Acrocyanosis after neoadjuvant ipilimumab plus nivolumab: a case report

Sirs,

The introduction of immunotherapy has marked the beginning of a new era in cancer treatment resulting in a long-term benefit in several cancer types including melanoma (1). Checkpoint inhibitors targeting the cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) or programmed cell death 1 (ligand) [PD-(L)1] have become standard of care for late stage disease. A wide variety of immune-related adverse events (irAE) has been described (2). Here we report a patient who presented with acrocyanosis after treatment with immune checkpoint inhibition (ICI).

In December 2018, a 55-year-old woman was diagnosed with stage III melanoma with an inguinal lymph node metastasis originating from a primary melanoma on her right calf. She had no medical history of cardiovascular or autoimmune diseases. At the Netherlands Cancer Institute she received two cycles neoadjuvant ipilimumab 1mg/ kg (CTLA-4 antibody) and nivolumab 3 mg/kg (PD-1 antibody) in the setting of a clinical study [NCT02977052], followed by excision of the lymph node metastasis after 6 weeks. She achieved complete pathological remission. Two months after the last ICI infusion she developed paresthesia in her right index finger. Within 10 days the symptoms gradually increased with episodes of blue discoloration resembling the Raynaud phenomenon to persisting acrocyanosis in both hands (Fig. 1A). On suspicion of an irAE, treatment with 1 mg/kg prednisolone and 30 mg nifedipine was initiated. A skin biopsy of her right index finger showed no abnormalities. Her symptoms worsened and prednisolone treatment was intensified to 2 mg/kg.

The patient was referred to the Vasculitis Expertise Center of the Amsterdam University Medical Centre one week later. By then, additional splinter bleeds at the nailfolds were present. Laboratory assessment showed no abnormalities (Table I). Cardiac embolisms were excluded and Doppler ultrasound showed normal signals in both arms and hands (not shown). The nifedipine dose was raised to 30 mg twice daily. Due to side effects and a lack of clinical response, the corticosteroid dosage was tapered to 30mg per day. Unfortunately, the pain in the fingers progressed and the treating oncologist started mycophenolate mofetil (MMF) and aspirin. Ten days later, tacrolimus 5 mg twice daily was added. The biopsied skin area and another papercut wound showed necrotic tissue (Fig. 1B). To avoid further ischaemia the patient was admitted for intravenous iloprost while continuing the immunosuppression. During 5 subsequent days of iloprost infusions the wounds



Fig. 1. A: Start of acrocyanosis 8 weeks after the last infusion of checkpoint inhibitors.
B: Development of ischaemic areas on the right hand after skin biopsy (index finger) and a paper cut (ring finger).
C: Resolution of acrocyanosis after treatment and development of vitiligo 6 months after the last infusion with checkpoint inhibitors.

Table I. Lab results at the time of presentation.

Sedimentation rate 2 0-30 mm/U C-reactive protein 0.7 0-5 mg/L Haemoglobin 9.0 7.5-10 mmo/L Thrombocytes 392 150-400 10E9/L Leukocytes 12.3 (H) 4.0-10.5 10E9/L Eosinophils 0.00 0-0.5 10E9/L Basophils 0.01 0-0.2 10E9/L Neutrophils 11.54 (H) 1.8-7.2 10E9/L Lymphocytes 0.47 (L) 1.5-4.0 10E9/L Monocytes 0.12 0.1-1.0 10E9/L Monocytes 0.12 0.1-1.0 10E9/L Immature granulocytes 0.15 (H) <0.06 10E9/L Potassium 4.5 3.5-4.5 mmol/L Calcium 2.32 2.2-2.6 mmol/L Albumin 44 35-50g /L Creatinine 77 65-95 μmol/L eGFR 75 >60 mL/min/1.73m² Urea 7.3 (H) 2.1-7.1 mmol/L Bliirubin 8 0-17 μmol/L Alk, fosf. 79 40-120 U/L 37°C Gamma-GT	Variable	Value	Reference
Haemoglobin 9.0 7.5–10 mmol/L	Sedimentation rate	2	0-30 mm/U
Thrombocytes 392 150-400 10E9/L Leukocytes 12.3 (H) 4.0-10.5 10E9/L Eosinophils 0.00 0-0.5 10E9/L Basophils 0.01 0-0.2 10E9/L Neutrophils 11.54 (H) 1.8-7.2 10E9/L Lymphocytes 0.47 (L) 1.5-4.0 10E9/L Monocytes 0.12 0.1-1.0 10E9/L Immature granulocytes 0.15 (H) <0.06 10E9/L	C-reactive protein	0.7	0-5 mg/L
Thrombocytes 392 150-400 10E9/L Leukocytes 12.3 (H) 4.0-10.5 10E9/L Eosinophils 0.00 0-0.5 10E9/L Basophils 0.01 0-0.2 10E9/L Neutrophils 11.54 (H) 1.8-7.2 10E9/L Lymphocytes 0.47 (L) 1.5-4.0 10E9/L Monocytes 0.12 0.1-1.0 10E9/L Immature granulocytes 0.15 (H) <0.06 10E9/L	Haemoglobin	9.0	7.5-10 mmol/L
Eosinophils 0.00 0-0.5 10E9/L		392	150-400 10E9/L
Basophils 0.01 0-0.2 10E9/L Neutrophils 11.54 (H) 1.8-7.2 10E9/L Lymphocytes 0.47 (L) 1.5-4.0 10E9/L Monocytes 0.12 0.1-1.0 10E9/L Immature granulocytes 0.15 (H) <0.06 10E9/L	Leukocytes	12.3 (H)	4.0-10.5 10E9/L
Neutrophils	Eosinophils	0.00	0-0.5 10E9/L
Lymphocytes 0.47 (L) 1.5-4.0 10E9/L Monocytes 0.12 0.1-1.0 10E9/L Immature granulocytes 0.15 (H) <0.06 10E9/L	Basophils	0.01	0-0.2 10E9/L
Monocytes 0.12 0.1-1.0 10E9/L Immature granulocytes 0.15 (H) <0.06 10E9/L	Neutrophils	11.54 (H)	1.8-7.2 10E9/L
Immature granulocytes O.15 (H) <0.06 10E9/L	Lymphocytes	0.47 (L)	1.5-4.0 10E9/L
Potassium 4.5 3.5-4.5 mmol/L Calcium 2.32 2.2-2.6 mmol/L Albumin 44 35-50 g/L Creatinine 77 65-95 μmol/L eGFR 75 >60 mL/min/1.73m² Urea 7.3 (H) 2.1-7.1 mmol/L Bilirubin 8 0-17 μmol/L Alk.fosf. 79 40-120 U/L 37°C Gamma-GT 19 0-40 U/L 37°C ASAT 19 0-40 U/L 37°C ALAT 21 0-34 U/L 37°C LDH 251 (H) 0-247 U/L 37°C Ferritin 88 20-250 μg/L Folic acid 15.5 5.2-34.8 mmol/L Cryoglobulins Negative 0-0.1 g/L Vitamin B12 708 (H) 150-700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9-1.8 g/L Complement C4 0.23 0.1-0.4 g/L TSH 0.16 (L) 0.5-5 mE/L<	Monocytes	0.12	0.1-1.0 10E9/L
Calcium 2.32 2.2-2.6 mmol/L Albumin 44 35-50 g/L Creatinine 77 65-95 μmol/L eGFR 75 >60 mL/min/1.73m² Urea 7.3 (H) 2.1-7.1 mmol/L Bilirubin 8 0-17 μmol/L Alk fosf. 79 40-120 U/L 37°C Gamma-GT 19 0-40 U/L 37°C ASAT 19 0-40 U/L 37°C ALAT 21 0-34 U/L 37°C LDH 251 (H) 0-247 U/L 37°C Ferritin 88 20-250 μg/L Folic acid 15.5 5.2-34.8 nmol/L Cryoglobulins Negative 0-0.1 g/L Vitamin B12 708 (H) 150-700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-pβ2-glycoprotein Negative Negative Negative Negative Complement C3 0.95 0.9-1.8 g/L Complement C4 0.23 0.1-0.4 g/L TSH 0.16 (L) 0.5-5 mE/L <tr< td=""><td>Immature granulocytes</td><td>0.15 (H)</td><td><0.06 10E9/L</td></tr<>	Immature granulocytes	0.15 (H)	<0.06 10E9/L
Albumin 44 35–50 g/L Creatinine 77 65–95 μmol/L eGFR 75 >60 mL/min/1.73m² Urea 7.3 (H) 2.1–7.1 mmol/L Bilirubin 8 0–17 μmol/L Alk.fosf. 79 40–120 U/L 37°C Gamma-GT 19 0–40 U/L 37°C ASAT 19 0–40 U/L 37°C ALAT 21 0–34 U/L 37°C LDH 251 (H) 0–247 U/L 37°C Ferritin 88 20–250 μg/L Folic acid 15.5 5.2–34.8 nmol/L Cryoglobulins Negative 0–0.1 g/L Vitamin B12 708 (H) 150–700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9–1.8 g/L Complement C4 0.23 0.1–0.4 g/L TSH 0.16 (L) 0.5–5 mE/L Free T4 18.3 12.0	Potassium	4.5	3.5-4.5 mmol/L
Creatinine 77 65–95 μmol/L eGFR 75 >60 mL/min/1.73m² Urea 7.3 (H) 2.1 –7.1 mmol/L Bilirubin 8 0 –17 μmol/L Alk fosf. 79 40 –120 U/L 37°C Gamma-GT 19 0 –40 U/L 37°C ASAT 19 0 –40 U/L 37°C ALAT 21 0 –34 U/L 37°C LDH 251 (H) 0 –247 U/L 37°C Ferritin 88 20 –250 μg/L Folic acid 15.5 5 .2–34.8 mmol/L Cryoglobulins Negative 0 –0.1 g/L Vitamin B12 708 (H) 150 –700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0 .95 0 .9–1.8 g/L Complement C4 0 .23 0 .1–0.4 g/L TSH 0 .16 (L) 0 .5–5 mE/L Free T4 18.3 12.0 –22.0 pmol/L	Calcium	2.32	2.2-2.6 mmol/L
eGFR 75 >60 mL/min/1.73 m² Urea 7.3 (H) 2.1–7.1 mmol/L Bilirubin 8 0–17 μ mol/L Alk,fosf. 79 40–120 U/L 37°C Gamma-GT 19 0–40 U/L 37°C ASAT 19 0–40 U/L 37°C ALAT 21 0–34 U/L 37°C LDH 251 (H) 0–247 U/L 37°C Erritin 88 20–250 μ g/L Folic acid 15.5 5.2–34.8 mmol/L Cryoglobulins Negative 0–0.1 g/L Vitamin B12 708 (H) 150–700 μ gmol/L Anti-nuclear antibodies Negative Complement C3 0.95 0.9–1.8 g/L Complement C4 0.23 0.1–0.4 g/L TSH 0.16 (L) 0.5–5 mE/L Free T4 18.3 12.0–22.0 μ mol/L 12.0–22.0 μ mol/L 12.0–22.0 μ mol/L 15.5 15.5 5.2–34.5 mmol/L 15.5 5.2 5.2 5.2 5.2 5.2 5.2 5.2 5.2 5.2	Albumin	44	35-50 g/L
Urea 7.3 (H) 2.1–7.1 mmol/L Bilirubin 8 0–17 μmol/L Alk.fosf. 79 40–120 U/L 37°C Gamma-GT 19 0–40 U/L 37°C ASAT 19 0–40 U/L 37°C ALAT 21 0–34 U/L 37°C LDH 251 (H) 0–247 U/L 37°C Ferritin 88 20–250 μg/L Folic acid 15.5 5.2–34.8 mmol/L Cryoglobulins Negative 0–0.1 g/L Vitamin B12 708 (H) 150–700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-plospholipids Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9–1.8 g/L Complement C4 0.23 0.1–0.4 g/L TSH 0.16 (L) 0.5–5 mE/L Free T4 18.3 12.0–22.0 pmol/L	Creatinine	77	65–95 µmol/L
Bilirubin 8 $0-17 \mu \text{mol/L}$ Alk.fosf. 79 $40-120 \text{ U/L } 37^{\circ}\text{C}$ Gamma-GT 19 $0-40 \text{ U/L } 37^{\circ}\text{C}$ ASAT 19 $0-40 \text{ U/L } 37^{\circ}\text{C}$ ALAT 21 $0-34 \text{ U/L } 37^{\circ}\text{C}$ LDH 251 (H) $0-247 \text{ U/L } 37^{\circ}\text{C}$ Ferritin 88 $20-250 \mu \text{g/L}$ Folic acid 15.5 $5.2-34.8 \text{ mmol/L}$ Cryoglobulins Negative $0-0.1 \text{ g/L}$ Vitamin B12 708 (H) $150-700 \text{ pmol/L}$ Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9-1.8 g/L Complement C4 0.23 0.1-0.4 g/L TSH 0.16 (L) 0.5-5 mE/L Free T4 18.3 12.0-22.0 pmol/L	eGFR	75	>60 mL/min/1.73m ²
Alk.fosf. 79 40–120 U/L 37°C Gamma-GT 19 0–40 U/L 37°C ASAT 19 0–40 U/L 37°C ALAT 21 0–34 U/L 37°C LDH 251 (H) 0–247 U/L 37°C Ferritin 88 20–250 μg/L Folic acid 15.5 5.2–34.8 nmol/L Cryoglobulins Negative 0–0.1 g/L Vitamin B12 708 (H) 150–700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-β2-glycoprotein Negative Negative Complement C3 0.95 0.9–1.8 g/L Complement C4 0.23 0.1–0.4 g/L TSH 0.16 (L) 0.5–5 mE/L Free T4 18.3 12.0–22.0 pmol/L	Urea	7.3 (H)	2.1-7.1 mmol/L
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ALAT 21 0-34 U/L 37°C LDH 251 (H) 0-247 U/L 37°C Ferritin 88 20-250 μg/L Folic acid 15.5 5.2-34.8 nmo/L Cryoglobulins Negative 0-0.1 g/L Vitamin B12 708 (H) 150-700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9-1.8 g/L Complement C4 0.23 0.1-0.4 g/L TSH 0.16 (L) 0.5-5 mE/L Free T4 18.3 12.0-22.0 pmol/L	Gamma-GT	19	0-40 U/L 37°C
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Folic acid 15.5 5.2–34.8 nmol/L Cryoglobulins Negative 0–0.1 g/L Vitamin B12 708 (H) 150–700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9–1.8 g/L Complement C4 0.23 0.1–0.4 g/L TSH 0.16 (L) 0.5–5 mE/L Free T4 18.3 12.0–22.0 pmol/L	LDH	251 (H)	0-247 U/L 37°C
Folic acid 15.5 5.2–34.8 nmol/L Cryoglobulins Negative 0–0.1 g/L Vitamin B12 708 (H) 150–700 pmol/L Anti-nuclear antibodies Negative Negative Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9–1.8 g/L Complement C4 0.23 0.1–0.4 g/L TSH 0.16 (L) 0.5–5 mE/L Free T4 18.3 12.0–22.0 pmol/L	Ferritin	88	$20-250 \mu g/L$
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Scleroderma immunoblot Negative* Negative Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9-1.8 g/L Complement C4 0.23 0.1-0.4 g/L TSH 0.16 (L) 0.5-5 mE/L Free T4 18.3 12.0-22.0 pmol/L	Vitamin B12	708 (H)	150-700 pmol/L
Anti-β2-glycoprotein Negative Negative Anti-phospholipids Negative Negative Complement C3 0.95 0.9-1.8 g/L Complement C4 0.23 0.1-0.4 g/L TSH 0.16 (L) 0.5-5 mE/L Free T4 18.3 12.0-22.0 pmol/L	Anti-nuclear antibodies	Negative	
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Free T4 18.3 12.0–22.0 pmol/L		0.16 (L)	ē.
ž.	Free T4		12.0-22.0 pmol/L
	TPO antibodies	<30	•

*Scleroderma immunoblot included antibodies against Scl-70, centromere (A and B), RNA polymerase III (11 and 155 kD), fibrillarin, NOR90, Th/To, PM-Sc100, PM-Sc175, Ku, PDGFR and Ro-52.

showed slow recovery. Three weeks later the symptoms had strongly improved. Nailfold capillaroscopy at that time showed no abnormalities (not shown). During the two months thereafter the immunosuppressive drugs were tapered while nifedipine treatment was continued. The acrocyanosis and wounds disappeared whereas a mild pain in her fingers lasted for 16 weeks longer before disappearing completely. Up until now, this patient is free of recurrence from her melanoma.

Letters to the Editors

The development of acrocyanosis and/or digital ischaemia in the setting of ICI is described before but seems rare (3-8). The aetiology remains unclear because most cases – including ours – showed no biochemical markers of inflammation, nor histological signs of immune cell tissue infiltration and/or damage. This contradicts the majority of systemic autoimmune diseases where usually inflammation can be found and/or autoantibodies are present.

Acral ischaemia has previously been described as a paraneoplastic phenomenon (9) but since our patient achieved complete response and is still relapse-free, it is unlikely that the observed symptoms are a result of a paraneoplastic syndrome.

The optimal treatment for ICI-induced acrocyanosis/digital ischaemia remains unclear. Results from treatment with prostacyclin analogues or calcium channel blockers are controversial (4, 5). High-dose (methyl) prednisolone might be considered, however several cases report no efficacy (4, 5). Similar to our case, addition of MMF to prednisolone and iloprost treatment resulted in complete resolution of symptoms in another case. Thus, aggressive immune suppression combined with atrial vasodilation and platelet aggregation inhibition might be a promising therapy option for steroid-refractory acrocyanosis.

In summary, acrocyanosis as an ICI induced irAE is a rare but serious complication, which seems to require more intense therapy than other irAEs (10). It should be managed multidisciplinary to avoid digital necrosis and amputation.

Ethics approval

The patient gave written informed consent for the anonymous publication of her case, including photographs of her hands.

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Competing interests: none declared.

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