuya Atsumi<sup>2</sup>, Masayuki Noguchi<sup>3</sup>, John Chiorini<sup>1</sup>.

<sup>1</sup>Molecular Physiology and Therapeutics Branch, National Institute of Dental and Craniofacial Research, National Institutes of Health, USA. <sup>2</sup>Department of Rheumatology, Endocrinology and Nephrology, Faculty of Medicine and Graduate School of Medicine, Hokkaido University, Japan. <sup>3</sup>Division of Cancer Biology, Institute for Genetic Medicine Hokkaido University, Japan.

**Background.** Environmental insults (*e.g.*, viral or bacterial infection) may induce the accumulation of misfolded proteins and activation of the unfolded protein response (UPR) pathway. Triggering of the UPR can result in endoplasmic reticulum (ER) stress, autophagy or cell death. Recent studies reported that salivary glands (SG) in primary SS (pSS) patients are under ER stress. To understand gland dysfunction in pSS we performed microarray analysis that revealed overexpressed UPR genes in minor salivary glands (MSGs) including lysosome associated membrane protein 3 (LAMP3). LAMP3, a UPR-responsive gene and inducible member the LAMP family (*e.g.*, LAMP1/2), is expressed in several cancers, neurodegenerative disorders, and inflammatory conditions; its role in pSS has not been defined. Given the link of the UPR in pSS, we assessed LAMP3 overexpression in cellular function.

**Methods.** Changes in epithelial gene expression were measured by microarray analysis of complimentary RNA (cRNA) isolated from MSGs of female patients with pSS. Results were compared with those obtained from the MSGs of sex-matched healthy volunteers or patients with an unrelated autoimmune disease (*e.g.*, IgG4 disease). Changes in autophagy, ER stress, and apoptosis were studied in the context of two SG-derived cell lines, HSG and A253 cells, by treatment with either chloroquine (CQ), cycloheximide (CHX), c-abl inhibitor (imatinib), Cathepsin D (CTSD) inhibitor (pepstatin A), or pan-caspase inhibitor (Z-VAD-FMK).

**Results.** Microarray analysis of differentially express genes indicated that LAMP3 expression was significantly increased in the MSGs of pSS patients when compared with healthy individuals or IgG4 disease patients. Expression of LAMP3 impeded cell growth relative to control cells and led to the accumulation of LC3-LAMP3 co-localized vesicles. When the effect of LAMP3 on autophagy was examined, LAMP3 decreased reporter protein degradation and increased the LC3II:LC3I ratio independent of treatment with CQ. Although no change in ER stress was measured via ATF expression, LAMP3 expression reduced p-crkl:crkl, increased degradation of LAMP1, and increased cytoplasmic CTSD. This data supports that LAMP3 expression alters autophagic flux leading to stalled autophagy. In addition, the LAMP3 induced an increase in cytoplasmic CTSD, resulting in activation of the BID-caspase 3 pathway and an increase in apoptosis.

**Conclusions.** Markers of autophagy and apoptosis have long been associated with pSS but a fundamental mechanistic understanding of the gene expression changes associated with this change in state has not been identified. Our study of LAMP3 expression in SG cells signifies a connection between these two observations and suggests increased LAMP3 expression can trigger stalled autophagy via degradation of c-abl and induced degradation of LAMP1. Furthermore, the change in lysosomal integrity results in the release of CTSD into the cytoplasm, initiating cellular apoptosis via the BID-Caspase 3 pathway independent of ER stress markers. Given significantly increased expression in pSS, inhibition of LAMP3 expression may represent a novel therapeutic strategy in the treatment of pSS.

# P-142

#### Do I sound dry?: voice analysis and affecting factors in patients with primary Sjögren's syndrome

Jennifer Lee<sup>1</sup>, Yong-sug Choi<sup>2</sup>, Sang-Yeon Kim<sup>2</sup>, Ji-Won Kim<sup>1</sup>, In-Chul Nam<sup>2</sup>, Young-Hoon Joo<sup>2</sup>, Young-Hak Park<sup>2</sup>, Dong-II Sun<sup>2</sup>, Sung-Hwan Park<sup>1</sup>. <sup>1</sup>Division of Rheumatology, Department of Internal Medicine, Seoul St Mary's Hospital, College of Medicine, The Catholic University of Korea, Seoul, South Korea. <sup>2</sup>Department of Otolaryngology-Head and Neck Surgery, College of Medicine, The Catholic University of Korea.

**Objective.** Saliva plays an important role in voice acuity, speech, and articulation. Therefore, we hypothesized that the reduced salivary rate might affect voice of patients with primary Sjögren's syndrome (pSS). The purpose of this study was to investigate whether patients with pSS have voice impairment compared to controls and to determine the affecting factors.

**Methods.** Subjects who complained of voice related symptoms underwent acoustic analysis, aerodynamic study, voice handicap index (VHI) questionnaire from September 2016 to January 2017. In cases of pSS patients, various disease-related parameters such as disease duration, EULAR Sjögren's syndrome disease activity index (ESSDAI), EULAR Sjögren's syndrome patients reported index (ESSPRI) were obtained by retrospective chart review.

**Results.** Fifty-four pSS patients and 52 controls were recruited. The subjects were all female, and mean age was  $53.9\pm9.73$ . VHI score was significantly higher in patients group ( $17.11\pm17.286$  vs  $9.35\pm10.518$ , p=0.006). However, the results of acoustic analysis and aerodynamic study were not different between the two groups, except that the proportion of subjects with abnormal mean flow rate (MFR) value in aerodynamic study was higher in pSS patients (68% vs. 50%, p=0.052), although it did not reach statistical significance. Disease-related parameters were available in 47 pSS patients. High VHI score was associated with low quality of life measured by EQ5D (spearman's rho=-0.39, p=0.007). Patients with abnormal MFR value showed higher physician global assessment ( $23.03\pm14.79$  vs.  $10.63\pm13.995$ , p=0.008), xerostomia inventory (XI) score ( $40.32\pm6.493$  p=0.006 vs.  $34.06\pm8.012$ , p=0.006), and higher ESR ( $30.06\pm20.797$  vs.  $17.75\pm12.835$ , p=0.016). However, ESSPRI, ESSDAI, salivary flow rate, or schirmer's test was not associated with voice related parameters.

**Conclusion.** Patients with pSS show higher VHI score, which was associated with low quality of life. MFR value tends to be abnormal in pSS patients and correlated with physician global assessment and XI score.

### P-143

### Orofacial myofunctional status and temporomandibular disorder symptoms in patients with Sjögren syndrome

Mariana Cristina Zanin<sup>1,2</sup>, Denny Marcos Garcia<sup>1,2</sup>, Eduardo Melani Rocha<sup>1,3</sup>, Ana Carolina Fragoso Motta, Cláudia Maria de Felício<sup>1,2,\*</sup>.

<sup>1</sup>Department of Ophthalmology, Otorhinolaryngology and Head and Neck Surgery School of Medicine of Ribeirão Preto, University of São Paulo – USP, Ribeirão Preto (SP), 14049-900, Brazil. <sup>2</sup>Craniofacial Research Support Center, University of São Paulo – USP – Ribeirão Preto (SP), Brazil. <sup>3</sup>Research Core for Ocular Physiopathology and Therapeutics.

**Purpose.** Sjögren's syndrome (SS) induces difficulty in chewing, swallowing, which is attributed to saliva deficit. However, the orofacial myofunctional and the frequency of temporomandibular disorder (TMD) and its relationship with the patients' perception of impairment is unknown in SS. In the present study, we investigated whether SS negatively affects the stomatognathic system and functions and whether the patients' perception of their functional limitations and TMD symptoms is related to orofacial myofunctional performance.

**Methods.** 19 women with SS based on the American-European Criteria and 20 healthy volunteers were compared by the orofacial myofunctional evaluation with scores protocol (OMES), Iowa Oral Performance Instrument (IOPI) model 2.2, electromyography (EMG) of masticatory muscles, and the Eating Assessment Tool (EAT-10). TMD were compared by pain to palpation of muscles and temporomandibular joint (TMJ), noise and range of jaw movements, using a digital caliper, and Jaw Functional Limitation Scale (JFLS-20).

**Results.** The orofacial myofunctional condition was worse in SS group. SS patients presented lower scores of all categories of OMES protocol (p<0.0001), tongue strength (p<0.001) and muscular activities of the temporalis and masseter measured by EMG (p<0.01 and 0.05, respectively). SS group manifested higher scores of muscles and TMJ pain, eating disorder (EAT-10) and JFLS-20 (p<0.0001).

**Conclusions.** The results showed that patients with Sjögren's Syndrome present impaired muscle and orofacial functions, TMJ signs/symptoms, eating and jaw limitations. Those disorders may prejudice to the disease control and must be addressed in the clinical evaluation to prevent nutritional and metabolic comorbidities in SS patients.

# **P-144**

# Relationship between caries and salivary flow rates in Sjögren's syndrome

Frederick B. Vivino<sup>1</sup>, Nicola Berman<sup>2</sup>, Joshua Baker<sup>1</sup>, Jonathan S. Dunham<sup>1</sup>, Andres Pinto<sup>3</sup>.

<sup>1</sup>University of Pennsylvania. <sup>2</sup>Lennox Hill Hospital. <sup>3</sup>Case Western Reserve University.

Background. Accelerated caries in Sjogren's (SS) subjects (Scts) is frequently attributed to reduced salivary flow (SF). Current guidelines for caries prophylaxis mention "stimulation of saliva flow" as a therapeutic option. Methods. Two hundred and twenty-five Scts with sicca symptoms underwent testing (excluding parotid sialography) between 2012-15 as per the 2002 AECG criteria and were classified as SS or non-SS (NSS). Evaluation included measurement of whole mouth unstimulated and parotid stimulated SF and dental examination or record review. Scts with incisal or cervical/ root caries were identified. For ordinal regression models, 3 categories of carious involvement were defined (0, 1-2 or >3 caries). Differences in the number of incisal, cervical/ root, & total caries in SS vs. NSS were compared. Statistical analysis was performed to evaluate differences in continuous & categorical variables and to identify clinical risk factors associated with caries. Because of a pre-hypothesized relationship between SF and total caries, measurements of unstimulated and stimulated salivary flow were included in multivariable models.

**Results.** Ninety-nine Scts were classified as SS and 126 as NSS. SS Scts had a significantly higher mean and median number of incisal, cervical/ root and total caries vs. NSS (Figure 1). Clinical evaluation is summarized in Table I. No significant differences in unstimulated whole or parotid stimulated SF (Table II), patient demographics, symptom prevalence, use of anticholinergic drugs, secretagogues, tooth loss, or dental visits in the last year were observed between the 2 groups.

A focus score >1/4 mm<sup>2</sup> was the only risk factor associated with total caries in SS. Within the SS cohort, there was no significant correlation between the total number of carious surfaces and low unstimulated whole or parotid stimulated SF.

**Conclusions.** Sjögren's Scts have a greater risk of caries compared to individuals with salivary hypofunction from other causes. While most SS Scts exhibit reduced SF, the rate itself may not be the sole determinant of caries risk. The overall risk may also be influenced by host factors and/or quantitative & qualitative differences in sialochemistry related to the underlying inflammatory process. Further research is needed to optimize strategies for caries prophylaxis in SS.



Fig. 1. Differences in the number of total, incisal, and cervical/root caries among subjects with Sjögren's syndrome and non-Sjögren's xerostomia.

	Sjögren's	Other sicca	p-value
Number of subjects	99	126	
Age	54.1 (13.8)	51.5 (13.1)	0.14
Female	87.8%	89.8%	0.64
Currently smoking	5.0%	5.4%	0.90
Duration dry mouth sx	40 (17,93)	36 (12,91)	0.55
Duration of dry eye sx	48 (22,115)	48 (14,120)	0.55
ANA or RF+	64%	23%	< 0.001
SSA+	63%	9.1%	< 0.001
Schirmer's <5mm/5 min	44%	37%	0.27
Focus score ≥1/4mm <sup>2</sup>	1 (1.1)	0 (0.0)	0=.001
Anticholinergic drugs	53%	56%	0.65
Cholinomimetic drugs	24%	17%	0.19
Antimalarial drugs	34%	18.6%	0.008
Number of incisal caries	0 (0,2)	0 (0,0)	0.004
Number of cervical/ root caries	0 (0,2)	0(0,1)	0.05
Total caries	1 (0,4)	0(0,2)	0.02
Any caries (%)	56%	44%	0.08
Dentures	11%	9.3%	0.67
Missing teeth	58%	50%	0.21
Oral Exam in last year	82%	85%	0.50
Dentist Visit in last year	80%	85%	0.29
Unstimulated whole mouth SFR*	0.62 (0.26, 1.16)	0.75 (0.28, 1.40)	0.33
% Abnormal USFR	12%	9%	0.51
Parotid stimulated SF*	0.001 (0.0.002)	0.001 (0.0.003)	0.96
% Abnormal stimulated SF	68%	75%	0.24

\*(ml/min).

Table II. Factors associated with greater total caries among all subjects and those diagnosed with Sjögren's Syndrome.

	All patients		Sjögren's patients	
	B (95% CI)	р	B (95%) CI	р
Age	1.02 (1.00, 1.04)	0.02	1.03 (1.00, 1.06)	0.07
Focus score >1/4 mm <sup>2</sup>	2.16 (1.28, 3.64)	0.004	2.88 (1.05, 7.93)	0.04
Anticholinergic drugs	1.62 (0.96, 2.73)	0.07	1.24 (0.56, 2.73)	0.59
Low parotid stimulated Saliyary Flow	0.99 (0.54, 1.75)	0.92	1.28 (0.54, 3.04)	0.57
Low whole mouth unstimulated Salivary flow	0.94 (0.55, 1.58)	0.80	1.03 (0.45, 2.33)	0.95

\*also tested and not-significant: sex, duration of xerostomia, duration of dry eyes, abnormal ocular staining, RF, ANA, SSA, abnormal Schirmer's test, and current smoking status.

# P-145

# Oral and dental symptoms and findings in patients with Sjögren's syndrome

Åslaug Haugsdal<sup>1,2</sup>, Haris Causevic<sup>3,4</sup>, Hannah Cecilie L. Bye<sup>2</sup>, Eli Nese Hopperstad<sup>2</sup>, Daniel Hammenfors<sup>3,4</sup>, Mihaela C. Marthinussen<sup>5</sup>, Malin V. Jonsson<sup>2,3</sup>.

<sup>1</sup>Department of Biological Sciences, University of Bergen, Norway. <sup>2</sup>Department of Clinical Dentistry – Section for Oral and Maxillofacial Radiology, University of Bergen, Norway. <sup>3</sup>Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, Norway. <sup>4</sup>Department of Rheumatology, Haukeland University Hospital, University of Bergen, Norway. <sup>5</sup>Oral Health Center of Expertise/Western Norway, Hordaland, Norway.

**Background.** A prominent feature of Sjögren's syndrome (SS) is chronic autoimmune inflammation of the salivary glands, manifested by oral dryness, major salivary gland swelling and a predisposition to dental caries and oral microbial infections. The aim of this study is to learn more about the dryness aspect of today's patients with SS.

**Methods.** Patients were recruited from the Department of Rheumatology, Haukeland University Hospital, University of Bergen, Norway (August-November 2017). As part of clinical follow-up encompassing major salivary gland ultrasonography (SGUS) and unstimulated (UWS) and stimulated sialometry (SWS) patients (n=15) were invited to a free dental check-up with radiographic examination (panoramic and two bite-wing images).

Results. By November 2017, thirteen patients had attended the dental ap pointment. Pathological SGUS (SGUS+) was determined in 7/13 patients; 5/7 and 3/6 had pathological levels of unstimulated (UWS+) and stimulated whole saliva (SWS+), respectively. UWS and SWS levels correlated (n=12, r=0.581, p=0.048). Xerostomia was reported by 12/13 patients; 9/12 and 5/11 with UWS+ and SWS+, respectively. In 4/9 and 3/5 patients with xerostomia, SGUS+ co-occurred with UWS+ and SWS+. In patients with xerostomia, hyposalivation was linked to problems eating dry food in 9/10 patients (p=0.007), two patients with altered taste and 4/5 patients having to get up to drink at night. No patients were current smokers, but previous smoking was reported by 2/8 patients with xerostomia. Patients with hyposalivation and xerostomia presented with dry lips in 7/9 cases, 5/7 manifested angular cheilitis, and 4/5 patients had increased facial mimics. All patients used fluoride toothpaste; 2/12 patients with xerostomia and hyposalivation used additional oral fluoride rinse, 5/6 used fluoride tablets, 7/9 used dental floss. Tooth picks were used by 8/9 patients, and one patient used interdental brushes. For relief of oral dryness, 3/5 patients with xerostomia and hyposalivation had found a product such as gel or spray that they were content using. Using Challacombe's scale for the clinical assessment of oral dryness, 6/13 had no dryness, 5/13 had mild dryness, and 2/13 had moderate dryness. Challacombe's score correlated with stimulated whole saliva levels (n=12, r=0.636, p=0.26) and with SGUS score (n=13, r=0.626, p=0.022) and (n=13, r=0.719, p=0.006), parotid and submandibular glands, respectively. Gingivitis was observed in 9/12 patients with xerostomia, and dental erosion in 10/12 patients. The number of decayed, missing, and filled teeth (DMFT) correlated with Challacombe's dryness score (n=13, r=0.615, p=0.25). Surprisingly, quality of life was somewhat higher in patients with hyposalivation compared to the patients with normal salivary secretion (p=0.050)

**Conclusions.** Oral dryness is common in patients with SS and linked to pathological SGUS changes, hyposalivation and caries experience. Patients with SS may benefit from additional information on caries prophylactic measures such as fluorides and regular oral health supervision.

#### **P-146**

# Is Sjögren's syndrome sialadenitis similar to salivary gland impairment in Lupus erythematosus?

Sheyla B. Bologna<sup>1</sup>; Giovanna Florezi<sup>1</sup>; Wanessa S. Cavalcante<sup>2</sup>, Sandra G. Pasoto<sup>3</sup>, Silvia V. Lourenço<sup>1</sup>.

<sup>1</sup> Department of Stomatology, Dental School, University of Sao Paulo, Sao Paulo-SP, Brazil. <sup>2</sup> Department of Dermatology, Medical School, University of Sao Paulo, Sao Paulo-SP, Brazil. <sup>3</sup> Department of Rheumatology, Medical School, University of Sao Paulo, Sao Paulo-SP, Brazil.

Disclosure: FAPESP 2014/15214-9; FAPESP 2017/11806-7

**Background.** Xerostomia is the subjective feeling of dry mouth mainly caused by salivary gland damage or dysfunction (1). Inflammatory diseases

such as Sjögren's syndrome (SS) are the main causes of this condition and the presence of this symptom in patients with lupus erythematosus (LE) is usually referred as secondary Sjögren's syndrome (2). Although these diseases share many clinical and laboratory aspects, some authors consider that both disorders as different conditions (3, 4). Based on this query, the aim of the present study was to analyze and compare morphological findings of minor salivary glands in patients with xerostomia diagnosed with primary SS or LE.

Methods. For this study 23 minor labial salivary gland (MLSG) from patients diagnosed with primary SS according to the American-European Consensus Criteria for Sjögren's syndrome and 23 MLSG from patients diagnosed with LE according to the criteria proposed by the American College of Rheumatology were submitted to histopathological analysis. The morphological aspects of all cases were examined using a conventional optical microscope equipped with a digital camera for photographic registration. The results were statistically analyzed using the two-tailed Fisher exact test. Results. There were 23 SS patients (21 were female and 2 male) and within LE group, 20 patients were female and 3 male. The most prevalent subset of LE was systemic LE (13/56.51%), followed consecutively by discoid LE (7/30.44%) and subacute cutaneous LE (3/13.05%). In pSS, the histopathological examination revealed a focal moderate/intense lymphocytic sialadenitis and excretory duct impairment with focus score ≥1 in all cases; vasculitis was detected in 19 cases. In LE, generalized mild/moderate sialadenitis was found in all cases. Moreover, in 22 LE salivary gland specimens hyalinization and thickening of ductal basement membrane were detected as well as perivascular inflammatory infiltrate, which was observed in 21cases: spongiosis with no ductal aggression was detected in 18 cases of LE. The morphological changes were distinct amongst the groups and this was statistically significant (p<0.0001). There was no difference within LE subsets. Conclusion. These results indicate that in each disorder MLSG has its specific morphological changes which leads to xerostomia and the impairment of MLSG in LE is probably due to a lupus sialadenitis whereas their MLSG present similar histopathological characteristics to others LE lesions.

#### References

- 1. VILLA A, CONNELL CL, ABATI S: Diagnosis and management of xerostomia and hyposalivation. *Ther Clin Risk Manag* 2015; 11: 45-51.
- BAER AN, MAYNARD JW, SHAIKH F, MAGDER LS, PETRI M: Secondary Sjögren's syndrome in systemic lupus erythematosus defines a distinct disease subset. J Rheumatol 2010; 37(6): 1143-9.
- SCHEINFELD N: Sjögren syndrome and systemic lupus erythematosus are distinct conditions. *Dermatol Online J* 2006; 12(1).
- FERNANDES JD, MENTA M, NICO S, AOKI V, BOLOGNA S, ROMITI R et al.: Xerostomia in Sjögren's syndrome and lupus erythematosus: a comparative histological and immunofluorescence study of minor salivary glands alterations. J Cutan Pathol 2010; 37(37): 432-8.

#### P-147

#### Development and validation of questionnaires to assess healthcare utilization and access in cohorts of patients with primary Sjögren's syndrome at the diagnosis and during the disease course

Chiara Seghieri<sup>1</sup>, Tommaso Grillo Ruggeri<sup>1</sup>, Chiara Baldini<sup>2</sup>, Luca Quartuccio<sup>3</sup>, Roberta Priori<sup>4</sup>, Elena Bartoloni<sup>5</sup>, Francesco Carubbi<sup>6</sup>, Saviana Gandolfo<sup>3</sup>, Salvatore De Vita<sup>3</sup>, Stefano Bombardieri<sup>2</sup>.

<sup>1</sup>Sant'Anna School of Advanced Studies, <sup>2</sup>Rheumatology Unit, University of Pisa, <sup>3</sup>Rheumatology Unit, University of Udine, <sup>4</sup>Rheumatology Unit, Sapienza University of Rome, <sup>5</sup>Rheumatology Unit, University of Perugia, <sup>6</sup>Rheumatology Unit, University of L'Aquila, Italy.

**Background/Purpose.** The geographic variation in healthcare service utilization and quality across and within countries is well documented and normally linked to differences in population health and needs. However, some of the variation may be unwarranted and driven by factors other than health needs, such as provider discretion, the availability and distribution of resources, financing and reimbursement models (Wennberg 2011). In this study, we identified the need for instruments to collect comparable data in Europe to establish practice profiles in the diagnosis, management and treatment of patients with Primary Sjögren's Syndrome (pSS).

**Methods.** We describe the development and preliminary validation of questionnaires to pSS to collect information on access to and intensity of treatments and services (*e.g.* diagnostic testing, hospitalizations, specialist visits), patients' satisfaction with the care received and socio-demographic data (*e.g.* age, sex, education level). A short questionnaire is also administered to specialists treating the selected pSS to collect data on the organization of their clinical centers.

Results. The pilot version of the two questionnaires were administered to 50 pSS in the clinical centre of Pisa (Italy) and counted 21 and 32 closedended questions, respectively. A narrative-based medicine section was also included in the questionnaires to collect stories about patients' care pathways and their health status before the diagnosis. Three questionnaires out of 47 were returned incomplete. Mean (SD) age was 60 (12.2) years and 96% of the sample was female. The majority of the respondents had a primary or secondary school (59%). Construct validity was supported by the questionnaire's ability to discriminate between groups with different levels of activity of the disease and socio-demographic characteristics. Disease activity was significantly associated with frequency of rheumatologic visits and diagnostic tests (p < 0.001). The total number of specialists involved in the care other than the rheumatologist varies significantly among patients. As expected, the most frequently involved were the ophthalmologist (90%) and the dentist (58%). Additionally, patients with lower education have attended on average less specialists than those with a high school or university degree (p < 0.001). Conclusions. preliminary results confirm that the questionnaire is a valid instrument to assess patterns of care for pSS in terms of access and utilization and in relation to clinical and socio-demographic characterictics of patients. Further analysis are ongoing in other clinical centers to verify the generalizability and additional psychometric properties of the instrument.

#### P-148

# Real world data using EULAR outcome assessment tools for primary Sjögren's syndrome (pSS) patients

Hargreaves B<sup>1</sup>, Howard Tripp N<sup>1,2</sup>, Kidd E<sup>2</sup>, Ng WF<sup>1,2</sup> <sup>1</sup>Institute of Cellular Medicine, Newcastle University, UK. <sup>2</sup>Rheumatology department, Newcastle upon Tyne Hospitals NHS Foundation Trust, UK.

**Background.** The EULAR Sjögren's Syndrome Study Group has developed standardised outcome assessment tools for pSS for clinical trials and for routine clinical assessment. While there has been a growing body of data on the use of these outcome assessment tools, data on its application in routine clinical practice remains sparse.

**Methods.** EULAR Sjögren's Syndrome Disease Activity Index (ESSDAI) and EULAR Sjögren's Syndrome Patient Reported Index (ESSPRI) were collected from patients attending a pSS clinic from a single UK centre. Anonymised data from 2009 to 2017 were analysed for patterns of change over time.

Results. 506 patients used the service during the study period with 369 having more than 1 clinic visit. For the ESSPRI, data was available for 346 patients from more than 2 visits; 113 patients had an ESSPRI at least 1 point higher at the last compared to the first visit (ESSPRI-worse group). 56 patients had an ESSPRI at least 1 point lower (ESSPRI-improved group) and 176 patients had a change of less than 1 in ESSPRI between the first and last visits (ESSPRI-unchanged group). The ESSPRI-worse group had a lower mean initial ESSPRI (4.59), compared to the ESSPRI-unchanged and ESSPRI-Improved group (6.27 and 6.29 respectively). There were no significant differences in age, sex and anti-Ro positivity between the three groups; age, percentage female and percentage anti-Ro+ were 60 years, 89%, 66% (ESSPRI-worse); 60 years; 86%, 60% (ESSPRI-unchanged); 62 years, 91%, 73% (ESSPRI-improved). For the ESSDAI, data was available for 356 patients from more than 2 visits; 57 had an ESSDAI at least 3 points higher at the last visit (ESSDAI-worse group); 68 patients had an ESSDAI at least 3 points lower (ESSDAI-improved); and 230 patients had a difference of less than 3 in ESSDAI between the first and last visits (ESSDAI-unchanged). The mean ESSDAI at first visit was significantly higher in the ESSDAI-improved group (9.2) compared to the ESSDAI-unchanged and ESSDAI-worse groups (3.1 and 2.4 respectively). There was no difference in age between the 3 ESSDAI groups. The ESSDAI-worse group had more female subjects and less anti-Ro positivity (93% & 54% respectively) compared to the ESSDAIimproved (84% & 59%) and ESSDAI-unchanged 86% & 65%) groups

**Conclusions.** ESSPRI and ESSDAI changed over time. Worsening ESSPRI was associated with lower baseline score whereas improvement in ESS-DAI was associated with higher baseline score. Although the relationships between treatment and change in outcome scores have not been analysed, given the lack of proven treatment available for pSS, our data may reflect natural history of the disease course rather than treatment effects.

# P-149

#### Development of high systemic activity in primary Sjögren syndrome: analysis of 1487 Spanish patients (GEAS-SS Registry)

A. Flores-Chávez<sup>1</sup>, S. Retamozo<sup>1</sup>, R. Solans<sup>2</sup>, G. Fraile<sup>3</sup>, B. Maure<sup>4</sup>, C. Feijoo<sup>5</sup>, R. Pérez-Alvarez<sup>6</sup>, FJ. Rascón<sup>7</sup>, M. Zamora<sup>8</sup>, M. López-Dupla<sup>9</sup>, MA Duarte-Millán<sup>10</sup>, M. Ripoll<sup>11</sup>, P. Guisado-Vasco<sup>12</sup>, E. Fonseca<sup>13</sup>, G. de-la-Red<sup>14</sup>, A. Chamorro<sup>15</sup>, E. Sanchez Vizcaíno<sup>16</sup>, B. Pinilla<sup>17</sup>, M.J. Soto-Cárdenas<sup>18</sup>, M. Akasbi<sup>19</sup>, P. Fanlo<sup>20</sup>, A. Gato<sup>21</sup>, I. Jiménez-Heredia<sup>22</sup>, B. De-Miguel<sup>23</sup>, S. Arteaga <sup>1.24</sup>, B. Kostov<sup>25</sup>, P. Brito-Zerón<sup>1.16</sup>, M. Ramos-Casals<sup>1</sup>, on behalf of the SS Study Group GEAS-SEMI\*

<sup>1</sup>Laboratory of Autoimmune Diseases Josep Font, IDIBAPS, Department of Autoim-mune Diseases, ICMiD, Hospital Clínic, Barcelona. <sup>2</sup>Autoimmune Diseases Unit, Department of Internal Medicine, Hospital Vall d'Hebron, Barcelona. 3Department of Internal Medicine, Hospital Ramón y Cajal, Madrid. 4Department of Internal Medicine, Complejo Hospitalario Universitario, Vigo. 5Department of Internal Medicine, Hospital Parc Taulí, Sabadell. 6Department of Internal Medicine, Hospital Alvaro Cunqueiro, Vigo, <sup>7</sup>Department of Internal Medicine, Hospital Son Espases, Palma de Mallorca <sup>8</sup>Department of Internal Medicine, Hospital Virgen de las Nieves, Granada. <sup>9</sup>Department of Internal Medicine, Hospital Joan XXIII, Tarragona. <sup>10</sup>Department of Internal Medicine, Hospital de Fuenlabrada, Fuenlabrada. <sup>11</sup>Department of Internal Medicine, Hospital Infanta Sofía, Madrid. <sup>12</sup>Department of Internal Medicine, Complejo Hos-pitalario Ruber Juan Bravo, Madrid. <sup>13</sup>Department of Internal Medicine, Hospital de Cabueñes, Gijón. 14Department of Internal Medicine, Hospital Esperit Sant, Santa Coloma de Gramenet.<sup>15</sup>Department of Internal Medicine, Hospital de Salamanca, Salamanca. <sup>16</sup>Unit of Clinical Investigation, Hospital CIMA-Sanitas, Barcelona. <sup>17</sup>Department of Internal Medicine, Hospital Gregorio Marañón, Madrid. <sup>18</sup>Department of Medicine, University of Cádiz, Cádiz. <sup>19</sup>Department of Internal Medicine, Hospital Infanta Leonor, Madrid. 20 Department of Internal Medicine, Hospital Virgen del Camino, Pamplona.<sup>21</sup>Department of Internal Medicina, Hospital de Albacete, Albacete.<sup>22</sup>Department of Internal Medicine, Hospital de Sagunto, Valencia. <sup>23</sup>Department of Inter-nal Medicine, Hospital 12 de Octubre, Madrid. <sup>24</sup>Universidad de Antioquía, Medellín, Colombia. <sup>25</sup>Transverse group for research in primary care, IDIBAPS, Barcelona.

**Background.** To characterize high systemic involvement in primary Sjögren syndrome (SS) in a large cohort of Spanish patients using the EULAR-SS disease activity index (ESSDAI).

Methods. The GEAS-SS Study Group was formed in 2005 with the aim of collecting a large series of Spanish patients with primary SjS. Systemic involvement was characterized using ESSDAI definitions for the 12 domains, and patients scoring as high in the domains that specifically contain high activity (lymphadenopathy, articular, cutaneous, pulmonary, renal, peripheral nervous system, central nervous system and muscular) were identified and analysed.

Results. Of the 1487 patients included in the Registry, 186 (12.5%) presented with 197 systemic features classified as high according to the corresponding organ-by-organ ESSDAI domains. There were 159 women and 27 men, with a mean age of 59,06 yrs (range 15-89 yrs) at the time of the diagnosis of high activity. High systemic activity was scored in the lymphadenopathy (n=53), peripheral nervous system (n=31), central nervous system (n=25), pulmonary (n=24), renal (n=21), articular (n=16), skin (n=16), hematological (n=7) and muscular (n=4) domains. Men were overrepresented in the muscular (33%), lymphadenopathy (19%) and cutaneous (19%) domains. The domains diagnosed at younger ages included muscular (mean age of 45 yrs), CNS (49,7 yrs), articular (54,3 yrs) and hematological (55,4 yrs), while cutaneous (63,2 yrs) and pulmonary (67,1 yrs) were diagnosed at older ages. The main clinical syndromes responsible of high systemic activity were lymphoma (n=53), ataxic neuronopathy (n=14), pulmonary fibrosis (n=12), diffuse leukocytoclastic vasculitis (n=10), severe polyneuropathy (n=10), myelitis (n=8), meningitis (n=7), multiple mononeuritis (n=7), membranous/membranoproliferative glomerulonephritis (n=6) and severe thrombocytopenia (n=5). Notably, 13 (81%) out of the 16 patients presenting with high activity in the articular domain (>6 joints involved) were finally diagnosed with rheumatoid arthritis during the follow-up

**Conclusions.** Primary Sjögren syndrome at diagnosis is presenting with high systemic activity in 12.5% of cases; half the cases were related to either the development of lymphoma or severe neurological features. Nearly all the patients presenting with a severe polyarthritis were finally diagnosed with associated rheumatoid arthritis.

#### How the different systemic organ involvements are overlapped in patients with primary Sjögren syndrome: analysis using a mathematical model

P-150

Soledad Retamozo<sup>1,2,3</sup>, Belchin Kostov<sup>4</sup>, Margit Zeher<sup>5</sup>, Kathy Sivils<sup>6</sup>, Thomas Mandl<sup>7</sup>, Raphaele Seror<sup>8</sup>, Xiaomei Li<sup>9</sup>, Chiara Baldini<sup>10</sup>, Xavier Mariette<sup>8</sup>, Jacques-Eric Gottenberg<sup>11</sup>, Debashish Danda<sup>12</sup>, Roberta Prioril<sup>3</sup>, Luca Quartuccio<sup>14</sup>, Gabriela Hernandez-Molina<sup>15</sup>, Berkan Armagan<sup>16</sup>, Aike A. Kruize<sup>17</sup>, Seung-Ki Kwok<sup>18</sup>, Marie Wahren-Herlenius<sup>19</sup>, Sonja Praprotnik<sup>20</sup>, Damien Sene<sup>21</sup>, Elena Bartoloni<sup>22</sup>, Maureen Rischmueller<sup>23</sup>, Roser Solans<sup>24</sup>, Yasunori Suzuki<sup>25</sup>, David Isenberg<sup>26</sup>, Valeria Valim<sup>27</sup>, Piotr Wiland<sup>28</sup>, Gunnel Nordmark<sup>29</sup>, Guadalupe Fraile<sup>30</sup>, Hendrika Bootsma<sup>31</sup>, Takashi Nakamura<sup>32</sup>, Roberto Giacomelli<sup>33</sup>, Valerie Devauchelle-Pensec<sup>34</sup>, Benedikt Hofauer<sup>35</sup>, Michele Bombardieri<sup>36</sup>, Virginia Fernandes Moça Trevisani<sup>37</sup>, Daniel Hammenfors<sup>38</sup>, Sandra G. Pasoto<sup>39</sup>, Steven E. Carsons<sup>40</sup>, Tamer A Gheita<sup>41</sup>, Fabiola Atzeni<sup>42</sup>, Jacques Morel<sup>43</sup>, Cristina Vollenveider<sup>44</sup>, Pilar Brito-Zerón<sup>1,45</sup>, Manuel Ramos-Casals<sup>1</sup>, on behalf of the Sjogren Big Data Consortium.

Sjögren Syndrome Research Group (AGAUR), Laboratory of Autoimmune Diseases Josep Font, IDIBAPS-CELLEX, Department of Autoimmune Diseases, ICMiD, University of Barcelona, Hospital Clínic, Barcelona, Spain. <sup>2</sup>Hospital Privado Universitario de Córdoba. Instituto Universitario de Ciencias Biomédicas de Córdoba (IUCBC) Córdoba- Argentina. 3Instituto De Investigaciones En Ciencias De La Salud (INIC SA), Universidad Nacional de Córdoba (UNC), Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) - Córdoba - Argentina. <sup>4</sup>Primary Care Research Group, IDIBAPS, Centre d'Assistència Primària ABS Les Corts, GESCLINIC, Barcelona, Spain. <sup>5</sup>Division of Clinical Immunology, Faculty of Medicine, University of Debrecen, Debrecen, Hungary. 6Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA. 7Department of Rheumatology, Malmö University Hospital, Lund University, Lund, Sweden. 8Center for Immunology of Viral Infections and Autoimmune Diseases, Assistance Publique -Hôpitaux de Paris, Hôpitaux Universitaires Paris-Sud, Le Kremlin-Bicêtre, Université Paris Sud, INSERM, Paris, France. 9Department of Rheumatology and Immunology, Anhui Provincial Hospital, Hefei, China. 10Rheumatology Unit, University of Pisa, Pisa, Italy. <sup>11</sup>Department of Rheumatology, Strasbourg University Hospital, Univer-sité de Strasbourg, CNRS, Strasbourg, France. <sup>12</sup>Department of Clinical Immunology & Rheumatology, Christian Medical College & Hospital, Vellore, India. <sup>13</sup>Department of Internal Medicine and Medical Specialties, Rheumatology Clinic, Sapienza University of Rome, Rome, Italy. 14Clinic of Rheumatology, Department of Medical and Biological Sciences, University Hospital "Santa Maria della Misericordia", Udine, Italy. <sup>13</sup>Immunology and Rheumatology Department, Instituto Nacional de Ciencias Médicas y Nutrición Salvador Zubirán. México City, Mexico. <sup>16</sup>Department of Internal Medicine, Division of Rheumatology, Hacettepe University Faculty of Medicine, Ankara, Turkey. 17Department of Rheumatology and Clinical Immunology, University Medical Center Utrecht, Utrecht, The Netherlands. <sup>18</sup>Seoul St. Mary's Hospital, The Catholic University of Korea, Seoul, South Korea. <sup>19</sup>Department of Medicine, Solna, Unit of Experimental Rheumatology, Karolinska Institutet, and Karolinska University Hospital, Stockholm, Sweden. 20 Department of Rheumatology, University Medical Centre, Ljubljana, Slovenia.<sup>21</sup> Service de Médecine Interne 2, Hôpital Lariboisière, Université Paris VII, Assistance Publique-Hôpitaux de Paris, 2, Paris, France. <sup>22</sup>Rheumatology Unit, Department of Medicine, University of Perugia, Perugia, Italy, <sup>23</sup>Department of Rheumatology, School of Medicine, The University of Western Australia, Crawley, Australia.24Department of Internal Medicine, Hospital Vall d'Hebron, Barcelona, Spain. <sup>25</sup>Division of Rheumatology, Kanazawa University Hospital, Kanazawa, Ishikawa, Japan. 26Centre for Rheumatology, Division of Medicine, University College London, London, UK. 27Department of Medicine, Federal University of Espírito Santo, Vitória, Brazil. 28 Department of Rheumatology and Internal Medicine, Wroclaw Medical Hospital, Wroclaw, Poland. 29Rheumatology, Department of Medical Sciences, Uppsala University, Uppsala, Sweden. <sup>30</sup>Department of Internal Medicine, Hospital Ramón y Cajal, Madrid, Spain. <sup>31</sup>Department of Rheumatology & Clinical Immunology, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands. <sup>32</sup>Department of Radiology and Cancer Biology, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan. <sup>33</sup>Clinical Unit of Rheumatology, University of l'Aquila, School of Medicine, L'Aquila, Italy. <sup>34</sup>Rheumatology Department, Brest University Hospital, Brest, France. <sup>35</sup>Hals-Nasen-Ohrenklinik und Poliklinik, Technische Universität München, München, Gernany. <sup>36</sup>Centre for Experimental Medicine and Rheumatology, Queen Mary University of London, London, UK. <sup>37</sup>Federal University of São Paulo, Sao Paulo, Brazil. <sup>38</sup>Department of Clinical Science, University of Bergen; and Department of Rheumatology, Haukeland University Hospital, Bergen, Norway. <sup>39</sup>Rheumatology Division, Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo (USP), São Paulo, Brazil. <sup>40</sup>Division of Rheumatology, Allergy and Immunology Winthrop-University Hospital, Stony Brook University School of Medicine, Mineola, NY, USA. <sup>41</sup>Rheumatology Department, Kasr Al Ainy School of Medicine, Cairo University, Cairo, Egypt. <sup>42</sup>IRCCS Galeazzi Orthopedic Institute, Milan, Italy. 43Department of Rheumatology, Teaching hospital and University of Montpellier, Montpellier, France. <sup>44</sup>German Hospital, Buenos Aires, Argentina. <sup>45</sup>Autoimmune Diseases Unit, Department of Medicine, Hospital CIMA- Sanitas, Barcelona, Spain.

**Background.** To analyse the degree of overlap of the different systemic organ involvements in patients with primary Sjögren syndrome (SjS) in a large international cohort using a mathematical model.

**Methods.** We have quantified at diagnosis systemic involvement (defined according to the 10 domains of the ESSDAI score) in patients included in the Big Data Sjögren Registry. We quantified how many patients had activity in more than one domain, how many patients had involvement of an isolated domain and we measured the degree of overlap between the main pairs and triplets of associated groups of active domains. Using a mathematical model, we represented the overlap between groups by means of Venn diagrams.

Results. 9118 patients were included in the Registry with ESSDAI values available at the time at diagnosis: 5758 (63%) had activity in at least one ES-SDAI domain. Among them, 3167 (55%) had only one active domain, 1532 (27%) two, 628 (11%) three and 431 (7%) four or more active domains. Three domains showed activity at diagnosis in at least 10% of patients: articular (37.9%), glandular (21.4%) and pulmonary (10.4%). The frequency of patients having concomitant activity in other organs varied widely in each domain: among patients with articular involvement, 54% of cases had another domain active, a figure that was 68% for patients with glandular involvement; 88% of patients with muscular involvement had concomitant activity in other organs, 78% of those with peripheral neuropathy and 74% of those with renal involvement. Numerically, the most frequent associations among two different domains were between articular and glandular (n=944), pulmonary (n=487), constitutional (n=464) or cutaneous (n=410) domains, while the most frequent associations among 3 organs were between articular/glandular plus pulmonary (n=224), constitutional (n=216) or lymphadenopathy (n=173) domains. Mathematical model identified the highest degree of overlap among the different domains for the following pairs: % of patients with muscular activity who had concomitant glandular activity ( $\Delta 30.2\%$  with respect to the expected frequency of glandular activity), those with lymphadenopathy who had concomitant glandular activity ( $\Delta 22.9\%$ ), and those with constitutional who had concomitant articular  $(\Delta 21.6\%)$  or glandular  $(\Delta 21.1\%)$  activities.

**Conclusions.** Primary Sjögren syndrome is presenting at diagnosis as a systemic disease in two thirds of cases; in more than half of cases, there was only one active domain, although 7% of patients presented with a multisystemic disease affecting 4 or more different organs. An enhanced degree of overlap in comparison with the expected values found for the total cohort was identified in the concomitant activity for the clusters glandular-lymphadenopathy-muscular and constitutional-articular-glandular (Fig. 1 a-b).



# P-151

#### Predictors of fatigue in 608 patients from the United Kingdom Primary Sjögren's Syndrome Registry

Katie L Hackett<sup>1,2,3</sup>, Kristen Davies<sup>1</sup>, Rebecca Bragg<sup>1,4</sup>, Sheryl Mitchell<sup>2</sup>, Samira T Miyamoto<sup>5</sup>, Dennis Lendrem<sup>1</sup>, Wan-Fai Ng<sup>1,2</sup> on behalf of the UK primary Sjögren's Syndrome Registry\*.

<sup>1</sup>Musculoskeletal Research Group, Institute of Cellular Medicine & NIHR Biomedical Research Centre for Ageing and Chronic Diseases, Newcastle upon Tyne, UK. <sup>2</sup>Newcastle upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. <sup>3</sup>Department of Social Work, Education & Community Wellbeing, Northumbria University, Newcastle upon Tyne, UK. <sup>4</sup>Queen Elizabeth Hospital, Gateshead, UK. <sup>3</sup>Universidade Federal do Espirito Santo, Vitoria and Universidade Federal de São Paulo, São Paulo, Brazil.

**Background.** Fatigue is a dominant symptom for 70% of primary Sjögren's syndrome (PSS) patients. Many PSS patients are likely to have comorbidities associated with fatigue and/or take drowsiness inducing medication.

This study reports on fatigue symptoms of a large cohort of 608 patients diagnosed with PSS using the American European Consensus Group criteria. We identify predictors of fatigue in patients with assignable causes (AC) and in those with no assignable cause for their fatigue.

**Methods.** We calculated physical and mental fatigue scores from the Profile of Fatigue and Discomfort tool. We collected measures of mood (Hospital Anxiety and Depression Scale (HADs)), daytime sleepiness, dryness (1-10 scale and Schirmer), pain (1-10) symptoms (ESSPRI), disease activity (ESSDAI), BMI and comorbidity and polypharmacy scores (CPS).

We allocated patients with AC of fatigue (comorbidities associated with fatigue, those taking drowsiness inducing medications and/or a HADs anxiety or depression score  $\geq 8$ ) to the AC group. We named the remaining cohort the CORE group. We made comparisons between the two groups and conducted a multiple regression analysis to identify potential predictors of physical and mental fatigue. We constructed a dummy variable permitting us to test directly for differences between the AC and CORE groups.

**Results.** 21% of patients were taking medication associated with drowsiness and 33% of patients had comorbidities associated with fatigue. 465 patients had AC for their fatigue. We saw no differences between groups for age, ESSDAI or BMI. The AC group had significantly greater symptom burden (physical fatigue, mental fatigue, pain, depression, and daytime sleepiness all p<0.0001 and dryness (p=0.0012)).

The main predictors of physical fatigue were pain, depression and dryness (all p=<0.0001). Whilst the relationships between pain and depression were slightly different (p=0.0023 and p=0.0269 respectively), the relationships with the main predictors were broadly similar in the two groups.

Depression, daytime sleepiness and pain were the main predictors of mental fatigue (all p<0.0001). There was a significant difference in the relationship with depression between the groups (p<0.031) but again, the relationships with the main predictors of mental fatigue were similar for both groups.

**Conclusion.** Large numbers of PSS patients have AC for their fatigue, including drowsiness inducing medication and comorbidities. While pain and depression are both predictors of physical and mental fatigue, dryness is an important predictor of physical but not mental fatigue. Reviewing medications, treating comorbidities (including depression) and addressing pain may all have a positive impact on PSS fatigue.

# P-152

# Depressive symptoms, fatigue, and dry eye in patients with Sjögren's syndrome

Sezen Karakus, MD<sup>1</sup>, Alan N. Baer, MD<sup>2</sup>, Esen K. Akpek, MD<sup>1,2</sup>. <sup>1</sup>Ocular Surface Diseases and Dry Eye Clinic, The Wilmer Eye Institute, Johns Hopkins University, Baltimore, Maryland, USA. <sup>2</sup>The Johns Hopkins Jerome L. Greene Sjögren's Syndrome Center, Baltimore, MD, USA.

**Background.** The well-known association of depression and dry eye is poorly understood particularly in patients with underlying Sjögren's syndrome (SS). In this study, we aimed to investigate the associations between depressive symptoms, physical and visual fatigue, and the severity of dry eye, in patients with SS related dry eye versus non-SS dry eye and compared to normal individuals.

**Methods.** Thirty-six dry eye patients with SS (SS related dry eye), 34 dry eye patients without SS (non-SS dry eye), and 34 controls aged 50 years and older were recruited from the multidisciplinary Jerome L. Greene SS

Center at Johns Hopkins University, Baltimore, Maryland. The 2012 American College of Rheumatology criteria were used for SS classification. All participants completed a mood questionnaire (Part D of the General Health Questionnaire) evaluating their depressive symptoms followed by assessments of visual fatigue, physical fatigue, and cognition (Mini-Mental Status Exam). After completing an Ocular Surface Disease Index (OSDI) questionnaire to evaluate dry eye symptoms, a battery of objective dry eye measures including tear film break-up time, tear osmolarity, Schirmer's test, and ocular surface staining were performed to quantify dry eye.

Results. Demographic characteristics including age and female proportion and severity of objective dry eye measures were comparable between SS related and non-SS dry eye patients. Patients with SS related dry eye had significantly higher depression scores compared to patients with non-SS dry eye and controls (3.7 vs. 1.4 vs. 1.0, p=0.03). Prevalence of physical fatigue was similar between SS related and non-SS dry eye patients (75% vs. 56%, p=0.09) and higher in both groups compared to controls (12%, p<0.001). Visual fatigue scores were higher in both SS- related and non-SS dry eye groups compared to controls, although the difference was not statistically different (2.8 vs. 2.5 vs. 1.5, p=0.09). OSDI scores were also similar between SS and non-SS related dry eye patients and greater in both groups compared to controls (39.8 vs. 40.1 vs. 4.8, p<0.001). After adjusting for age, sex, and cognitive status, there was a significant association between having SS-related dry eye and greater depression scores (3.5, 95%CI=1.1-6.0, p=0.005) and visual fatigue scores (1.6, 95%CI=0.5-2.7, p=0.005) compared to controls. However, no significant association was found between non-SS dry eye and depression and visual fatigue scores compared to controls (p>0.05). Physical fatigue was associated with both SS-related and non-SS dry eye, although the Odds Ratio (OR) was greater for SS related dry eye (OR=37.1, 95%CI=7.6 to 182.0, p<0.001 and OR=9.9, 95%CI=2.7 to 36.1, p=0.001). Greater OSDI scores correlated with presence of physical fatigue (rho=0.48, p>0.001) and greater visual fatigue scores (r=0.44, p < 0.001) but not with greater depression scores (p < 0.05).

**Conclusions.** Patients with SS related dry eye experience greater depressive symptoms and physical fatigue compared to normal individuals. Both physical and visual fatigue correlate with more severe dry eye symptoms. However, depressive symptoms seem to be more related to having SS rather than severe dry eye.

## P-153

#### Correlation between sleeping quality with fatigue, quality of life and disease activity (ESSDAI) in patients with primary Sjögren's syndrome

Luciana Paula Dardin<sup>1</sup>, Ana Beatriz Andreo Garcia<sup>1</sup>, Consuelo Bueno Diniz Adán<sup>2</sup>, Fania Cristina dos Santos<sup>3</sup>, Marco Tulio de Mello<sup>4</sup>, Virginia Fernandes Moça Trevisani<sup>1,5</sup>.

<sup>12</sup>Emergency Medicine and Evidence-Based Medicine Department, Sao Paulo Federal University (UNIFESP) – Paulista School of Medicine (EPM), Sao Paulo, Brazil.
<sup>2</sup>Ophthalmology Department of São Paulo Federal University (UNIFESP) Paulista School of Medicine (EPM), Sao Paulo, Brazil. <sup>3</sup>Geriatrics and Gerontology Discipline, Sao Paulo Federal University (UNIFESP) – Paulista School of Medicine (EPM), Sao Paulo, Brazil. <sup>4</sup>Ceriatrics and Gerontology Discipline, Sao Paulo, Brazil. <sup>4</sup>Physical Education School, Minas Gerais Federal University, Minas Gerais, Brazil. <sup>5</sup>Rheumatology Discipline, Santo Amaro University (UNISA) – School of Medicine, Sao Paulo, Brazil.

**Introduction.** Fatigue is a prominent and disabling feature in the majority of primary Sjögren's Syndrome (pSS) patients. Although the presence of sleep disorders in pSS patients has been previously confirmed in other studies, its relationship with fatigue and disease activity needs to be further assessed.

**Objectives.** To assess the sleep quality in pSS patients using the Pittsburg sleep quality index (PSQI) and actigraphy (PSQI), and its relationship with fatigue, quality of life, and disease activity (ESSDAI).

**Methods.** 50 pSS patients and 50 controls with sleep disorders according to the PSQI were included. The pSS patients used an actigraph for 15 days and filled in the following questionnaires and tools: PSQI, Profile of fatigue and discomfort (PROFAD), Visual Analog Scale (VAS pain and VAS fatigue), EULAR Sjögren's Syndrome Patients Reported Index (ESSPRI), Restless Legs Syndrome International Rating Scale (EIGSPI), Mini-Mental State Examination (MMSE), and EULAR Sjögren's Syndrome disease activity index (ESSDAI).

**Results.** The pSS patients were on average aged 56.4 years, had suffered from the disease for 12 years, and 100% presented fatigue. The ESSDAI showed that 40 patients (80%) demonstrated low disease activity and 10 (20%) moderate activity. Controls were on average aged 56.5 years, and all

presented sleep disorders, (PSQI average total score 10.5). The actigraphy in pSS patients demonstrated means of 26.2 minutes for sleep latency, 48.2 minutes for nightly awakenings, 89.7% for sleep efficiency, and 398.5 minutes (approximately 6.5 hours) of sleep. There was a positive and significant correlation between PSQI and the following variables: VAS pain (p=0,020), VAS fatigue (p=0,006), PROFAD total score (p=0.005), and fatigue and mental ESSPRI (p=0,012, p=0,009). There was a positive correlation between the number of nightly awakenings assessed using actigraphy and disease activity by ESSDAI (p=0.012). A comparison between groups demonstrated a significant difference regarding the VAS fatigue outcome, (p=0.011), where the syndrome group presented higher scores than the control group.

**Conclusion.** This study showed a positive and significant correlation between sleep disorders with pain, fatigue, and disease activity. It may also be concluded that other factors independent of sleep have influence on fatigue in pSS patients, since when compared with the controls with sleep disorders, the pSS patients maintained higher scores for fatigue.

### P-154

# Effectiveness of Resistance Exercise in women with primary Sjögren's syndrome (pSS): a randomized trial

Paulo A Minali<sup>1</sup>, Luciana Dardin<sup>1</sup>, Carolina FMGP Mota<sup>1</sup>, Virginia FM Trevisani<sup>1.2</sup>.

<sup>1</sup>Evidence-Based Medicine Discipline, Sao Paulo Federal University (UNIFESP) –Sao Paulo, Brazil. <sup>2</sup>Rheumatology Discipline, Santo Amaro University (UNISA) – Sao Paulo, Brazil.

**Introduction.** Primary Sjögren Syndrome (pSS) is an autoimmune inflammatory systemic disease that affects exocrine glands and less frequently internal organs. Inflammation can affect any system, including the musculoskeletal system, resulting in reduction in physical functions with consequent decrease in muscle strength, aerobic capacity, joint mobility, and static balance. In addition, psychosocial impairment with potential worsening of quality of life and functional capacity (FC) is described.

**Objective.** Analyze the effectiveness of resistance exercise on daily motor behavior and FC in women with pSS.

**Methods.** Fifty nine patients were randomized; of which 51 completed the study (26 assigned to the exercise group - EXG and 25 to the control group - CG). The EXG participated in a 16 week supervised exercise program, including 2 sessions per week and 3 sets of 10 maximal repetitions per exercise. Both before and at the end of the intervention protocol four variables were evaluated: the daily motor activity index (DMAI) was analyzed by an actigraph, which records body movements; the FC by the Fullerton Functional Fitness Test that consists of a sequence of 6 tests that mimic the neuromotor and cardiorespiratory needs involved in the activities of daily life; the disease activity (ESSDAI questionnaire); and quality of life (SF-36 questionnaire).

**Results.** After the intervention period in the EXG group, all FC parameters showed improvement over basal and final times, except the upper limb (UL) flexibility test (p=0.896): UL strength (p<0.001), lower limb (LL) strength (p<0.001), lower limb (LL) strength (p<0.001), LL flexibility (p=0.001), aerobic capacity (p<0.001), agility (p=0.002). A similar situation occurred with the SF-36 where all domains improved, except for the emotional aspect (p=0.710): functional capacity (p<0.001), limitation by physical aspects (p=0.005), pain (p<0.001), and mental health (p<0.001). There was no change in the DMAI (p=0.2) or ESSDAI (p=0.284). In the CTRL group, the only variable that showed a difference when compared to baseline was aerobic capacity of the FC test, presenting a tendency to improve (p=0.05).

**Conclusion.** Thirteen of the 17 variables evaluated showed improvement after the resistance exercise program. The intervention protocol did not change the daily motor behavior (DMAI), flexibility parameter of UL of the FC test, or the domain emotional aspects of the quality of life questionnaire. Resistance exercise did not worsen disease activity (ESSDAI). In this sense, we can conclude that the program that included resistance exercise was effective in improving functional capacity and quality of life in women with pSS.

# P-155

# Pilocarpine spray for the treatment of xerostomia: a randomized, double-blind, placebo-controlled trial

Ana Carolina Fragoso Motta, Vanessa Tonetto Marques, Fernanda Fortes Cabral, Tábata Larissa dos Santos Pólvora, Leandro Dorigan de Macedo, Lara Maria Alencar Ramos Inocenttini, Maira Peres Ferreira Duarte, Fabiola Reis Oliveira, Eduardo Melani Rocha, Osvaldo de Freitas, Camila Tirapelli. School of Dentistry of Ribeirão Preto, Ribeirão Preto Medical School, School of Pharmaceutical Sciences of Ribeirão Preto, University of São Paulo, Brazil.

**Background.** Xerostomia secondary to hyposalivation is a significant burden for patients with Sjögren's Syndrome (SS), since it may affect taste, speech, swallowing, and cause oral discomfort and burning. Management of xerostomia remains an unresolved topic for SS patients. Oral pilocarpine has showed to be very effective for controlling xerostomia in several clinical trials, but with variable side effects. Topical formulation could be an approach for getting local effect with minimal side effects. The aim of this study was to evaluate the effectiveness of pilocarpine spray as a treatment for xerostomia in patients with SS.

**Methods.** This was a randomized, double-blind, crossover placebo-controlled clinical trial (NCT02982577) of patients with SS complaining of xerostomia. Patients were randomly assigned to either placebo or pilocarpine (5%) spray and instructed to use 3 times a day for 4 weeks (equivalent concentration to 5mg of oral pilocarpine). Patients used pilocarpine spray or placebo for one month, with a one-month washout period between treatments. Outcomes measures were salivary flow (Unstimulated Whole Saliva Flow - UWSF), xerostomia (Xerostomia Inventory - XI) and quality of life (Oral Health Impact Profile - OHIP-14), which were performed at baseline, one hour (only UWSF) and at one month of treatment.

**Results.** Twenty-four patients were enrolled and randomized to receive pilocarpine (n=12) or placebo spray (n=12). Pilocarpine spray significantly increased UWSF after one hour of using the spray (Wilcoxon test; p=0.01), but no changes were observed at one month. Xerostomia symptoms (XI) improved at one month of therapy (Friedman test; p<0.05), and OHIP-14 scores declined 1 month after using pilocarpine spray related to functional limitation dimension (Friedman test; p=0.03). Sprays were well tolerated and no clinical side effects were observed.

**Conclusions.** Topical application of pilocarpine spray is effective and safe for managing xerostomia in patients with SS, and should be considered as a therapeutic option.

# P-156

# A new treatment method of severe keratoconjunctivitis sicca with designed soft contact lens filled with cyclosporin A

Safonova T.N.

Research Institute of Ophthalmic Diseases, Russian Federation.

**Background.** The aim is to improve the treatment methods of severe forms of keratoconjunctivitis sicca (KCS) with the help of the newly designed soft contact lens (SCL).

**Material and methods.** We created an original SCL made of silicone hydrogel material with individual optical power and multiple grooves filled with 0.05% Cyclosporin A (CyA).

This SCL saturated with CyA was examined in 31 patients (4 male and 27 female (43 eyes)) aged from 32 to 67 years with severe KCS, who were constantly wearing them for 7-14 days. Additionally, they instilled artificial tears with hyaluronic acid without preservatives up to 6 times a day. Standard ophthalmic examination, Schirmer test, break-up time, tests with the vital dye, which was assessed by the Oxford scale, osmometry of tears (TearLab System, USA), pH-metry tears on test basis with a litmus test of high sensitivity, seeding content conjunctival cavity of the microflora, the measurement of the width of the optic fissure were carried out, lesion index of the ocular surface (OSDI) was determined. Results of treatment were monitored after 1 week, 1, 3, 6, 12 months from beginning of SCL wearing. Results. Wearing SCL saturated with CyA, facilitated complete epithelialization of the cornea and helped to reduce inflammation from 1 week up to 1 month (Table I). This significantly reduced time of treatment of severe forms of KSC compared with standard therapy. After clinical improvement achievement CyA eyedrops were instilled twice a day. On this treatment instillation of artificial tears decreased to 3-4 times a day.

**Conclusion.** This new treatment method of severe forms of KCS was found on application of originally designed therapeutic SCL which provided output of CyA 0.05% on ocular surface, while instilling artificial tears without preservatives. The results have shown high efficiency of this method in terms of reducing corneal epithelialization and relieving inflammation on the ocular surface.



Table I. Dynamics of parameters on treatment.

	Before treatment	1 week	1 mo	3 mo	6 mo	12 mo
Vision	0,2-0,9 ( <i>Me</i> =0,5)	0,5-1,0 ( <i>Me</i> =0,8)*	0,8-1,0 ( <i>Me</i> =0,9)*	0,8-1,0 ( <i>Me</i> =0,9)*	0,8-1,0 ( <i>Me</i> =0,9)	0,8-1,0 ( <i>Me</i> =0,9)
Width of the optic fissure ( <i>M</i> ± <i>SD</i> ), mn	5,0±0.7	9,5±0.5*	9,0±1,0*	9,0±q,0*	9,1±0.8*	9,2±0,7*
Schirmer test (M±SD), mm	3,4±1,1	3,7±1,2	4,8±1,4	6,1±0,6*	7,1±1,0*	7,9±1,0*
Osmometry of tears, mOsms/L	316-381 ( <i>Mc</i> =318,0)	275-316 ( <i>Mc</i> =300,5)*	275-306 ( <i>Mc</i> =275,8)*	275-305 ( <i>Mc</i> =275,5)	275-302 *( <i>Mc</i> =275,5)	275-302 *( <i>Mc</i> =275,0)*
pH of tears (M±SD)	6,5±0,2	6,5±0,2	6,5±0,2	6,5±0,2	6,5±0.4	6,5±0,4
Tests with the drye (M±SD), ball	3,1±0,5	1,5±0,5*	0,5±0,2*	0,5±0,2*	0,4±0,1*	0,4±0,1*
OSDI ( <i>M</i> ± <i>SD</i> ), ball	86,8±10,3	38,7±17,2*	38,7±17,2*	17,2±5,5*	15,8±4,7*	15,8±4,0*

\*(p<0,05).

### P-157

# Dry eye therapeutics: approvals finally, pipeline bursting, novel delivery, new mechanisms

Sheppard, John.

Eastern Virginia Medical School, Virginia Eye Consultants, Norfolk, VA United States.

Dry eye disease (DED) therapy has entered a new golden era of innovation, funding, marketing and awareness. Clinical choices will empower prescribers, benefit patients, challenge payers and foster awareness.

**Approvals.** Topical Liftegrast 0.1% preservative free solution (Xiidra, Shire) welcomes the first addition to topical DED therapeutics in 12 years. Novel molecular engineering created an active totally unique to ocular surface application. A successful launch prompted growth in numerous ocular surface markets, including Restasis.

Neurostimulation (True Tear, Allergan) introduces a totally new mechanism of action thought to be complimentary to existing allopathic surgical and pharmaceutical therapeutics. The handheld device emulates natural tripartite tear production through afferent Trigenminal nerve stimulation, controlled by the patient.

**Pipeline.** There are over 40 entities in Phase I through Phase III evaluation for DED therapy. These agents include semi-fluorinated alkane or nanoparticle vehicle delivery of cyclosporine, an enhanced mucus penetrating nanoparticle vehicle with loteprednol etabonate for treatment of episodic dry eye, epithelial sodium channel blocking agent, neuropeptide agonist, topical pure hypochlorous acid, and an aldehyde sequestration agent.

Topical biologics include recombinant human serum albumin (rHSA), a

naturally occurring secretagogue (lacritin), and a large surface-active endogenous complex mucinous glycoprotein (lubricin).

**Delivery.** New methods for drug delivery include a wide variety of topical vehicles: emulsions, mucus penetrating particles, semifluorinated alkanes, nanoparticles, and gels. Iontophoresis can delivery charged particles to the cornea, sclera and anterior segment. Neurostimulation is delivered through a single use biodegradable polymer for nasal mucosal interface. Thermal pulsation therapy provides deep Meibomian gland warming, massage and extirpation. Intense pulsed light (IPL) and electrolytic probing (Maskin) liberate posterior lamellar lid secretions.

**Novel mechanisms.** include LFA1-ICAM blockade, anterior ethmoidal nerve electrostimulation, ENaC blockade, and topical minocycline induced physiochemical changes in Meibomian secretion content.

Landscape. Payers will see more challenges as numerous new entities enter the marketplace after the intensely costly approval process. Despite costs, early intervention, lifestyle and advocational efficiency, decreased absenteeism, and reduced morbidity from ulceration, keratitis, pain, refractive error will undoubtedly benefit society through reduced clinic visits, fewer wasted empirical prescriptions, and lower medication costs overall.

#### P-158

#### Superior inhibition of disease-relevant inflammatory mediators and cell biology by combined leftunomide and hydroxychloroquine supports rationale for combination therapy in pSS patients

E. van der Heijden<sup>1,2</sup>, S. Hartgring<sup>1,2</sup>, A. Kruize<sup>2</sup>, T. Radstake<sup>1,2</sup>, J. van Roon<sup>1,2</sup>. <sup>1</sup>Laboratory of Translational Immunology, <sup>2</sup>Dept. of Rheumatology & Clinical Immunology, UMC Utrecht, Utrecht, the Netherlands.

**Background.** T and B-cell-driven immunity is critically involved in immunopathology of pSS. Recently we demonstrated synergistic T and B-cell activation upon T cell triggering and TLR7/9-driven B cell activation in pSS patients, accompanied by synergistic induction of immunogloulins and IFN $\gamma$ - and IL-17- producing T-cells. In addition, TLR7/9-expressing activated pDCs associated with increased type I IFNs and IFN-inducible genes are increased pSS patients. Several studies have shown that the DMARDs leflunomide and hydroxycholoroquine inhibit immune activation in pSS but only show moderate efficacy. However, LEF and HCQ target different pathways with overlapping, but also potentially additive mechanisms, where LEF primarily targets T and B-cells and HCQ TLR7/9-driven B-cell and pDC activation. Hence, the additive effects of LEF and HCQ were assessed in vitro on CD4 T- and B-cell activation and production of interferons  $\alpha$  and  $\gamma$ , Th-related cytokine CXCL13, as well as IgG and IgM. Inhibition was compared to IFN $\alpha$  receptor (IFNAR) blockade.

**Methods.** PBMCs of healthy individuals (n=9) and of pSS patients (n=8) were cultured with antigen (SEB) and TLR9L, in presence or absence of LEF, HCQ and their combination in clinical relevant concentrations. Anti-IFNAR antibody was tested at the maximal inhibitory concentration. Proliferation of T and B-cells and release of IFN- $\alpha$ , IFN- $\gamma$ , CXCL13, IgG and IgM were measured.

**Results.** SEB/TLR9 robustly induced T and B-cell activation, IFN $\gamma$ , IFN $\alpha$ , CXCL13, IgG and IgM production (all *p*<0.001). LEF dose dependently inhibited B and T-cell proliferation, IFN, CXCL13 and Ig production. HCQ dose dependently inhibited B-cell proliferation, IFN- $\alpha$ , CXCL13, and immunoglobulin production. T-cell proliferation and IFN- $\gamma$  production were inhibited by HCQ only at higher concentrations. At several suboptimal concentrations LEF and HCQ additively inhibited T-cell proliferation both in healthy individuals and in pSS patients. Significant additive effects were seen for all outcome measures except IFN- $\alpha$ . Since IFN $\alpha$  was already robustly inhibited by HCQ alone (eg.for pSS 90% at 3.3 µM, *p*<0.001), only trends towards additive effects were observed. Interestingly, anti-IFNAR treatment strongly inhibited IFN $\alpha$  production and CXCL13 production (both *p*<0.001), but did not inhibit T and B cell proliferation or IFN $\gamma$  production.

**Conclusion.** More potent than IFNAR blockade LEF and HCQ robustly inhibited proliferation of T and B-cells, cytokine production and immuno-globulin production with clear additive efficacy in both healthy individuals as in pSS patients. These data support the potential surplus value of combination therapy with LEF and HCQ for patients with pSS.

### P-159

Multicenter, Phase 2 exploratory study evaluating the safety and efficacy of DEXTENZA for the treatment of dry eye disease

Sheppard JD<sup>1</sup>, Torkildsen GL<sup>2</sup>, Metzinger JL<sup>3</sup>, Stein L<sup>4</sup>, Ousler G<sup>4</sup>, Goldstein  $MH^3$ .

<sup>1</sup>Virginia Eye Consultants, Norfolk, VA. <sup>2</sup>Andover Eye Associates, Andover, MA. <sup>3</sup>Ocular Therapeutix, Inc., Bedford, MA. <sup>4</sup>Ora, Inc., Andover, MA.

**Purpose.** To evaluate the safety and efficacy of DEXTENZA<sup>™</sup> as a 30-day sustained release drug (dexamethasone) insert compared to placebo (PV) when placed in the canaliculus of the eyelid in subjects who exhibit signs and symptoms of dry eye disease.

Methods. This was a prospective, multicenter, randomized, parallel-arm, double-masked, vehicle controlled feasibility study to evaluate the safety and efficacy of DEXTENZA compared to PV for the treatment of subjects exhibiting signs and symptoms of dry eye disease. Initially, the effect of the PV was established during a 6-week insert-only phase to establish baseline data; subsequently, subjects were randomized 1:1 to receive DEXTENZA or PV. The insert was placed in the inferior vertical canaliculus of both eyes of each subject. During the Treatment Phase, subjects underwent follow-up visits at ≥60 minutes post-insertion and at post-insertion Days 3, 15 and 30. Subjects were exited from the study when the Investigator confirmed patency in both eyes. A broad range of efficacy and safety measures were collected, due to the exploratory nature of this study, including corneal fluorescein staining and conjunctival Lissamine green staining, questionnaires (ODS, OSDI, VAS and SPEED), Schirmer testing, and tear-film breakup time. Retention, ease of use and ease of visualization were also evaluated, in addition to other exploratory and safety assessments.

**Results.** A total of 43 subjects (DEXTENZA, 22; PV, 21) were randomized. Subjects in the DEXTENZA group showed statistically significant improvements in corneal fluorescein staining compared to subjects in the PV at Days 15 and 30 for the inferior region and at Day 30 for the total corneal region. Conjunctival staining results were significant for the DEXTENZA group for total conjunctival staining and for 4 of 6 regions. The severity and frequency of dry eye symptoms decreased over the course of the study in both treatment groups as assessed by the SPEED questionnaire. Slight discomfort (ODS) and moderate discomfort (VAS, 29% to 38%) were reported by subjects, with similar scores in both groups. Product insertion and product visualization were rated as easy for most subjects. No significant differences occurred between the groups for the presence of the insert on Days 60 and 90. The insert was not visualized in any subjects by Day 90. No serious adverse events related to DEXTENZA were reported.

**Conclusion.** DEXTENZA showed moderate improvements in the clinical signs of dry eye disease. Use of DEXTENZA may be safe and effective for the reduction in signs of dry eye disease; further, larger-scale Phase 3 studies are warranted.

# P-160

# Evaluation of Pilocarpine treatment in xerostomia by pulsed doppler color ultrasonography: ECHOPILO Study

T. Depinoy, A. Saraux, J-O. Pers, S. Boisramé, D. Cornec, T. Marhadour, D. Guellec, V. Devauchelle-Pensec, S. Jousse-Joulin.

**Objective.** Assessment of the evolution of major salivary gland structural involvement is an important issue in the follow-up of patients complaining of xerostomia. The objective of our study was to evaluate the vascularization of parotid glands (PG) using Pulsed Doppler color ultrasonography (USPD) in patients treated by pilocarpine.

**Methods.** We prospectively included patients with objective dry mouth syndrome who attended the Sjögren's Syndrome Multidisciplinary Consultation at Brest University Hospital. The vascularization was assessed by the RI at the left parotid before and after stimulation with lemon. Only patients with pathological RI (<0.8) were included in order to observe RI evolution after pilocarpine. A pulsed doppler USPD of the salivary glands with measure of the vascularization by RI for the parotid glands was carried out by the same operator (SJJ). A dental consultation with measure of salivary flows before and after stimulation was performed. These examinations were performed at baseline and after 3 months of treatment with pilocarpine at 4 mg 3 times daily.

**Results.** Among the 19 patients included, 11 received pilocarpine treatment for the whole 3 months period, 6 of the 8 remaining patients stopped the pilocarpine due to side effects. Among the 11 patients with a follow-up evaluation at 3 months, 5 had primary Sjögren's syndrome according to the American-European's classification criteria. The differences of RI before and after lemon stimulation was on average of -0.04 at baseline and -0.04 at M3. The sum of ultrasound's grades average of the four glands was 3.47 at M0 and 4.18 at M3. The non-simulated salivary flow was on average of 1.96 mL/mn at M0 and 5.23 mL/mn at M3, whereas the average of stimulated salivary flow was 2.84 mL/mn at M0 and 8.51 mL/mn at M3. None of these observed differences were statistically significant: RI before and after lemon stimulated (p=0.26) and stimulated salivary flow (p=0.139). Concerning the 3 patients with Sjögren's syndrome, the differences in RI before and after lemon stimulation was lower probably due to a lower initial RI.

**Conclusion.** Preliminary results showed no significant differences between the 4 gland's grade, ultrasound's RI and salivary flow before and after three months of pilocarpine's treatment. The study was marked by a large number of pilocarpine's discontinuation (31 %) due to adverse effects. The vascularisation of salivary glands could be an opportunity to follow our patient with Sjögren's syndrome or with xerostomia but more studies are needed to prove the interest of this procedure.

#### References

- 1. CORNEC *et al.*: «Contribution of Salivary Gland Ultrasonography to the Diagnosis of Sjögren's Syndrome».
- JOUSSE-JOULIN et al.: «Ultrasound Assessment of Salivary Glands in Patients with Primary Sjögren's Syndrome Treated with Rituximab».

#### **P-161**

#### Sjögren's Syndrome Foundation Clinical Practice Guidelines for pulmonary involvement in Sjögren's

Scofield RH<sup>1</sup>, Lee AS<sup>2</sup>, Baraf HSB<sup>3</sup>, Gupta N<sup>4</sup>, Lynch J<sup>5</sup>, Meehan R<sup>6</sup>, Moua T<sup>7</sup>, St. Clair EW<sup>8</sup>, Dunleavy K<sup>9</sup>, Carteron N<sup>10</sup>, Carsons SE<sup>11</sup>, Hammitt KM<sup>12</sup> on behalf of the Sjögren's Syndrome Foundation.

<sup>1</sup>Oklahoma Medical Research Foundation; University of Oklahoma Health Sciences Center; US Department of Veterans Affairs Medical Center, <sup>2</sup>Mayo Clinic, Jacksonville, FL, <sup>3</sup>Arthritis and Rheumatism Associates; George Washington University School of Medicine; University of Maryland School of Medicine; <sup>4</sup>University of Cincinnati, <sup>5</sup> \*National Jewish Health, <sup>7</sup>Mayo Clinic, Rochester, MN, <sup>8</sup>Duke University Medical Center, <sup>9</sup>George Washington University Cancer Center; <sup>10</sup>University of California San Francisco, <sup>11</sup>New York University School of Medicine, Winthrop-University Hospital, <sup>12</sup>Sjogren's Syndrome Foundation.

**Background.** Management guidelines for Sjögren's patients remain a major unmet need, particularly for potentially severe extraglandular (systemic) manifestations. To meet this patient and clinician need, the Sjögren's Syndrome Foundation (SSF) launched a major initiative to develop Clinical Practice Guidelines for Sjögren's in 2012. Goals include: improving quality and consistency of care, creating guidance documents for US clinicians, obtaining broad acceptance of guidelines by key professional and government organizations, educating payers and identifying gaps in evidence to spur much needed research. Following 4 recent publications by the SSF on initial Clinical Practice Guidelines under the rheumatology/systemic, oral, and ocular topics, Phase 2 is now well underway. This phase aims to comprehensively collate scientific evidence and expert opinion on the management of pulmonary manifestations of Sjögren's.

**Methods.** Guideline members developed a rigorous and transparent process based on American College of Rheumatology quality of care standards and protocols developed by other professional organizations such as the American Society of Clinical Oncology. A unique aspect of these guidelines is the inclusion of multiple specialists. The Pulmonary Topic Review Group (TRG) includes an equal number of rheumatologists and pulmonologists as well as an oncologist with expertise in Sjögren's. The process involved drafting and ranking clinical questions, defining the literature search and study criteria, and outcome measures to be addressed *a priori* to execution of the search. Members then identified eligible abstracts, and a minimum of 2 TRG members extracted data into tables, which included study characteristics and evidence and quality assessment. Recommendations will be drafted and finalized via a Delphi-type consensus process involving more than 30 medical specialists.

**Results.** Areas of pulmonary coverage were outlined and include screening and diagnosis; upper airway disease (xerostomia, dysphagia, laryngopharyngeal reflux, vocal cord, obstructive sleep apnea); lower airway disease (xerotrachea, bronchiectasis, bronchiolitis, obstructive lung diseases including COPD and asthma); interstitial lung disease; lymphoproliferative disease (including bronchus- associated lymphoid tissue lymphoma/ non-Hodgkin lymphoma, amyloid, and nodular lymphoid hyperplasia); vascular lung disease; lung transplant; and peri-operative and other non-medication management. Almost 200 informational and clinical questions were drafted. **Conclusions.** Clinically relevant questions for the development of Clinical Practice Guidelines for pulmonary manifestations in Sjögren's were developed by multi-specialty teams of Sjögren's experts across the U.S. Literature searches, data extraction, and recommendation development are underway. Recommendations will utilize evidence in Sjögren's when available and evidence from closely related diseases and expert opinion when little to no evidence in Sjögren's is available. Areas for future research will be delineated with a focus on patient-perceived areas of unmet need.

Disclosures. The SSF Clinical Practice Guidelines are fully supported by the Sjögren's Syndrome Foundation with no pharmaceutical support. No compensation was paid to any author other than salary to SSF staff. **RH Scofield**, Abbvie; **A Lee**, Intermune, Gilead; **HSB Baraf**, Abbvie, Daiichi Sankyo, Gilead, GlaxoSmithKline, Novartis, Pfizer, R-Pharm, UCB, United Healthcare; **N Gupta**, None; **J Lynch**, None; **R Meehan**, Sanofi Genzyme, Fidia; **T Moua**, Boehringer Ingelheim; **EW St. Clair**, Bristol-Myers Squibb; **K Dunleavy**, None; **N Carteron**, Bristol-Myers Squibb, Genentech Roche, Regeneron; **SE Carsons** Novartis, Roche; **KM Hammitt**, None.

#### P-162

#### Sjögren's Syndrome Foundation Clinical Practice Guidelines for mucosal management and treatment and use of secretagogues in Sjögren's

Al-Hashimi I<sup>1</sup>, Papas A<sup>2</sup>, Alevizos I<sup>3</sup>, Brennan M<sup>4</sup>, Navazesh M<sup>5</sup>, Pinto A<sup>6</sup>, Stewart C<sup>7</sup>, Sweier D<sup>8</sup>, Tanzer J<sup>9</sup>, Vivino F<sup>10</sup>, Shiboski C<sup>11</sup>, Zero DT<sup>12</sup>, Carsons SE<sup>13</sup>, Hammitt KM<sup>14</sup> on behalf of the Sjögren's Syndrome Foundation. <sup>1</sup>University of Texas Southwestern Medical Center, <sup>2</sup>Tufts University School of Dental Medicine, <sup>3</sup>National Institute of Dental and Craniofacial Research, National Institutes of Health, <sup>4</sup>Carolinas Medical Center, <sup>3</sup>University of Southern California, <sup>6</sup>Case Western Reserve, <sup>7</sup>University of Florida School of Dentistry, <sup>8</sup>University of Michigan School of Dentistry, <sup>9</sup>Retired, University of Connecticut School of Dental Medicine, <sup>10</sup>University of Pennsylvania, Penn Presbyterian Medical Center, <sup>11</sup>University of Connetistry School of Medicine; Winthrop-University Hospital, <sup>14</sup>Sjögren's Syndrome Foundation.

**Background.** Oral complications in Sjögren's can lead to painful oral infection/lesions/irritation, rampant caries and loss of teeth, in addition to difficulty eating and talking, which compromise the quality of life of patients with Sjögren's. There is a clear unmet need for managing the lack of saliva and its consequences in Sjögren's. The Sjögren's Syndrome Foundation (SSF) launched a major initiative to address this need in 2012, and clinical practice guidelines for caries prevention were published in 2016 in the *Journal of the American Dental Association (JADA)* and included in the *Rheumatic Diseases Clinics of North America*. Phase 2 oral guidelines are now well underway and include coverage of mucosal management and treatment and use of secretagogues in Sjögren's. Guidelines on caries restoration and management and parotid gland swelling also will be developed.

**Methods.** Guidelines members developed a rigorous and transparent process based on standards set by the American Dental Association, American College of Rheumatology and American Society of Clinical Oncology. The methodology utilizes a Delphi-type consensus process and includes drafting clinical questions and defining literature search parameters and study criteria *a priori* to execution of the search. Eligible abstracts then are identified, and a minimum of 2 guidelines members extract the data on study characteristics, evidence and quality assessment. Recommendations will be drafted and finalized with input from at least 30 oral healthcare professionals who were not involved in the guidelines development.

**Results.** The Mucosal Management and Treatment Topic Review Group (TRG) drafted clinical questions for its outline covering oral symptoms of mucosal pain and/or inflammation in Sjögren's, including prevalence and identification of oral cancer. Prevention and management of burning mouth; oral candidiasis; lichenoid reactions or hypersensitivity-induced mucositis; and oral trauma associated with oral dryness were discussed. The Secretagogues TRG's clinical questions explore whether studies have demonstrated that sialogogues improve salivary flow and composition; reduce the occurrence of candida, other or al infections and burning mouth; and translate to better outcomes for the incidence of caries, periodontal disease, taste disturbance; hoarseness, chronic cough, and subjective and objective dryness and discomfort.

**Conclusions.** Initial clinically relevant questions for Clinical Practice Guidelines for oral mucosal management and use of secretagogues for dry

mouth in Sjögren's were established by Sjögren's experts across the U.S. Literature searches, data extraction, and recommendation development is underway. Recommendations utilize evidence from Sjögren's studies when available and expert opinion when little to no evidence in Sjögren's literature is available. Areas for future research will be delineated with a focus on patient-perceived areas of unmet need.

Disclosures. The SSF Clinical Practice Guidelines are fully supported by the Sjögren's Syndrome Foundation with no pharmaceutical support. No compensation was paid to any author other than salary to SSF staff. I Al-Hashimi, None; A Papas, Biogen, GSK, MGI Pharma, Novarits, Parion; I Alevizos, Medimmune; M Brennan, Medimmune, Dermtreat; M Navazesh, None; A Pinto, CAP trials; C Stewart, None; D Sweirs, None; J Tanzer, Advantage Dental; FB Vivino, Biogen, Immco Diagnostics, Novartis; C Shiboski, Pasteur Institute; DT Zero, GlaxoSmithKline, Johnson & Johnson, C3 Jian, Noveome Therapeutics, Novartis; SE Carsons, Novartis, Roche; KM Hammitt, None.

#### P-163

#### Sjögren's Syndrome Foundation Clinical Practice Guidelines for neurological involvement in Sjögren's

Mandel S<sup>1</sup>, Vivino FB<sup>2</sup>, Fox RI<sup>3</sup>, Deboo A<sup>4</sup>, Birnbaum J<sup>5</sup>, Bloch D<sup>6</sup>, Brasington R<sup>7</sup>, Brown ES<sup>8</sup>, De Sousa E<sup>9</sup>, Gelfand J<sup>10</sup>, Gronseth G<sup>11</sup>, Lange DJ<sup>12</sup>, Lawrence-Ford T<sup>13</sup>, Lewis J<sup>14</sup>, Liao YJ<sup>15</sup>, Maitz E<sup>16</sup>, Maitz S<sup>17</sup>, Noaiseh G<sup>18</sup>, Pavlakis P<sup>19</sup>, Sarka G<sup>20</sup>, Sicotte N<sup>21</sup>, Varadhachary A<sup>22</sup>, Wallace DJ<sup>23</sup>, Wilson JW<sup>24</sup>, Winter WC<sup>25</sup>, Scofield RH<sup>26</sup>, Carteron N<sup>27</sup>, Carsons SE<sup>28</sup>, Hammitt KM<sup>29</sup> on behalf of the Sjögren's Syndrome Foundation.

<sup>1</sup>Hofstra Northwell, <sup>2</sup>University of Pennsylvania, Penn Presbyterian Medical Center, <sup>3</sup>Scripps Memorial Hospital, <sup>4</sup>Lewis Katz School of Medicine at Temple University, <sup>5</sup>The Johns Hopkins University School of Medicine, <sup>6</sup>Partners Healthcare; Massachusetts General Hospital; Harvard Medical School, <sup>7</sup>Washington University in St. Louis, <sup>8</sup>University of Texas Southwestern Medical Center, <sup>9</sup>University of Oklahoma, <sup>10</sup>University of California San Francisco, <sup>11</sup>University of Kansas, <sup>12</sup>Hospital for Special Surgery; Cornell, <sup>13</sup>North Georgia Rheumatology Group; Philadelphia College of Medicine; Emory University, <sup>14</sup>University of Virginia, <sup>15</sup>Stanford University, <sup>16</sup>Widener University, <sup>17</sup>PhD-candidate, Chestnut Hill College, <sup>18</sup>University of Pittsburgh Medical Center, <sup>19</sup>Hospital for Special Surgery, <sup>30</sup>UCLA, CSUN, <sup>21</sup>Cedars-Sinai/David Geffen School of Medicine at UCLA, <sup>22</sup>Washington University in St. Louis, <sup>22</sup>Cedars-Sinai/David Geffen School of Medicine at UCLA, <sup>24</sup>Retired; Rheumatology, <sup>25</sup>Charlottesville Neurology and Sleep Medicine; Martha Jefferson Hospital Sleep Medicine Center, <sup>26</sup>Oklahoma Medical Research Foundation; University of Oklahoma Health Sciences Center; US Departnet of Veterasity School of Medicine; Winthrop-University Hospital, <sup>28</sup>Sjögren's Syndrome Foundation.

**Background.** Standardization of the diagnosis and treatment of neurologic manifestations of Sjögren's remains an unmet clinical need. A multi-disciplinary task force of rheumatologists, neurologists and other specialists has been formed to develop consensus recommendations concerning the neurological features of the disease.

**Methods.** Guidelines members in rheumatology and neurology were organized into central (CNS) and peripheral nervous system (PNS) Topic Review Groups (TRGs). Additional specialists include neuro-ophthalmology, vasculitis, psychiatry, neuro-psychology, and sleep medicine. The initial guidelines development process is based on American College of Rheumatology and American Society of Clinical Oncology procedures, including drafting and ranking of clinical questions, defining literature search parameters and determining outcome measures to be addressed prior to the literature review. Members then will identify eligible abstracts and a minimum of 2 TRG members extract data on evidence and study quality. Recommendations will be drafted and finalized via the Delphi method. The American Academy of Neurology appointed representatives to both TRGs.

**Results.** PNS coverage areas were based upon both the neuro-anatomic localization to the motor, sensory and autonomic nerves and the etiology involving demyelinating, axonal and antibody-based pathologies. These include cranial neuropathies; axonal or sensory motor or sensory neuropathies; small fiber neuropathies (length-dependent or non-length dependent); ataxic sensory neuropathies/large fiber ganglionopathies; mononeuritis multiplex/multiple mononeuropathies; and autonomic neuropathies. CNS coverage areas were based upon the clinical syndromes involving encephalopathies, seizures, strokes, and myelopathies and etiologies including demyelinating and vasculopathic pathology. Included are vasculiti; optic neuritis; vestibular/auditory, olfactory, taste; autoimmune encephalitis; myelitis or other demyelinating syndromes; psychiatric (anxiety, depression, psychosis); cognitive dysfunction; and impaired sleep.

**Conclusions.** The initial clinically relevant questions for Clinical Practice Guidelines for neurological manifestations in Sjögren's were developed by multi-specialty teams of Sjögren's experts across the U.S. Literature searches, data extraction, and recommendations for the diagnosis and treatment of

neurological manifestations of Sjögren's are based on published studies related to Sjögren's or related diseases. When results of clinical investigation are not available, consensus expert opinion will be used as an initial step to standardize management and treatment.

Disclosures: The SSF Clinical Practice Guidelines are fully supported by the Sjögren's Syndrome Foundation with no pharmaceutical support. No compensation was paid to any author other than salary to SSF staff. S Mandel, None; FB Vivino, Biogen, Immco Diagnostics, Novartis; RI Fox, Abbvie, Celgene, Novartis; A Deboo, None; J Birnbaum, None; D Bloch, Amgen, Baxter, Biogen, Collagene, Foundation Medicine, Lab Corp, Novartis, Roche; R Brasington, Amgen, Celgene, GlaxoSmithKline, Mallinckrodt, Novartis, Pfizer; ES Brown, None; E De Sousa, None; J Gelfand, Genentech, MedDay, Eli Lilly, Zosano, ENeura; G Gronseth, None; DJ Lange, None; T Lawrence-Ford, Genesis Pure, Abbvie, Amgen, Bristol-Myers Squibb, Eli Lilly, Genentech, GlaxoSmithKline, Irko, Janssen, Pfizer, Questcor, Takeda, UCB; J Lewis, Lupus Research Alliance; YJ Liao, None; E Maitz, None; S Maitz, None; G Sarka, None; N Sicotte, None; A Varadhachary, Cassidy Schade LLP, Hughes-Socol-Piers Resnick, WUSTL Hospitalist; DJ Wallace, None; JW Wilson, None; WC Winter, None; RH Scofield, Abbvie; N Garteron, Bristol-Myers Squibb, Genentech Roche, Regeneron; SE Carsons, Novartis, Roche; KM Hammitt, None.

#### **P-164**

#### A summary of key findings from the Sjögren's Syndrome Foundation's National Patient Survey

Taylor S<sup>1</sup>, Champigny M<sup>1</sup>, Hammitt KM<sup>1</sup>, Makara M<sup>1</sup>, Sankar V<sup>2</sup>, Forstot SL<sup>3</sup>, Hurley P<sup>4</sup>, Niewold TB<sup>5</sup>, Carteron N<sup>6</sup>.

<sup>3D</sup>, Hurley L, Hurley L, Hurley H, Hurley H, Hurley H, Harvard Medical School, <sup>2</sup>Corneal Consultants of Colorado; University of Colorado School of Medicine, <sup>4</sup>American Society of Clinical Oncology, <sup>3</sup>New York University School of Medicine, <sup>6</sup>University of California San Francisco.

**Background.** Sjögren's is a systemic autoimmune disease that affects the entire body. The purpose of this major national patient survey was to gain an understanding from adults who have been diagnosed with Sjögren's about the impact of the disease on their quality of life, including the physical, emotional and financial burdens.

**Methods.** The Sjögren's Syndrome Foundation (SSF) conducted the *Living* with Sjögren's survey between May 11 and July 11, 2016. Participants were recruited by Harris Poll from a pool of 9,252 active SSF patient members. The survey was conducted among adults aged 18 years or older who reported having been clinically diagnosed with Sjögren's by a physician or other medical professional. The survey asked closed-ended questions about patient experiences with Sjögren's and the impact it has on their quality of life.

Results. There were 3,072 survey responses (33% response rate), 2,963 of which were included in the analyses. Survey respondents were 96% female and 4% male; 32% were aged 60 years or less. On average, respondents saw 4.6 different healthcare professionals annually and used 8.8 medications and treatments to help manage their Sjögren's symptoms. Nearly all respondents (96%) indicated they wished that additional treatments for Sjögren's were available. The most common symptoms experienced by survey respondents on a weekly or more frequent basis were dry mouth (92%), dry eyes (92%), fatigue (80%), dry or itchy skin (76%), and morning stiffness (69%). Respondents reported having been diagnosed by a health care provider with an average of five other health conditions, the most frequent of which were GERD (45%), Raynaud's (38%), neuropathy (38%), and sinusitis (33%). Most respondents (71%) reported Sjögren's interferes with their daily activities, and 79% and 54% indicated the disease has led them to make at least one day-to-day change at home or at work, respectively. Sjögren's was identified as adding a significant emotional burden to the lives of 74% of respondents (80% of patients >= 60 years of age vs 71% of > 60 years.). A negative impact on relationships with friends and family (63%), sex life (59%), spouse and partner relations (55%) and caring for children (19%) was also reported. 66 percent of respondents indicated Sjögren's adds a significant financial burden to their life. Those aged 60 years or less were more likely to view Sjögren's as a significant burden in their lives compared to those older than 60 (72% vs. 63%).

**Conclusions.** The findings from this national survey highlight the complexity of Sjögren's and its impact on patients and their families. Sjögren's is burdensome to patients and affects their well-being and quality of life in a variety of ways, including physical, emotional, and financial challenges. Patients reported the need for new treatment options to treat the various manifestations of Sjögren's. The findings from this survey will help to inform and support future SSF efforts to increase public and professional awareness of Sjögren's and encourage research into new treatments and a cure. Disclosures: The SSF Clinical Practice Guidelines are fully supported by the Sjögren's Syndrome Foundation with no pharmaceutical support. No compensation was paid to any author other than salaries to SSF staff. S Taylor, None; M Champigny, None; KM Hammitt, None; M Makara, None; V Sankar, Medimmune; SL Forstot, Allergan, Kala, Aldeyra, Tear Solutions, CSL-USA, Senju USA, CXLO; P Hurley, None; TB Niewold, EMD Serono, Janssen; N Carteron, Bristol-Myers Squibb, Genentech Roche, Regeneron.

#### P-165

#### Is elastography a new tool to differentiate Sjögren syndrome to sicca syndrome: results of the ELSA (elastography of salivary glands) study

S. Jousse-Joulin, L. Bressollette, T. Depinoy, V. Devauchelle-Pensec, D. Cornec, T. Marhadour, G. Carvajal, D. Guellec, A.Saraux.

**Background/Purpose.** Ultrasonography (US) has been developed in salivary glands (SG) and particularly in primary Sjögren syndrome (pSS) for 10 years. However, the training curve is long and the reliability is not completely achieved (1). A new tool procedure has been developed to study the elasticity of the tissue parenchyma using elastography and could be implemented in the evaluation of SGUS pSS patients.

The objective was to evaluate SGUS using grey scale (GS) and the elasticity salivary glands parenchyma using elastography to differentiate pSS from sicca syndrome (SS) patients in a longitudinal consultation of sicca syndrome in Doppler Unit of Brest (France).

**Methods.** 63 patients complaining of sicca syndrome were enrolled in the ELSA study. At inclusion, all patients underwent a standardized workup including a clinical evaluation, laboratory tests, SG histology, SGUS and elastography. Immunological testing included: anti-nuclear antibodies, anti-SSA (recognizing both Ro 52 and 60 kDa), anti SSB, native anti-DNA, ANCA, rheumatoid factors and anti-CCP. A Toshiba Applio machine was used to evaluate in GS SGUS parenchyma and elastography -elastometry. The 6 main SG were examined in US according to previous scoring (2) elastography (with color map) and elastometry (in cm/sec) and: left paroti d gland (USLSLG), right submandibular gland (USRSMG), right sublingual gland (USRSMG), right sublingual gland (USRSLG).

**Results.** 15 patients fulfilled the AECG criteria. The clinical characteristics of the patients in terms of ocular dryness and shirmer test were not significant between pSS and Sicca patients. There was a significant difference (p=0.04) concerning oral dryness (p=0.04), immunological datas: Antinuclear antibodies and anti SSA and minor salivary glands biopsy (p=0.007). We found significant differences between the 2 groups in GS for the USLPG (p=0.021) and USLSMG (p=0.05).Evaluation of sublingual glands using US in grey scale and elastometry of the 6 SG showed no significant differences between the two groups.

**Conclusion.** Grey scale SGUS seems to be more sensitive to differentiate pSS to Sicca patients compared to elastometry. US of sublingual glands showed no involvement in the echostructural damage in pSS patients and suggest not examining these glands in the USpSS evaluation. Elastometry has been described to be a new tool and might be added as a new imaging technique to ultrasonography in pSS patients (3). However, ELSA study results showed no differences between pSS and Sicca patients. We need to follow the evolution of the Sicca population with US in GS and elastometry to detect potential echostrural parenchymal damage which might not yet be present at inclusion.

#### References

- JOUSSE-JOULIN S, et al.: Is salivary gland ultrasonography a useful tool in Sjögren's syndrome? A systematic review. *Rheumatology* (Oxford) 2016 May; 55(5): 789-800.
- CORNEC D et al.: Contribution of salivary gland ultrasonography to the diagnosis of Sjögren's syndrome: toward new diagnostic criteria? Arthritis Rheum 2013 Jan; 65(1): 216-25.
- SAMIER-GUÉRIN A et al.: Can ARFI elastometry of the salivary glands contribute to the diagnosis of Sjögren's syndrome? Joint Bone Spine 2016 May; 83(3): 301-6.

### P-166

Major salivary gland ultrasonography as diagnostic tool of primary Sjögren's syndrome and lymphoma

Rodionova EB, Palshina SG, Burceva MV, Misiyuk A, Khvan UI, Saphonova TN, Vasiliev VI, Radenska-Lopovok SG, Torgashina AV, Sokol EV, Chalcev BD.

Nasonova research institute of Rheumathology, Research intitute of Opthalmic disease, Sechenova Medical university, Moscow Russian Federation.

**Objectives.** Ultrasonography of major salivary glands (USG) in patients with primary Sjögren's syndrome (pSS) is very popular non - invasive test and has potential source of classification criteria for this disease. USG was evaluated as an alternative to sialography and in comparison with parotid and minor salivary biopsy to pSS.

Methods. 44 patients with pSS (ACR-EULAR criteria) underwent USG, parotid sialography, stimulated saliva flow test (SFT) <2,5ml/5 min. Among them 16 had MALT- lymphoma development (group 2) and underwent parotid biopsy. During the diagnostic of pSS, primary major salivary glands lymphoma group was detected (MALT - lymphoma type -1, follicular lymphoma-2, Hodgkin type -1) - 3 group (4 cases). Parenchymal echogenicity, homogeneity, hypoechogenic and hyperechogenic areas and clearness of salivary gland border were scored according to the Hocevar scoring system (cut-off - 15). Total ultrasound score was counted in 1 group mediana - 22 (20, 75-26), 2 group- mediana - 15(13, 75-19, 25), 3 group - mediana -19,5 (19,75-21). Statistical analyses were performed using MEDCALC program. Pearson's chi-squared statistic had been used. The association between USG and SFT, parotid sialography, anti - SSA/Ro, MSG and parotid gland biopsy was analysed in two groups: pSS - 1 group (n=28) and pSS/MALT - 2 group (n=16). Mann-Whitney U tests were used to evaluate differences in total ultrasound score between patients in 3 groups.

**Results.** In the 1 pSS-group we found medium correlation between USG and SFT (p<0,05) and MSG biopsy (p>0,05), poor correlation with parotid sialography (p>0,05) and anti – SSA/Ro (p>0,05). In the pSS/ MALT - 2 group very strong correlation was detected between USG and parotid gland (p<0,01) biopsy, strong correlation with parotid sialography (p>0,05), and medium correlation with SFT (p>0,05). Poor correlation had been found with anti – SSA/Ro (p<0,05) and MSG (p<0,05) biopsy. Total ultrasound score was not significantly higher between first/ second group and group with primary salivary glands lymphoma (p>0,05).

**Conclusion.** Our data suggest that major salivary ultrasonography is the useful method for detecting late changes in pSS and complications such as MALT-lymphoma.

#### P-167

#### Longitudinal changes in sonographic salivary gland alterations in patients with Sjögren's syndrome

Benedikt Hofauer MDa, Naglaa Mansour MDa, Andreas Knopf MDa. Otorhinolaryngology / Head and Neck Surgery, Klinikum rechts der Isar, Technical University Munich, Ismaningerstr. 22, 81675 Munich, Germany.

**Background.** The role of sonography in the evaluation of salivary gland alterations in patients with suspected Sjögren's syndrome (SjS) and its benefit with regard to the diagnosis is part of continuing investigation. With most studies concentrating on the contribution to the diagnosis, only few reports exist on longitudinal changes in the sonomorphology of salivary glands in patients with SjS. Studies have shown that systemic and local therapeutic interventions are able to influence the results of different sonographic modalities. Despite intense research, many questions in ultrasound of salivary glands remain unanswered. The aim of the presented investigation is the examination of sonographic salivary gland alterations during the five-year follow up of patients with SjS.

Methods. Patients with SjS diagnosed according to the AECG classification criteria were included in this study. The ESSPRI was applied for the evaluation of patient's symptoms and the ESSDAI for systemic features. During 2011 and 2016 the sonoelastographic alterations of the salivary glands in patients with SjS were evaluated. Acoustic Radiation Force Impulse (ARFI) imaging (=shear wave velocity), Real Time Tissue Elastography (RTTE) and Virtual Touch Tissue Imaging (VTTI) were applied in addition to B-Mode sonography (BMUS). Results of BMUS, RTTE and VTTI were graded with appropriate scoring systems.

**Results.** Fifty patients diagnosed with SjS were included (45 female, age: 56 years). In 2011, the mean ESSPRI score was 8.3 (SD=4.6) and the mean ESSDAI score was 5.6 (SD=7.5). Initially, the sonographic evaluation of the parotid gland (PG) resulted in a mean score of 1.6 (SD=0.6) and in the submandibular gland (SMG) of 1.7 (SD=1.0), the mean ARFI value of PG was 2.99m/s (SD=0.93) and the mean ARFI value of the SMG was 2.15m/s (SD=0.57).

Clinical examination and sonoelastographic evaluation was repeated after five years in 2016, revealing a mean ESSPRI score of 6.1 (SD=3.2, p=0.002) and a mean ESSDAI score of 4.6 (SD=7.0, p<0.001). The mean sonographic score of the PG was 1.4 (SD=0.7, p<0.001) and of the SMG was 1.9 (SD=1.0, p=0.034). There was a decline in the sonographic score of the PG of 0.27 (SD=0.5) and increase of the SMG of 0.1 (SD 1.1) on average. After five years a significant decline of ARFI values could be observed in the PG (2.33m/s, SD=0.70, p<0.001) while no significant changes of the ARFI of the SMG could be observed. Results of RTTE and VTTI did not change significantly. The mean time interval between onset of first symptoms and first sonographic examination in 2011 was 57.3 months (SD=60.8).

**Conclusion.** The five-year sonoelastographic follow up of salivary gland alterations in patients with SjS revealed a decline in the severity of sono-graphic alterations of the parotid gland in BMUS and ARFI imaging, indicating a certain capability for modulation of salivary gland affection in SjS.

### P-168

# Salivary gland ultrasound in patients with Sjögren's syndrome – A comparison between bed-side, video and still-image scoring

Daniel S. Hammenfors<sup>1,2,3</sup>, Haris Causevic<sup>1,2,3</sup>, Johan G. Brun<sup>1,2</sup>, Roland Jonsson<sup>2,3</sup>, Malin V. Jonsson<sup>3,4</sup>.

<sup>1</sup>Department of Clinical Science – Section for Rheumatology, University of Bergen, Norway. <sup>2</sup>Department of Rheumatology, Haukeland University Hospital, University of Bergen, Norway. <sup>3</sup>Broegelmann Research Laboratory, Department of Clinical Science, University of Bergen, Norway. <sup>4</sup>Department of Clinical Dentistry – Section for Oral and Maxillofacial Radiology, University of Bergen, Norway.

**Background.** Major salivary gland ultrasound (SGUS) has been shown to increase sensitivity and specificity of classification criteria for primary Sjögren's syndrome. However, several different scoring systems and methods and lack of international standards for evaluation makes it difficult to integrate SGUS in daily clinical practice. In this study we aimed to compare bed-side, video and still-image scoring, to determine inter- and intra-observer variations.

**Methods.** One clinical investigator (HC) performed SGUS on consecutive patients previously diagnosed with pSS (n=32). Glands were examined using a GE Logic E9 with a linear transducer (6-15 MHz). All glands were examined in longitudinal and transversal planes, and scored bed-side (real-time). In the same session, representative videos and still-images for both longitudinal and transversal planes, from all four glands, were obtained and stored. Videos and images were stored and anonymized for blinded randomized evaluation by three experts in SGUS (HC,DSH, MVJ). In cases where two videos and/or images were available from the same gland and projection, the highest score was applied.

**Results.** Mean age of the patients was 56 years, and the female:male ratio 7:1. Schirmers I-test was positive in 16/31 patients, and 24/32 were anti-Ro/SSA positive. Focus score had been determined in 20 patients, and 15 patients had a focus score of 1 or more. The 2016

ACR-EULAR 2016 classification criteria was fulfilled by 24/32 patients.

Live-score correlated with all image and video scorings for HC, except for the left submandibular transversal video evaluation. The best correlation for the right parotid was the longitudinal video (.793), for the left parotid it was the transversal image (.786), and the longitudinal video (.753).

For the submandibular glands, the best correlation was the longitudinal videos; right gland (.699) and left gland (.621).

When comparing inter-observer correlation, Spearman correlation coefficient ranged from .370 to .934 (Table I), with the overall best results obtained for video of the right and left parotid longitudinal projection, and still-image of the right submandibular longitudinal projection.

#### 14th International Symposium on Sjögren's Syndrome

Table I. Inter-observes correlation (Spearman correlation).

	Video	Still-image	
Right parotid longitudinal	.807831838	-691742757	
Right parotid transverse	.764769780	.658765912	
Left parotid longitudinal	.760843864	.702727866	
Left parotid transverse	.724735795	.739824831	
Right submandibular longitudinal	.718780791	.736828934	
Right submandibular transverse	.670743831	.741767849	
Left submandibular longitudinal	.718743870	.573748811	
Left submandibular transverse	.603721739	.370642800	

**Conclusions.** Both intra- and inter-observer variations were good. In general, bed-side scoring correlated better with longitudinal video as compared to still-images. If possible, we suggest scoring to be performed bed-side, with a video as documentation and possibility to monitor disease progression.

#### P-169

#### Diagnostic and predictive evaluation using salivary gland ultrasonography in primary Sjögren's syndrome

Kyung-Ann Lee, Sang-Heon Lee, and Hae-Rim Kim.

Division of Rheumatology, Department of Internal Medicine, Konkuk University Medical Center, Konkuk University School of Medicine, Seoul, Korea.

**Background.** We aimed to assess the diagnostic accuracy of salivary gland ultrasonography (SGUS) as a single test for the detection of primary Sjögren's syndrome (pSS) and examine the prognostic factors for severe structural damage of the salivary glands based on SGUS score.

**Methods.** Patients with pSS (n=94) and idiopathic sicca syndrome (n=44) were evaluated using the SGUS 0-48 scoring system, which comprises five parameters: parenchymal echogenicity, homogeneity, hypoechoic areas, hyperechogenic reflections, and clearness of posterior borders. The salivary gland volume and intraglandular power Doppler signal (PDS) were also assessed. A multivariate linear regression analysis was performed to determine the factors associated with SGUS score.



Fig. 1. Representative images showing salivary gland ultrasonography in primary Sjögren's syndrome.

(A.D) Paroidi ultrasonographic grade of homogeneity. (A) Normal homogenous paroidi gland (Grade 0). (B) Mild inhomogeneous paroidi gland (Grade 1). (C) Evident inhomogeneous paroidi gland (Grade 2). (D) grossy inhomogeneous paroidi gland (Grade 3). (E-H) Submandibular ultrasonographic grade of homogeneity. (E) Grade 0. (F) Grade 1. (G) Grade 2. (H) Grade 3.



**Results.** Patients with pSS showed a significantly higher SGUS score than controls [median (IQR): 24.5 (13.0) vs 6 (3.75), p<0.001]. An SGUS cut-off of ≥14 had a sensitivity of 80.9% and a specificity of 95.5% for the diagnosis of pSS. There were no significant differences in the measured volumes and PDS between pSS patients and controls. The SGUS score correlated with unstimulated salivary flow rate (USFR), serum rheumatoid factor and IgG. Double seropositivity with anti-Ro/SS-A and anti-La/SS-B ( $\beta$ =6.060, p=0.001) and USFR ( $\beta$ =-1.913, p<0.001) were independently associated with the SGUS score.

**Conclusions.** The SGUS scoring system is a valuable diagnostic method for pSS. Double seropositivity of anti-Ro/SS-A and La/SS-B is an independent predictive factor for structural damage of the salivary glands.

### P170

#### Clinical comparison of salivary gland ultrasound to minor salivary gland biopsy in the diagnosis of Sjögren's syndrome

Lauren Meyerson Long, RN, BSN<sup>1</sup>, Margaret Beach, PA-C<sup>1</sup>, Blake Warner, DDS, PhD, MPH<sup>1</sup>, Ilias Alevizos, DMD, MMSc<sup>1</sup>.

<sup>1</sup>Salivary Dysfunction Unit, National Institute of Dental and Craniofacial Research, National Institutes of Health, USA.

**Introduction.** In recent years, there has been an increase in the use and perceived clinical utility of salivary gland ultrasonography (SG-US) in the evaluation of patients for Sjögren's syndrome (SS). SG-US core items and scoring methodology was established by the European League of Rheumatism (EULAR) in 2017. The panel identified 10 SG-US characteristics that demonstrated reliability when considering a diagnosis of SS. The minor salivary gland biopsy is a well-established component for the objective evidence to support a diagnosis of SS and is heavily weighted by the most recent iteration of the American College of Rheumatology/EULAR classification criteria. Many clinical settings do not routinely have capability to provide or perform salivary gland biopsies but may have access to US. Thus, it is important to determine the clinical significance of US findings for diagnostic purposes. The objective of this project was to determine the correlation of SG-US findings with surgical pathology scores.

**Methods.** Clinical data from fifty (n=50) consecutive previously undiagnosed subjects evaluated for salivary gland dysfunction at the NIDCR/Sjögren's Syndrome Clinic were reviewed. All subjects received a minor salivary gland biopsy and SG-US. Clinical findings were assessed for correlation between SG-US findings consistent with changes seen in SS and patients whose minor salivary gland biopsies demonstrated a focus score (FS) of at least 1 lymphocytic focus per 4mm<sup>2</sup> ( $1 \ge$  focus/4mm<sup>2</sup>) of salivary gland tissue. The data were divided into 3 subject groups: 1) those with normal SG-US findings and no foci, 2) those with a focus of at least 1 and normal or indeterminate SG-US findings, and 3) those with at least 1 focus and SG-US findings consistent with SS. Salivary function across these groups using whole unstimulated and glandular [*e.g.*, parotid and submandibular/sublingual] unstimulated and stimulated were compared using Kruskal-Wallis at a *p*-value of 0.05 as significant.

Results. Twenty-two of the 50 (44%) of the subjects satisfied diagnostic cri-

Posters

teria for a diagnosis of SS following routine work up, including both SG-US and minor salivary gland biopsy. Approximately 27% of the 22 subjects (6 of 22) had positive US and FS consistent with SS, while 50% (11 of 22) had FS consistent with SS, but normal or indeterminate US scores. Analyses of stimulated and unstimulated salivary gland function between FS and SG-US are being assessed.

**Conclusions.** Minor salivary gland biopsy remains an important tool in the diagnosis of SS. Here, we present a pilot analysis of our ongoing efforts on the implementation of SG-US in the diagnostic armamentarium of SS. Based on our findings, US imaging alone is not a reliable diagnostic predictor for SS, but can provide clinical information on salivary gland function. Although these results support the use of SG-US scoring as an adjunctive measure when considering a diagnosis of SS, it is possible that additional findings may be apparent when using a larger sample size, and correlations with other clinical characteristics might be established.

# P-171

# Ultrasound imaging of salivary glands correlates with labial gland biopsy scores in Sjögren 's syndrome

Shabnum Ali<sup>\*1</sup>, Jackie Brown<sup>2</sup>, Rose Ngu<sup>2</sup>, Troy Daniels<sup>3</sup>, John Greenspan<sup>3</sup>, Edward Odell<sup>4</sup>, Penelope J Shirlaw<sup>1</sup>, Kuldipsinh G Gohil<sup>1</sup>, Bruce Kirkham<sup>5</sup>, Genevieve Larkin<sup>6</sup>, Stephen J Challacombe<sup>1</sup>.

<sup>5</sup>Rheumatology, King's College London and Guy's & St Thomas' NHS Foundation Trust, UK. <sup>6</sup>Ophthalmology King's College Hospital. <sup>3</sup>Oral Pathology UCSF.

**Objectives.** Ultrasound imaging (US) is a non-invasive, reproducible method for assessing the major salivary glands and results can be quantified. The purpose of this study was to determine the relationship in Sjögren's Syndrome (SS) between US scoring of major glands and focal lymphocytic scores (FS) of minor salivary glands

Methods. Biopsies of labial-glands and Ultrasounds of major-glands were performed on 199 patients (mean age 55 years, 18 male, 181 female) attending the Sjögren's Clinic at Guy's Hospital. Patients were categorised into 3 groups: those diagnosed with SS according to the American-European classification-criteria, those with non-specific sialadenitis (sialadenitis-group), and those with normal biopsy results (control-group). FS of labial-glands were assessed by two calibrated histopathologists. Ultrasounds were carried out using a single Philips-iU22<sup>®</sup> Ultrasound, and a disease severity-score determined. **Results.** There was a highly significant positive correlation between FS (mean=1.8, SD=2.4) and US scores (mean=2.7, SD=2.9) (n=199 r= 0.708 p<0.0001) in the whole series. For the SS group (mean FS score= 3.6, SD=2.5; mean US score=5.1, SD=2.4) there was a significant positive correlation between FS and US scores (r=0.458 n=87 p<0.0001). For the Sialadenitis-group (mean FS score=0.4, SD=0.6; mean US score=1.1, SD=1.7) there was a significant correlation between FS and US scores (r=0.394 n=66 p<0.002). No obvious correlation between FS and US scores (r=0.026 n=46) existed for the control-group (mean FS=0.2, SD=0.5; mean US score=0.5, SD=1.3).

**Conclusion.** A highly significant positive correlation was found between ultrasound scores of major salivary glands and focus scores of minor salivary (labial) glands.

**Relevance.** This is the first large-scale study to analyse the relationship between US scores of major salivary glands, and Focus Scores of minor salivary glands. Ultrasound analysis should prove to be an extremely important investigation in both cross-sectional and longitudinal studies of Sjögren's syndrome.

# P-172

#### Ultrasonography of salivary glands in primary Sjögren's syndrome and association with symptoms of dryness, disease activity and biopsy of minor salivary glands

Milic V<sup>1</sup>, Radunovic G<sup>1</sup>, Damjanov N<sup>1</sup>.

<sup>1</sup>Institute of Rheumatology, School of Medicine, University of Belgrade, Serbia.

**Objective.** To analysis ultrasonography (US) changes of salivary glands (SG) in patients with primary Sjögren's syndrome (pSS) and association US findings with symptoms of dryness, disease activity and biopsy of minor salivary glands (MSG).

Methods. This study included 205 pSS patients (mean age 53.9±11.5, disease duration 5.6 years) and 87 healthy controls (mean age 52.3±14.7). All pSS patients fulfilled the AECG diagnostic criteria. The disease activity was evaluated by EULAR SS disease activity index (ESSDAI), Sjögren's Syndrome Disease Damage Index (SSDDI) and EULAR Sjögren's syndrome patient reported index (ESSPRI). Parotid and submandibular glands on both sides were assessed by US for size, parenchymal echogenicity and inhomogeneity, posterior glandular border and presence of intraglandular lymph nodes. Inhomogeneity of the SG were graded according to the De Vita scoring system. The global SGUS score (0-6) was the sum of the scores of each pair of SG. Statistical analysis was performed by SPSS v19. Data were compared using *t*-test,  $\chi^2$  test and Mann-Whitney U test. The association US of SG adjusted for potential confounders as independent variables was examined using univariate linear regression analyses. All statistical analyses were performed using SPSS, Version 19.0.

Results. Xerophtalmia and xerostomia were presented in 185/205 (90.2%) and 186/205 (91.2%), respectively. According to ESSDAI, the majority of pSS patients 88/205 (43%) had moderate disease activity. Seventy-eight per cent of pSS patients were anti-SSA positive, 44% anti-SSB positive. Biopsy of MSG was positive in 140/172 (81.4%) pSS patients. US abnormalities were established in 197 (96%) pSS patients and in 16 (18%) controls (p<0.0001). Pathological sizes of SG were more frequently in pSS patients than controls, 111 (54.2%) vs. 3 (3.4%) patients, respectively (p < 0.0001). The echogenicity of the SG was pathological in 142 (69.3%) pSS patients and in only 5 (5.7%) control group (p<0.0001). The pathological glandular border was frequently in pSS patients than control group, 48 (23.4%) vs. 2 (2.3%), (p<0.0001). No differences were detected between the two groups of patients for enlarged intraglandular lymph nodes. Most of pSS patients had pathological inhomogeneity, 197/205 (96.1%) vs. 16/85 (18.4%) in control group (p<0.0001). The median SGUS was significantly higher in pSS patients in comparison with control group [median (range) 4(0-6) vs. 0(0-2), p<0.0001]. After adjustment for potential confounders variables, dry mouth ( $\beta$ =1.542; p=0.04), ESSDAI  $(\beta=1.203; p=0.008)$ , SSDDI  $(\beta=4.768; p=0.02)$  and biopsy of MSG  $(\beta=2.735;$ p=0.05) were significantly associated with advanced US changes of SG. Dry eyes and ESSPRI were not associated with US (p>0.05).

Conclusions. Our findings confirm that most of established pSS patients had pathological US features. Degree of xerostomia, objective disease activity and damage and biopsy of MSG had predictive value for advanced US change of salivary glands.

## P-173

#### Ultrasound of salivary gland in primary Sjögren syndrome with clinical relation in Taiwan

Yen-Fu Chen<sup>1</sup>, Yao-Fan Fang<sup>1</sup>, Y.F. Chen, M.D<sup>2</sup>.

<sup>1</sup>Division of Rheumatology, Allergy and Immunology, Department of Internal Medicine, Chang Gung Memorial Hospital, Tao-Yuan County, Taiwan. <sup>2</sup>Division of Rheu matology, Allergy, Chang Gung Memorial Hospital Tao-Yuan County, Taiwan

Objective. To investigate the ultrasound of salivary gland in Primary Sjögren syndrome and its clinical relation in Taiwanese patients.

Methods. Prospective cohort study of newly diagnosis of Primary Sjögren syndrome since 2017 around 28 patients in Chang Gung Memorial Hospital and follow up by every 3 months.

Results: The mean ages were 50.5 years respectively. Sicca symptoms mean duration were 1.3 years. ESSPRI (EULAR Sjögren syndrome patient report index) were 14.6. ESSDAI (EULAR Sjogren syndrome disease index) were 4.46. Of the 28 patients, one patient later found a MALT lymphoma due to our early arrangement of ultrasound of salivary gland with further biopsy. Ultrasound of the salivary gland score mostly showed grade 1 or grade 2.



**Image 1.** Ultrasound images of right parotid gland illustrating varying grades of pathological changes. Grades 0–3 were used when evaluating US examinations of each patients salivary gland. Below is grade 2.

Table I. 28 Primary Sjögren syndrome patient.

Male	3.5% (1/28)		
Age (yrs)	50.5 yrs		
Sicca duration (yrs)	1.3 yrs		
Anti-SSA	291.5 (AU/ml)	<100	
Anti-SSB	178.4 (AU/ml)	<100	
RF-IgM	25.6 (IU/mL	<6	
IgG	1735.6 (mg/dL)	700~1600	
IgG4	65.6 (mg/dL)	3~201	
C3	117.5	90~180	
C4	65.6	10~40	
ESSPRI	14.6	0~30	
ESSDAI	4.4	0~123	
Ultrasound score	Grade 1 or Grade 2		

Conclusions: This study underlines the ultrasound of salivary gland maybe a diagnostic tool of Sjögren's syndrome, but not relation with clinical severity of Primary Sjögren syndrome with ESSDAI and ESSPRI. Reference

1. HAMMENFORS DS, BRUN JG, JONSSON R, JONSSON MV: Clin Exp Rheumatol 2015; 33: 56-62

# P-174

#### Effectiveness and safety ov vellow fever vaccine in patients with primary Sjögren's syndrome

V. Valim<sup>1</sup>, S.A. Gouvea<sup>1</sup>, S.M.B. Lima<sup>2</sup>, A.C.C. Azevedo<sup>3</sup>, A.T. Carvalho<sup>3</sup>, V.P.M. Pascoal<sup>4</sup>, S.T. Miyamoto<sup>1</sup>, A.L.S. Souza<sup>1</sup>, P.C.M. Rocha<sup>1</sup>, L. Balarini<sup>1</sup>, E.V. Serrano<sup>1</sup>, R.H. Duque<sup>1</sup>, M.B.R.O. Gavi<sup>1</sup>, V.G. Dinis<sup>1</sup>, L.C. Caser<sup>1</sup>, L.D. Moura<sup>1</sup>, A.D. Pinto<sup>1</sup>, A.P.N.B. Lima<sup>1</sup>, L.F.S. Pinto-Neto<sup>5</sup>, E.T.L. Polito<sup>1</sup>, T.B. Clemente<sup>1</sup>, E.S. Magalhaes<sup>1</sup>, M.M. Thebit<sup>1</sup>, M.F. Bissoli<sup>1</sup>, O.A. Martins-Filho<sup>4</sup>. <sup>1</sup>Universidade Federal do Espírito Santo (UFES), Vitória, ES, Brasil. <sup>2</sup>Instituto de Tecnologia em Imunobiológicos (BIOMANGUINHOS), Fundação Oswaldo Cruz (FI-OCRUZ), Rio de Janeiro, RJ, Brasil. <sup>3</sup>Fundação Oswaldo Cruz (FIOCRUZ), Rio de Janeiro, RJ, Brasil. 4Centro de Pesquisas René Rachou, Fundação Oswaldo Cruz (FI-OCRUZ), Belo Horizonte, MG, Brasil. <sup>5</sup>Escola de Medicina Santa Casa Misericórdia (EMESCAM), Vitória, ES, Brasil.

Background. Yellow fever is a viral haemorrhagic fever transmitted by mosquitoes. The live attenuated vaccine is highly immunogenic and offers prolonged and generally safe protection in immunocompetent patients. The aim of this study is to evaluate the immunogenicity and safety of the vaccine in patients with primary Sjögren syndrome (PSS) in the short and long term.

Methods. This is a phase IV controlled prospective study, including 50 pa tients with PSS and 29 healthy controls. All of them were vaccinated for the first time by the 17DD YF vaccine (Biomanguinhos, Brasil) after individual clinical evaluation. Patients who presented CD4 <200 cells/mm3, neoplasia, HIV, primary immunodeficiency, using cyclophosphamide, prednisone >20mg/d, azathioprine >2mg/kg/d, chlorambucil, mycophenolate or biological therapy were excluded. Patients were evaluated at baseline, after 3, 4, 5, 6, 7, 14 and 30 days of the vaccination for viremia and humoral response. Plaque reduction neutralization test (PRNT) was measured at baseline and after 28 days. Disease activity was evaluated at baseline and after 6 months. Serum and cell samples were frozen at -70°C. The analyses of viremia and PRNT will be processed by Instituto Fiocruz-BH and Fundação Biomanguinhos-RJ. Results. The mean age was 53 (±15.5) years, ESSDAI 1.2 (±2.4), ESS-PRI 3.6 (±2.9), PCR 3.3 (±4.2), C3 174.2 (±266.2), C4 26.3 (±10.7) and IgG 1555.8 (±769.5). Most patients showed disease under control before and after 6 months (p=0.556). Forty four percent of the patients were using hydroxychloroquine, 18% methotrexate, 4% leflunomide, 2.3%, sulfasalazine and 2% corticosteroid. Nine patients had adverse events from the 21 who were using only one medication and all 4 patients who were using 2 medications had an adverse event, and the patient who was using 3 medications did not have any adverse event (p=0.066). There was no difference between patients and controls in the occurrence of adverse events (46% vs 27.6%, p=0.106). The main local symptoms were: pain (16%), nodule (2%), oedema (4%) and heat (2%). The main systemic adverse events were: malaise (24%), myalgia (20%), headache (12%), arthralgia (12%), low back pain (10%), weakness in limbs (10%), productive cough (10%), dry cough (8%), pruritus (8%), abdominal pain (6%), nausea (6%), vomit (6%), fever (4%), dyspnoea (4%) e diarrhoea (2%).

Conclusion. The yellow fever vaccine is safe in patients with PSS with low disease activity and under low immunossupression. It is necessary to await the studies of vaccine kinetics for further conclusions.

# P-175

# Activation of RANKL system in Sjögren's syndrome related lymphoma

Eleni Palli, Adrianos Nezos, Clio P. Mavragani.

Department of Physiology, School of Medicine, National and Kapodistrian University of Athens, Athens, Greece.

Introduction/Objective. Sjögren's syndrome (SS) is an autoimmune exocrine disease characterized by chronic dysfunction and destruction of the salivary and lacrimal glands, resulting in mucosal dryness. Approximately 5-10% of patients are identified as a high-risk group for the development of lymphoproliferative disease, most commonly B-non Hodgkin's lymphoma. Previous studies support the activation of the receptor activator of nuclear factor kappa-B ligand (RANKL) pathway, as an important contributor in both systemic autoimmune diseases and haematological malignancies. The purpose of this study was to investigate the role of RANKL pathway in SSrelated lymphoma.

Methods. RANKL and osteoprotegerin (OPG) titers were determined in sera derived from 66 SS patients and 12 sicca controls (SC) by ELISA. In addition, mRNA expression levels of RANKL and OPG were determined in cDNA isolated from minor salivary gland tissues of 34 SS patients, 19 patients with SS complicated with lymphoma and 11 SC using RT-PCR. In order to identify potential associations of the components of the RANKL/RANK/OPG system with disease related characteristics, demographics, clinical and serological features of SS patients were recorded after thorough chart review. For statistical analysis SPSS 24.0 software has been implemented.

**Results.** SS patients displayed increased RANKL and OPG serum titers, as well as salivary gland tissue RANKL mRNA levels compared to SC. At salivary gland tissue level, OPG mRNA levels were decreased in SS patients associated with lymphoma compared to both SS and SC groups. Patients with higher serum OPG levels displayed significantly higher erythrocyte sedimentation rate (ESR) values, and a trend toward increased rheumatoid factor titers and Tarpley scores in minor salivary gland biopsies.

**Conclusion.** Decreased OPG and increased RANKL mRNA expression levels in salivary gland tissues derived from SS patients complicated by lymphoma compared to controls might imply an important role of deregulated RANKL pathway in SS related lymphomagenesis.

### P-176

# Interleukin-22 and associated genes are targets of microRNAs dysregulated in primary Sjögren's syndrome associated lymphoma

Jessica R Tarn<sup>1</sup>, Gunnel Nordmark<sup>2</sup>, Colin Gillespie<sup>3</sup>, Shereen Al-Ali<sup>1,9</sup>, Katherine James<sup>6,7</sup> Sheryl Mitchell<sup>4</sup>, Simon Bowman<sup>5</sup>, Bridget Griffiths<sup>4</sup>, UK primary Sjögren's syndrome registry<sup>1</sup>, Svein Joar Auglaend Johnsen<sup>8</sup>, Katrine Norheim<sup>8</sup>, Carin Backlin<sup>2</sup>, Lars Rönnblom<sup>2</sup>, Maija-Leena Eloranta<sup>2</sup>, Eva Baecklund for the Autolymphoma Study Group <sup>2</sup>, Roald Omdal<sup>8</sup>, David Alan Young<sup>1</sup> and Wan-Fai Ng<sup>1,4,10</sup>.

<sup>1</sup>Institute of Čellular Medicine, Newcastle University, Newcastle upon Tyne, UK. <sup>2</sup>Department of Medical Sciences, Rheumatology and Science for Life Laboratory, Uppsala University, Uppsala, Sweden. <sup>3</sup>School of Mathematics and Statistics, Newcastle University, Newcastle upon Tyne, UK. <sup>4</sup>Newcastle upon Tyne Hospitals NHS Foundation Trust, Newcastle upon Tyne, UK. <sup>5</sup>University Hospital Birmingham, Birmingham, UK. <sup>6</sup>Institute for Cell and Molecular Biosciences, Newcastle University, Newcastle upon Tyne, UK. <sup>8</sup>Stavanger University Hospital, Stavanger, Norway. <sup>9</sup>Department of biology, College of Science, University of Basrah, Basrah, Iraq. <sup>10</sup>NIHR Newcastle Biomedical Research Centre, Newcastle University, Newcastle upon Tyne, UK.

**Background.** The role of microRNAs (miRNAs) in primary Sjögren's syndrome (PSS) and PSS-related lymphoma remains unclear despite the association of miRNAs with several rheumatologic disorders and with malignancies.

**Methods.** We profiled miRNA expression in blood samples of PSS patients with untreated and treated lymphoma from 3 independent and international cohorts with 49 lymphoma and 58 non-lymphoma patients in total. Analysis of putative targets of the differentially expressed miRNAs was performed *in silico*. Target gene-miRNA interactions were assessed by luciferase reporter assay. **Results.** We identified 18 differentially expressed miRNAs (DEmiRs) between PSS patients who had lymphoma and those without lymphoma that

were common between the 3 independent cohorts. These 18 DEmiRs (including miR-15, miR-17, miR-21, miR-103, miR-130, miR-374, miR-454 and miR-532) belong to 8 different miRNA families. The miRNAs from 6 of these miRNA families are predicted *in silico* to target Th17, Th22 and  $T_{\rm FH}$  cell associated genes. We demonstrated by luciferase assays that 6 of the predicted targets of the DEmiRs (*IL22, STAT3, AHR, CXCR5, ICOS* and *RORC*) are likely to have direct interactions with the DEmiRs.

**Conclusion.** We detected 18 differentially expressed miRNAs between PSS patients with and without current or historical lymphoma. Based on *in vitro* reporter assays these miRNAs are likely to regulate 6 genes encoding homing receptors for T-cells, transcription factors and inflammatory cytokines. These miRNA-gene interactions may reinforce the cycle of chronic inflammation, B-cell development and lymphoid neogenesis. Down-regulation of these miRNAs could result in dis-inhibition of the target genes, which in turn promote tertiary lymphoid organ development and subsequent lymphomagenesis (Figure. 1). Further *in vivo* work is required to confirm these miRNA-gene interactions within the context of lymphomagenesis.



Fig. 1. Hypothesis of mIRNA involvement in tertiary lymphold organ formation in the context of PSS associated lymphoma.

#### **P-177**

#### TREX1 variants in Sjögren's syndrome related lymphomagenesis

Adrianos Nezos<sup>1</sup>, Panagiota Makri<sup>2</sup>, Mary K. Crow<sup>3</sup>, Clio P Mavragani <sup>1.24</sup>. <sup>1</sup>Department of Physiology, School of Medicine, National University of Athens, Athens, Greece. <sup>2</sup>Department of Pathophysiology, School of Medicine, National University of Athens, Athens, Greece. <sup>3</sup>Mary Kirkland Center for Lupus Research, Hospital for Special Surgery, Weill Medical College of Cornell University, New York, NY, USA. <sup>4</sup>Joint Academic Rheumatology Program, National and Kapodistrian University of Athens, School of Medicine, Athens, Greece.

We aimed to explore whether Three-prime Repair Exonuclease 1 (TREX1) genetic variants could influence the risk of primary Sjogren's syndrome (SS) and SS-related lymphoma. Three TREX1 variants (rs11797, rs3135941 and rs3135945) were evaluated in 229 SS, 89 SS-lymphoma (70 SS-MALT and 19 SS non-MALT) and 240 healthy controls (HC) by PCR-based assays. In available 52 peripheral blood and 26 minor salivary gland (MSG) SS samples, mRNA expression of type I interferon (IFN) related genes and TREX1 was determined by real-time PCR. Significantly decreased prevalence of rs11797 A minor allele was detected in SS patients complicated by non-MALT lymphoma compared to HC (OR [95% CI]: 0.4 [0.2-0.9], p-value: 0.02). SS patients carrying the rs11797 AA genotype had increased prevalence of anti-Ro/SSA autoantibodies and type I IFN related gene mRNA expression in MSG tissues. These data support genetically related dampened type I IFN production as an additional mechanism for SS-related lymphomagenesis.
#### Posters

#### 14th International Symposium on Sjögren's Syndrome

P-178

Thymic stromal lymphopoietin expression and function in lymphoproliferation and lymphoma in primary Sjögren's syndrome

Saviana Gandolfo<sup>1</sup>, Cinzia Fabro<sup>1</sup>, Michela Bulfoni<sup>2</sup>, Sabino Russi<sup>3</sup>, Luca Quartuccio<sup>1</sup>, Domenico Sansonno<sup>3</sup>, Carla Di Loreto<sup>2</sup>, Daniela Cesselli<sup>2</sup>, Salvatore De Vita<sup>1</sup>.

<sup>1</sup>Rheumatology Clinic, University of Udine, Department of Medical Area DAME, Udine, Italy. <sup>2</sup>Institute of Anatomic Pathology, University of Udine, Department of Medical Area DAME, Udine, Italy. <sup>3</sup>Department of Biomedical Sciences and Human Oncology, University of Bari, Italy.

**Background.** Thymic stromal lymphopoietin (TSLP) is an epithelial lymphopoietic cytokine mainly expressed at interfaces between the body and the environment, showing immunoregulatory properties and acting as a B cell growth factor. Two isoforms of TSLP have been described, showing opposite biological actions. The short form (sfTSLP) is constitutively expressed at body barriers, where it regulates the immune tolerance. Conversely, the expression of the long TSLP isoform (lfTSLP) is inducible and upregulated in several inflammatory diseases.

Recent studies highlighted the contribution of TSLP to the immunopathology of human autoimmune and systemic inflammatory disorders and of both solid and hematological malignancies.

**Objective.** to investigate TSLP expression and function in primary Sjögren's syndrome (pSS) patients, stratified according to the lymphoproliferative status, from fully benign (fbSS) to myoepithelial sialadenitis (MESA) and to B-cell non-Hodgkin lymphoma (NHL).

**Methods.** based on our preliminary TSLP serological data in 79 pSS patients and 100 controls (21 non-autoimmune sicca-syndrome, nSS; 79 sex and age-matched healthy blood donors, HBDs), TSLP expression was also studied in salivary glands (SG) biopsies from 38 pSS patients (13 fbSS; 13 MESA; 12 NHL) and from 13 nSS by RT-PCR, immunohistochemistry and immunofluorescence. TSLP *in vitro* functional effects were also evaluated on peripheral B lymphocytes collected from 5 patients in each pSS subgroup, and from 5 HBDs.

**Results.** serum studies showed significantly higher TSLP levels in pSS compared to controls, significantly increasing from fbSS to MESA and to NHL. In SG biopsies, TSLP-positive B lymphocytes increased with the progression of lymphoproliferation, maximally in NHL, consistently with the detection of inducible lfTSLP mRNA only in MESA and NHL. Constitutive sfTSLP mRNA levels also increased in pSS compared to controls, but with no statistical difference among pSS subgroups.

TSLP promoted a significant B-cell activation and immunoglobulin production by peripheral B lymphocytes in pSS-related MESA and NHL.

**Conclusions.** a pathogenetic role of TSLP is herein suggested in pSS for the first time. TSLP, which promotes B-cell expansion, progressively increases from benign to malignant B-cell lymphoproliferation in pSS. With the progression of lymphoproliferation, the B-cells, rather than the salivary epithelium, appear as the major TSLP source, in its long isoform. Further studies and the analysis of TSLP as a biomarker in pSS are worthwhile.

# P179

# CXCL13 serum levels are associated with lymphoma risk in primary Sjögren's syndrome

Traianos EY<sup>1</sup>,\*, Locke J<sup>1</sup>,\*, Tarn JR<sup>1</sup>, Bowman S<sup>2</sup>, UK primary Sjögren's syndrome registry, Lendrem D<sup>1</sup> and Ng WF<sup>1,3</sup>. <sup>1</sup>Institute of Cellular Medicine, Newcastle University, UK. <sup>2</sup>Univesity Hospital Bir-

<sup>1</sup>Institute of Cellular Medicine, Newcastle University, UK. <sup>2</sup>Univesity Hospital Birmingham, UK. <sup>3</sup>NIHR Newcastle Biomedical Research Centre. \*Denote joint first authorship.

**Background.** Primary Sjögren's syndrome (pSS) is an autoimmune disease characterized by an increased risk for non-Hodgkin lymphoma (NHL) development. CXCL13 is a chemokine known to be associated with the homing of B cells within ectopic germinal centres (GC) and with reactive lymphoid proliferation in pSS. Considering that the presence of GC is a histological risk factor in pSS associated lymphoma, in this study we examined the role of CXCL13 in predicting NHL.

Methods. Serum CXCL13 levels were measured using the enzyme-linked immunosorbent assay (ELISA) in 114 female pSS patients included in the United Kingdom Primary Sjögren's Syndrome Registry (UKPSSR) cohort,

and 30 controls. Sixteen patients had lymphoma and the rest were classified into high-, moderate-, and low-risk groups based on the lymphoma risk score proposed by Fragkioudaki *et al.* (1). CXCL13 levels were compared between pSS patients and controls and between pSS patients with lymphoma and those without using Mann-Whitney U test. Multivariate regression analyses were performed to determine the relationships between CXCL13 and the Fragkioudaki's lymphoma risk score, disease duration, the EULAR Sjögren's syndrome (SS) disease activity index (ESSDAI) score, C4 levels, anti-Ro/SSA antibodies, and B-cell biomarker.

**Results.** CXCL13 serum levels were found to be associated with pSS as compared to controls (p<0.0001), but no difference was found in CXCL13 serum levels between patients with lymphoma and those without (p=0.4268). The predictors for Fragkioudaki's lymphoma risk score were combined light chain (CMBY), CXCL13 and B-cell activating factor (BAFF) (p=0.0295, p=0.0327, and p=0.0346 respectively) on multivariate analysis. High values of CMBY, CXCL13 and BAFF were positively corelated with the risk score. There was no significant relationship between CXCL13 levels and ESSDAI score, disease duration, C4 levels, anti-Ro/SSA antibodies, beta-2 microglobulin and kappa to lambda ratio.

**Conclusion.** Serum CXCL13 level was an independent determinant of Fragkioudaki's lymphoma risk score. However, CXCL13 levels were not associated with lymphoma occurrence although the sample size of the lymphoma group was relatively small. A larger replication study is ongoing. Our findings may indicate the role of CXCL13 in supporting antigen-activated polyclonal B-cell expansion in GC which predispose to lymphoma development, but may not play a key role in the proliferation or survival of malignant B cells.

#### References

 FRAGKIOUDAKIS, MAVRAGANIC, MOUTSOPOULOS H: Predicting the risk for lymphoma development in Sjögren syndrome. *Medicine* 2016; 95(25): e3766.







Fig. 2. CXCL13 serum levels in association with low-, moderate- and high- pSS associated lymphoma risk groups. \*\*\*: p<0.0001.

# 14th International Symposium on Sjögren's Syndrome

# **P-180**

Single cell RNA sequencing of B cell subsets from parotid glands of patients with Sjögren's syndrome identifies the expression profile of epithelium-associated FcRL4<sup>+</sup> B cells

Gwenny M. Verstappen<sup>1</sup>, John A. Ice<sup>2</sup>, Hendrika Bootsma<sup>1</sup>, Sarah A. Pringle<sup>1</sup>, Erlin A. Haacke<sup>1,3</sup>, Frederik K.L. Spijkervet<sup>4</sup>, Christopher J. Lessard<sup>2,5</sup>, Frans G.M. Kroese<sup>1</sup>.

<sup>1</sup>Department of Rheumatology and Clinical Immunology, University of Groningen, University Medical Center Groningen, The Netherlands. <sup>2</sup>Arthritis and Clinical Immunology Research Program, Oklahoma Medical Research Foundation, Oklahoma City, OK, USA. <sup>3</sup>Department of Pathology and Medical Biology, University of Groningen, University Medical Center Groningen, The Netherlands. <sup>4</sup>Department of Oral and Maxillofacial Surgery, University of Groningen, University Medical Center Groningen, The Netherlands. <sup>3</sup>Department of Pathology, University of Oklahoma Health Sciences Center, Oklahoma City, OK, USA.

**Background.** A subset of B cells expressing the inhibitory Fc receptor-like protein 4 (FcRL4) is found in salivary gland lesions of patients with primary Sjögren's syndrome (pSS). FcRL4+ B cells are associated with ductal epithelial cells forming lymphoepithelial lesions (LEL), in particular within parotid glands. Furthermore, FcRL4 is expressed by mucosa-associated lymphoid tissue (MALT) lymphoma B cells. We aimed to investigate, by single cell and bulk RNA sequencing, how the transcriptome of FcRL4+ B cells differs from FcRL4-negative naïve and memory B cells in salivary gland tissue of pSS patients. We hypothesize that FcRL4+ B cells contribute to LEL formation and are prone to lymphomagenesis.

**Methods.** Parotid gland biopsies of 5 pSS patients without MALT lymphoma and 1 pSS patient with MALT lymphoma were obtained. Single cell suspensions were prepared by mechanical disruption and enzymatic digestion. The cells were incubated with anti-CD19, anti-CD27 and anti-FcRL4 antibodies and sorted into single cell and 'bulk' (5 cells per well) based on the following definitions: CD19+CD27-FcRL4- ('naïve'), CD19+CD27+FcRL4- (memory) and CD19+FcRL4+ (FcRL4+). Library preparation was done using an in-house SMARTseq2 protocol and sequencing was done on an Illumina HiSeq2500. Expression data were analyzed using Seurat or DESeq packages in R.

**Results.** Samples from 4 pSS patients and the MÅLT lymphoma patient passed quality control and were included. A total of 206 single cells and 450 cells in bulk were included in the analysis. Genes identified by differential expression were subjected to gene pathway analysis. Both in single cell and bulk analysis, multiple genes coding for integrins, such as *ITGAX* (CD11c) were significantly upregulated in FcRL4+ B cells. Gene Ontology pathways that showed the highest upregulation in FcRL4+ B cells (both single cell and bulk) were receptor binding, GTPase and protein kinase pathways. Analysis of bulk samples further revealed that expression levels of *CXCR3*, *NFKB1*, *JAK2* and *LYN*, among others, were significantly upregulated in FcRL4+ B cells, compared with either naïve or memory B cells. *LCK* was, for example, downregulated. Interestingly, preliminary data from comparison of single CD27-negative 'naïve' B cells of the MALT-lymphoma patient with non-lymphoma patients showed upregulation of *MX1*, *OAS1*, *ISG15*, *BTK* and *TNF*, among others.

**Conclusions.** FcRL4+ B cells in salivary glands of pSS patients show upregulation of genes involved in homing and cell adhesion, consistent with their tissue location close to the epithelium. FcRL4+ B cells also show increased upregulation of genes that promote inflammation. These cells exhibit all characteristics of chronically stimulated CD11c+ memory B cells, and we speculate that these cells contribute significantly to the epithelial damage seen in the glandular tissue of pSS patients.

# P-181

#### Numbers of B-lymphocytes increase when formation of lymphoepithelial lesions progresses in salivary glands of primary Sjögren's syndrome patients

Martha S. van Ginkel<sup>1</sup>, Erlin A. Haacke<sup>1,2</sup>, Hendrika Bootsma<sup>1</sup>, Suzanne Arends<sup>1</sup>, Jolien F. van Nimwegen<sup>1</sup>, Gwenny M. Verstappen<sup>1</sup>, Fred K.L. Spijkervet<sup>3</sup>, Arjan Vissink<sup>3</sup>, Bert van der Vegt<sup>2</sup>, Frans G.M. Kroese<sup>1</sup>. Departments of <sup>1</sup>Rheumatology and Clinical Immunology, <sup>2</sup>Pathology and Medical Biology, and <sup>3</sup>Oral and Maxillofacial Surgery, University of Groningen and University Medical Center Groningen, Groningen, the Netherlands.

Background. Primary Sjögren's syndrome (pSS) is a common chronic systemic autoimmune disease characterized by dry mouth and dry eyes. The major histopathological finding in the salivary glands of pSS patients are periductal lymphocytic infiltrates (foci). In association with these infiltrates, lymphoepithelial lesions (LELs) may be formed. LELs are defined by infiltration of lymphocytes within the epithelium with hyperplasia of ductal cells. Although it has been suggested that B-lymphocytes predominate in LELs, the relative role of B- and T- lymphocytes in LEL formation is not known. Since LELs are a characteristic finding in salivary glands of pSS patients, quantification of B- and T-lymphocytes in the different stages of LEL development will give more insight in their formation as well as in the pathogenesis of pSS. Methods. The study population consisted of 15 pSS patients, who fulfilled the ACR-EULAR criteria and underwent salivary gland biopsies of both the parotid and labial gland. Patients were not treated with biologicals and 13 patients were not using other immunosuppressive therapy. Focus score, and presence and severity of LELs were evaluated on HE stained sections. Severity of lesions was scored from stage 0 to stage 3 (stage 0: lymphocytic ductal infiltration without hyperplasia of the epithelium; stage 1: lymphocytic ductal infiltration and <50% of hyperplastic epithelium; stage 2: lymphocytic ductal infiltration and  $\geq 50\%$  of hyperplastic epithelium; stage 3: lymphocytic ductal infiltration and fully circumferentially hyperplastic epithelium with occluded lumen). Numbers of B- and T-lymphocytes within LELs were counted (10 ducts per biopsy) by using Image J cell counter on serial CD20 and CD3 stained sections. High molecular weight cytokeratin staining was used to identify ductal borders. Generalized estimating equations (GEE) were used to analyse the numbers of B- and T-lymphocytes and B/T ratio's over the different stages of severity of LELs.

Results. B- and T-lymphocytes can both infiltrate within the ductal epithelium forming LELs in the salivary glands of pSS patients. T-lymphocytes were present in all LELs, scattered through the whole ductal epithelium. Whereas B-lymphocytes were found in clusters, mostly located in the hyperplastic area of the ductal epithelium. With higher severity of LELs, the numbers of B- and T-lymphocytes increased significantly, in both the parotid and labial gland. The numbers of B-lymphocytes increased relatively more with higher severity of LELs than T- lymphocytes. This has led to an increased intraepithelial B/T ratio in more pronounced LELs. This increased B/T ratio was even more pronounced in parotid than in labial glands. In both glands, there was a predominance of T-lymphocytes in lower LEL stages. In more severe LEL stages, B-lymphocytes outnumbered the T-lymphocytes. Conclusion. Given the relative increase in the number of B-lymphocytes with higher severity of LELs as well as their close association with proliferating ductal epithelial cells, we conclude that B-lymphocytes play a major role in the hyperplasia of the ductal epithelium.

#### **P-182**

# Usefulness of 18F-FDG positron emission tomography (PET) for lymphoma diagnosis in patients with primary Sjögren's syndrome

Jeremy Keraen, Estelle Blanc, Florent Besson, Veronique Leguern, Magalie Meyer, Julien Henry, Rakiba Belkhir, Gaétane Nocturne, Xavier Mariette, Raphaèle Seror.

Rheumatology and nuclear medicine departments, Hopital Bicetre, Université Paris-Sud, Le Kremlin-Bicêtre, France. Nuclear medicine department, Centre Chirurgical Marie Lannelongue, Université Paris-Sud, Le Plessis-Robinson, France. Internal medicine department, Hospital Cochin, Paris Descartes University, Paris, France.

**Background.** Primary Sjögren's syndrome (pSS) is the autoimmune disease having the highest risk of lymphoma. The differential diagnosis between benign and malignant lymphoproliferation is sometimes difficult. Among imaging procedures, 18F-FDG PET could be useful for that purpose. The objectives were to compare 18F-FDG PET results between patients with and without lymphoma to identify PET pattern associated with lymphomas in pSS.

**Methods.** Retrospective study conducted in 2 centers including pSS patients (ACR/EULAR 2016 criteria) who undergo 18F-FDG PET. PET abnormalities were compared between patients with and without lymphoma. Two independent readers analyzed PET blind to lymphoma diagnosis. ESS-DAI-PET score previously described by Cohen et al. was calculated.

**Results.** 45 patients were included; 15 had lymphoma: MALT (n=12), nodal marginal zone with plasmacytic differentiation (n=2), diffuse large B-cell (n=1). Patients with lymphoma had more frequently parotid gland swelling (67% vs 20 %, p=0.003) and higher ESSDAI score (24 [13.5-29] vs 9 [5-20], p=0.003), even after exclusion of lymphoma item (19 [11-27] vs 9 [5-20], p=0.003). Compared to non-lymphoma patients, mean size (45.5 [38-56] mm vs. 40 [37-41] mm; p=0.048) and maximum standardized uptake value

#### Posters

(SUVmax) of the parotid glands (5.6 [5-6.9] vs 3.8 [3.2-4.4]; p=0.001) were higher in lymphoma patients. 53.3% of patients with lymphoma and 43.3% without lymphoma had lymph node FDG uptake, but neither their number nor their repartition or mean SUV differ between them.

Pulmonary uptake was observed in 6 (40%) patients with lymphoma and 6 (20%) without lymphoma (p=0.17). But in lymphoma patients, this uptake was focal in 5 (33.3%) patients (nodules or condensation) and in only one (3.3%) patient without lymphoma (p=0.01). Remaining patients had interstitial FDG uptake. Mean ESSDAI-PET score (4 [2-4] vs. 2 [1-3] p=0.04) and SUVmax at any site (6.3 [5.6-7.3] vs. 4.2 [3.7-5.9] p=0.02) were significantly higher in lymphoma group. A SUVmax of parotid gland >4.6 was highly suggestive of lymphoma with a sensitivity of 73% and a specificity of 87%. At any site, a SUVmax ≥6 had a sensitivity of 66.7% and a specificity of 76.7% for the diagnosis of lymphoma. 20 patients had PET guided biopsy of a hypermetabolic lesion that conducted to lymphoma diagnosis in 7 (40%) cases. After chemotherapy for lymphoma, PET was available for 10 patients: complete regression of hypermetabolic lesions was observed in 6 patients (60%), and decreased uptake intensity in the 4 remaining patients. Conclusion. Some of the systemic manifestation of pSS (lung, lymph nodes and salivary glands) can be assessed by 18F-FDG PET. Lymph nodes hypermetabolism is common with a similar frequency in patients with and without lymphoma. Nevertheless, some 18F-FDG PET abnormalities are associated with lymphoma diagnosis: SUV max at any site  $\geq 6$ , SUV max of parotid glands  $\geq 5$  and focal nodular hypermetabolic lung lesions. Finally, PET can be helpful to guide biopsy toward the most hypermetabolic structure for diagnosing lymphoma.

#### P-183

Long-term results of R-CHOP therapy in patients with diffuse large B cell lymphoma (DLBCL) associated with Sjögren's syndrome (SS)

Efstathia K. Kapsogeorgou, PhD<sup>1</sup>, Dimitrios C. Ziogas, MD<sup>1</sup>, Aristea Papageorgiou, MD<sup>1</sup>, Athanasios G. Tzioufas, MD<sup>1</sup>, Michael Voulgarelis, MD<sup>1</sup>. <sup>1</sup>Department of Pathophysiology and Academic Joint Rheumatology Program, School of Medicine, National and Kapodistrian University of Athens, Greece.

**Background.** Development of DLBCL in SS is a major outcome, since it is the second most common type of lymphoma in SS, with the higher risk of adverse events during follow-up and significantly poorer outcome for patient survival compared to the other common SS-associated lymphomas. First line treatment is R-CHOP (rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone), which was empirically based on the therapeutic experience of the DLBCLs of the general population with similar 5-year survival rates. In this study, we evaluated the 10-year survival of SS-DLBCLs treated with R-CHOP.

**Methods.** Twelve cases with DLBCL from total 92 patients with SS-associated lymphomas that are included in our institution database were retrospectively studied. Eleven patients received R-CHOP and one CHOP. Democraphic, clinical, laboratory and histological features of SS disease and lymphoma were recorded and analyzed by appropriate statistical approaches.

Results. Patients with SS-associated DLBCL were monitored for a longterm follow-up since their SS- and DLBCL-diagnosis. Median follow-up from the diagnosis of SS was 38 years and a total of 211.15 person-years at risk, was reached and median follow-up from the diagnosis of DLBCL was 5.88 years and a total of 77.70 person-years at risk, was reached. During this follow-up, 6 patients died, but only 3 deaths were lymphoma-related. Seven patients achieved a complete remission (CR; CR rate: 58.33%) and 6 of them remained in CR and are still alive with a median duration of CR response reaching to 11.08 years (range: 0.01-13.92 years). In all patients that presented an event during follow up, this was observed during the first five years. From the diagnosis of SS, the 5-year and the 10-year OS% was 91.60% and 82.50%, respectively. From the diagnosis of DLBCL, the 5-year and 10-year EFS% was 33.3% and 22.22%, respectively and the median EFS was 3.26 (range: 0.01-13.92) years. The 5-year and 10-year OS% was 58.33% and 48.61%, respectively, while the median OS was 5.88 (range: 0.01-13.92) years.

**Conclusions.** The survival rates of SS-associated DLBCLs are similar to those described (10-year OS%: 43.5%) by the first study of Groupe d'Etude des Lymphomes de l'Adulte (GELA) investigating the addition of rituximab to CHOP and support the use of R-CHOP as treatment of choice.

## **P-184**

#### Cigarette smoking is a risk factor for developing primary Sjögren's syndrome with Ro/SSA and La/SSB autoantibodies

Albin Björk<sup>1</sup>, Johannes Mofors<sup>1</sup>, Marika Kvarnström<sup>1</sup>, Helena Forsbladd'Elia<sup>2</sup>, Sara Magnusson Bucher<sup>3</sup>, Jan Hillert<sup>4</sup>, Per Eriksson<sup>5</sup>, Thomas Mandl<sup>6</sup>, Gunnel Nordmark<sup>7</sup>, Lars Alfredsson<sup>8</sup>, Marie Wahren-Herlenius<sup>1</sup>.

<sup>1</sup>Department of Medicine, Karolinska Institutet, Karolinska University Hospital, Sweden. <sup>2</sup>Department of Public Health and Clinical Medicine, Rheumatology, Umeå University, Sweden. <sup>3</sup>Department of Rheumatology, Faculty of Medicine and Health, Örebro University, Sweden. <sup>4</sup>Department of Clinical Neuroscience, Karolinska Institutet, Karolinska University Hospital, Sweden. <sup>5</sup>Department of Clinical Experimental Medicine, Linköping University, Sweden. <sup>6</sup>Department of Rheumatology, Skåne University Hospital, Sweden. <sup>7</sup>Department of Medical Sciences, Uppsala University, Sweden. <sup>8</sup>Institute of Environmental Medicine, Karolinska Institutet, Sweden.

**Background.** Cigarette smoking is a well-established risk factor for several systemic autoimmune disorders, including rheumatic diseases. However, only a limited number of studies have investigated the effect of smoking on the risk of developing primary Sjögren's syndrome (pSS), reporting contradictory results. This may relate to factors such as them being conducted before development of current classification criteria, including few patients or using sicca controls for comparison. The aim of this study was therefore to investigate the impact of smoking on the development of pSS in a large, clinically well characterized cohort of patients with pSS.

**Methods.** A case-control study using prevalent cases of pSS classified according to the American-European Consensus Criteria (606 cases and 5,925 population controls) was performed. Smoking habits prior to diagnosis were obtained from questionnaire data and cases and controls were classified into ever-smokers or never-smokers. The impact of ever- smoking on pSS was assessed by calculating odds ratios (OR) with 95% confidence intervals (CI) employing logistic regression. Estimates were adjusted for age, sex, time period and area of residence.

**Results.** Ever-smokers had an increased risk of developing pSS (OR 1.3, 95% CI 1.1-1.5). The risk of ever-smokers to develop pSS was somewhat higher for men (OR 2.0, 95% CI 1.0–4.1, compared to OR 1.2, 95% CI 1.0–1.5 for women). Stratifying the analysis according to Ro/SSA and La/SSB autoantibody positivity revealed that the increased risk of pSS associated with smoking was limitd to individuals with autoantibodies; both in pSS positive for Ro/SSA and La/SSB autoantibodies (OR 1.5, 95% CI 1.2–2.0) as well as in Ro/SSA and/or La/SSB positive pSS (1.4, 95% CI 1.2–2.0) as well as in Ro/SSA and/or La/SSB positive pSS (1.4, 95% CI 1.1–1.7), but not in pSS negative for these autoantibodies (OR 1.1, 95% CI 0.8–1.6). **Conclusions.** We observed a significantly increased risk for ever-smokers of both sexes to develop pSS in the largest study to date. The increased risk was only evident for development of Ro/SSA and/or La/SSB positive pSS, and not for pSS negative for these autoantibodies. The data indicate that underlying genetic factors predisposing for autoantibody positivity may be of importance for the increased risk.

# P-185

#### Tobacco smoking effects on clinical, serological and histological manifestations of Sjögren's syndrome, a cross-sectional study of the OASIS cohort

Alexandre Dumusc<sup>1,2</sup>, Francesca Barone<sup>3</sup>, Rachel Brown<sup>2</sup>, Timothy Bates<sup>2</sup>, Andrea Richards<sup>4</sup>, Ana Poveda<sup>4</sup>, Jon Higham<sup>4</sup>, Saaeha Rauz<sup>3</sup>, Simon J Bowman<sup>2</sup>, Benjamin Fisher<sup>2,3</sup>.

<sup>1</sup>University Hospital Lausanne, Lausanne, Switzerland. <sup>2</sup>University Hospitals Birmingham NHS Foundation Trust, Birmingham, UK. <sup>3</sup>University of Birmingham, Birmingham, UK, <sup>4</sup>Birmingham Dental Hospital, Birmingham, UK.

**Background.** Previous studies showed that tobacco smoking in patients with primary Sjögren's syndrome (pSS) is negatively associated with a histopathological focus score and anti-Ro/anti-La serological positivity (1). We aimed to evaluate the association of tobacco smoking history with clinical, serological and histological features of patients with pSS.

**Methods.** Cross-sectional study of patients at the time of their inclusion in the OASIS cohort between 2014 and March 2017. This UK prospective research cohort includes patients with suspected pSS or known pSS.

Patients included in the analysis in the pSS group all fulfilled the ACR-EULAR (2016) classification criteria for pSS. We excluded patients with secondary Sjögren's syndrome. Characteristics of pSS patients with and

# 14th International Symposium on Sjögren's Syndrome

without smoking history were compared, including clinical, histological and serological data. These patients were also compared with patients with sicca symptoms without pSS included in the cohort.

For statistical analysis, we used unpaired t test, Mann-Whitney test, Fisher's exact test and Chi-square test when appropriate.  $P \le 0.05$  was considered statistically significant.

**Results.** Among the 174 patients included in the cohort, 83 fulfilled the ACR-EULAR 2016 classification criteria for pSS and 59 presented with sicca symptoms without pSS. The rate of current/previous smoking was respectively 5%/22% in the pSS group and 11%/32% in the sicca-non pSS group; 71%/58% of the patients with pSS/sicca-non pSS never smoked (p=0.16).

Patients with pSS who ever smoked, compared to those who never smoked, had a higher body mass index (BMI) (30.4±5.5 versus 26.9±5.3, kg/m<sup>2</sup> p=0.01) and a higher ESSPRI (8.0 [5.7-8.7] versus 6.0 [4.8-7.5], median [IQR], p=0.02) which is a patient reported outcome assessing fatigue, pain and dryness. They also had a higher pulmonary domain score of ESSDAI  $(0.29\pm0.64$  versus  $0.06\pm0.25$ , p=0.02) which did not impact the total ES-SDAI score which assesses systemic disease activity. Anti-Ro/La positivity, rheumatoid factor and IgG levels did not differ between both groups. Furthermore, tear and saliva production, assessed by Schirmer's test and unstimulated whole saliva flow rate, was similar in both groups. Histopathological data from minor salivary gland biopsies taken from patients with pSS who ever smoked (n=38) and never smoked (n=33) were compared. We did not observe differences between groups in terms of focus score, number of lymphoid aggregates, plasma cell infiltrates, fibrosis severity and presence of germinal centres on H&E or indicated by CD21+ immunostaining. Conclusion. Patients with pSS who ever smoked showed a higher BMI, ES-SPRI and pulmonary domain score of ESSDAI compared with patients with pSS who never smoked at inclusion in our cohort. In contrast with published data, they did not differ in terms of auto-antibody profile and minor salivary gland histological features.

References

 STONE DU, FIFE D, BROWN M et al.: Effect of Tobacco Smoking on The Clinical, Histopathological, and Serological Manifestations of Sjögren's Syndrome. PLoS One 2017; 12(2): e0170249.

#### P-186

#### No association between smoking status and serum cytokine levels in patients with primary Sjögren's syndrome

Peter Olsson<sup>1,2</sup>, Kristin Skogstrand<sup>3</sup>, Anna Nilsson<sup>1,4</sup>, Elke Theander<sup>1</sup>, Gunnar Houen<sup>5</sup>, Thomas Mandl<sup>1,2</sup>.

<sup>1</sup>Department of Clinical sciences, Malmö, Rheumatology, Lund University, Malmö, Sweden. <sup>2</sup>Department of Rheumatology, Skåne University Hospital, Lund University, Malmö, Sweden. <sup>3</sup>Department of Congenital Disorders, Center for Neonatal Screening, Statens Serum Institut, Copenhagen, Denmark. <sup>4</sup>Department of Rheumatology, Linköping University Hospital, Linköping, Sweden. <sup>5</sup>Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark.

**Background.** Smoking is reported to affect a number of diseases including rheumatic diseases. Epidemiological studies have previously found a negative association between primary Sjögren's syndrome (pSS) and smoking. The aim of this study was to examine whether smoking status affects serum cytokine expression and other markers of disease activity in pSS patients. **Method.** 51 consecutive pSS patients and 33 population controls were in-

cluded in the study. Clinical and standard laboratory parameters were registered. Twenty-four and 27 patients were ever smokers and never smokers respectively. Serum cytokines (IL-1 $\beta$ , IL-2, IL-3, IL-4, IL-6, IL-8, IL-10, IL-12, IL-17, IL-18, IL-33, IFN- $\alpha$ , IFN- $\gamma$ , TNF- $\alpha$ , EGF, BAFF, Fas-ligand, RANTES, TGF- $\beta$ 1) were assessed using the MesoScale U-plex platform.

**Results.** IL-6, IL-12, IL-17 and IL-18 were significantly increased in pSS patients in comparison to controls. However, no differences in cytokine levels were found when comparing ever and never smoking pSS patients. Furthermore, no associations between smoking status and the ESSDAI total score, IgG levels, or complement levels were found.

**Conclusion.** In this study, no differences in cytokine levels were found between ever and never smoking pSS patients despite the negative association between pSS diagnosis and smoking in epidemiological studies. A possible explanation includes a local effect of smoking on salivary glands rather than a systemic effect by cigarette smoke.

## P-187

# Influence of smoking and obesity on the risk of developing primary Sjögren's syndrome: a population-based cohort study

Divi Cornec, MD, PhD<sup>6.7</sup>, Luisa Servioli, MD<sup>1.2</sup>, Gabriel Maciel, MD<sup>1.2</sup>, Carlotta Nannini, MD<sup>1.3</sup>, Cynthia S. Crowson, MS<sup>1.4</sup>, Sara J. Achenbach, MS<sup>4</sup>, Eric L. Matteson, MD, MPH<sup>1.5</sup>, Alvise Berti, MD<sup>6.8</sup>.

<sup>1</sup>Division of Rheumatology, Mayo Clinic College of Medicine and Science, Rochester, MN, USA. <sup>2</sup>Autoimmune diseases Department, Medical Clinic 1. Hospital Maciel, 25 de Mayo 172, Montevideo, Uruguay 11000. <sup>3</sup>Department of Rheumatology, Hospital of Prato, Prato, Italy. <sup>4</sup>Division of Biomedical Statistics and Informatics, Department of Health Sciences Research, Mayo Clinic College of Medicine and Science, Rochester, MN, USA. <sup>5</sup>Division of Epidemiology, Department of Health Sciences Research, Mayo Clinic College of Medicine and Science, Rochester, MN, USA. <sup>6</sup>Division of Epidemiology, Department of Health Sciences Research, Mayo Clinic College of Medicine, Mayo Clinic College of Medicine and Science, Rochester, MN, USA. <sup>7</sup>Rheumatology Department, Brest Teaching Hospital, Brest, France. <sup>8</sup>Immunology, Rheumatology, Allergy and Rare Diseases Department, San Raffaele Scientific Institute, Milan; and Rheumatology Department, Santa Chiara Hospital, Trento; both in Italy.

**Objective.** Cigarette smoking and obesity have been identified as risk factors for developing several autoimmune diseases, and may be protective for others. We aimed to explore the role of these risk factors in primary Sjögren's syndrome (pSS).

Methods. A cohort of Olmsted County, Minnesota residents diagnosed with pSS between January 1, 2000 and December 31, 2015 was identified based on individual medical record review. Each of the cases was matched to 3 age- and sex-matched comparators without pSS randomly selected from Olmsted County residents, indexed to the date of pSS diagnosis. Smoking status was divided into three categories of current smoker, ex-smoker and never smoker. The body weight and height closest to date of diagnosis/ index date (±1 year) were used. Obesity was defined as a body mass index (BMI)≥30/kg/m<sup>2</sup>.

**Results.** 106 incident cases of pSS and 318 controls were identified. The odds ratio (OR) of pSS comparing current smokers with never smokers adjusted for age and sex was 0.34 (95% confidence interval (CI): 0.14, 0.85; p<0.05), while the age- and sex-adjusted OR for former smokers compared to never smokers were 1.27 (95% CI, 0.80, 2.03). The OR of pSS comparing obese subjects with non-obese subjects was 0.79 (95% CI, 0.48, 1.30), while the OR of pSS for BMI analyzed as a continuous variable was 0.97 (95% CI, 0.94, 1.01).

**Conclusions.** In this population-based study, current smokers have a lower risk of developing pSS while BMI does not affect this risk.

### **P-188**

#### Primary Sjögren's syndrome increases the risk of cardiovascular events, with the highest risk in SSA/SSB autoantibody positive patients

Johannes Mofors<sup>1</sup>, Linnea Westermark<sup>2</sup>, Albin Björk<sup>1</sup>, Marika Kvarnström<sup>1</sup>, Helena Forsblad- d'Elia<sup>3</sup>, Sara Magnusson Bucher<sup>4</sup>, Per Eriksson<sup>5</sup>, Thomas Mandl<sup>6</sup>, Marie Wahren-Herlenius<sup>1</sup>, Gunnel Nordmark<sup>2</sup>.

<sup>1</sup>Department of Medicine, Karolinska Institutet, Karolinska University Hospital, Sweden. <sup>2</sup>Department of Medical Sciences, Uppsala University, Sweden. <sup>3</sup>Department of Public Health and Clinical Medicine, Rheumatology, Umeå University, Sweden. <sup>4</sup>Department of Rheumatology, Faculty of Medicine and Health, Örebro University, Sweden. <sup>6</sup>Department of Clinical Experimental Medicine, Linköping University, Sweden.

**Background.** An increased risk of cardiovascular disease is well-established in autoimmune diseases such as rheumatoid arthritis and systemic lupus erythematosus. Studies examining cardiovascular morbidity in primary Sjögren's syndrome (pSS) are however few and inconclusive to date. In the present study, we therefore examined the incidence of cardiovascular events in patients with pSS using population-based national health registers. **Methods.** We established a cohort of clinically well-characterized cases of pSS fulfilling the American-European Consensus Group criteria, (n=979, 93% females) and used controls from the Swedish population matched on age, sex and area of residence (n=9,790). Data including ICD10 codes were extracted from the National Patient Register to identify cardiovascular events occurring after pSS diagnosis. Cox proportional hazard modeling was used to calculate Hazard Ratios (HR) and 95% Confidence Intervals (CI) of the events in patients.

#### Posters

**Results.** The mean age at pSS diagnosis was 54.4 years. Patients with pSS had a significantly increased risk of myocardial infarction (5.6%) compared with controls (3.2%), (HR 1.8, 95% CI 1.4-2.4), occurring at a mean of 9.8 years after pSS diagnosis. The risk was even higher in pSS patients positive for SSA and/or SSB autoantibodies (n=679 pSS, versus n=6790 controls) (HR 2.0, 95% CI 1.4-2.9). A non-significant trend of increased risk was observed in pSS patients without these autoantibodies (n=282 pSS, versus n=2820 controls) (HR 1.5, 95% CI 0.9-2.5).

Stratifying the SSA and/or SSB positive pSS for gender, the increased risk was significant in females (HR 2.0, 95% CI 1.4-3.0) with only a trend in males (HR 1.9, 95% CI 0.8-4.5). pSS could not be reliably associated with an increased risk of cerebral infarction, occurring in 4.2% of pSS patients, compared with 3.2% among controls (HR 1.3, 95% CI 0.9-1.8). However, pSS patients positive for SSA and/or SSB autoantibodies had a small increased risk of cerebral infarction (HR 1.4, 95% CI 1.0 – 2.1) while the risk in antibody negative pSS was in parity with the general population (HR 1.1, 95% CI 0.6 – 2.1). In affected patients, the cerebral infarction occurred after a mean of 12 years post diagnosis.

Venous thromboembolism was also more frequent in pSS compared with controls and deep venous thrombosis was observed in 4.4% of patients at a mean of 7 years after pSS diagnosis, versus 1.9% in controls (HR 2.3, 95% CI 1.7-3.3). Pulmonary embolism presented in 2.6% of patients at a mean of 10 years after pSS diagnosis, versus 1.1% in controls (HR 2.3, 95% CI 1.5-3.6). In pSS positive for SSA and/or SSB, the risk was even more striking or deep venous thrombosis (HR 2.7, 95% CI 1.8-4.0) and pulmonary embolism (HR 2.6, 95% CI 1.5-4.3).

**Conclusions.** We observed a significantly increased risk for both arterial and venous cardiovascular events occurring after pSS diagnosis, compared to matched population controls. Patients with pSS positive for SSA and/or SSB autoantibodies were at higher risk, suggesting that autoimmune disturbances influence the risk of cardiovascular complications.

#### P-189

#### Ethnicity of Sjögren's syndrome in the USA

Rohan Sharma, MBBS, Astrid Rasmussen, MD, PhD, Lida Radfar, DDS, David Lewis, DDS, Kiely Grundahl, BS, C. Erick Kaufman, MD, Joan A. Merrill, Kaustubh S. Chaudhari, MBBS, Kristi Koelsch, PhD, Kathy L Silvis, Christopher J. Lessard, R. Hal Scofield.

Arthritis & Clinical Immunology Program, Oklahoma Medical Research Foundation, Departments of Medicine and Pathology, University of Oklahoma Health Sciences Center, Medical Service, Oklahoma City Department of Veterans Affairs Medical Center; Department of Oral Diagnosis and Radiology, College of Dentistry, University of Oklahoma Health Sciences Center, Oklahoma City, USA.

**Background.** We undertook this study to describe the racial and ethnic diversity of primary Sjögren's syndrome (pSS) compared to SLE, which is known to be more common and severe in Black Americans than White Americans.

**Methods.** Individuals with sicca were evaluated by a rheumatologist, ophthalmologist and a dentist. History and physical examination with stimulated and unstimulated salivary flow (WUSF) and a lip biopsy, ocular surface staining with Lissamine green and fluorescein, an unanaesthetised Schirmer's test as well as general laboratory tests were performed. Subjects were classified using ACEG and ACR criteria for pSS. We compared the non-Hispanic Black pSS to the non-Hispanic White pSS subjects with one to four age and sex match in terms of clinical and serological manifestations. We also compared pSS subjects with non-Sjögren's Sicca (nSS) and those in a SLE cohort followed in the same facility. P values were corrected for multiple comparisons. Due to low representation of other ethnic groups in our study population we only considered non-Hispanic Blacks and non-Hispanic Whites in the study.

**Results.** We classified 327 subjects in the clinic as pSS of which 201 were considered for the study. Among these 187 (92.1%) were self-identified as White, while only 14 (6.9%) were self-identified as Black. There were 7 (3.05%) Blacks and 223 (96.95%) Whites in nSS group. Among the SLE subjects, there were 106 (29.5%) Black and 253 (61.5%) as White. Thus, we found that black Americans were 5 times more likely to have SLE compared to pSS( $\chi$ 2=36.17, p<0.00001, OR=5.45), while there was no such difference when compared to pSS subjects with Nss(control group) ( $\chi$ 2=2.76, p=0.0966, OR=0.41). We also compared the ethnic make-up of the pSS to the population and found no difference, while among SLE patients Black Americans were several times more common than in the general popula-

tion. Concerning the classification criteria, we found that Black subjects had higher incidence of corneal erosion as evident by Lissamine green test (p=2.7x10-4 by Fisher's exact test). We also evaluated pSS subjects for systemic manifestations. Black subjects were found to have higher incidence of parotid gland enlargement (p=0.01 by Fisher's exact test) and hypergammaglobulinemia IgG type (p=0.02 by Fisher's exact test).

**Conclusions.** In contrast to SLE, Black Americans were enriched among subjects with pSS. This is likely due to a biological difference in the risk of the two diseases between these ethnic groups. However, socioeconomic differences leading to health care disparity may be responsible for some of the difference in representation among subjects with these diseases. We found minor differences in the manifestations of pSS between Black and White Americans, suggesting pSS is not more severe in Blacks. Similar to risk of disease, this is in contrast to the situation in SLE where Black Americans have more severe disease.