
Biochemical markers in osteoarthritis with lessons learned from osteoporosis

M.A. Karsdal, K. Henriksen, A.C. Bay-Jensen

Nordic Bioscience, Herlev, Denmark.

Morten Asser Karsdal, PhD

Kim Henriksen, PhD

Anne Christine Bay-Jensen, PhD

Please address correspondence to:

Prof. Morten Asser Karsdal,

Nordic Bioscience,

Herlev Hovdgade 207,

2730 Herlev, Denmark.

E-mail: mk@nordicbio.com

Received on and accepted on September 11, 2019.

Clin Exp Rheumatol 2019; 37 (Suppl. 120): S73-S87.

© Copyright CLINICAL AND

EXPERIMENTAL RHEUMATOLOGY 2019.

Key words: osteoarthritis, biomarkers, osteoporosis

ABSTRACT

Osteoarthritis (OA) is a disease of the whole joint, including synovium, bone and cartilage. OA is a slow degenerative and very heterogeneous disease, with both varying levels of disease activity and progression. Biomarkers are urgently needed to assist drug developers in selecting and developing the projects with the highest chance of success. Biomarkers for enrichment of clinical studies, early efficacy as well as other diagnostic tools are needed. Osteoporosis and OA have many similarities. In osteoporosis an armamentarium of treatments are now available with high clinical efficacy and well-described effects on biomarkers. Possibly, lessons learned from the osteoporosis field in the use of biomarkers may be applied in the OA field, from both technical and scientific perspectives. To help guide the way, the FDA has recently published the BEST guidelines, to facilitate obtaining a common vocabulary to assist biomarker researchers. In the current review, we will review the biomarkers of OA, with emphasis on bone, cartilage and synovial biomarkers, and draw clear perspectives to the use of biomarkers for drug development and clinical practice in the osteoporosis field.

Introduction

Osteoarthritis (OA) is the most common form of arthritis and one of the leading causes of disability in the world, with more than 10% of the elderly population having symptomatic disease (1, 2). The hallmark of the disease is joint pain and progressive degeneration of articular cartilage involving remodelling of all joint tissues (bone, synovium, ligaments) with subsequent JSN (3). Several lines of evidence suggest that the structural integrity of articular cartilage is dependent on normal subchondral bone turnover, synovial inflammation,

intact chondrocyte function and physiological biomechanical stresses (4).

In addition, cartilage and bone metabolism may be partly linked, particularly subchondral bone turnover and its interaction with articular cartilage (4). This suggests that biomarkers of OA may be from several different compartments, as we consider OA a whole joint disease.

Osteoporosis and osteoarthritis share many similarities that may enable use and interpretation of biomarkers in the OA field. Both are slowly progressive diseases which pose a range of drug development challenges, particularly in Phase II dose-finding studies, and even in Phase 1b decision-enabling studies (5).

In osteoporosis, an armamentarium of treatments are available and the relationship between potential efficacy and delta biomarker change is well understood (6). These challenges remain to be addressed for OA, to optimise drug development as well as personalised and precision medicine (7, 8).

In osteoporosis, the standard biomarkers of bone resorption (CTX-I & NTX-I) (9-11) as well as bone formation (PINP, osteocalcin & BSAP) (10, 12, 13), provide optimal assessment tools when calculating the tissue balance (14). Interestingly, osteoporosis is a disease with both increased bone formation and bone resorption, as exemplified in figure 1, extracted from (15-17) and key data from (18). In contrast, the OA field is still exploring the diagnostic and prognostic capacity of biomarkers allowing deconstruction of the tissue turnover, and thereby diagnosis, prognosis and monitoring response to treatment.

The osteoporosis field has long benefited from a host of sensitive and reliable methods, which have documented utility as surrogate markers of pharmacodynamic effects targeting the bone com-

Competing interests: the authors all hold stock in Nordic Bioscience.

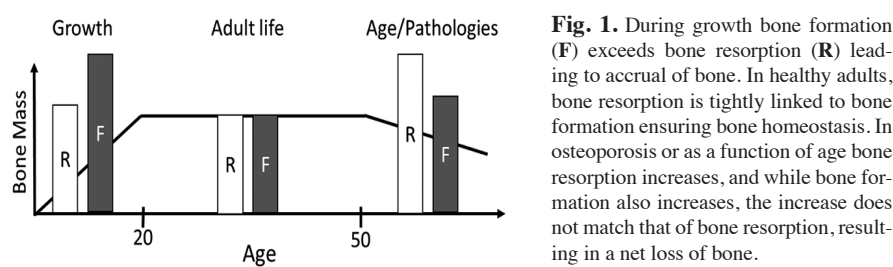


Fig. 1. During growth bone formation (F) exceeds bone resorption (R) leading to accrual of bone. In healthy adults, bone resorption is tightly linked to bone formation ensuring bone homeostasis. In osteoporosis or as a function of age bone resorption increases, and while bone formation also increases, the increase does not match that of bone resorption, resulting in a net loss of bone.

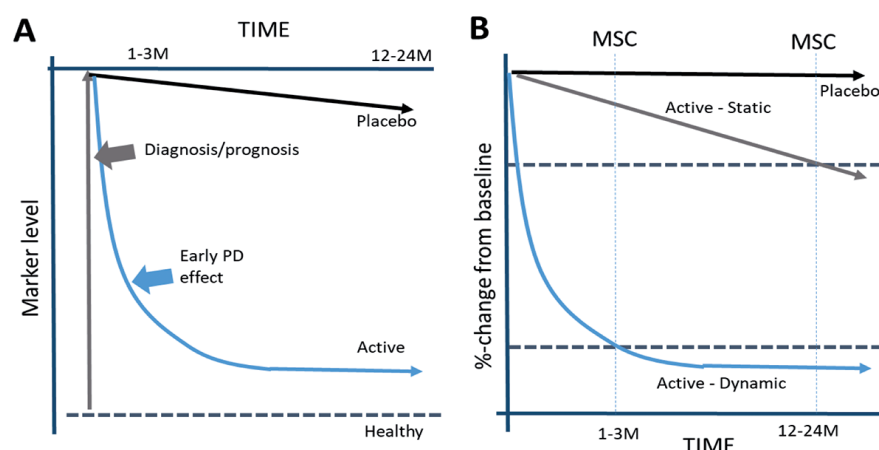


Fig. 2. Different applications of biomarkers.

A: Using a biomarker to identify the subgroup of a disease population that progresses (Diagnosis/Prognosis), and the same or an alternative biomarker to monitor the early PD response to a given treatment. **B:** For drug development the increased magnitude of the dynamic biochemical marker provides a clear advantage compared to a static biomarker such as imaging by MRI, DXA or alike. Modified from (22). MSC, minimal significant change.

partment (12). These biomarkers have provided drug developers with a series of advantages leading to rapid and effective development of drugs, and the osteoporosis market has, accordingly, a series of efficacious drugs that clearly sets it apart from a field such as OA, where fast and efficacious markers are still being investigated and developed (7, 19-21).

Figure 2 presents the ability of the dynamic bone markers to provide an indication of early efficacy, as opposed to the classical and more static biomarkers. As illustrated, the time up to a minimal significant change (MSC) using imaging, such as joint space width (JSW) assessed by x-ray, is often 12-24 months, whereas the biochemical markers often show immediate responses resulting in significant changes within 1-3 months. Such biomarkers allow drug developers more confidence and information to invest in the project with the best possibility for success. This type of biomarker is what is needed for the OA field.

Common confounders in the assessment of biomarkers

Measurement of biomarkers is in many cases dependent on a whole range of parameters (23, 24). Among these are assay technology, fasting status, time of sample, type of sample (plasma, serum etc.), handling of the sample, the population (males/females, age, disease status, treatments) and many more. These parameters have been extensively discussed in several papers (23, 24), including in the rheumatology field (23), and for the ease of reading we have included them in Table I, which is modified from (23).

Overall, there is an ever-expanding portfolio of biomarkers applied in a plethora of diseases, both for identification of patients, identification of fast progressors and for monitoring responses to therapy. With this in mind, there is a continuous need to ensure that these tools are as comparable as possible.

Biomarker nomenclature

Biomarkers are utilised for assessment

of many different parameters in clinical studies. Hence, having a glossary ensuring broad understanding of the clinical relevance and thereby the application of the biomarkers is critical. In this regard, the FDA and NIH have proposed the BEST Resource, which captures these aspects in a simple and easy to follow format. The Osteoarthritis Research Society International has proposed the BIPED criteria (25); assessing Burden of disease, Investigative, Prognostic, Efficacy of intervention and Diagnostics use. However, the BEST definitions include a more detailed glossary around biomarker functions and surrogate endpoints, such as pharmacodynamics and safety, and thereby ensures both consistency and common understanding of issues related to use/interpretation of biomarker data from clinical trials (26).

Introduction to the bone field

Bone remodelling in healthy adults is a process occurring at all time points, and it is key to safeguarding bone integrity (17). At the cellular level, the bone resorbing osteoclasts and the bone forming osteoblasts, which work in synchrony to ensure calcium homeostasis and repair of damage occurring in the bone remodelling units, conduct bone remodelling (17).

In many cases, pathologies in bone arise from imbalance in bone remodelling, as exemplified by post-menopausal osteoporosis, as well as a host of other metabolic bone disorders (17, 27). These disorders are characterised by increased bone resorption, followed by increased bone formation, albeit to an insufficient extent. This results in loss of bone, and potentially osteoporosis and a high risk of fractures which are known to be detrimental to both quality of life and life expectancy (17).

Post-menopausal osteoporosis is the most common bone pathology, occurring in as many as 1/3 of all women (28). In parallel, a larger number of men also experience bone loss as a function of age and cessation of gender hormone production (28). Osteoporosis, whether in men, women or induced by glucocorticoids, is a low bone mass disorder, characterised by thinning of the bone structures, leading to increased risk of

Table I. Common parameters influencing the evaluation of novel non-invasive biomarkers.

BIOLOGICAL PARAMETERS	Food intake	Diurnal variation	Seasonal variation	Disease activity	Medical condition and treatments
SAMPLING PARAMETERS	Needle gauge, site of draw, handling	Shipping and storage condition	Freeze-thaw cycles	Matrix (serum, plasma, urine, CSF, saliva etc.)	Anti-coagulant (EDTA, heparin citrate)
ANALYTE FEATURES	Total protein	Protein fragment	Active enzyme	Latent enzyme	Biological role
ASSAY FORMAT	Competitive	Sandwich	Mono- or poly-clonal antibody	Multiplex or singular	Sample volume
ASSAY PARAMETERS	Analyte recovery and precision	Buffer robustness	Measurement-range	Sensitivity	Specificity and accuracy
STUDY PARAMETERS	Patient population and confounders	Mode of action	Duration of study	Onset of action	Number of samples

Adapted and updated from references (23) and (24).

fragility fractures. With an aging world population (28, 29) osteoporosis is becoming a global problem.

WHO defines osteoporosis based on DXA scans of the lumbar spine. The mean T-score in the lumbar spine of young adults (Young Adult Mean (YAM)) serves as the reference, and having a YAM between -1.0 and -2.5 indicates low bone mass (osteopenia), while a T-score below -2.5 of YAM is osteoporosis (29).

The major challenge using bone mineral density (BMD) as the definition of os-

teoporosis is the poor ability of BMD to predict the risk of fractures; a phenomenon underscored by a large amount of fractures occurring in individuals who are not in the osteoporosis category of BMD T-scores (30, 31). The predictive ability of BMD is markedly improved when utilising changes BMD over time; however, changes in BMD are small and rather slow, and the time to a predictive change is quite long, *i.e.* more than a year in many cases (30, 31).

To account for the rather poor predictive ability of BMD, the FRAX® al-

gorithm was developed (32). FRAX® includes additional risk factors for fractures, such as age, gender, prior fractures, BMD, family history of fractures, BMI, smoking, alcohol, rheumatoid arthritis and glucocorticoid use (32). Unfortunately, implementing FRAX® has not improved the fracture risk prediction dramatically (33). An important and interesting point of discussion relating to FRAX® is the lack of use of bone turnover markers (BTMs) in the algorithm. The decision not to include them is likely driven by the rather variable nature of the BTMs, a point of discussion in the coming sections (34).

Table II. BEST resource for biomarkers (26).

Biomarker	Definition
Diagnostic biomarker	<ul style="list-style-type: none"> • Detect or confirm presence of the medical condition of interest. • Identify individuals with a subtype of the medical condition of interest.
Monitoring biomarker	<ul style="list-style-type: none"> • Monitoring status of a medical condition by repeated measurements. • Assessing possible effect of exposure to a drug or an environmental agent.
Pharmacodynamic/response biomarker	<ul style="list-style-type: none"> • Display if a biological response has occurred after exposure to a drug or an environmental agent
Predictive biomarker	<ul style="list-style-type: none"> • Identify those subjects who are more prone than similar subjects, to experience a favorable or unfavourable effect after exposure to a drug or environmental agent.
Prognostic biomarker	<ul style="list-style-type: none"> • Identify probability of a clinical event, disease recurrence or progression in patients with the medical condition of interest.
Safety biomarker	<ul style="list-style-type: none"> • Measure before and/or after exposure to a drug or environmental agent to assess possible toxicity as an adverse effect
Susceptibility/Risk biomarker	<ul style="list-style-type: none"> • Assessing the potential for developing a medical condition in a subject who does not currently have any symptoms

Bone turnover markers:

what they can and cannot do

As mentioned previously, the challenge with BMD is the slow rate of changes, which limits the ability of using it to identify those who lose bone fast and those who respond well to treatment, thereby pin-pointing the individuals who are in most need of treatment and those who should be enrolled in clinical trials of anti-osteoporotic drugs (12, 35). These points underscored the need for being able to measure the bone turnover balance with a higher sensitivity, *i.e.* through the use of biochemical markers that reflect bone resorption and bone formation rates, and as such provide higher resolution information about the changes in bone, and thereby help identify patients in need (12, 35).

In relation to bone turnover markers, collagen type I turnover has proven highly relevant. Collagen type I is, by

far, the most abundant protein in the bone matrix, and accordingly early biomarker development focused on monitoring changes in type I collagen synthesis and degradation (36, 37). Importantly, studies of collagen type I formation and degradation biomarkers have shown that monitoring these processes provides a unique ability to measure the actual bone turnover rate. When these assessments of rate are combined with the bone status provided by BMD measurements, it presents the possibility to identify those in need of treatment, *i.e.* fast bone losers (38-40).

For monitoring bone resorption, collagen type I degradation by the bone resorbing osteoclasts is a key process. Accordingly, studies have shown that CTX-I, which is a collagen type I fragment generated by the osteoclast protease cathepsin K, is a very sensitive measurement of bone resorption (12, 41). Accordingly, CTX-I has been shown to respond to anti-resorptive therapies, including bisphosphonates, denosumab, oestrogen, SERMs (selective oestrogen modulator), cathepsin K inhibitors, as well as other compounds at various stages of clinical development (17, 42).

The major breakthrough for CTX-I was a study of the bisphosphonate, alendronate (43, 44). The key results are shown in Figure 3, and they highlight the CTX-I and BMD changes as a function of alendronate dose. The data clearly illustrate a fast and large suppression of CTX-I, early after onset of treatment. In contrast, changes in BMD occurred more slowly, and the magnitude of change was markedly smaller (Fig. 3). Most importantly, the BMD change induced by alendronate was predicted by the early CTX-I alterations, clearly highlighting the utility of CTX-I alteration in terms of assisting drug development, both at the level of dose selection, as well as the overall chances of obtaining an efficacious drug on bone resorption.

Existing bone markers and their utility

Three major categories of bone markers are of relevance: (1) bone resorption markers, *i.e.* the activity of the bone resorbing osteoclasts, (2) markers that reflect the number, but not the

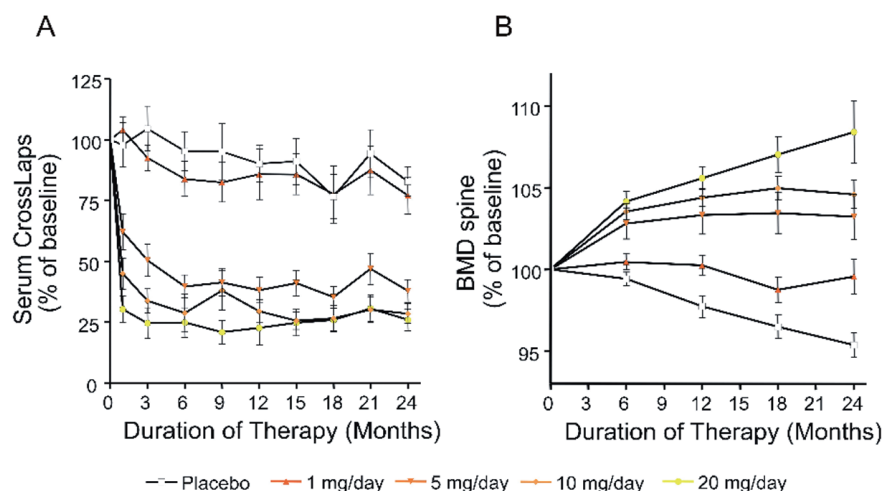


Fig. 3. (A & B) Dose-response in biochemical marker CTX-I in response to alendronate in a phase II clinical trial. The sensitivity of response of the biochemical marker CTX-I compared to that of the “gold standard” BMD provides evidence of efficacy faster and in a smaller study population. Modified from (43) and discussed in (11).

activity, of osteoclasts, and (3) bone formation markers providing insights into the activity of the osteoblasts. For overview, we have summarised the bone turnover biomarkers which are still of clinical utility in Table III, including a brief description of their use with the appropriate references.

A few biomarkers not falling into the three above-described categories exist. These include assessment of the levels of the bone formation inhibitors, DKK1 and sclerostin, as well as the pro-osteoclastic cytokine RANKL (12). In bone, these biomarkers still do not have a clear-cut additional value on top of the classical biomarkers. On the other hand, in rheumatic diseases, including osteoarthritis and rheumatoid arthritis, they may still be of quite some interest, although more studies are needed.

The bone field has benefitted substantially from having truly well-characterised biomarkers reflecting bone resorption and bone formation (10, 17). Utilising these biomarkers has allowed drug development for osteoporosis by identification of novel drug targets and candidates. The BTMs have also shown significant clinical utility as markers of bone safety as a function of various drugs, include glitazones, anti-psychotics and more (41). However, the most significant contribution of the BTMs has been to ramp up the speed of drug development through the implementa-

tion of earlier indications of efficacy, or lack thereof, on BMD and subsequently fracture reduction in the clinical studies.

The utility of BTMS for diagnosis and prognosis

While CTX-I is one of the most, if not the most sensitive marker of bone resorption, it is not a suitable tool for measurement of BMD in patients, as the correlation between CTX and BMD, at any given time point, is modest (41, 60). Overall, there is a slight elevation of the BTMs following menopause, which corresponds to the overall increase in bone turnover, both at the level of bone resorption and bone formation; however, the increases are not large enough to provide a stand-alone diagnostic value at any given time point (35, 52). There are indications that the inverse relationship between bone turnover markers and bone mineral density becomes stronger with age, an association that is particularly good for bone resorption markers (61, 62).

In terms of prognostic ability, the BTMs, due to their direct relation to the activities of the bone cell populations, have a clear-cut value, as underscored by the large studies called EPIDOS and OFELY (63-66). In EPIDOS and OFELY, the relationship between baseline levels of the BTMs and fracture risk was investigated, and the BTMs provide an independent ability to predict fracture risk in these populations

Table III. Biomarkers in the bone field and their application according to BEST (26).

*Recommended by The International Osteoporosis Foundation (IOF) and the International Federation of Clinical Chemistry and Laboratory Medicine (IFCC) as the biomarkers to use in clinical studies (35).

Biomarker	Process	Description	BEST classification	Ref.
α CTX-I	Bone resorption particularly in cases with very high bone turnover, such as Paget's, Osteolytic metastases, OA and RA.	A c-terminal crosslinked fragment of collagen type I generated by cathepsin K cleavage during osteoclast-mediated bone resorption of newly synthesised collagen	Prognosis, Pharmacodynamic	(45)
β CTX-I*	Bone resorption	A C-terminal crosslinked and isomerised fragment of collagen type I generated by cathepsin K cleavage during osteoclast-mediated bone resorption of mature collagen	Prognosis, Pharmacodynamic, Safety	(12, 41, 46, 47)
NTX	Bone resorption	An N-terminal crosslinked fragment of type I collagen generated by cathepsin K during osteoclastic bone resorption	Prognosis, Pharmacodynamic	(48)
TRACP 5b	Osteoclast number	TRACP 5b (Tartrate resistant acid) is an enzyme produced specifically by the osteoclasts	Pharmacodynamic	(10, 49, 50)
Cathepsin K	Osteoclast number	An enzyme produced specifically by the osteoclasts. Present in both a pro and active form	Pharmacodynamic	(10, 51)
PINP* PICP	Bone formation	The propeptides are trimeric, globular peptides enzymatically released from newly synthesised pre-pro-collagen prior to incorporation of the collagen molecule into the extracellular bone matrix	Prognosis, Pharmacodynamic, Safety	(35, 46, 47, 52)
BSAP	Bone formation	BASP (bone specific alkaline phosphatase) an enzyme specifically produced by osteoblasts	Pharmacodynamic	(35, 52)
Osteocalcin	Bone formation	OC is a 49-amino acid bone-specific protein produced by the osteoblast and inserted into the bone matrix	Pharmacodynamic	(35, 52)
RANKL	Not clear yet	RANK-L (receptor nuclear activator of NFkappabeta- ligand)Pro-osteoclastic cytokine produced by osteoblasts, primarily expressed on the cell surface	Not clear	(53)
ICTP	Matrix destruction in cases of high turnover, such as Paget's, Osteolytic metastases and RA. Not bone resorption	Matrix Metallo Proteinase cleaved type I collagen generated by many cell types including osteoclasts	Pharmacodynamic	(54)
Sclerostin	Not clear yet	Inhibitor of wnt signalling and thereby bone formation produced by the osteocytes	Not clear	(55-58)
DKK1	Not clear yet	DKK1(Dickkopf-related protein) Inhibitor of wnt signalling and thereby bone formation	Not clear	(59)

of elderly or post-menopausal women. Furthermore, in combination with other risk factors for future fractures, such as BMD and/or prior fractures, the prognostic capacity was enhanced (63-67), albeit with some fracture sites being better predicted than others (64, 65, 68).

However, as mentioned before, the ability of the BTMs, and particularly CTX-I to reflect the cellular activity in bone, *i.e.* the bone resorptive activity of the osteoclasts assessed by CTX-I, is what sets the BTMs apart in terms of drug development (13, 41).

BTMS as pharmacodynamic markers

As alluded to earlier in this review, one aspect of the BTMs has aided the osteoporosis field more than anything and that is their ability to monitor efficacies of intervention (41). This holds true whether it is the suppression of bone resorption by bisphosphonates or the induction of bone formation by PTH analogues, as well as for the other osteoporosis drugs and drug candidates (41). In early clinical trials, the BTMs are used to provide an early indication of treatment efficacy for osteoporosis,

as illustrated by the earlier mentioned alendronate example, but also by studies of the cathepsin K inhibitor odanacatib (69-71). In these studies, the BTMs are applied to provide surrogate measures of the BMD changes, as well as to provide an indication of fracture risk reduction, already at the early stages of development, thereby allowing go-no-go decisions based on these data (13).

As an illustration of the utility and the magnitude of the responses measured using BTMs, Table IV provides an overview of the therapy-induced

changes in CTX-I and PINP, which are the markers recommended by the IOF-IFCC (35), and as such the ones applied in most trials.

For CTX-I there is a very clear relationship to the suppression of bone resorption, with the most potent resorption inhibitors, such as denosumab, virtually eliminating CTX-I from the circulation, clearly indicating specificity for bone resorption (72, 73). On the other hand, less potent anti-resorptives, such as raloxifene suppress CTX-I to a lower extent (74), but also have a less pronounced effect on BMD, once again underscoring the strong relationship between changes in CTX-I and changes in BMD. An important aspect in addition to the suppression of CTX-I by the anti-resorptives, is the extent of the suppression of bone formation also induced by anti-resorptives (17, 41). This is illustrated by the prominent suppression of the bone formation marker PINP by bisphosphonates and denosumab (17, 41), which is consequent to the tight coupling of bone formation to bone resorption during bone remodelling.

PINP has also been applied extensively during the development of bone anabolic drugs, such as PTH and PTHrP analogues, and romosozumab (75-78). In these studies, clear PINP responses were observed as a function of the drugs, and when combining the changes of PINP with alterations in CTX-I, a good indication of the expected BMD changes was obtained (75-78).

As indicated earlier, one important point about the BTM responses needs to be underscored: suppression in BTM observed after three months of treatment is predictive of long term change in BMD (43, 44). Additionally, the amplitude of the BTM response (30–100% change in the majority of studies) clearly exceeds that amplitude of BMD (1–10%) and fracture rates (1–5%), and therefore the number of study subjects in early trials can be reduced, allowing faster and more efficient determination of whether it is a drug candidate worth pursuing (11). Finally, the ability to monitor both bone resorption and bone formation simultaneously and frequently, also allowed investigation of novel drug candidates with alternative modes

of action, *i.e.* drugs that only inhibit bone resorption without effects on bone formation (79-81), or drugs with a pure anabolic effect, as opposed to the PTH analogues which augment overall bone turnover (75-78). At present, there are drugs only inhibiting bone resorption, while BTM studies of the PTHrP analogues abaloparatide and the anti-sclerostin antibody have shown that a purer bone anabolic effect can be obtained with these drugs (75-78). These data again highlight the importance of having these types of biomarkers available during drug development to ensure successful development and differentiation from other drugs.

The safety aspect: BTMS as indicators of bone adverse effects

During clinical development of several different types of drugs for indications ranging from cancer and rheumatoid arthritis, through type 2 diabetes to viral infections and psychosis, warning signals in terms of reports of increased fracture rates have been presented (46, 47, 99, 100-105). While the mode of action underlying the increased fracture rates in some cases is poorly understood, it has become clear that the increased fracture rates reside in detrimental effects of the drugs on bone turnover (46, 47, 99, 100-105).

In many cases, the detrimental effects of these drugs on bone can be monitored using BTMs, as has clearly been shown for glitazones and glucocorticoids (99, 100). While the benefits of these drugs often clearly exceed the increased risk of fractures, application of the BTMs could allow deselection of those losing bone the fastest, as illustrated by studies of glitazones (78, 106), and as such complications can be reduced by treating subjects at risk of bone loss with other alternatives (13). Furthermore, the BTMs have also been applied during the development of non-bone harming TZDs, such as the partial PPAR gamma agonist balaglitazone, again highlighting the potential of BTMs for monitoring adverse bone responses during drug development (47). More recently, measurements of BTMs have been applied in the development of FGF-21 analogues, where they indi-

cate a modest reduction in bone formation accompanied by a minor increase in CTX-I (107), and as such can help guide the future development of drugs with the same mode of action, as described for Pegbelfermin (105, 108).

Limitations

The main limitation in the interpretation and utility of the BTMs is variation. Importantly, it is well known how to control the majority of the variation (24, 23,109). A series of studies shed light on the impact of diurnal variation and food intake on the BTMs, and these clearly showed that it is essential to collect blood samples for BTM analyses in the fasting state and in the morning, as this circumvents the impact of these parameters on the read-out (35, 109).

At the individual level, the application of BTMs is still rather flawed unless the samples are collected longitudinally, with multiple samples collected over time and thereby studying the fluctuation of the BTMs between individuals exposed to various stimuli. This has been neatly demonstrated in clinical studies involved in the optimisation of dosing time and frequency of oral calcitonin (79, 80, 110, 111), where alterations were provided a way of monitoring response to treatment at specific time points, but also compliance with the treatment (11, 35).

There are two primary types of variation: The one that cannot be controlled but needs to be carefully reported: age and gender, menopause, diseases and drugs, fractures, prolonged bedrest, and others (see Table I) (35, 109, 112). As mentioned previously, circadian rhythm and food intake, which, in addition to a host of technical parameters, such as needle gauge, tubes and location of the blood draw also need to be controlled to obtain good BTM data (35, 109, 112).

Cartilage turnover markers

Joint destruction has been suggested to follow a pattern of inertia accelerating disease progression (113). This poses high demands on the understanding of the clinical representation in the interpretation of image-based biomarkers. Highly important, as depicted in Figure 4, the level of a biomarker may be

Table IV: Summary of the changes in bone resorption and bone formation marker measured in drug treatment studies using the IOF-recommended markers CTX-I and PINP (35).

MOA	Treatment	CTX*	PINP*
Anti-resorptive	Nasal calcitonin	-10%	(82)
	Oral calcitonin#	-20%	(83, 84)
	Alendronate	-71 – -81%	-64 – -70% (74, 85, 86)
	Risedronate	-55%	-48% (87)
	Ibandronate	-58% – -73%	ND (87-89)
	Zoledronate	-58%	-59% (90)
	Denosumab	-70 – -75%	-50 – -60% (72, 73)
	HRT (s.c. pellet)	-40%&	-35% (91)
	Raloxifene	-21 – -28%	-34% (92, 93)
	Strontium Ranelate	-12%	-6.3% (94, 95)
	Odanacatib#	-72%	-40% (70, 71)
	ONO-5334	-41%	-27% (96)
Anabolic	PTH(1-34)	5%	111-135% (74, 75)
	PTH(1-84)	10-100%	90-150% (97, 98)
	Abaloparatide	20%	95% (78)
	Romosozumab	-50%	+180% (76, 77)

The table was inspired by (12). *The responses are presented as ranges depending on different doses, treatment strategies, and different cohorts. &uNTX, not CTX-I.

low or high independent of the disease status if the biomarker is related to disease activity such as the tissue turnover biomarkers described in Table III (114). In direct alignment, a bathtub analogy may be applied. If we are to predict how much water is in a bathtub tomorrow, we need three measures. How much water is in it today (this could be an image of the knee); how much water is running in (this could be a cartilage formation biomarker such as PRO-C2 or PIIANP); and finally how much water is running out (this could be a cartilage degradation biomarker such as CTX-II, C2C, or C2M) (115, 116). With this balance and refinement, an increased understanding is achieved. The level of a

biomarker does not necessarily need to correlate to the status of the disease, that is KL score, but rather to the velocity in which the disease is progressing (117). In addition, as a direct consequence, we possibly need to combine the imaging and soluble biomarker modalities rather than making simple correlations (118). Cartilage consists mainly of type II and X collagen, as well as other minor collagens (119) in addition to a host of non-collagenous proteins such as COMP and HA (120) and aggrecan which is the other main macromolecule next to type II collagen. During cartilage formation of degradation, cartilage turnover changes, either in response to a pathological insult or to pharma-

cotherapy. Cartilage consists of type II collagen (60–70% of the dry weight) and proteoglycans (10%) of which aggrecan is the most abundant (119). The key mediators of cartilage degradation include matrix metalloproteinases (MMPs) and the closely related aggrecanases, which are members of the ADAM-TS family (a disintegrin and metalloproteinase with thrombospondin motifs) (121). Aggrecan is degraded by both MMPs and aggrecanases, whereas type II collagen is degraded mainly by MMPs. Since type II collagen is the most abundant protein in cartilage, several different degradation fragments of type II collagen have been identified for non-invasive and objective assessment of joint pathology (122).

If the delicate balance between cartilage formation and cartilage degradation is slightly tilted toward a loss of cartilage, an overall thinning of cartilage over time may be the result. With joint disease, this imbalance is accelerated leading to a measurable net loss of cartilage (123).

Table V provides a list of biomarkers used in direct research setting in the rheumatology field .

Cartilage degradation

C-terminal telopeptide of type II collagen - CTX-II

The turnover of type II collagen is slow; however, in joint degenerative pathologies such as OA and RA an overproduction of proteases leads to increased type II collagen degradation (118). When MMPs act on type II collagen, small fragments of the protein are released into the synovial fluid and later into serum and excreted in urine, where it may be targeted as biochemical markers (150).

CTX-II is a cartilage degradation marker which has shown to be the best prognostic biomarker in the field (125, 117). Elevated CTX-II has, in multiple clinical studies, shown to be diagnostic and prognostic for OA. Reijman *et al.* showed that high levels of CTX-II were associated with increased risk (odds ratio of 5 in the upper quartile) of both knee and hip OA (117). In addition, elevated levels were highly predictive with an odds ratio of more than 8 for radiographic progression measured by joint

HETEROGENESITY IN PROGRESSION, DISEASE ACTIVITY AND STATUS

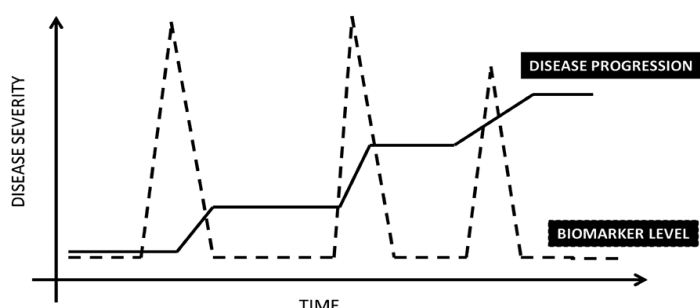


Fig. 4. The level of disease activity biomarkers may be independent of the status and level of disease, and as such clinical data and biomarker data may not always be interpreted as simple correlations, but rather in combination with synergy as demonstrated by Dam and colleagues (118). As OA progression is higher in some intervention, levels of prognostic biomarkers may be elevated during these periods, but low in period of slow progression.

Table V. An objective list of biomarkers used in preclinical and clinical setting reflecting either the bone, cartilage or synovium component of the joint. Modified from Bay-Jensen *et al.* (123)

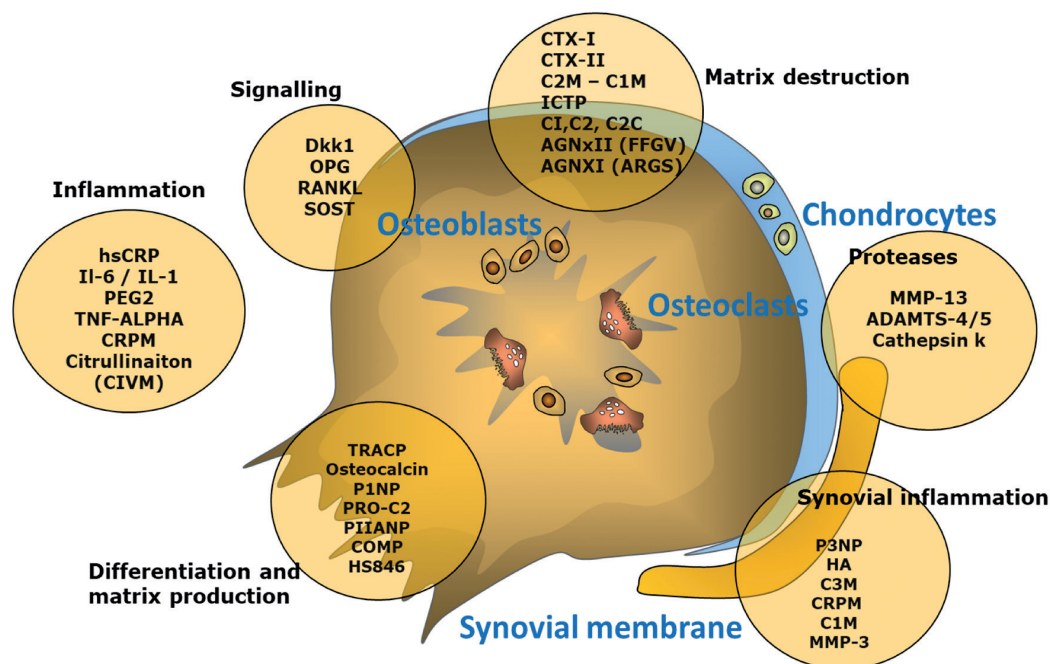
Biomarker/Protein	Description/Understanding	Selected references
BONE		
Alpha CTX-I	Cathepsin K degraded newly formed type I collagen – subchondral bone turnover	Associated with subchondral bone turnover, JSN and osteophyte progression (124, 125).
CTX-I	Old type I collagen degraded by Cathepsin K degraded type I collagen	FNIH, CTX-I was associated with disease progression (126).
NTX	Cathepsin K degraded type I collagen	FNIH, NTX was associated with disease progression (126).
Osteocalcin	Bone formation	FNIH, osteocalcin was borderline associated with disease progression (126).
CARTILAGE		
ARGS /NITEGE	Aggrecanase mediated degradation of aggrecan.	Serum and urine ARGS associated with OA (127) and response to therapy in cartilage explants (128).
C2C	MMP-mediated degradation of type II collagen.	C2C concentrations were correlated with CTX-II, ARGS, osteocalcin, osteopontin and IL-8, but not structural joint injury by MRI (129).
C2M	MMP-mediated degradation of type II collagen.	C2M was associated with KL-2 score and levels of chronic inflammation (130).
C-Col10	Type X collagen turnover.	C-Col10 was elevated in patients with significant OA (131).
Coll2-1	Protease-mediated degradation of type II collagen.	Curcumin treatment reduced Coll-2-1 serum levels (132).
Coll2-1 -NO2	Protease-mediated degradation of nitrosylated type II collagen.	Baseline levels were negatively associated with incidence of knee OA (133).
COMP	Cartilage oligomeric matrix protein turnover/degradation.	CTX-II and COMP were related to progression of OA and (134).
CTX-II	Protease-mediated degradation of type II collagen.	CTX-II was associated with progression of OA(117), and diagnosis, and responded to therapy(135-137).
Fib3-1 / -2	Protease-mediated degradation of Fibulin 3.	Fib3-1, Fib3-2 and Fib3-3 were associated with incidence of clinical knee OA (138).
PRO-C2	The pro-peptide of type II collagen – cartilage formation.	PRO-C2 was induced by different treatments in <i>ex vivo</i> cultures, and predictive of structural progression (139, 140).
PIIANP	Type IIA collagen formation.	FNIH, PIIANP was associated with structural progression (126).
INFLAMMATION		
VICM	Macrophage activity – inflammation.	VICM was shown to be associated with radiographic progression of ankylosis spondylitis (141).
C1M & C3M	MMP mediated type I and III collagen. Inflammation mediated tissue degradation, associated with synovitis.	C1M was associated with radiographic progression in RA(142) and both were shown to be associated with synovitis (143) and respond to efficacious but not non-efficacious therapy (144).
MMP-3	Both total and active MMP-3 assays are available.	MMP-3 is highly produced by the inflamed synovium and response to anti-inflammatory treatments(145).
CRPM	During tissue inflammation, CRP produced in the liver, binds to inflammatory cells in the tissue and is metabolised into smaller fragments, of which one is CRPM (146).	CRPM was elevated under inflammatory condition in OA (130) and associated to progression, and response to anti-inflammatory treatments (147), predicting efficacy.
C4M	Basement membrane remodelling, associated with blood vessels.	Prognostic for progression in RA (148) and elevated in a range of inflammatory conditions (149).

space narrowing (117). These data have been confirmed in two other studies by Saberi *et al.* and Valdes *et al.*, who also showed a significant risk of having OA and of progression (134). In a large Japanese OA cohort, CTX-II was found to

be correlated with radiographic severity (117, 118, 151), as well as in the GARP study by Meulenbelt and colleagues to be directly associated with the number of skeletal sites (spine, hip, hands and knees) affected by radiographic OA

(152). Furthermore, there are several studies showing that CTX-II may act as a marker of response (110), and finally CTX-II levels have been shown to be affected by several different treatments (111).

Fig. 5. Biomarkers of different tissues (bone, cartilage and synovium), as well as different processes; inflammation, signalling and cell differentiation/ matrix production. Each biomarker may be used for a specific purpose. Modified and reproduced with permission from (184).



Aggrecanase degradation of aggrecan – ARGS

Aggrecan is the most abundant proteoglycan of the articular cartilage, and a significant amount of attention has been devoted to identification of pathophysiological relevant degradation fragments as well as developing antibodies and assay towards those (127, 153, 154). There is a suite of literature available on the “degradome” of aggrecan, of which some of these fragments may have more pathobiological relevance than others, and of which some may even have signalling capabilities (155). The ADAMTS-4/5 generated fragment with the N-terminus ARGS has received the most attention (156), and albeit an array of other aggrecan biomarkers are available which may prove useful in the future (11) ARGS has proven to be the most robust of the tested biomarkers. ARGS measured in synovial fluid has been shown to be associated with improvements in KOOS symptoms and pain in patients with anterior cruciate ligament (ACL) trauma with concomitant articular cartilage injuries (154, 157). The link between ARGS and pain has recently been suggested to be mediated through the TLR receptor pathway, in which a high induction of IL-6 leads to a vicious cycle of inflammation, protease production, joint destruction and more TLR activation (155, 158).

Two different ARGS assays (156, 159) have been reported, but further validation work is required to understand the pathophysiological relevance in OA, as well as upgrading their technical performance. Important for the current context, the ARGS assay has been applied in early drug development, in preclinical and clinical settings as well as human and bovine cartilage explants (121, 127, 160-162).

Cartilage oligomeric protein - COMP

COMP may well be the most used biomarker in the OA field, albeit with very varying data (163). For example, in a traumatic OA study no association was found between traumatic knee OA severity and concentrations of COMP (129, 164, 165), whereas a study in women showed that the highest levels of COMP were associated with increased risk of radiographic OA (166). The most used COMP assays do not discriminate between COMP degradation and turnover, and as COMP is controlling the fibrillar formation of collagens, a list of publications is arising in which COMP is associated with different collagen diseases, such as fibrosis of the lung and liver (167). Recently, a new COMP assay was presented which measures a specific COMP fragment – COMPneo (168, 169). Preliminary data showed that this biomarker was indeed

released from human articular cartilage when stimulated with catabolic factors, which warrants further investigations.

Cartilage formation

Biomarkers of cartilage formation are urgently needed, and a range of biomarkers are becoming available such as CS846 (139) and PIIANP (170), CPII (PIICP) (171) and PRO-C2 (172). PRO-C2 quantifies the propeptide of type IIB collagen and in contrast, PIIANP (171, 173), PRO-C2 reflects formation of adult form of type II collagen. Data are emerging, and latest from the FNIH, that these biomarkers may be valuable in finding progressors (126).

Synovitis

The synovium is becoming increasingly investigated in OA and has been debated to be part of an inflammatory endotype (122, 130). The synovium is the structure surrounding the joint cavity. It is composed of two resident cell types: fibroblast-like synoviocytes (FLS) and macrophage-like synoviocytes (174). Synovitis is the inflammatory condition, which clinically is manifested by local warmth, swelling and tenderness in joint diseases such as RA, OA and SpA (175). Synovitis involves expansion of resident synoviocytes, fibrosis and infiltration of mononuclear cells resulting in an enlarged synovium with

increased cell numbers. Synovitis may not be the initiator of the disease, but at some point may become the driver of disease (176, 177), and could constitute an endotype that would warrant a targeted therapy.

CRP is often used as a biomarker for inflammation, although it is a simple acute phase reactant produced in the liver, not conferring any tissue specificity (146). In addition, erythrocyte sedimentation rate (ESR), IL-1 β , IL-6, TNF- α and fibrinogen are also used as quantitative measurement of inflammation, however with limited success likely due to the non-joint related activity and production of these biomarkers, as well as an extreme variation, limiting the clinical applicability (175). As these simple inflammatory biomarkers do not reflect synovitis or joint tissue inflammation directly, another avenue to explore could be biomarkers associated with tissue turnover, possibly inflammation-driven. Some biomarkers associated with structural proteins are highly correlated with CRP; these include MMP-mediated type I and III collagen degradation markers C1M and C3M (175), and citrullinated and MMP-degraded vimentin (VICM) (147, 149, 178). C1M was associated with a phenotype of OA with high remodelling, KL radiographic grade, and high CRP and CRPM (130), and in other studies shown to be predictive of progression in RA (142), but most importantly, C1M was shown to provide dose resolution in response to anti-inflammatory treatment in RA (147), that was associated with clinical efficacy. These data combined may suggest that tissue inflammation biomarkers can provide the prognostic and efficacy of intervention capacities that standard CRP does not. When CRP is deposited in the tissue, it is metabolised by MMPs, resulting in smaller fragments of that protein (175). One such fragment is CRPM (146). CRPM, was recently shown to be associated with OA progression, with a small variation as compared to CRP.

Figure 5 reflects how several different biomarkers have been developed and used for OA, focusing on each of the three key tissues of the joint; bone, cartilage and synovium. How different

biomarkers are associated with these three tissues is illustrated in Figure 5, modified with permission from (180)

The biomarker consortia: beginning of success and a much-needed joint effort

Recently two biomarker consortia have been established and have begun to report data. The FNIH biomarker initiative in OA and APPROACH. Very importantly, the FNIH/OAI cohorts have identified a set of biomarkers to be associated with prognostic for structural progression, which included the biomarkers CTX-I, NTX, alpha-CTX-I, CTX-II and PIIANP (126). In addition, a concerted effort to identify reference range of these biomarkers, which is much needed in clinic research, has been completed (181). In addition, a second wave of research has now been undertaken in this consortium, in which clinical studies are used to validate these findings and better investigate the suite of biomarkers to be used in clinical studies. Such data are much awaited.

In addition, the APPROACH consortium is well underway with the exact mission of delivering non-invasive biomarkers for patient endotyping and drug development tools. These data are highly needed in the field, and yet to be publicly available.

The future

There is an imminent need for reliable biomarkers in the OA field. At least 3 different classes of biomarkers are urgently needed to support drug development and delineate the clinical trajectory of patients.

Efficacy

Biomarkers which at an early timepoint carry the promise and hope of clinical efficacy, such as an early delta change in CTX-I associated with increased BMD in the osteoporosis field.

Endotype

All OA patients may not respond to the same intervention, and there is a need for identification of the OA patients which are fast progressors and respond to a given treatment at the same time. A potential breakthrough was recently

found in the UK biobank with more than 3,500,000 patients analysed (186). Three genes associated with cartilage formation and repair were identified to be associated with OA. FGF-18, GDF-5 and TGF-beta (182). Interestingly, all these growth factors stimulate cartilage formation as measured by PRO-C2 in cartilage explants cultures (139, 183), and recently PRO-C2 was shown in 3 independent cohorts to be associated with structural disease progression. Whether this could be a low repair endotype which should receive additional attention, needs a further investigation.

Clinical trial enrichment

The landscape of clinical trial design in the OA field is changing. As OA has been acknowledged as a serious disease by the FDA the use of accelerated approval is now a possibility, much like the field of drug development to treat NASH and cancer has seen major advances. This means that biomarker enrichment for outcome which is either TJR or TKR, may be needed (184). Preliminary data was presented at the OARSI conference, in which CTX-II was prognostic for TJR in two combined phase III clinical studies. Additional data need to be presented.

References

1. CROSS M, SMITH E, HOY D *et al.*: The global burden of hip and knee osteoarthritis: estimates from the global burden of disease 2010 study. *Ann Rheum Dis* 2014; 73:1323-30.
2. LOTZ M, MARTEL-PELLETIER J, CHRISTIANSEN C *et al.*: Value of biomarkers in osteoarthritis: current status and perspectives. *Ann Rheum Dis* 2013; 72: 1756-63.
3. SIEBUHR AS, HE Y, GUDMANN NS *et al.*: Biomarkers of cartilage and surrounding joint tissue. *Biomark Med* 2014; 8: 713-31.
4. KARSDAL MA, BAY-JENSEN AC, LORIES RJ *et al.*: The coupling of bone and cartilage turnover in osteoarthritis: Opportunities for bone antiresorptives and anabolics as potential treatments? *Ann Rheum Dis* 2014; 73.
5. KARSDAL MA, MICHAELIS M, LADEL C *et al.*: Disease-modifying treatments for osteoarthritis (DMOADs) of the knee and hip: lessons learned from failures and opportunities for the future. *Osteoarthritis Cartilage* 2016; 24: 2013-21.
6. KARSDAL MA, QVIST P, CHRISTIANSEN C *et al.*: Optimising antiresorptive therapies in postmenopausal women: why do we need to give due consideration to the degree of suppression? *Drugs* 2006; 66: 1909-18.
7. KARSDAL MA, CHRISTIANSEN C, LADEL C *et al.*: Osteoarthritis--a case for personalized

- health care? *Osteoarthritis Cartilage* 2014; 22: 7-16.
8. KARS DAL MA, BAY-JENSEN A-C, HENRIKSEN K *et al.*: Rheumatoid arthritis: a case for personalized health care? *Arthritis Care Res* (Hoboken) 2014; 66: 1273-80.
9. GARNERO P, FERRERAS M, KARS DAL MA *et al.*: The type I collagen fragments ICTP and CTX reveal distinct enzymatic pathways of bone collagen degradation. *J Bone Miner Res* 2003;18.
10. HENRIKSEN K, TANKO LB, QVIST P *et al.*: Assessment of osteoclast number and function: Application in the development of new and improved treatment modalities for bone diseases. *Osteoporos Int* 2007;18: 681-85
11. KARS DAL MA, HENRIKSEN K, LEEMING DJ *et al.*: Biochemical markers and the FDA critical Path: How biomarkers may contribute to the understanding of pathophysiology and provide unique and necessary tools for drug development. *Biomarkers* 2009; 14: 181-201.
12. HENRIKSEN K, LEEMING DJ, CHRISTIANSEN C *et al.*: Use of bone turnover markers in clinical osteoporosis assessment in women: Current issues and future options. *Womens Health* 2011; 7: 689-98.
13. HENRIKSEN K, BOHREN KM, BAY-JENSEN AC *et al.*: Should biochemical markers of bone turnover be considered standard practice for safety pharmacology? *Biomarkers* 2010; 15: 195-204.
14. KARS DAL MA, SCHETT G, EMERY P *et al.*: IL-6 receptor inhibition positively modulates bone balance in rheumatoid arthritis patients with an inadequate response to anti-tumor necrosis factor therapy: biochemical marker analysis of bone metabolism in the tocilizumab RADIATE study (NCT00106522). *Semin Arthritis Rheum* 2012;42:131-39.
15. HENRIKSEN K, KARS DAL MA, MARTIN TJ: Osteoclast-derived coupling factors in bone remodeling. *Calcif Tissue Int* 2014; 94: 88-97.
16. KARS DAL MA, MARTIN TJ, BOLLERSLEV J *et al.*: Are nonresorbing osteoclasts sources of bone anabolic activity? *J Bone Miner Res* 2007; 22: 487-94.
17. HENRIKSEN K, BOLLERSLEV J, EVERTS V *et al.*: Osteoclast activity and subtypes as a function of physiology and pathology—implications for future treatments of osteoporosis. *Endocr Rev* 2011; 32: 31-63.
18. IKI M, AKIBA T, MATSUMOTO T *et al.*: Reference database of biochemical markers of bone turnover for the Japanese female population. Japanese Population-based Osteoporosis (JPOS) Study. *Osteoporos Int* 2004; 15: 981-91.
19. BAY-JENSEN A-C, THUDIUM CS, GUALILLO O *et al.*: Biochemical marker discovery, testing and evaluation for facilitating OA drug discovery and development. *Drug Discov Today* 2018; 23: 349-58.
20. CONNELLY JJ, CHEREPANOVA OA, DOSS JF *et al.*: Epigenetic regulation of COL15A1 in smooth muscle cell replicative aging and atherosclerosis. *Hum Mol Genet* 2013; 22: 5107-20.
21. KRAUS VB, BLANCO FJ, ENGLUND M *et al.*: Call for standardized definitions of osteoarthritis and risk stratification for clinical trials and clinical use. *Osteoarthritis Cartilage* 2015; 23.
22. KARS DAL MA, HENRIKSEN K, LEEMING DJ *et al.*: Novel combinations of Post-Translational Modification (PTM) neo-epitopes provide tissue-specific biochemical markers—are they the cause or the consequence of the disease? *Clin Biochem* 2010; 43: 793-804.
23. KARS DAL MA, WOODWORTH T, HENRIKSEN K *et al.*: Biochemical markers of ongoing joint damage in rheumatoid arthritis—current and future applications, limitations and opportunities. *Arthritis Res Ther* 2011;13: 215.
24. HENRIKSEN K, O'BRYANT SE, HAMPEL H *et al.*: The future of blood-based biomarkers for Alzheimer's disease. *Alzheimers Dement* 2014; 10: 115-31.
25. BAUER DC, HUNTER DJ, ABRAMSON SB *et al.*: Classification of osteoarthritis biomarkers: a proposed approach. *Osteoarthritis Cartilage* 2006; 14: 723-27.
26. Group F-NBW. BEST (Biomarkers, Endpoints, and other Tools) Resource. Food and Drug Administration (US) 2016.
27. HENRIKSEN K, NEUTZSKY-WULFF A V, BONEWALD LF *et al.*: Local communication on and within bone controls bone remodeling. *Bone* 2009; 44: 1026-33.
28. SEEMAN E, DELMAS PD: Bone quality – the material and structural basis of bone strength and fragility. *N Engl J Med* 2006; 354: 2250-61.
29. ARCEO-MENDOZA RM, CAMACHO P: Prediction of fracture risk in patients with osteoporosis: a brief review. *Womens Health* (Lond Engl) 2015; 11: 477-82; quiz 483-4.
30. MILLER PD, SIRIS ES, BARRETT-CONNOR E *et al.*: Prediction of fracture risk in postmenopausal white women with peripheral bone densitometry: evidence from the National Osteoporosis Risk Assessment. *J Bone Miner Res* 2002; 17: 2222-30.
31. SCHUIT SCE, VAN DER KLIFT M, WEELAEAM *et al.*: Fracture incidence and association with bone mineral density in elderly men and women: the Rotterdam Study. *Bone* 2004; 34: 195-202.
32. KANIS JA, HARVEY NC, JOHANSSON H *et al.*: FRAX Update. *J Clin Densitom* 2017; 20: 360-67.
33. HILLIER TA, CAULEY JA, RIZZO JH *et al.*: WHO absolute fracture risk models (FRAX): do clinical risk factors improve fracture prediction in older women without osteoporosis? *J Bone Miner Res* 2011; 26: 1774-82.
34. MCCLOSKEY E V, VASIKARAN S, COOPER C *et al.*: Official Positions for FRAX® clinical regarding biochemical markers from Joint Official Positions Development Conference of the International Society for Clinical Densitometry and International Osteoporosis Foundation on FRAX®. *J Clin Densitom* 2011; 14: 220-22.
35. VASIKARAN S, EASTELL R, BRUYÈRE O *et al.*: Markers of bone turnover for the prediction of fracture risk and monitoring of osteoporosis treatment: a need for international reference standards. *Osteoporos Int* 2010; 22: 391-420.
36. DANIELS SJ, LEEMING DJ, ESLAM M *et al.*: ADAPT: An algorithm incorporating PRO-C3 accurately identifies patients with NAFLD and advanced fibrosis. *Hepatology* 2019; 69: 1075-86.
37. VIGUET-CARRIN S, GARNERO P, DELMAS PD: The role of collagen in bone strength. *Osteoporos Int* 2006; 17: 319-36.
38. CHRISTIANSEN C, RIIS BJ, RØDBRO P: Prediction of rapid bone loss in postmenopausal women. *Lancet* 1987; 1: 1105-8.
39. CHRISTIANSEN C: Selection of postmenopausal women for estrogen therapy. *Postgrad Med* 1989; Spec no:10-2; discussion 33-43.
40. RIIS BJ, HANSEN MA, JENSEN AM *et al.*: Low bone mass and fast rate of bone loss at menopause: equal risk factors for future fracture: a 15-year follow-up study. *Bone* 1996; 19: 9-12.
41. HENRIKSEN K, CHRISTIANSEN C, KARS DAL MA: Role of biochemical markers in the management of osteoporosis. *Climacteric* 2015; 18 (Suppl. 2): 10-8.
42. SONDERGAARD BC, HENRIKSEN K, WULF H *et al.*: Relative contribution of matrix metalloproteinase and cysteine protease activities to cytokine-stimulated articular cartilage degradation. *Osteoarthritis Cartilage* 2006;14: 738-48
43. RAVN P, CLEMMESSEN B, CHRISTIANSEN C: Biochemical markers can predict the response in bone mass during alendronate treatment in early postmenopausal women. Alendronate Osteoporosis Prevention Study Group. *Bone* 1999; 24: 237-44.
44. RAVN P, HOSKING D, THOMPSON D *et al.*: Monitoring of alendronate treatment and prediction of effect on bone mass by biochemical markers in the early postmenopausal intervention cohort study. *J Clin Endocrinol Metab* 1999; 84: 2363-68.
45. CLOOS PAC, FLEDELIUS C: Collagen fragments in urine derived from bone resorption are highly racemized and isomerized: a biological clock of protein aging with clinical potential. *Biochem J* 2000;345:473.
46. GRUNTMANIS U, FORDAN S, GHAYEE HK *et al.*: The peroxisome proliferator-activated receptor-gamma agonist rosiglitazone increases bone resorption in women with type 2 diabetes: a randomized, controlled trial. *Calcif Tissue Int* 2010; 86: 343-49.
47. HENRIKSEN K, BYRJALSEN I, QVIST P *et al.*: Efficacy and safety of the PPARγ partial agonist balaglitazone compared with pioglitazone and placebo: a phase III, randomized, parallel-group study in patients with type 2 diabetes on stable insulin therapy. *Diabetes Metab Res Rev* 2011; 27: 392-401.
48. OKABE R, INABA M, NAKATSUKA K *et al.*: Significance of serum CrossLaps as a predictor of changes in bone mineral density during estrogen replacement therapy; comparison with serum carboxyterminal telopeptide of type I collagen and urinary deoxypyridinoline. *J Bone Miner Metab* 2004; 22: 127-31.
49. RISSANEN JP, SUOMINEN MI, PENG Z *et al.*: Secreted tartrate-resistant acid phosphatase 5b is a Marker of osteoclast number in human osteoclast cultures and the rat ovariectomy model. *Calcif Tissue Int* 2008; 82: 108-15.

50. RISSANEN JP, YLIPAHKALA H, FAGERLUND KM *et al.*: Improved methods for testing antiresorptive compounds in human osteoclast cultures. *J Bone Miner Metab* 2009; 27: 105-9.
51. MUÑOZ-TORRES M, REYES-GARCÍA R, MEZQUITA-RAYA P *et al.*: Serum cathepsin K as a marker of bone metabolism in postmenopausal women treated with alendronate. *Maturitas* 2009; 64: 188-92.
52. LEEMING DJ, ALEXANDERSEN P, KARS DAL MA *et al.*: An update on biomarkers of bone turnover and their utility in biomedical research and clinical practice. *Eur J Clin Pharmacol* 2006; 62: 781-92.
53. FINDLAY DM, ATKINS GJ: Relationship between serum RANKL and RANKL in bone. *Osteoporos Int* 2011; 22: 2597-602.
54. LEEMING DJ, HE Y, VEIDAL SS *et al.*: A novel marker for assessment of liver matrix remodeling: An enzyme-linked immunosorbent assay (ELISA) detecting a MMP generated type I collagen neo-epitope (C1M). *Biomarkers* 2011; 16: 616-28.
55. CEJKA D, JÄGER-LANSKY A, KIEWEG H *et al.*: Sclerostin serum levels correlate positively with bone mineral density and microarchitecture in haemodialysis patients. *Nephrol Dial Transplant* 2012; 27: 226-30.
56. MÖDDER UI, HOEY KA, AMIN S *et al.*: Relation of age, gender, and bone mass to circulating sclerostin levels in women and men. *J Bone Miner Res* 2011; 26: 373-79.
57. DRAKE MT, SRINIVASAN B, MÖDDER UI *et al.*: Effects of parathyroid hormone treatment on circulating sclerostin levels in postmenopausal women. *J Clin Endocrinol Metab* 2010; 95: 5056-62.
58. MÖDDER U IL, CLOWES JA, HOEY K *et al.*: Regulation of circulating sclerostin levels by sex steroids in women and in men. *J Bone Miner Res* 2011; 26: 27-34.
59. BUTLER JS, MURRAY DW, HURSON CJ *et al.*: The role of Dkk1 in bone mass regulation: correlating serum Dkk1 expression with bone mineral density. *J Orthop Res* 2011; 29: 414-8.
60. BAUER DC, SKLARIN PM, STONE KL *et al.*: Biochemical markers of bone turnover and prediction of hip bone loss in older women: the study of osteoporotic fractures. *J Bone Miner Res* 1999; 14: 1404-10.
61. STEPAN JJ: Prediction of bone loss in postmenopausal women. *Osteoporos Int* 2000; 11 (Suppl. 6): S45-54.
62. DELMAS PD, EASTELL R, GARNERO P *et al.*: The use of biochemical markers of bone turnover in osteoporosis. Committee of Scientific Advisors of the International Osteoporosis Foundation. *Osteoporos Int* 2000; 11 (Suppl. 6): S2-17.
63. SORNAY-RENDU E, MUNOZ F, GARNERO P *et al.*: Identification of osteopenic women at high risk of fracture: the OFELY study. *J Bone Miner Res* 2005; 20: 1813-39.
64. GARNERO P, SORNAY-RENDU E, CLAUS-TRAT B *et al.*: Biochemical markers of bone turnover, endogenous hormones and the risk of fractures in postmenopausal women: the OFELY study. *J Bone Miner Res* 2000; 15: 1526-36.
65. GARNERO P, HAUSHERR E, CHAPUY MC *et al.*: Markers of bone resorption predict hip fracture in elderly women: the EPIDOS Prospective Study. *J Bone Miner Res* 1996; 11: 1531-38.
66. GARNERO P, DARGENT-MOLINA P, HANS D *et al.*: Do markers of bone resorption add to bone mineral density and ultrasonographic heel measurement for the prediction of hip fracture in elderly women? The EPIDOS prospective study. *Osteoporos Int* 1998; 8: 563-69.
67. BRUYERE O, COLLETTE J, DELMAS P *et al.*: Interest of biochemical markers of bone turnover for long-term prediction of new vertebral fracture in postmenopausal osteoporotic women. *Maturitas* 2003; 44: 259-65.
68. GERDHEM P, IVASKA KK, ALATALO SL *et al.*: Biochemical markers of bone metabolism and prediction of fracture in elderly women. *J Bone Miner Res* 2004; 19: 386-93.
69. REID I, EISMAN J, BONE H *et al.*: Odanacatib in the treatment of postmenopausal women with low bone mineral density: 3-year continued therapy and resolution of effect. *Bone* 2010; 46: S31-32.
70. EISMAN JA, BONE HG, HOSKING DJ *et al.*: Odanacatib in the treatment of postmenopausal women with low bone mineral density: three-year continued therapy and resolution of effect. *J Bone Miner Res* 2011; 26: 242-51.
71. BONE HG, MCCLUNG MR, ROUX C *et al.*: Odanacatib, a cathepsin-K inhibitor for osteoporosis: a two-year study in postmenopausal women with low bone density. *J Bone Miner Res* 2010; 25: 937-47.
72. CUMMINGS SR, SAN MARTIN J, MCCLUNG MR *et al.*: Denosumab for prevention of fractures in postmenopausal women with osteoporosis. *N Engl J Med* 2009; 361: 756-65.
73. MILLER PD, BOLOGNESE MA, LEWIECKI EM *et al.*: Effect of denosumab on bone density and turnover in postmenopausal women with low bone mass after long-term continued, discontinued, and restarting of therapy: a randomized blinded phase 2 clinical trial. *Bone* 2008; 43: 222-29.
74. ARLOT M, MEUNIER PJ, BOIVIN G *et al.*: Differential effects of teriparatide and alendronate on bone remodeling in postmenopausal women assessed by histomorphometric parameters. *J Bone Miner Res* 2005; 20: 1244-53.
75. GLOVER SJ, EASTELL R, MCCLOSKEY EV *et al.*: Rapid and robust response of biochemical markers of bone formation to teriparatide therapy. *Bone* 2009; 45: 1053-58.
76. PADHI D, JANG G, STOUCH B *et al.*: Single-dose, placebo-controlled, randomized study of AMG 785, a sclerostin monoclonal antibody. *J Bone Miner Res* 2011; 26: 19-26.
77. MCCLUNG MR, BROWN JP, DIEZ-PEREZ A *et al.*: Effects of 24 months of treatment with romosozumab followed by 12 months of denosumab or placebo in postmenopausal women with low bone mineral density: a randomized, double-blind, phase 2, parallel group study. *J Bone Miner Res* 2018; 33: 1397-406.
78. COSMAN F, MILLER PD, WILLIAMS GC *et al.*: Eighteen months of treatment with subcutaneous abaloparatide followed by 6 months of treatment with alendronate in postmenopausal women with osteoporosis: Results of the ACTIVExtend Trial. *Mayo Clin Proc* 2017; 92: 200-10.
79. KARS DAL MA, BYRJALSEN I, HENRIKSEN K *et al.*: A pharmacokinetic and pharmacodynamic comparison of synthetic and recombinant oral salmon calcitonin. *J Clin Pharmacol* 2009; 49: 229-34.
80. KARS DAL MA, BYRJALSEN I, RIIS BJ *et al.*: Investigation of the diurnal variation in bone resorption for optimal drug delivery and efficacy in osteoporosis with oral calcitonin. *BMC Clin Pharmacol* 2008; 8: 12.
81. SCHALLER S, HENRIKSEN K, SVEIGAARD C *et al.*: The Chloride Channel Inhibitor NS3736 prevents bone resorption in ovariectomized rats without changing bone formation. *J Bone Miner Res* 2004; 19: 1144-53.
82. CHESNUT CH, SILVERMAN S, ANDRIANO K *et al.*: A randomized trial of nasal spray salmon calcitonin in postmenopausal women with established osteoporosis: the prevent recurrence of osteoporotic fractures study. PROOF Study Group. *Am J Med* 2000; 109: 267-76.
83. HENRIKSEN K, BYRJALSEN I, ANDERSEN JR *et al.*: A randomized, double-blind, multicenter, placebo-controlled study to evaluate the efficacy and safety of oral salmon calcitonin in the treatment of osteoporosis in postmenopausal women taking calcium and vitamin D. *Bone* 2016; 91: 122-29.
84. TANKÓ LB, BAGGER YZ, ALEXANDERSEN P *et al.*: Safety and efficacy of a novel salmon calcitonin (sCT) technology-based oral formulation in healthy postmenopausal women: acute and 3-month effects on biomarkers of bone turnover. *J Bone Miner Res* 2004; 19: 1531-38.
85. HANNON RA, CLOWES JA, EAGLETON AC *et al.*: Clinical performance of immunoreactive tartrate-resistant acid phosphatase isoform 5b as a marker of bone resorption. *Bone* 2004; 34: 187-94.
86. ROSEN CJ, HOCHBERG MC, BONNICK SL *et al.*: Treatment with once-weekly alendronate 70 mg compared with once-weekly risendronate 35 mg in women with postmenopausal osteoporosis: a randomized double-blind study. *J Bone Miner Res* 2005; 20: 141-51.
87. EMKEY R, DELMAS PD, BOLOGNESE M *et al.*: Efficacy and tolerability of once-monthly oral ibandronate (150 mg) and once-weekly oral alendronate (70 mg): additional results from the Monthly Oral Therapy with Ibandronate for Osteoporosis Intervention (MO-TION) study. *Clin Ther* 2009; 31: 751-61.
88. DELMAS PD, RECKER RR, CHESNUT CH *et al.*: Daily and intermittent oral ibandronate normalize bone turnover and provide significant reduction in vertebral fracture risk: results from the BONE study. *Osteoporos Int* 2004; 15: 792-28.
89. DELMAS PD, ADAMI S, STRUGALA C *et al.*: Intravenous ibandronate injections in postmenopausal women with osteoporosis: one-year results from the dosing intravenous administration study. *Arthritis Rheum* 2006; 54: 1838-46.
90. BLACK DM, DELMAS PD, EASTELL R *et al.*: Once-yearly zoledronic acid for treatment

- of postmenopausal osteoporosis. *N Engl J Med* 2007; 356: 1809-22.
91. PEREDA CA, HANNON RA, NAYLOR KE *et al.*: The impact of subcutaneous oestradiol implants on biochemical markers of bone turnover and bone mineral density in postmenopausal women. *BJOG* 2002; 109: 812-20.
92. MEUNIER PJ, VIGNOT E, GARNERO P *et al.*: Treatment of postmenopausal women with osteoporosis or low bone density with raloxifene. Raloxifene Study Group. *Osteoporos Int* 1999; 10: 330-36.
93. ETTINGER B, BLACK DM, MITLAK BH *et al.*: Reduction of vertebral fracture risk in postmenopausal women with osteoporosis treated with raloxifene: results from a 3-year randomized clinical trial. Multiple Outcomes of Raloxifene Evaluation (MORE) Investigators. *JAMA* 1999; 282: 637-45.
94. MEUNIER PJ, SLOSMAN DO, DELMAS PD *et al.*: Strontium ranelate: dose-dependent effects in established postmenopausal vertebral osteoporosis—a 2-year randomized placebo controlled trial. *J Clin Endocrinol Metab* 2002; 87: 2060-66.
95. QUESADA-GÓMEZ JM, MUSCHITZ C, GÓMEZ-REINO J *et al.*: The effect of PTH(1-84) or strontium ranelate on bone formation markers in postmenopausal women with primary osteoporosis: results of a randomized, open-label clinical trial. *Osteoporos Int* 2011; 22: 2529-37.
96. EASTELL R, NAGASE S, SMALL M *et al.*: Effect of ONO-5334 on bone mineral density and biochemical markers of bone turnover in postmenopausal osteoporosis: 2-year results from the OCEAN study. *J Bone Miner Res* 2014; 29: 458-66.
97. BLACK DM, BILEZIKIAN JP, ENSRUD KE *et al.*: One year of alendronate after one year of parathyroid hormone (1-84) for osteoporosis. *N Engl J Med* 2005; 353: 555-65.
98. BLACK DM, BOUXSEIN ML, PALERMO L *et al.*: Randomized trial of once-weekly parathyroid hormone (1-84) on bone mineral density and remodeling. *J Clin Endocrinol Metab* 2008; 93: 2166-72.
99. DOVIO A, PERAZZOLO L, OSELLA G *et al.*: Immediate fall of bone formation and transient increase of bone resorption in the course of high-dose, short-term glucocorticoid therapy in young patients with multiple sclerosis. *J Clin Endocrinol Metab* 2004; 89: 4923-28.
100. GREY A, BOLLAND M, GAMBLE G *et al.*: The peroxisome proliferator-activated receptor-gamma agonist rosiglitazone decreases bone formation and bone mineral density in healthy postmenopausal women: a randomized, controlled trial. *J Clin Endocrinol Metab* 2007; 92: 1305-10.
101. STELLBRINK H-J, ORKIN C, ARRIBAS JR *et al.*: Comparison of changes in bone density and turnover with abacavir-lamivudine versus tenofovir-emtricitabine in HIV-infected adults: 48-week results from the ASSERT study. *Clin Infect Dis* 2010; 51: 963-72.
102. MOTYL KJ, DICK-DE-PAULA I, MALONEY AE *et al.*: Trabecular bone loss after administration of the second-generation antipsychotic risperidone is independent of weight gain. *Bone* 2012; 50: 490-98.
103. BILEZIKIAN JP, WATTS NB, USISKIN K *et al.*: Evaluation of bone mineral density and bone biomarkers in patients with type 2 diabetes treated with canagliflozin. *J Clin Endocrinol Metab* 2016; 101: 44-51.
104. WATTS NB, BILEZIKIAN JP, USISKIN K *et al.*: Effects of canagliflozin on fracture risk in patients with type 2 diabetes mellitus. *J Clin Endocrinol Metab* 2016; 101: 157-66.
105. CHARLES ED, NEUSCHWANDER-TETRI BA, PABLO FRIAS J *et al.*: Pegbelfermin (BMS-986036), PEGylated FGF21, in patients with obesity and type 2 diabetes: results from a randomized phase 2 study. *Obesity* (Silver Spring) 2019; 27: 41-49.
106. BILEZIKIAN JP, JOSSE RG, EASTELL R *et al.*: Rosiglitazone decreases bone mineral density and increases bone turnover in postmenopausal women with type 2 diabetes mellitus. *J Clin Endocrinol Metab* 2013; 98: 1519-28.
107. KIM AM, SOMAYAJI VR, DONG JQ *et al.*: Once-weekly administration of a long-acting fibroblast growth factor 21 analogue modulates lipids, bone turnover markers, blood pressure and body weight differently in obese people with hypertriglyceridaemia and in non-human primates. *Diabetes Obes Metab* 2017; 19: 1762-72.
108. SANYAL A, CHARLES ED, NEUSCHWANDER-TETRI BA *et al.*: Pegbelfermin (BMS-986036), a PEGylated fibroblast growth factor 21 analogue, in patients with non-alcoholic steatohepatitis: a randomised, double-blind, placebo-controlled, phase 2a trial. *Lancet* 2019; 392: 2705-17.
109. HANNON R, EASTELL R: Preanalytical variability of biochemical markers of bone turnover. *Osteoporos Int* 2000; 11 (Suppl. 6): S30-44.
110. KARS DAL M, BYRJALSEN I, BAY-JENSEN A *et al.*: Biochemical markers identify influences on bone and cartilage degradation in osteoarthritis - The effect of sex, Kellgren-Lawrence (KL) score, Body Mass Index (BMI), oral salmon calcitonin (sCT) treatment and diurnal variation. *BMC Musculoskelet Disord* 2010; 11: 125.
111. KARS DAL MA, BYRJALSEN I, HENRIKSEN K *et al.*: The effect of oral salmon calcitonin delivered with 5-CNAC on bone and cartilage degradation in osteoarthritic patients: a 14-day randomized study. *Osteoarthritis Cartilage* 2010; 18: 150-59.
112. QVIST P, CHRISTGAU S, PEDERSEN BJ *et al.*: Circadian variation in the serum concentration of C-terminal telopeptide of type I collagen (serum CTx): effects of gender, age, menopausal status, posture, daylight, serum cortisol, and fasting. *Bone* 2002; 31: 57-61.
113. FELS ON D, NIU J, SACK B *et al.*: Progression of osteoarthritis as a state of inertia. *Ann Rheum Dis* 2013; 72: 924-29.
114. BIHLET AR, KARS DAL MA, BAY-JENSEN A-C *et al.*: Clinical drug development using dynamic biomarkers to enable personalized health care in COPD. *Chest* 2015; 148: 16-23.
115. KARS DAL MA, HENRIKSEN K, LEEMING DJ *et al.*: Biochemical markers and the FDA Critical Path: how biomarkers may contribute to the understanding of pathophysiology and provide unique and necessary tools for drug development. *Biomarkers* 2009; 14: 181-202.
116. KARS DAL MA, BAY-JENSEN AC, LEEMING DJ *et al.*: Quantification of 'end products' of tissue destruction in inflammation may reflect convergence of cytokine and signaling pathways-implications for modern clinical chemistry. *Biomarkers* 2013;18.
117. REIJMAN M, HAZES JM, BIERMA-ZEINSTRAS M *et al.*: A new marker for osteoarthritis: cross-sectional and longitudinal approach. *Arthritis Rheum* 2004; 50: 2471-78.
118. DAM EB, LOOG M, CHRISTIANSEN C *et al.*: Identification of progressors in osteoarthritis by combining biochemical and MRI-based markers. *Arthritis Res Ther* 2009; 11: R115.
119. LUO Y, SINKEVICIUTE D, HE Y *et al.*: The minor collagens in articular cartilage. *Protein Cell* 2017; 8: 560-72.
120. EYRE DR: The collagens of articular cartilage. *Smith Arthritis Rheum* 1991; 21: 2-11.
121. KARS DAL MA, MADSEN SH, CHRISTIANSEN C *et al.*: Cartilage degradation is fully reversible in the presence of aggrecanase but not matrix metalloproteinase activity. *Arthritis Res Ther* 2008; 10: R23.
122. KARS DAL MA, BAY-JENSEN AC, HENRIKSEN K *et al.*: The pathogenesis of osteoarthritis involves bone, cartilage and synovial inflammation: May estrogen be a magic bullet? *Menopause Int* 2012; 18: 139-46.
123. BAY-JENSEN AC, REKER D, KJELGAARD-PETERSEN CF *et al.*: Osteoarthritis year in review 2015: Soluble biomarkers and the BIPEd criteria. *Osteoarthritis Cartilage* 2016; 24: 9-20.
124. HUEBNER JL, BAY-JENSEN AC, HUFFMAN KM *et al.*: Alpha C-telopeptide of type I collagen is associated with subchondral bone turnover and predicts progression of joint space narrowing and osteophytes in osteoarthritis. *Arthritis Rheumatol* (Hoboken, NJ) 2014; 66: 2440-49.
125. KRAUS VB, COLLINS JE, HARGROVE D *et al.*: Predictive validity of biochemical biomarkers in knee osteoarthritis: data from the FNIH OA Biomarkers Consortium. *Ann Rheum Dis* 2017; 76: 186-95.
126. KRAUS VB, COLLINS JE, HARGROVE D *et al.*: Predictive validity of biochemical biomarkers in knee osteoarthritis: data from the FNIH OA Biomarkers Consortium. *Ann Rheum Dis* 2017; 76: 186-95.
127. GERMASCHESKI FM, MATHENY CJ, LARKIN J *et al.*: Quantitation OF ARGS aggrecan fragments in synovial fluid, serum and urine from osteoarthritis patients. *Osteoarthritis Cartilage* 2014; 22: 690-97.
128. KARS DAL MA, SUMER EU, WULF H *et al.*: Induction of increased cAMP levels in articular chondrocytes blocks matrix metalloproteinase - Mediated cartilage degradation, but not aggrecanase-mediated cartilage degradation. *Arthritis Rheum* 2007; 56: 1549-58.
129. KUMAHASHI N, SWARD P, LARSSON S *et al.*: Type II collagen C2C epitope in human synovial fluid and serum after knee injury - associations with molecular and structural markers of injury. *Osteoarthritis Cartilage* 2015; 23: 1506-12.

130. SIEBUHR AS, PETERSEN KK, ARENDT-NIELSEN L *et al.*: Identification and characterisation of osteoarthritis patients with inflammation derived tissue turnover. *Osteoarthritis Cartilage* 2014; 22: 44-50.
131. HE Y, SIEBUHR AS, BRANDT-HANSEN NU *et al.*: Type X collagen levels are elevated in serum from human osteoarthritis patients and associated with biomarkers of cartilage degradation and inflammation. *BMC Musculoskelet Disord* 2014; 15: 309.
132. HENROTIN Y, GHARBI M, DIERCKXSENS Y *et al.*: Decrease of a specific biomarker of collagen degradation in osteoarthritis, COL2-1, by treatment with highly bioavailable curcumin during an exploratory clinical trial. *BMC Complement Altern Med* 2014; 14: 159.
133. LANDSMEER ML, RUNHAAR J, HENROTIN YE *et al.*: Association of urinary biomarker COL2-1NO with incident clinical and radiographic knee OA in overweight and obese women. *Osteoarthritis Cartilage* 2015; 23: 1398-404.
134. SABERI HOSNIJEH F, SIEBUHR AS, UITTERLINDEN AG *et al.*: Association between biomarkers of tissue inflammation and progression of osteoarthritis: Evidence from the Rotterdam study cohort. *Arthritis Res Ther* 2016; 18: 81.
135. KARSDAL MA, BYRJALSEN I, ALEXANDERSEN P *et al.*: Treatment of symptomatic knee osteoarthritis with oral salmon calcitonin: Results from two phase 3 trials. *Osteoarthritis Cartilage* 2015; 23: 532-43.
136. KARSDAL MA, BYRJALSEN I, LEEMING DJ *et al.*: The effects of oral calcitonin on bone collagen maturation: Implications for bone turnover and quality. *Osteoporos Int* 2008;19: 1355-61.
137. KARSDAL MA, BYRJALSEN I, HENRIKSEN K *et al.*: A pharmacokinetic and pharmacodynamic comparison of synthetic and recombinant oral salmon calcitonin. *J Clin Pharmacol* 2009; 49.
138. RUNHAAR J, SANCHEZ C, TARALLA S *et al.*: Fibulin-3 fragments are prognostic biomarkers of osteoarthritis incidence in overweight and obese women. *Osteoarthritis Cartilage* 2016 ;24: 672-78.
139. REKER D, KJELGAARD-PETERSEN CF, SIEBUHR AS *et al.*: Sprifermin (rhFGF18) modulates extracellular matrix turnover in cartilage explants ex vivo. *J Transl Med* 2017; 15: 250.
140. GUDMANN NS, WANG J, HOIELT S *et al.*: Cartilage turnover reflected by metabolic processing of type II collagen: A novel marker of anabolic function in chondrocytes. *Int J Mol Sci* 2014; 15: 18789-803.
141. BAY-JENSEN AC, KARSDAL MA, VASSILIADIS E *et al.*: Circulating citrullinated vimentin fragments reflect disease burden in ankylosing spondylitis and have prognostic capacity for radiographic progression. *Arthritis Rheum* 2013; 65: 972-80.
142. SIEBUHR AS, BAY-JENSEN AC, LEEMING DJ *et al.*: Serological identification of fast progressors of structural damage with rheumatoid arthritis. *Arthritis Res Ther* 2013; 15: R86.
143. KJELGAARD-PETERSEN CF, PLATT A, BRADDOCK M *et al.*: Translational biomarkers and ex vivo models of joint tissues as a tool for drug development in rheumatoid arthritis. *Arthritis Rheumatol* 2018; 70: 1419-28.
144. BAY-JENSEN AC, PLATT A, BYRJALSEN *et al.*: Effect of tocilizumab combined with methotrexate on circulating biomarkers of synovium, cartilage, and bone in the LITHE study. *Semin Arthritis Rheum* 2014; 43: 470-78.
145. SIEBUHR AS, KJELGAARD-PETERSEN CF, SUN S *et al.*: Suppression of active, but not total MMP-3, is associated with treatment response in a phase III clinical study of rheumatoid arthritis. *Clin Exp Rheumatol* 2018; 36: 94-101.
146. SKJØT-ARKIL H, SCHETT G, ZHANG C *et al.*: Investigation of two novel biochemical markers of inflammation, matrix metalloproteinase and cathepsin generated fragments of C-reactive protein, in patients with ankylosing spondylitis. *Clin Exp Rheumatol* 2012; 30: 371-79.
147. BAY-JENSEN AC, PLATT A, SIEBUHR AS *et al.*: Early changes in blood-based joint tissue destruction biomarkers are predictive of response to tocilizumab in the LITHE study. *Arthritis Res Ther* 2016; 18: 13.
148. GUDMANN NS, JUNKER P, JUHL P *et al.*: Type IV collagen metabolism is associated with disease activity, radiographic progression and response to tocilizumab in rheumatoid arthritis. *Clin Exp Rheumatol* 2018; 36: 829-35.
149. MORTENSEN JH, GODSKESSEN LE, JENSEN MD *et al.*: Fragments of citrullinated and MMP-degraded vimentin and MMP-degraded type III collagen are novel serological biomarkers to differentiate crohn's disease from ulcerative colitis. *J Crohns Colitis* 2015; 9: 863-72.
150. LEEMING DJ, BAY-JENSEN AC, VASSILIADIS E *et al.*: Post-translational modifications of the extracellular matrix are key events in cancer progression: Opportunities for biochemical marker development. *Biomarkers* 2011; 16: 193-205.
151. DAM EB, BYRJALSEN I, KARSDAL MA *et al.*: Increased urinary excretion of C-telopeptides of type II collagen (CTX-II) predicts cartilage loss over 21 months by MRI. *Osteoarthritis Cartilage* 2009; 17: 384-89.
152. MEULENBELT I, KLOPPENBURG M, KROON HM *et al.*: Urinary CTX-II levels are associated with radiographic subtypes of osteoarthritis in hip, knee, hand, and facet joints in subject with familial osteoarthritis at multiple sites: the GARP study. *Ann Rheum Dis* 2006; 65: 360-65.
153. LARSSON S, LOHMANDER LS, STRUGLICS A: An ARGS-aggrecan assay for analysis in blood and synovial fluid. *Osteoarthritis Cartilage* 2014; 22: 242-49.
154. STRUGLICS A, LARSSON S, KUMAHASHI N *et al.*: Changes in cytokines and aggrecan ARGS neopeptide in synovial fluid and serum and in c-terminal crosslinking telopeptide of type II collagen and N-terminal crosslinking telopeptide of type I collagen in urine over five years after anterior cruciate ligame. *Arthritis Rheumatol* (Hoboken, NJ) 2015; 67: 1816-25.
155. LEES S, GOLUB SB, LAST K *et al.*: Bioactivity in an aggrecan 32-mer fragment is mediated via toll-like receptor 2. *Arthritis Rheumatol* (Hoboken, NJ) 2015; 67: 1240-49.
156. MADSEN SH, SUMER EU, BAY-JENSEN A-C *et al.*: Aggrecanase- and matrix metalloproteinase-mediated aggrecan degradation is associated with different molecular characteristics of aggrecan and separated in time ex vivo. *Biomarkers* 2010; 15: 266-76.
157. WASILKO SM, TOURVILLE TW, DESARNO MJ *et al.*: Relationship between synovial fluid biomarkers of articular cartilage metabolism and the patient's perspective of outcome depends on the severity of articular cartilage damage following ACL trauma. *J Orthop Res* 2016; 34: 820-27.
158. MILLER RE, ISHIHARA S, TRAN PB *et al.*: An aggrecan fragment drives osteoarthritis pain through Toll-like receptor 2. *JCI Insight* 2018; 3.
159. SUMER EU, QVIST P, TANKO LB: Matrix Metalloproteinase and aggrecanase generated aggrecan fragments: implications for the diagnostics and therapeutics of destructive joint diseases. *Drug Dev Res* 2007; 68: 1-13.
160. LARKIN J, LOHR TA, ELEFANTE L *et al.*: Translational development of an ADAMTS-5 antibody for osteoarthritis disease modification. *Osteoarthritis Cartilage* 2015; 23: 1254-66.
161. MILLER RE, TRAN PB, ISHIHARA S *et al.*: Therapeutic effects of an anti-ADAMTS-5 antibody on joint damage and mechanical allodynia in a murine model of osteoarthritis. *Osteoarthritis Cartilage* 2016; 24: 299-306.
162. MILLER RE, BELMADANI A, ISHIHARA S *et al.*: Damage-associated molecular patterns generated in osteoarthritis directly excite murine nociceptive neurons through Toll-like receptor 4. *Arthritis Rheumatol* (Hoboken) 2015; 67: 2933-43.
163. HUA B, OLSEN EHN, SUN S *et al.*: Serological biomarkers detect active joint destruction and inflammation in patients with haemophilic arthropathy. *Haemophilia* 2017; 23.
164. SWÄRD P, FROBELL R, ENGLUND M *et al.*: Cartilage and bone markers and inflammatory cytokines are increased in synovial fluid in the acute phase of knee injury (hemarthrosis)--a cross-sectional analysis. *Osteoarthritis Cartilage* 2012; 20: 1302-8.
165. SWÄRD P, STRUGLICS A, ENGLUND M *et al.*: Soft tissue knee injury with concomitant osteochondral fracture is associated with higher degree of acute joint inflammation. *Am J Sports Med* 2014; 42: 1096-102.
166. ASLAM I, PERJAR I, SHI XA *et al.*: Associations between biomarkers of joint metabolism, hand osteoarthritis, and hand pain and function: the Johnston County Osteoarthritis Project. *J Rheumatol* 2014; 41: 938-44.
167. KARSDAL MA, NIELSEN SH, LEEMING DJ *et al.*: The good and the bad collagens of fibrosis - Their role in signaling and organ function. *Adv Drug Deliv Rev* 2017; 121: 43-56.
168. KIVIRIKKO KI, MYLLYLÄ R: Posttranslational enzymes in the biosynthesis of collagen: intracellular enzymes. *Methods Enzymol* 1982; 82 Pt A: 245-304.
169. ÅHRMAN E, LORENZO P, HOLMGREN K *et al.*: Novel cartilage oligomeric matrix protein (COMP) neopeptides identified in synovial

- fluids from patients with joint diseases using affinity chromatography and mass spectrometry. *J Biol Chem* 2014; 289: 20908-16.
170. BAY-JENSEN A. C., ANDERSEN TL, CHARNIBEN TABASSI N *et al.*: Biochemical markers of type II collagen breakdown and synthesis are positioned at specific sites in human osteoarthritic knee cartilage. *Osteoarthritis Cartilage* 2008; 16: 615-23.
171. ROUSSEAU J-C, SANDELL LJ, DELMAS PD *et al.*: Development and clinical application in arthritis of a new immunoassay for serum type IIA procollagen NH2 propeptide. *Methods Mol Med* 2004;101:25-37.
172. LUO Y, HE Y, REKER D *et al.*: A novel high sensitivity type II collagen blood-based biomarker, PRO-C2, for assessment of cartilage formation. *Int J Mol Sci* 2018; 19: 3485.
173. SHARIF M, KIRWAN J, CHARNI N *et al.*: A 5-yr longitudinal study of type IIA collagen synthesis and total type II collagen degradation in patients with knee osteoarthritis - Association with disease progression. *Rheumatology* 2007;46:938-43.
174. KJELGAARD-PETERSEN C, SIEBUHR AS, CHRISTIANSEN T *et al.*: Synovitis biomarkers: Ex vivo characterization of three biomarkers for identification of inflammatory osteoarthritis. *Biomarkers* 2015; 20: 547-56.
175. SIEBUHR AS, BAY-JENSEN AC, JORDAN JM *et al.*: Inflammation (or synovitis)-driven osteoarthritis: an opportunity for personalizing prognosis and treatment? *Scand J Rheumatol* 2016; 45: 87-98.
176. KRAUS VB, BLANCO FJ, ENGLUND M *et al.*: Call for standardized definitions of osteoarthritis and risk stratification for clinical trials and clinical use. *Osteoarthritis Cartilage* 2015; 23: 1233-41.
177. KARSDAL MA, CHRISTIANSEN C, LADEL C *et al.*: Osteoarthritis - a case for personalized health care? *Osteoarthritis Cartilage* 2014; 22: 7-16.
178. MORTENSEN JH, MANON-JENSEN T, JENSEN MD *et al.*: Ulcerative colitis, Crohn's disease, and irritable bowel syndrome have different profiles of extracellular matrix turnover, which also reflects disease activity in Crohn's disease. *PLoS One* 2017; 12: e0185855.
179. CURRAN ME, ATKINSON DL, EWART AK *et al.*: The elastin gene is disrupted by a translocation associated with supravalvular aortic stenosis. *Cell* 1993; 73: 159-68.
180. LOTZ M, MARTEL-PELLETIER J, CHRISTIANSEN C *et al.*: Value of biomarkers in osteoarthritis: Current status and perspectives. *Ann Rheum Dis* 2013; 72: 1756-63.
181. KRAUS VB, HARGROVE DE, HUNTER DJ *et al.*: Establishment of reference intervals for osteoarthritis-related soluble biomarkers: The FNIH/OARSI OA Biomarkers Consortium. *Ann Rheum Dis* 2017; 76: 179-85.
182. TACHMAZIDOU I, HATZIKOTULAS K, SOUTHAM L *et al.*: Identification of new therapeutic targets for osteoarthritis through genome-wide analyses of UK Biobank data. *Nat Genet* 2019; 51: 230-36.
183. GIGOUT A, GUEHRING H, FROEMEL D *et al.*: Sprifermin (rhFGF18) enables proliferation of chondrocytes producing a hyaline cartilage matrix. *Osteoarthritis Cartilage* 2017; 25: 1858-67.
184. BAGER CL, KARSDAL M, BIHLET A *et al.*: Incidence of total hip and total knee replacements from the prospective epidemiologic risk factor study: considerations for event driven clinical trial design. *BMC Musculoskelet Disord* 2019; 20: 303.