

FcγRIIa polymorphism in patients with rheumatoid arthritis

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ABSTRACT

Objective. Polymorphism of phagocyte IgG receptor FcγRIIa may modulate immune complex mediated inflammation, particularly when immune complex contain IgG.

Methods. FcγRIIa genotyping in 82 patients with rheumatoid arthritis (RA) and 148 healthy subjects was performed using the polymerase chain reaction technique with allele specific primers.

Results. No significant relation between FcγRIIa genotypes and susceptibility to RA was observed, but extraarticular complications with high frequency were revealed in patients with R/R131 genotype.

Conclusion. The results suggest that the FcγRIIa polymorphism is not a risk factor for RA.

Introduction

The chronic tissue-destructive process in patients with rheumatoid arthritis (RA) has been attributed to an ongoing immune response in which immune complexes play an important role. Immune complexes containing IgG may be a major pathogenic factor, responsible for the initiation and persistence of the inflammatory cascade and its resulting destructions in the cartilage (1, 2). IgG complexes interact with synovial cells via cellular receptors for IgG. The most widely distributed class of IgG Fc receptors is receptor Fc RIIa, which mediates a variety of physiological functions upon clustering by immune complexes. An allelic polymorphism in the human Fc RIIa receptor gene that consists of a single base G-A substitution at nucleotide 494 results in an amino acid change from arginine R to histidine H at position 131 (3). This substitution alters the affinity of the receptor for at least three IgG subclasses: murine IgG₁, human IgG₂ and IgG₃. The polymorphism was defined by differences in the binding of murine IgG₁ anti CD3mAB to Fc RIIa of human monocytes and T cell. The allelic form with arginine as amino acid 131 expressed high affinity for murine IgG₁ whereas the form with histidine showed low affinity (4). Recent data suggest that this polymorphism may be relevant to Fc RIIa function. Polymorphonucle-

ar neutrophils homozygous for H/H131 show greater capacity to phagocytose bacteria or erythrocytes opsonized with IgG₂ than those homozygous for R/R131 (5). This polymorphism may also be implicated in the susceptibility to heparin-induced thrombocytopenia and lupus erythematosus, especially lupus nephritis (6, 7).

The aim of the study was to determine the Fc RIIa polymorphism in patients with rheumatoid arthritis

Materials and methods

Materials

We examined 82 patients (56 women, 26 men, age 16-79 years, mean 54.2) with rheumatoid arthritis diagnosed according to the criteria of American College of Rheumatology. The disease duration was 1-20 years (mean 14.3). Patients were recruited from the outpatient and inpatient populations of the Department of Rheumatology, University Hospital in Szczecin, Poland. All subjects were Caucasian from the Pomeranian region of Poland.

Subjects involved in the study underwent routine biochemical blood analysis and when required anticardiolipin antibodies, antinuclear antibodies and immunologic complexes. In all patients the x-ray of chest, hands and feet and when required other joint were performed. The evaluation of subjects included physical examinations with attention to pattern of joint involvement, presence of nodules and other extraarticular features such as vasculitis, anaemia, sicca syndrome, amyloidosis, organ involvement, and laboratory features such as erythrocyte sedimentation rate (ESR) and rheumatoid factor (RF). The subcutaneous nodules without other extraarticular manifestations in 19 patients were diagnosed. The group with other extraarticular manifestations included 15 patients: 3 with anaemia, 2 with anaemia and nodules, 1 with vasculitis and nodules, 6 with vasculitis, 1 with vasculitis and amyloidosis, 2 with sicca syndrome and amyloidosis. Amyloidosis was diagnosed by histomorphology (skin- and bowel- or duodenum biopsy), vasculitis by histomorphology (skin biopsy) and angiogram.

The group with severe joint manifestations included patients with more than 6 swollen joints and radiologically diagnosed erosions. In the group with severe RA was 15 patients with particularly severe disease progress, resistant to second-line therapy. The control group consisted of 148 healthy subjects (86 women and 62 men, age 19–75 years, mean 48.7). The study was approved by the local ethics committee and written informed consent was obtained from all subjects.

Isolation of genomic DNA

Genomic DNA was extracted manually (precipitation with trimethylammonium bromide salts from leukocytes contained in 450 µL of venous blood with ethylenediaminetetraacetic acid (EDTA) as an anticoagulant. DNA was then precipitated in 95% ethanol dissolved in distilled water, and stored at -20°C until analysis (8). The chemicals for the extraction were purchased from Sigma Company (St Louis, Mo).

FcγRIIA genotyping

We performed 25 µL PCRs containing 2.5 µL of genomic DNA (approx. 100 ng), 2.5 µL of 10 x PCR buffer (contains 15 mM MgCl₂, Gibco BRL), 200 µM of each dNTP (Gibco BRL), and 0.5 U of Taq polymerase (Gibco BRL). We used 0.5 µM H131-specific sense primer (5'-ATCCCAGAAATTCTCCCA-3') from the second extracellular domain or 0.5 µM R131-specific sense primers (5'-ATCCCAGAAATTCTCCCG-3' (all primers were synthesized by Gibco BRL) and 0.5 µM common antisense primer from an area of the downstream intron where the sequences for Fc RIIa, Fc RIIb, and Fc RIIc

diverge (5'-CAATTTTGCTGCTATGGGC-3'). The resulting fragment was 253 bp in length. As internal control, we used 0.125 µM human growth hormone (HGH)-I primer (5'-CAGTGCCTTCCCAACCATTCCCTTA-3') and 0.125 µM HGH-II primer (5'-ATCCACTCACGGATTTCTGTGTGTTTC-3'), which resulted in a 439-bp fragment. We used a thermal cycler (Mastercycler Eppendorf) to perform a hot-start PCR as follows: 5 minutes at 95°C, 10 cycles of 1 minute at 95°C, 2 minutes at 57°C, and 1 minute at 72°C; thereafter, to enhance the sensitivity, we used 22 cycles of 1 minute at 95°C, 2 minutes at 54°C, and 1 minute at 72°C and a final extension step for 5 minutes at 72°C. Each PCR analysis was performed with control samples for R/R, H/R and H/H genotypes. The PCR amplification products were separated on 1.5% agarose and visualized by using ethidium bromide (9).

Statistical analysis

Frequencies of Fc RIIa alleles were given with 95% confidence intervals (95%CI). The distribution of Fc RIIa alleles in patients with RA was compared with healthy subjects and statistically evaluated by use of the χ^2 test with Yates correction for small groups.

Results

The distribution of Fc RIIa genotypes in 148 Polish healthy subjects did not differ from that of Caucasian populations (Table I). The genotypes R/R131, R/H131 and H/H131 were found in 33 (22.3%), 71 (48.0%) and 44 (29.7%) subjects respectively and there was no difference in distribution between the male and female controls. For the total

group of 82 RA patients the distribution was: R/R131:19 subjects (23.2%); R/H131: 36 (43.9%); and H/H131: 27 (32.9%) and did not significantly differ from the control group (Table I).

Subcutaneous nodules without other extraarticular manifestations were detected among 6 (31.6%), 7 (19.5%), 6 (22.2%) of patients with genotypes R/R131, R/H131 and H/H131 respectively. The differences were not statistically significant (Table II). Other extraarticular manifestations were diagnosed in 6 (31.6%) patients with R/R131 genotype, in 5 (13.9%) with R/H131 and in 4 (14.8%) with genotype H/H131. The odds ratio for the risk of extraarticular complications in individuals with R/R131 genotype was 2.65 greater, (95% CI 0.88 – 9.63) than in subjects with genotype H/H131. The frequency of extraarticular involvement was not high enough to yield statistical significance. A larger cohort of RA patients with extraarticular manifestations will be required to confirm that association. Among subjects with R/R131 genotype magnitude joint involvement in 8 patients (42.1%) was diagnosed, but in subjects with genotypes H/R131 and H/H131 in 16 (44.4%) and 11 (40.7%) respectively. The risk of severe joint manifestations was similar in particular Fc RIIa genotypes (OR for R/R131 vs H/H131 - 1.06) (Table II). Moreover the correlation between Fc RIIa polymorphism and diseases activity and response under therapy was revealed. In subjects with R/R131 genotype 14 patients (73.7%) with active RA were found, in subjects with H/R131 26 (72.2%) in H/H131 19 (70.4%). The risk of active of RA not differed depending on Fc RIIa genotypes (OR for

Table I. Distribution of Fc RIIa genotypes in control group and RA patients.

	Fc RIIa R/R 131		Fc RIIa R/H 131		Fc RIIa H/H 131		p value*
	n	%	n	%	n	%	
Control group n - 148	33	(22.3)	71	(48.0)	44	(29.7)	NS
RA patients n - 82	19	(23.2)	36	(43.9)	27	(32.9)	NS

* RA patients vs control group.

Table II. The distribution of Fc RIIa genotypes in RA patients.

	Fc RIIa R/R 131		Fc RIIa R/H 131		Fc RIIa H/H 131		P value*	OR	95% CI
	n - 19		n - 36		n - 37				
	n	%	n	%	N	%			
Patients with nodulosis without other extra-articular manifestations	6	(31.6)	7	(19.5)	6	(22.2)	NS	1.62	0.56 – 7.48
Patients with extraarticular manifestations	6	(31.6)	5	(13.9)	4	(14.8)	NS	2.65	0.82 - 14.15
Patients with positive RA	14	(73.7)	21	(58.3)	15	(59.3)	NS	1.93	0.46 – 8.41
Patients with severe joint involvement	8	(42.1)	16	(44.4)	11	(40.7)	NS	1.06	0.27 – 4.11
Patients with active RA	14	(73.7)	26	(72.2)	19	(70.4)	NS	1.18	0.27 – 5.34
Patients with severe RA	4	(21.0)	7	(19.4)	4	(14.8)	NS	1.52	0.26 – 8.96

* Patients with R/R 131 genotype vs H/H 131.

R/R131 vs H/H131- 1.18).

There was also no correlation between Fc RIIa polymorphism and response of the disease under therapy. Among subjects with R/R131 genotype were 4 patients (21.0%) resistant to second-line therapy, in H/R131 and H/H131 7 (19.4%) and 4 (14.8%) respectively. The risk was similar for R/R131, H/R131 and H/H131 genotypes (OR for R/R131 vs H/H131 - 1.53).

Among patients with R/R131 genotype rheumatoid factor in 14 subject (73.7%) was diagnosed, for R/H131 genotype in 21 (58.3%), for H/H131 genotype in 16 (59.3%). There was no statistically significant difference in frequency of rheumatoid factor between studied groups (Table II).

Additionally, there was no correlation between the Fc RIIa polymorphism and age of first occurrence of clinical symptoms among patients with RA, what may suggest that specific Fc RIIa alleles are not associated with disease susceptibility (Table III).

Discussion

This study examined the hypothesis that Fc RIIa polymorphism may be a heritable factor influencing RA sus-

ceptibility and its extraarticular manifestations.

The distribution of Fc RIIa genotypes in control population was similar to that obtained in other European populations (10, 11). Comparing the distribution of Fc RIIa genotypes among RA patients with controls there was no significant difference between these two groups.

The Fc receptors for IgG constitute a family of hemopoietic cell surface molecules that can stimulate cellular responses upon binding of antibody-antigen complexes and mediate antibody-dependent phagocytosis (12, 13). Fc RIIa receptors play an essential role in the clearance of immune complexes (14). Impaired handling and removal of immune complexes by the mononuclear phagocyte system results in its deposition and inflammatory processes in organs and tissues. However, the underlining basis of such defective immune complex clearance and its relation to the observed clinical manifestation has not been defined. Down regulation of Fc RIIa expression and function effects on signal transduction and anti Fc antibodies have been implicated as factor in aberrant Fc functioning

(15, 16).

Indeed, there is increasing evidences that the Fc RIIa R/R131 genotype is a risk factor for the manifestation of immune complex mediated diseases. The impaired phagocytosis of immune complexes seen in patients with systemic lupus erythematosus can be correlated to the presence of R/R131 allele (17,18). The SLE patients with R/R131 genotype develop at a higher rate lupus nephritis, haematological abnormalities, sicca syndrome, antinuclear antibodies and hypocomplementemia (19). Additionally, Fc RIIa polymorphism is associated with heparin-induced thrombocytopenia (20, 21). Furthermore, the Fc RIIa polymorphism has an impact on the susceptibility to bacterial infections (5,22). Phagocytosis of IgG-opsonized bacteria is less effective in individuals with R/R131 allotype and these patients exhibit a higher susceptibility to infection by encapsulated bacteria. Accordingly, a low incidence of infection with encapsulated bacteria has been noted in the Japanese population where the prevalence of H/H131 allotype was observed (11, 23).

Several lines of evidence have impli-

Table III. The mean age of first occurrence of clinical symptoms among patients with RA according to Fc RIIa genotypes.

	Fc RIIa R/R 131		Fc RIIa R/H 131		Fc RIIa H/H 131		p value *
Mean value (years) ± SD	44.9	(± 8.66)	46.22	(± 15.21)	49.59	(± 10.75)	

*Patients with genotype R/R 131 vs H/H 131.

cated Fc receptors, in particular IgG binding receptors in RA pathogenesis (24). Fc RIII was detected on synovial intima in normal and arthritic human joints and on invading macrophages. An Fc RIII gene polymorphism has been correlated with RA susceptibility and extraarticular manifestations involvement (25, 26).

The results of our study suggest that Fc RIIa polymorphism does not represent a genetic risk factor for the RA occurrence and disease progress. Though the risk of some extraarticular manifestations was higher in subjects with R/R131 genotype, there were no correlations with joints manifestations, disease activity, response under therapy. It seems that RA pathogenesis may be associated with Fc RIII polymorphism but Fc RIIa polymorphism is involved particularly in immune complexes mediated diseases such as SLE.

References

1. REVEILLEJD: The genetic contribution to the pathogenesis of rheumatoid arthritis. *Curr Opin Rheumatol* 1998; 10: 187-200.
2. EDWARDS JCW, CAMBRIDGE G: Hypothesis. Rheumatoid arthritis: the predictable effect of small immune complexes in which antibody is also antigen. *Br J Rheumatol* 1998; 37: 126-30.
3. WARMERDAM PAM, Van de WINKEL JGJ, GOSSELIN EJ, CAPEL PJA: Molecular basis for a polymorphism of human Fc receptor II (CD32). *J Exp Med* 1990; 172: 19-25.
4. PARREN PWHI, WARMERDAM PAM, BOEIJE LCM et al.: On the interaction of IgG subclasses with the low affinity Fc RIIa (CD32) on human monocytes, neutrophils, and platelets. Analysis of a functional polymorphism to human IgG2. *J Clin Invest* 1992; 90: 1537-46.
5. SANDERS LAM, VAN DE WINKEL JGJ, RIJKERS GT et al.: Fc receptor IIA (CD32) heterogeneity in patients with recurrent bacterial respiratory tract infections. *J Infect Dis* 1994; 170: 854-61.
6. BURGESS JK, LINDEMAN R, CHESTERMAN CN, CHONG BH: Single amino acid mutation of Fc receptor is associated with the development of heparin-induced thrombocytopenia. *Br J Haematol* 1995; 91: 761-6.
7. SALMON JE, MILLARD S, SCHACHTER LA et al.: Fc RIIA alleles are heritable risk factors for lupus nephritis in African Americans. *J Clin Invest* 1996; 97: 1348-54.
8. GUSTINCICH S, MANFIOLETTI G, DEL SAL G et al.: A fast method for high-quality genomic DNA extraction from whole blood. *Biotechniques* 1991; 11: 298-301.
9. FLESCH BK, BAUER F, NEPPERT J: Rapid typing of the human Fc receptor IIA polymorphism by polymerase chain reaction amplification with allele-specific primers. *Transfusion* 1998; 38: 174-6.
10. JOUTSI L, JAVELA K, PARTANEN J, KEKOMÄKIR: Genetic polymorphism H131R of Fc receptor type IIA (Fc RIIA) in a healthy Finnish population and in patients with or without platelet-associated IgG. *Eur J Haematol* 1998; 61: 183-9.
11. OSBORNE JM, CHACKO GW, BRANDT JT, ANDERSON CL: Ethnic variation in frequency of an allelic polymorphism of human Fc RIIA determined with allele specific oligonucleotide probes. *J Immunol Methods* 1994; 173: 207-17.
12. SALMON JE, EDBERG JC, BROGLE NL, KIMBERLEY RP: Allelic polymorphisms of human Fc receptor IIA and Fc receptor IIIB. Independent mechanisms for differences in human phagocyte function. *J Clin Invest* 1992; 89: 1274-81.
13. INDIK Z, KELLY C, CHIEN P, LEVISON AI, SCHREIBER AD: Human Fc RII, in the absence of other Fc receptors, mediates a phagocytic signal. *J Clin Invest* 1991; 88: 1766-71.
14. LÖW A, HOTZE A, KRAPF FD, SCHRANZ W et al.: The non-specific clearance function of the reticuloendothelial system in patients with immune complex mediated diseases before and after therapeutic plasmapheresis. *Rheumatol Int* 1985; 5: 69-72.
15. BOROS P, MURYOI T, SPIERA H et al.: Autoantibodies directed against different classes of Fc R are found in sera of autoimmune patients. *J Immunol* 1993; 5: 218-24.
16. VAN DE WINKEL JGJ, CAPEL PJA: Human IgG Fc receptor heterogeneity: Molecular aspects and clinical implications. *Immunol Today* 1993; 14: 215-20.
17. ZUNIGA R, NG S, PETERSON MGE et al.: Low-binding alleles of Fc receptor types IIA and IIIA are inherited independently and are associated with systemic lupus erythematosus in Hispanic patients. *Arthritis Rheum* 2001; 44: 361-7.
18. MANGER K, REPP R, SPRIEWALD BM et al.: Fc receptor IIA polymorphism in Caucasian patients with systemic lupus erythematosus: association with clinical symptoms. *Arthritis Rheum* 1998; 41: 1181-9.
19. DUTTS A J, BOOTSMAN H, DERKSEN RHM et al.: Skewed distribution of IgG Fc receptor IIA (CD32) polymorphism is associated with renal disease in systemic lupus erythematosus patients. *Arthritis Rheum* 1995; 38: 1832-6.
20. CARLSSON LE, SANTOSO S, BAURICHTER G et al.: Heparin-induced thrombocytopenia: New insights into the impact of the Fc RIIA-RH131 polymorphism. *Blood* 1998; 92: 1526-31.
21. BRANDT JT, ISENHART CE, OSBORNE JM et al.: On the role of platelet Fc RIIa phenotype in heparin-induced thrombocytopenia. *Thromb Haemost* 1995; 74: 1564-72.
22. BREDIUS RG, DERKX BH, FIJEN CA et al.: Fc receptor IIA (CD32) polymorphism in fulminant meningococcal septic shock in children. *J Infect Dis* 1994; 170: 848-53.
23. MUSSER JM, KROLL JS, GRANOFF DM et al.: Global genetic structure and molecular epidemiology of encapsulated *Haemophilus influenzae*. *Rev Infect Dis* 1990; 12: 75-111.
24. JI H, OHMURA K, MAHMOOD U et al.: Arthritis critically dependent on innate immune system players. *Immunity* 2002; 16: 157-68.
25. WIKANINGRUM R, HIGHTON J, PARKER A et al.: Pathogenic mechanisms in the rheumatoid nodule: comparison of proinflammatory cytokine production and cell adhesion molecule expression in rheumatoid nodules and synovial membranes from the same patient. *Arthritis Rheum* 1998; 41: 17783-97.
26. MORGAN AW, GRIFFITHS B, PONCHEL F et al.: Fc receptor type IIIA is associated with rheumatoid arthritis in two distinct ethnic groups. *Arthritis Rheum* 2000; 43: 2328-34.