# Cryoglobulins: putative effectors of adaptive immune response

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### ABSTRACT

Cryoglobulinaemia consists of circulating monoclonal and/or polyclonal immunoglobulins with rheumatoid factor (RF) activity that precipitate at temperatures <37°C. Cryoglobulinaemic syndrome, characterised by clinical signs of systemic vasculitis, is associated with chronic infection of hepatitis C virus (HCV) and might evolve in B-cell malignancies. In about one third of all HCV infection cases, serum autoantibodies are commonly found. This is probably due directly to the transformation of infected B cells but, also, indirectly, to the viral chronic stimulation of a pool of autoreactive B cells. The pattern of IgG subclasses seems to contribute to the worsening progression of HCV infection into lymphoproliferative and/or autoimmune diseases. Many evidences showed that B cells circulating in patients with HCV-associated mixed cryoglobulinaemia (MC) are profoundly abnormal; moreover, in most of cases, normal B cells are replaced by expanded clonal B cells characterised by the low expression of CD21. After viral eradication, these cells persist in circulation and their occurrence does not correlate with serum cryoglobulins nor with vasculitis response or relapse. It is probably due to the persistence of monoclonal B cells producing RF, that in course of MC can be reactivated by circulating immune complexes, highly produced during infections or tumours. Here, we aimed to review current literature focusing the pathogenesis of MC referring to specificity and immunochemical characteristics of the immunoglobulins involved in cryoprecipitation.

### Introduction

Cryoglobulinaemia is defined as the presence of circulating immunoglobulins (Igs) that precipitate at cold tem-

peratures and re-dissolve on rewarming. Cryoglobulins (CGs) maintain their solubility when aggregated in optimal thermal condition. A reduction of temperature provokes steric modifications of molecules with exposition of non-polar residua, reduction of solubility and cryoprecipitation: it probably occurs because of the rapid formation of cold-insoluble IgM-IgG immune complexes or simply by a decreasing solubility, resulting from an unfavourable interaction between CGs and solvent at low temperatures (1). When the temperature rewarms at 37°C, molecules return to the initial conformation (2). Typically, CGs precipitate in the skin vessels of the extremities where the temperature more frequently drops below 37°C causing skin lesions and are responsible for renal injury by an immune-complex mechanism (3).

Reversible cryoprecipitability of proteins was observed by Heidelberger and Kendall as a concomitant feature of immune-complex formation (4). Wintrobe and Buell firstly pointed out the clinical importance of CGs in a patient with multiple myeloma (1, 5). The relevance of this phenomenon became apparent when clinical associations with vasculitis and nephritis were described, resembling those observed in experimental serum sickness.

According to the characteristics of the involved Igs, CGs have been classified by Brouet *et al.*: type I is formed of a single monoclonal Ig; type II of a monoclonal Ig and polyclonal Ig; in type III, only polyclonal immunoglobulins are found (6). Type II and III CGs cryoprecipitate, characterised by IgG-antigen/IgM-anti-IgG immune complexes, are often composed by autoantibodies (auto-Abs) and monoclonal RF IgM and/or IgG (7, 8).

Type I CGs occurs in patients with lymphoproliferative disorders such as multiple myeloma, lymphoma, or Waldestrom's macroglobulinaemia (1) producing symptoms of vasculitis or distal gangrene/necrosis. Type II and type III CGs are detected in patients with a wide variety of diseases, infective and autoimmune disorders, lymphoproliferative and chronic liver diseases, with the main symptoms of purpura, arthralgia, and Raynaud's phenomena. In 1992s, Musset et al. observed new microheterogeneity represented by the occurrence of two or more small monoclonal bands of IgM or IgG isotype in sera from patients with cryoglobulinaemia (9). Similar results have been described by Tissot, who characterised some trace amount of CGs with a pattern of polyclonal IgG associated with a mixture of polyclonal and monoclonal IgM (10). The inclusion of this new subtype of CGs, as a transitional step between type II CGs and type III CGs, focused greater attention on clinical, histopathological and follow-up aspects as well as on a more adequate diagnostic and therapeutic definitions, which differ along each and every moment of the evolutionary pathway of the disease (11, 12) (Table I).

CGs testing is often neglected by clinicians, despite its usefulness in patient management. Recent reports stressed the requirement of international guidelines, standardised pre-analytical protocols to maintain samples at right temperature procedures in the 3 phases of laboratory workup (pre-analytical, analytical and characterisation of the cryoprecipitate) (2, 3, 13, 14) (Fig. 1). Mixed cryoglobulinaemia (MC) is a multifactorial disorder, based on genetic, environmental, and immunogenic factors, occurring in combination in the pathogenesis of the disorder. HCV appears to have a preeminent aetiologic role in MC, since HCV infection can be found in 76-95% of patients with MC. The clonality and HCV specificity of Ig in cryoprecipitates from patients with HCV-related MC were investigated (8). Notably, type I CGs containing IgG3 isotype shows a greater tendency to aggregate. The IgG3 differ from the other subclasses of IgG for non-ionic and hydrophobic interaction that may favourite the auto aggregation (15).

This review is aimed to analyse the current literature focusing the pathogenesis of cryoglobulinaemia referring to specificity and immunochemical characteristics of the Igs involved in cryoprecipitation.

### Pathophysiology and biomarkers

Cryoprecipitation takes place thanks to different parameters as specific Ig levels, pH, ionic force, temperature, weak noncovalent interactions as well as the electrical charge that is dependent by the amino acid sequences and sugar moieties contained in Ig (16). Host predisposition and environmental triggers contribute to the pathogenesis. The result is the production of monoclonal and/or polyclonal immunoglobulins, interacting with each other to form immune complexes, most frequently associated with chronic infectious diseases as (HCV and HBV), autoimmune diseases, and B-cell lymphoproliferative disorders (16).

A genome-wide significant association with CGs-related vasculitis was identified with single nucleotide polymorphism near NOTCH4 and MHC class II genes (17). Chronic infection of T, B lymphocytes, and macrophages may be responsible for the proliferation of B lymphocytes producing circulating immune complexes composed of CGs and auto-Abs. Persistent antigenic stimuli could be responsible for the development of autoimmunity by molecular mimicry.

B cell proliferation is an important mechanism in the pathogenesis of cryoglobulinaemia that may induce an overproduction of polyclonal free light chains (FLC). Polyclonal circulating FLCs provide an index of total Ig synthesis and may be considered a biomarker of immune stimulation and inflammation, correlated with disease activity in different autoimmune conditions, an attractive novel target in precision medicine (18-20). Patients at risk of disease flare could be monitored with FLC, to allow early intervention and possibly reduce end-organ damage and mortality. Increased FLC is associated with a presence of CGs. Chronic HCV infection and viral replication may drive B-cell activation and dysfunction, resulting in FLC overproduction (21). CGs seem to be part of a progressive clonal selection process in which B-cells are stimulated to produce oligoclonal IgG3 with RF activity. The persistence of the antigenic stimulus elicits the production of polyclonal IgM-RF and subsequently the formation of oligoclonal IgG/polyclonal IgM containing CGs. In the last stage, a monoclonal IgM-RF clone is formed which may coexist with a monoclonal IgG3-RF clone (12).

The evidence that RF-IgG, RF-IgM and serum FLC increase whilst C4 is reduced, both in symptomatic and asymptomatic patients, suggests that, even in absence of MC symptoms, the low levels of CGs may represent a trigger of activation for immune system in course of HCV infection. Moreover, the finding of a positive correlation between raised levels of vascular endothelial growth factor with both free k and  $\lambda$  chains in HCV-MC patients could be suggestive that the strong viral-driven B activation would be worsening into lymphoproliferative disorders (22, 23).

# Cryoglobulinaemia in non-HCV patients

Current literature on non-HCV-associated cryoglobulinaemia is very limited and little is known about the immunological and serological profile of affected patients. The cryoglobulinaemic syndrome not associated with HCV infection is often found concomitantly with other infections, autoimmune diseases and B-cell lymphoproliferative disorders.

Type I cryoglobulinaemia is associated with the presence of B-cell malignancies, notably Waldenström macroglobulinaemia, myeloma and monoclonal gammopathy of undetermined significance (MGUS), where the capacity to form cryoprecipitates is a feature of the monoclonal protein. Although cryoglobulinaemia can be asymptomatic and CGs serum levels not always correlated with clinical severity, some patients with low levels of CGs may nonetheless show severe symptoms (21, 24). Saadoun *et al.* described clinical signs and outcomes of a group of patients

with non-HCV-related MC. The im-

	IG COMPONENT	CLINICAL SIGNS	DISORDERS	<b>BIOMARKER S</b>
CG Type I	<b>monoclonal</b> IgM (mainly)	<b>Cutaneous</b> purpura, acrocyanosis, necrosis, ulcers,	MGUS,	RF-IgM/IgG: +
	IgG2 (mainly), IgG3 IgA (rarely)	livedo reticularis	SM, Waldeström macroglobulinaemia	FLC
		Extra cutaneous peripheral neuropathy, renal and joint involvement	MM	
		Systemic vasculitis		
MC Type II	monoclonal (RF activity)	<b>Cutaneous</b> Meltzer's Triad	<b>Infectious</b> HCV, HBV, HIV	
	+ <b>polycional</b> IgM+IgG IgG+IgG IgA+IgG (rarely)	<b>Extra cutaneous</b> renal disease, membrano-proliferative glomerulonephritis. neuropathy intestinal ischaemia, alveolar haemorrhage, CNS involvement myocardium failure	Non-infectious Sjögren's syndrome, LES, RA, B-cell Lymphoma, NHL Solid Tumors Cold acquisicin Discass	AutoAb
			Cold-agglutinin Disease Essential MC (unknown cause)	ANA, ENA, AMA Hypo-complementemia ↓C3, ↓C4, ↓CH50
MC Type II	I polyclonal or olygoclonal (RF activity) + polyclonal (microeterogeneous) IgG+IgM	Cutaneous Extra cutaneous peripheral neuropathy, renal and joint involvement	Infectious HBV, HIV, Epstein-Barr, Cytomegalovirus Endocarditis, Spirochetes Fungal infections Parasitosys	<b>RF-IgM/IgG</b> Type II: ++ Type III: ++ <b>FLC</b> k: ++
	IgM+IgG+IgA IgA+IgG		Non-infectious LES, RA Intestinal diseases Biliary cirrhosis Solid Tumors Essential MC	λ: +

Table I. Key features of cryoglobulins and cryoglobulinaemic syndromes.

munological features consisted in low C4 and C3 complement level, RF activity, antinuclear antibodies (ANA), anti-SSA and anticardiolipin antibodies with different frequencies (25).

In many infectious diseases, cryoglobulinaemia has been considered as an epiphenomenon without a clinical significance and related to disease management. Recently, Eble et al. show a high prevalence of CGs level ≤0.05 g/L in clinical practice suggesting that non-HCV CGs level ≤0.05 g/L may be responsible for severe renal and neurological complications, leading to high morbidity and mortality in these patients (24). In patients with HBV-related MC the cryoglobulinaemic glomerulonephritis is the main renal lesion (26). In a multicentre study, the effect of HBV antiviral treatment on cryoglobulinaemic vasculitis, a rare complication occurring in HBV patients, was investigated. A decrease of cryocrit

level was observed at the end of treatment in all patients. Moreover, a reduction of RF and C4 levels was observed in the same cohort (27, 28). Overall, as concluded in a recent review on HBVrelated MC, nucleot(s)ide analogues represent a promising therapeutic option for HBV-related vasculitis (29).

### HCV cryoglobulinaemia

In about one third of HCV infected patients, serum auto-Abs are commonly found, mainly anti-smooth muscle (ASMA) and ANA, with an increased prevalence in females (30, 31). This is probably due to direct transformation of infected B cell but also to HCV chronic stimulation of a limited pool of autoreactive B cells (32, 33).

Lenzi *et al.* observed that in chronic HCV-patients, the anti-liver and kidney microsomal antibody type 1 (anti-LKM1) positivity correlates with the activity of liver disease, suggesting a role in the progression of liver damage (34). Gregorio *et al.* showed that the reactivity to the nuclear and smooth muscle peptides homologous to HCV antigens is strongly associated with chronic HCV infection, indicating that these auto-Abs are generated as a specific response to HCV infection and that represent a consequence of crossreactive immune responses (35).

Analysing HCV-patients with rheumatoid arthritis (RA) in comparison with non-HCV RA, we found an increased prevalence of ANA in HCV-RA group, whereas anti-mitochondrial antibodies, ASMA, anti-parietal cell antibodies and anti-LKM 1 (performed by indirect immunofluorescence) were negative. Moreover, ANA were observed above all in cryoprecipitates (82%) compared to the supernatant (48%). The ANA pattern was also different: in non-HCV-RA patients, the coarsely speckled pattern was the quite frequently occurred,

#### Cryoglobulins and immune response / U. Basile et al.



Fig. 1. The phases of laboratory workup: pre-analytical, analytical and characterisation of the cryoprecipitate.

completely absent among HCV-RA patients, while, the finely speckled pattern was mostly frequent in HCV-RA group. The occurrence of non-specific patterns in HCV-positive patients could be linked to a non-specific antibody response, possibly due to the persistent antigenic stimulus of the virus (7). We also found that presence of ANA seems strictly related to type II and type III CGs production, moreover ANA positive patients showed a higher mean percentage of cryocrit (8, 36).

Several factors, often interconnected, seem to contribute to the pathogenesis and progression of lymphoproliferative and/or autoimmune diseases in HCVinfected patients such as different IgG subclasses patterns (12). Human IgG include four subclasses: IgG1 (60-65% total IgG), IgG2 (20-25%), IgG3 (5-10%) and IgG4 (3-6%), which display different chemical and physical properties and whose clinical significance is not completely known. Little is known about the role of IgG subclasses in the pathophysiology of cryoglobulinaemia, but different studies displayed that IgG subclasses may characterised different syndromes and autoimmune disorders (37-39). The analysis of IgG subclasses distribution in patients with HCV-cryoglobulinaemia resulted in the enrichment of certain IgG subclasses, mainly IgG1 and IgG3 frequently monoclonal (40).

Serum distribution of IgG subclasses and their role in patients with cryoglobulinaemia has been evaluated since 1971 when Virella et al. reported that IgG3 may trigger concentration-dependent aggregates and the hypothesis of the high interactions towards similar molecules, possibly due to conformational reason, was confirmed by the prevalence of IgG3 proteins among IgG CGs (41). These data agree with the analysis of Ig components in murine CGs that confirmed the auto reactivity of IgG3 and showed the univocal presence of the IgG3 subclass and IgG3 monoclonal CGs with RF activity that might be responsible for extra-hepatic manifestations (42). Human and murine IgG3 display an equal capacity to self-assembly and demonstrate a role in the formation of cryoprecipitate. IgG3 induces effector functions acting as a potent pro-inflammatory antibody, therefore its short half-life may function to limit an abnormal inflammatory response (43).

In our study on naïve HCV patients without clinical signs of autoimmune/ lymphoproliferative disorders, we described a strong correlation between ANA positivity and IgG3 subclass and the prevalence of the finely speckled pattern in the IgG3-positive HCV-positive subgroup (36). In addition, most of the RF IgG3 positive patients were ANA positive. IgG3 is indicative of a RF activity against the IgG1-HCV complex, and their autoreactivity may indicate a persistent immune stimulation (36). RF is an autoantibody against the Fc

portion of human IgG enabling them to form immune complexes; this ability is important in the pathogenesis of cryoglobulinaemic vasculitis (33). RF was found in different diseases with immune dysregulation. Moreover, the presence of RF and a reduction of C4 can be observed in most MC patients (12). A cross-reactivity between the HCV, which acts as antigen, and RF IgM or IgG has been demonstrated; many reports showed that the persistence of the virus induces monoclonal (after a transition from oligoclonal) RF. The continuous stimulation of B cells by the virus can induce the development of malignant non-Hodgkin lymphomas as suggested from the evidence that the treatment of infection leads to clinical improvement and lymphoma regression in most patients (44-46) (Fig. 2).

The occurrence of RFs reactive production by HCV stimulus, in patients with genetic abnormalities, may represent the trigger for a neoplastic process leading to the development of lymphoma (47, 48). Sansonno et al. reported that B cells, isolated from the liver of infected patients, produced WA, a specific monoclonal RF bearing the WA cross-idiotype, in about 60% of cases of HCV-related type II MC (49, 50). Antiviral therapy induces the decline of WA B cells secreting WA monoclonal RF and cryoglobulinaemia, in parallel with the decrease of viraemia, suggesting that the production of WA monoclonal RF is driven by HCV (51). WA B cells can also be detected in asymptomatic patients with HCV infection; thus, this could be a marker to predict a cryoglobulinaemic vasculitis and to initiate early antiviral therapy (51).

In HCV patients, the measurement of serum FLC can be clinically useful above all as a predictive index of MC and for the diagnostic and prognostic value of the  $k/\lambda$  ratio. High free k serum levels were found in HCV positive patients with type II MC, characterised by a monoclonal component mounting light chains k (52-53), and their serum levels have recently been used for monitoring extrahepatic manifestations caused by HCV (18). The abnormal  $k/\lambda$  ratio correlates positively with the severity of clinical data associated





with the lymphoproliferative disorders related to HCV infection, and represents an early index of activation of the lymphoproliferative mechanism in the follow-up of patients with infection HCV (53).

Our group recently observed that FLC assessment could be used as a diagnostic marker of MC vasculitis/syndrome. Cryoglobulinaemia is associated exclusively with B cell proliferative diseases (33), due to different pathogenetic factor which leads to aberrant B cell functions. Pre-treatment FLC ratio values may be considered an important, cost-effective, and easily accessible parameter for predicting patient response to expensive treatments, such B cells depletion with anti-CD20 monoclonal antibody (54).

The pathogenetic correlation between MC and HCV suggests that the virus may be responsible of a multistep process that supports the progression for a wide range of extra-hepatic manifestations, independently from the development of infection and it may be responsible for the proliferation of B lymphocytes which trigger the production of circulating immune complexes composed of CGs and auto-Abs.

# Mixed cryoglobulinaemia and B cells

Many evidences showed that circulating B cells in HCV-MC patients are profoundly disturbed and in most of the cases an expansion of CD2110W Bcells replace the normal B cell pool (55-57). CD21<sup>low</sup> B-cells in MC are very similar to the CD2110w B-cells found in other immunological and chronic infectious diseases (58-61), characterised by B cell hyperactivation. Phenotypically, expanded CD2110w B-cells in MC resemble marginal zone B-cells by the co-expression of IgM and CD27 and are characterised by a peculiar pattern of homing receptors including: CD11c, a typical marker of dendritic cells; CXCR3, CCR7, CD62L, CXCR4, and CXCR5, and inhibitory receptors, as CD22, FcyRIIB (CD32b), CD72, CD85j, CD85k, CD95, LAIR-1, Siglec-6 and Fc receptor-like 4 (FCRL4), an inhibitory receptor that was first described in a subset of human tonsillar B cells with low CD21 expression (58, 62-64).

In 20% of MC patients clonal CD21<sup>low</sup> B-cells express a B cell receptor (BCR) encoded by the VH1-69 and Vk3-20 heavy and light variable genes. This BCR displays RF activity and is putatively directed to an unknown HCV antigen (65, 66).

CD21<sup>low</sup> B-cells resemble exhausted virus-specific T cells, described in HIV and HCV infections for the expression of inhibitory receptors and the low proliferative potential in response to different stimuli, leading to their definition as exhausted cells (67). Indeed, CD21<sup>low</sup> B-cells fail to proliferate after stimulation of TLR 7 and 9 and BCR, whereas it seems that co-stimulation with CD40L, BCR, in the presence of

IL-10 and IL-2 induces an increased proliferative response (56-58, 62). Stimulation of TLR9, however, enhances secretion of TNF $\alpha$  and RF antibodies by these cells suggesting partial unresponsiveness (68). In addition, CD21<sup>low</sup> B-cells appear to be anergic as they fail to flux calcium upon B-cell receptor triggering (69), display high constitutive expression of the active phosphorylated form of extracellular signal-regulated kinase (pERK) and are prone to die by apoptosis (57, 69-70). These functional signatures usually characterise murine B cells made anergic by continual BCR engagement by antigen (71). In accordance, dysregulated expression of apoptosis and anergy-related genes has been found in MC-derived clonal B cells and high expression of DEC1/STRA13, a negative regulator of B cell activation, seems to be a common trait (62).

Recently the occurrence of clonal B cells in MC after eradication of HCV, achieved with direct acting antivirals (DAA), has been studied (72). DAAs offer a unique opportunity to analyse the B cell functional properties after removal of chronic antigen stimulation as they lack the immunomodulatory effect, typical of interferon (IFN)-based therapies. It was shown that the anergic features of clonal B cells, as pERK overexpression and accelerated apoptosis, rapidly reverted after disengagement from HCV, while phenotypic and functional features of exhaustion persist in peripheral blood after sustained viral response for several months, suggesting that other extracellular signals might be responsible for their survival. In the long-term immunological and clinical outcomes of patients with mixed cryoglobulinaemic vasculitis (MCV) treated with DAAs, with or without non-Hodgkin's lymphoma (NHL), circulating B-cell clones persisted in most of patients with MCV or with MCV associated with indolent NHL for up to 2 years after the cure of HCV infection (73). The persistence of B-cell clones did not correlate with serum CGs or vasculitis response to therapy or relapse after viral eradication. In this view it is noteworthy that some MC patients, after HCV eradication, experience a vasculitis' relapse associated with the occurrence of infections, vaccination, and tumours (74, 75). It can be speculated that RF bearing monoclonal B cells in MC might be reactivated by circulating immune complexes produced in high quantities in condition such as infections or tumours. Therefore, as suggested by Kaplan et al., the persistence of clonal exhausted B cells in MC, might explain the relapse of MC vasculitis despite clearance of the chronic antigen stimulation provided by HCV (76). Minafò et al. recently investigated the Ig heavy and light chain variable regions of monoclonal B-cells from patients with HCV-associated lymphoproliferative disorders and found that distinctive light chain stereotyped signatures either endowed with homology with anti-HCV E2 antibodies or with RF sequences might help distinguishing HCV-dependent lymphoproliferative disorders from those autoantigen driven, accidentally occurring in HCV infected individuals (77). These observations strengthen the hypothesis that other stimuli, than infective ones, might contribute for the survival and transformation of B cell clones in HCV-associated MC.

# Mixed cryoglobulinaemia and therapeutic strategies

Considering the pathogenesis of MC, different therapeutic strategies have been developed in recent years and are available for patients suffering from this vasculitis. Some of these treatments are curative whereas other have a limited effect (78, 79). Effectiveness of IFN $\alpha$  was observed empirically in 1987 and after the demonstration of the pathogenic role of HCV in MC, treatment with IFN $\alpha$  became a rational therapeutic strategy, although sustained virologic response and vasculitis disappearance was observed in less than half of the patients (80). Recently the advent of new direct acting anti-viral changed the scenario of HCV-MC achieving sustained virologic response in almost 100% of the cases, together with vasculitis disappearance (81). After this initial enthusiasm, long-term follow-up of MC patients showed that vasculitis relapse was a possible, and not so rare

event (82) and the persistence of peripheral B cell clones might be involved in vasculitis reactivation (73, 74). In this view the addition of anti-CD20 treatment could be ideally the optimal therapeutic strategy as DAA achieve sustained virologic response and the depletion of B cells could prevent future relapse. Efficacy of rituximab, a B-cell depleting anti-CD20 monoclonal antibody, has been widely demonstrated both for HCV and essential MC, but its effect is limited, and retreatments are usually necessary (83). If circulating B cell clones reappear after B cell depletion and virus eradication has not been investigated so far.

Other therapeutic interventions include plasmapheresis that is usually used for life-threatening forms of vasculitis and allows the temporary removing of the cryoprecipitating proteins (84).

# Mixed cryoglobulinaemia, COVID-19 and endothelitiis

The viral aetiology in MC vasculitis is well defined in most of cases. Given the potential impact of inflammatory and immunopathological complications in severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) disease 2019 (COVID-19), here we speculated about a potential link between MC and COVID-19 based on very recent literature. Even if at now reports describing the occurrence of MC vasculitis in COVID patients are still lacking, at least two mechanisms can be envisaged to hypothesise a damage of Coronavirus on endothelial cells. Indirectly, as it has been reported that in COVID-19 vasculitis a life-threatening escalation of a dramatic immune dysregulated response, with a switch from a pathogenetic mechanism of type 2 T-helper immune response (humoral immunity) to a type III hypersensitivity (immune complex disease), takes place (85). The deposition of immune complexes induces an amplification of the inflammatory state evolving to cytokine storm syndrome. During SARS-CoV-2 infection, type III hypersensitivity might occur for an excess of soluble antigens determining an accumulation of immune complexes. These antigenantibody complexes may precipitate inside tissues, in blood vessels, inducing a severe inflammatory state by the action of complement anaphylatoxins (C3a and C5a) and tissue damage. The result of this process inside the walls of blood vessels is a vasculitis sustained by a mechanism of type III hypersensitivity, that is depicted by an acute necrotising vasculitis with neutrophilic infiltration, karyorrhexis and fibrinoid necrosis (85).

Directly, it has been demonstrated that SARS-CoV-2 can infect engineered human blood vessel organoids *in vitro* (86). The angiotensin converting enzyme 2 (ACE2) receptors, used by the SARS-CoV-2 to infect the host, are largely expressed by endothelial cells which traverse multiple organs; evidence of direct viral infection of the endothelial cells *in vivo* and diffuse endothelial inflammation (endothelitiis) in several organs has been reported in post mortem histological analysis of different tissues (87).

The combination of these two postulated mechanisms (supported by demonstrated evidence) let open the possibility that SARS-CoV-2 triggered dysregulated-immune response (that becomes independent from virus itself) may impair blood vessels resulting in a vasculitis (as observed in HCV-MC). The worsening progression is widely influenced by both endothelial-induced damage to vital organs and co-morbidities associated with underlying diseases.

### Conclusions

The dysregulation of immune system, involved in cryoprecipitation, was here reviewed. MC displays a double face being a lymphoproliferative and autoimmune disease as well, and CGs can be considered as the result of the adaptive immune response, induced by the persistence of endogenous (autoimmune diseases) or exogenous stimuli (virus).

Since their occurrences increase comorbidities and reduce survival, it is important to detect serum CGs in all patients with immunological alterations. The early diagnosis of a cryoglobulinemia and the worsening evolution from a mild form of vasculitis to a severe and life-threatening disease up to the development of non-Hodgkin lymphomas requires a careful analysis and integration of all clinical, laboratory and histological data. Even in the era of DAA that achieve HCV eradication in almost 100% of patients, relapse of MC vasculitis or non-response to treatment have been described (74). For these reasons, the deeply comprehension of underlying mechanisms together with the definition of biomarkers indicative for unresponsiveness to therapy or predictive for disease relapse are widely desired and might ameliorate patients tailored treatments.

The expansion of abnormal clonal B cells producing cryoglobulins could become independent to the trigger and virus eradication. Cryoglobulinaemia could represent a biological model of a new systemic disease induced by a virus as trigger of a dysregulated immune response that, in the end, becomes independent from the virus itself.

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#### References

- STONE MJ: Waldenstrom's macroglobulinemia: hyperviscosity syndrome and cryoglobulinemia. *Clin Lymphoma Myeloma* 2009; 9: 97-9.
- SARGUR R, WHITE P, EGNER W: Cryoglobulin evaluation: best practice? Ann Clin Biochem 2010; 47: 8-16.
- 3. DAMOISEAUX J: The diagnosis and classification of the cryoglobulinemic syndrome. *Autoimmun Rev* 2014; 13: 359-62.
- HEIDELBERGER M, KENDALL FE: A quantitative study of the precipitin reaction between type III pneumococcus polysaccharide and purified homologous antibody. J Exp Med 1929; 50: 809-23.
- 5. WINTROBE MM, BUELL MV: Hyperproteinemia associated with multiple myeloma: With report of a case in which an extraordinary hyperproteinemia was associated with

thrombosis of the retinal veins and symptoms suggesting Raynaud's disease. *Bull Johns Hopkins Hosp* 1933; 52: 156-65.

- BROUET JC, CLAUVEL JP, DANON F, KLEIN M, SELIGMANN M: Biologic and clinical significance of cryoglobulins. A report of 86 cases. *Am J Med* 1974; 57: 775-88.
- BASILE U, GULLI F, TORTI E et al.: Antinuclear antibody detection in cryoprecipitates: Distinctive patterns in hepatitis C virus-infected patients. *Dig Liver Dis* 2015; 47: 50-6.
- BASILE U, GULLI F, GRAGNANI L et al.: Different biochemical patterns in type II and type III mixed cryoglobulinemia in HCV positive patients. *Dig Liver Dis* 2018; 50: 938-43.
- MUSSET L, DIEMERT MC, TAIBI F et al.: Characterization of cryoglobulins by immunoblotting. Clin Chem 1992; 38: 798-802.
- TISSOT JD, SCHIFFERLI JA, HOCHSTRASSER DF *et al.*: Two-dimensional polyacrylamide gel electrophoresis analysis of cryoglobulins an identification of an IgM-associated peptide. *J Immunol Meth* 1994; 173: 63-75.
- RAMOS-CASALS M, STONE JH, CID MC, BOSCH X: The cryoglobulinaemias. *Lancet* 2012; 379: 348-60.
- 12. BASILE U, GULLI F, GRAGNANI L *et al.*: IgG3 subclass: A possible trigger of mixed cryoglobulin cascade in hepatitis C virus chronic infection. *Dig Liver Dis* 2017; 49: 1233-39.
- 13. BASILE U, TORTI E, DELL'ABATE MT *et al.*: Pre-analytical phase in cryoglobulin (CRG) detection: an alternative method for sample transport. *Clin Chem Lab Med* 2016; 54: e123-26.
- 14. GULLI F, SANTINI SA, NAPODANO C et al.: Cryoglobulin Test and Cryoglobulinemia Hepatitis C-Virus Related. *Mediterr J Hema*tol Infect Dis 2017; 9: e2017007.
- OTANI M, KUROKI A, KIKUCHI S *et al.*: Sialylation determines the nephritogenicity of IgG3 cryoglobulins. *J Am Soc Nephrol* 2012; 23: 1869-78.
- DESBOIS AC, CACOUB P, SAADOUNA D: Cryoglobulinemia: An update in 2019. *Joint Bone Spine* 2019; 86:707-13.
- ZIGNEGO AL, WOJCIK GL, CACOUB P et al.: Genome-wide association study of hepatitis C virus- and cryoglobulin-related vasculitis. *Genes Immun* 2014; 15: 500-5.
- BASILE U, GULLI F, GRAGNANI L et al.: Free light chains: Eclectic multipurpose biomarker. J Immunol Methods 2017; 451: 11-9.
- NAPODANO C, POCINO K, RIGANTE D et al.: Free light chains and autoimmunity. Autoimmun Rev 2019; 18: 484-92.
- GULLI F, NAPODANO C, MARINO M et al.: Serum immunoglobulin free light chain levels in systemic autoimmune rheumatic diseases. Clin Exp Immunol 2020; 199: 163-71.
- BASILE U, NAPODANO C, POCINO K et al.: Serological profile of asymptomatic HCV positive patients with low level of cryoglobulins. *Biofactors* 2019; 45: 318-25.
- BASILE U, MARINO M, GRAGNANI L et al.: Sentinel biomarkers in HCV positive patients with mixed cryoglobulinemia. J Immunol Methods 2020; 476: 112687.
- 23. GERI G, TERRIER B, SEMOUN O et al.:

### Cryoglobulins and immune response / U. Basile et al.

Surrogate markers of B cell non-Hodgkin's lymphoma in patients with hepatitis C virusrelated cryoglobulinaemia vasculitis. *Ann Rheum Dis* 2010; 69: 2177-80.

- 24. EBLE V, LEGALLICIER B, JOLY P *et al.*: Long term outcome of patients with low level of cryoglobulin (<0.05g/L). *Autoimmun Rev* 2016; 15: 440-6.
- 25. SAADOUN D, SELLAM J, GHILLANI-DALBIN P, CRECEL R, PIETTE JC, CACOUB P: Increased risks of lymphoma and death among patients with non-hepatitis C virus-related mixed cryoglobulinemia. *Arch Intern Med* 2006; 166: 2101-8.
- 26. LI SJ, XU ST, CHEN HP *et al.*: Clinical and morphologic spectrum of renal involvement in patients with HBV-associated cryoglobulinaemia. *Nephrology* (Carlton) 2017; 22: 449-55.
- 27. CACOUB P, SAADOUN D, BOURLIÈRE M et al.: Hepatitis B virus genotypes and extrahepatic manifestations. J Hepatol 2005; 43: 764-70.
- MAZZARO C, DAL MASO L, URRARO T et al.: Hepatitis B virus related cryoglobulinemic vasculitis: A multicentre open label study from the Gruppo Italiano di Studio delle Crioglobulinemie - GISC. *Dig Liver Dis* 2016; 48: 780-4.
- 29. MAZZARO C, DAL MASO L, VISENTINI M *et al.*: Hepatitis B virus-related cryogobulinemic vasculitis. the role of antiviral nucleot(s) ide analogues: a review. *J Intern Med* 2019; 286: 290-8.
- YANG DH, HO LJ, LAI JH: Useful biomarkers for assessment of hepatitis C virus infectionassociated autoimmune disorders. *World J Gastroenterol* 2014; 20: 2962-70.
- 31. CZAJA, AJ, MANNS MP, HOMBURGER HA: Frequency and significance of antibodies to liver/kidney microsome type 1 in adult with cronic hepatitis. *Gastroenterology* 1992; 103: 1290-5.
- 32. ZIGNEGO AL, GRAGNANI L, PILUSO A et al.: Virus-driven autoimmunity and lymphoproliferation: The example of HCV infection. Expert Rev Clin Immunol 2015; 11: 15-31.
- 33. ROCCATELLO D, SAADOUN D, RAMOS-CASALS M et al.: Cryoglobulinaemia. Nat Rev Dis Primers 2018; 4: 11.
- 34. LENZI M, BELLENTANI S, SACCOCCIO G et al.: Prevalence of non-organ-specific autoantibodies and chronic liver disease in the general population: a nested case-control study of the Dionysos cohort. *Gut* 1999; 45: 435-41.
- 35. GREGORIO GV, CHOUDHURI K, MA Y et al.: Mimicry between the hepatitis C viruspolyprotein and antigenic targets of nuclear and smooth muscle antibodiesin chronic hepatitis C virus infection. *Clin Exp Immunol* 2003; 133: 404-13.
- 36. GULLI F, BASILE U, GRAGNANI L et al.: Autoimmunity and lymphoproliferation markers in naïve HCV-RNA positive patients without clinical evidences of autoimmune/ lymphoproliferative disorders. *Dig Liver Dis* 2016; 48: 927-33.
- NAPODANO C, MARINO M, STEFANILE A et al.: Immunological Role of IgG Subclasses. Immunol Invest 2020; 11: 1-18.
- 38. GULLI F, BASILE U, GRAGNANI L et al .:

IgG cryoglobulinemia. *Eur Rev Med Pharmacol Sci* 2018; 22: 6057-62.

- 39. MARINO M, BASILE U, SPAGNI G et al.: Long-lasting rituximab-induced reduction of specific-but not total-IgG4 in MuSK-positive myasthenia gravis. *Front Immunol* 2020; 11: 613
- 40. WONG VS, EGNER W, ELSEY T, BROWN D, ALEXANDER GJ: Incidence, character, and clinical relevance of mixed cryoglobulinemia in patients with chronic hepatitis C virus infection. *Clin Exp Immunol* 1996; 104: 25-31.
- VIRELLA G: IgG subclasses in relation to viscosity and cryoglobulin syndromes. *Br Med J* 1971; 2: 322.
- 42. IZUI S, BERNEY T, SHIBATA T, FULPIUS T: IgG3 cryoglobulins in autoimmune MRL-Ipr/ Iprl mice: immunopathogenesis, theurapetic approaches and relevance to similar human diseases. Ann Rheum Dis 1993; 52: S48-54.
- VIDARSSON G, DEKKERS G, RISPENS T: IgG subclasses and allotypes: from structure to effector functions. *Front Immunol* 2014; 5: 520.
- 44. ARCAINI L, BESSON C, FRIGENI M *et al.*: Interferon-free antiviral treatment in B-cell lymphoproliferative disorders associated with hepatitis C virus infection. *Blood* 2016; 128: 2527-32.
- 45. ARCAINI L, VALLISA D, RATTOTTI S et al.: Antiviral treatment in patients with indolent B-cell lymphomas associated with HCV infection: a study of the Fondazione Italiana Linfomi. Ann Oncol 2014; 25: 1404-10.
- 46. ZIGNEGO AL, GIANNINI C, GRAGNANI L: HCV and lymphoproliferation. *Clin Dev Immunol* 2012; 2012: 980942.
- 47. GRAGNANI L, FOGNANI E, PILUSO A, ZIGNE-GO AL: Hepatitis C-associated B-cell non-Hodgkin lymphomas: The emerging role of miRNA-26b. J Hepatol 2013; 59: 1362-3.
- MOLL J, ISAILOVIC N, DE SANTIS M, SELMI C: Rheumatoid factors in hepatitis B and C infections: connecting viruses, autoimmunity, and cancer. *Isr Med Assoc J* 2019; 21: 480-86.
- 49. SANSONNO D, DE VITA S, IACOBELLI AR, CORNACCHIULO V, BOIOCCHI M, DAMMAC-CO F: Clonal analysis of intrahepatic B cells from HCV-infected patients with and without mixed cryoglobulinemia. *J Immunol* 1998; 160: 3594-601.
- 50. AGNELLO V, ZHANG QX, ABEL G, KNIGHT GB: The association of hepatitis C virus infection with monoclonal rheumatoid factors bearing the WA cross-idiotype: implications for the etiopathogenesis and therapy of mixed cryoglobulinemia. *Clin Exp Rheumatol* 1995; 13 (Suppl. 13): S101-4.
- 51. KNIGHT GB, GAO L, GRAGNANI L et al.: Detection of WA B cells in hepatitis C virus infection: a potential prognostic marker for cryoglobulinemic vasculitis and B cell malignancies. Arthritis Rheum 2010; 62: 2152-9.
- 52. DISPENZIERI A, KYLE R, MERLINI G et al.: International Myeloma Working Group guidelines for serum-free light chain analysis in multiple myeloma and related disorders. *Leukemia* 2009; 23: 215-24.
- 53. TERRIER B, SENE D, SAADOUN D et al.: Serum-free light chain assessment in hepatitis C virus-related lymphoproliferative disorders. Ann Rheum Dir 2009; 68: 89-93.

- 54. BASILE U, GRAGNANI L, PILUSO A et al.: Assessment of free light chains in HCV positive patients with mixed cryoglobulinemia vasculitis undergoing rituximab treatment. *Liver Int* 2015; 35: 2100-7.
- 55. CARBONARI M, CAPRINI E, TEDESCO T et al.: Hepatitis C virus drives the unconstrained monoclonal expansion of VH1-69-expressing memory B-cells in type II cryoglobulinemia: a model of infection-driven lymphomagenesis. J Immunol 2005; 174: 6532-39.
- 56. VISENTINI M, CONTI V, CRISTOFOLETTI C et al.: Clonal expansion and functional exhaustion of monoclonal marginal zone B cells in mixed cryoglobulinemia: the yin and yang of HCV-driven lymphoproliferation and autoimmunity. Autoimmun Rev 2013; 12: 430-5.
- 57. CHARLES ED, BRUNETTI C, MARUKIAN S et al.: Clonal B-cells in patients with hepatitis C virus-associated mixed cryoglobulinemia contain an expanded anergic CD21low B-cell subset. *Blood* 2011; 117: 5425-37.
- 58. RAKHMANOV M, KELLER B, GUTENBERG-ER S et al.: Circulating CD21low B cells in common variable immunodeficiency resemble tissue homing, innate-like B cells. Proc Natl Acad Sci USA 2009; 106: 13451-6.
- 59. SAADOUN D, TERRIER B, BANNOCK J et al.: Expansion of autoreactive unresponsive CD21-/low B cells in Sjögren's syndromeassociated lymphoproliferation. Arthritis Rheum 2013; 65: 1085-96.
- 60. THORARINSDOTTIR K, CAMPONESCHI A, JONSSON C et al.: CD21-/low B cells associate with joint damage in rheumatoid arthritis patients. Scand J Immunol 2019; 29: e12792.
- MOIR S, HO J, MALASPINA A et al.: Evidence for HIV- associated B-cell exhaustion in a dysfunctional memory B-cell compartment in HIV-infected viremic individuals. J Exp Med 2008; 205: 1797-805.
- 62. VISENTINI M, CAGLIUSO M, CONTI V et al.: Clonal B cells of HCV- associated mixed cryoglobulinemia patients contain exhausted marginal zone-like and CD21(low) cells overexpressing Stra13. Eur J Immunol 2012; 42: 1468-76.
- 63. ISNARDI I, NG YS, MENARD L *et al.*: Complement receptor 2/CD21- human naive B cells contain mostly autoreactive unresponsive clones. *Blood* 2010; 115: 5026-36.
- 64. EHRHARDT GR, HSU JT, GARTLAND L et al.: Expression of the immunoregulatory molecule FcRH4 defines a distinctive tissuebased population of memory B cells. J Exp Med 2005; 202: 783-91.
- 65. DE RE V, DE VITA S, MARZOTTO A *et al.*: Sequence analysis of the immunoglobulin antigen receptor of hepatitis C virus-associated non-Hodgkin lymphomas suggests that the malignant cells are derived from the rheumatoid factor-producing cells that occur mainly in type II cryoglobulinemia. *Blood* 2000; 96: 3578-84.
- 66. NG PP, KUO CC, WANG S *et al.*: B-cell receptors expressed by lymphomas of hepatitis C virus (HCV)-infected patients rarely react with the viral proteins. *Blood* 2014; 123: 1512-15.
- 67. DAY CL, KAUFMANN DE, KIEPIELA P et al.: PD-1 expression on HIV-specific T cells is associated with T-cell exhaustion and disease

# Cryoglobulins and immune response / U. Basile et al.

progression. Nature 2006; 443: 350-4.

- COMARMOND C, LORIN V, MARQUES C et al.: TLR9 signalling in HCV-associated atypical memory B cells triggers Th1 and rheumatoid factor autoantibody responses. J Hepatol 2019; 71: 908-19.
- 69. TERRIER B, JOLY F, VAZQUEZ T *et al.*: Expansion of functionally anergic CD21-/ low marginal zone-like B cell clones in hepatitis C virus infection-related autoimmunity. *J Immunol* 2011; 187: 6550-63.
- 70. VISENTINI M, MARRAPODI R, CONTI V et al.: Dysregulated extracellular signal-regulated kinase signaling associated with impaired B-cell receptor endocytosis in patients with common variable immunodeficiency. J Allergy Clin Immunol 2014; 134: 401-10.
- GAULD SB, BENSCHOP RJ, MERRELL KT, CAMBIER JC: Maintenance of B cell anergy requires constant antigen receptor occupancy and signaling. *Nat Immunol* 2005; 6: 1160-7.
- 72. DEL PADRE M, TODI L, MITREVSKI M et al.: Reversion of anergy signatures in clonal CD-2110w B-cells of mixed cryoglobulinemia after clearance of HCV viremia. *Blood* 2017; 130: 35-8.
- 73. VISENTINI M, DEL PADRE M, COLANTUONO S et al.: Long-lasting persistence of large Bcell clones in hepatitis C virus-cured patients with complete response of mixed cryoglobulinaemia vasculitis. *Liver Int* 2019; 39: 628-32.

- 74. VISENTINI M, QUARTUCCIO L, DEL PADRE M et al.: Late relapses of hepatitis C viruscured mixed cryoglobulinaemia associated with infection or cancer. *Rheumatology* (Oxford) 2018; 57: 1870-1.
- 75. SOLLIMA S, MILAZZO L, VASSALINI P, ANTI-NORI S, GALLI M: Recurrence of mixed cryoglobulinaemia vasculitis following influenza vaccination despite clearance of hepatitis C virus infection. *Clin Exp Rheumatol* 2018; 36: 161-2.
- 76. KAPLAN DE: Persistence of exhaustion in cured hep C. *Blood* 2017; 130: 4.
- 77. MINAFÒ YA, DEL PADRE M, CRISTOFOLETTI C, CAPRINI E, PEREZ M, ARANBURU A: A stereotyped light chain may shape virusspecific B-cell receptors in HCV-dependent lymphoproliferative disorders. *Genes Immun* 2020; Feb 18.
- DAMMACCO F, SANSONNO S: Therapy for Hepatitis C virus-related cryoglobulinemic vasculitis. N Engl J Med 2013; 369: 1035-45.
- 79. MAZZARO C, DAL MASO L, QUARTUCCIO L et al.: Long-term effects of the new direct antiviral agents (DAAs) therapy for HCV-related mixed cryoglobulinaemia without renal involvement: a multicentre open-label study. *Clin Exp Rheumatol* 2018; 36 (Suppl. 111): S107-14.
- 80. GRAGNANI L, FOGNANI E, PILUSO A *et al.*: Long-term effect of HCV eradication

in patients with mixed cryoglobulinemia: a prospective, controlled, open- label, cohort study. *Hepatology* 2015; 61: 1145-53.

- 81. GRAGNANI L, VISENTINI M, FOGNANI E et al.: Prospective study of guideline-tailored therapy with direct-acting antivirals for hepatitis C virus-associated mixed cryoglobulinemia. *Hepatology* 2016; 64: 1473-82.
- BONACCI M, LENS S, MARIÑO Z et al.: Long-term outcomes of patients with HCVassociated cryoglobulinemic vasculitis after virologic cure. *Gastroenterology* 2018; 155: 311-15.
- 83. COLANTUONO S, MITREVSKI M, YANG B et al.: Efficacy and safety of long-term treatment with low-dose rituximab for relapsing mixed cryoglobulinemia vasculitis. Clin Rheumatol 2017; 36: 617-23.
- DAMMACCO F, SANSONNO S: Therapy for Hepatitis C virus-related cryoglobulinemic vasculitis. N Engl J Med 2013; 369: 1035-45.
- RONCATI L, LIGABUE G, FABBIANI L et al.: Type 3 hypersensitivity in COVID-19 vasculitis. Clin Immunol 2020; 217: 108487.
- MONTEIL V, KWON H, PRADO P et al.: Inhibition of SARS-CoV-2 infections in engineered human tissues using clinical-grade soluble human ACE2. Cell 2020; 181; 905-13.
- 87. VARGA Z, FLAMMER AJ, STEIGER P et al.: Endothelial cell infection and endotheliitis in COVID-19. *Lancet* 2020; 395: 1417-8.