

# Searching for the “X factor” in Sjögren’s syndrome female predilection

L.G. Chatzis, A.V. Goules, A.G. Tzioufas

Department of Pathophysiology, School of Medicine, National and Kapodistrian University of Athens, and Institute for Autoimmune Systemic and Neurological Diseases, Athens, Greece.

Loukas G. Chatzis, MD  
Andreas V. Goules, MD, PhD  
Athanasios G. Tzioufas, MD

Please address correspondence to:  
Athanasios G. Tzioufas MD,  
Department of Pathophysiology,  
School of Medicine, National and  
Kapodistrian University of Athens,  
Mikras Asias Str. 75,  
115 27 Athens, Greece.

E-mail: agtzi@med.uoa.gr

Received on June 29, 2021; accepted in revised form on August 8, 2021.

*Clin Exp Rheumatol* 2021; 39 (Suppl. 133): S00-S00.

© Copyright CLINICAL AND EXPERIMENTAL RHEUMATOLOGY 2021.

**Key words:** Sjögren’s syndrome, female predilection, autoimmune disease, XCI process X chromosome

### ABSTRACT

*Sjögren’s syndrome is typified by a strong female predilection which is also observed in other systemic autoimmune diseases. Although many factors may be contributing to this phenomenon, the exact underlying mechanisms remain unclear. Apart from the traditionally considered hormonal and environmental factors, lately the role of sex chromosomes and especially of the X chromosome has drawn much attention. In the current review, we focus on the inherent genetic imbalance between the sex chromosomes and their influence and role on gender-discordant disease presentation. To compensate for this imbalance, nature has created a defective epigenetic mechanism to silence the second rich in immune related genes X chromosome. Genes escaping silencing, transfer the genetic imbalance into the transcriptional and protein level, contributing to gender differences as reflected in functions of the innate and adaptive immunity. Under this prism, recent research data on SS, regarding specific immune X-linked loci are being presented and analysed. The “X Factor” in the search for an explication of women’s predilection in autoimmunity, may lie behind these unique properties of the X chromosome.*

### Introduction

Autoimmune disorders (ADs) represent a range of organ specific or systemic diseases characterised by aberrant immune responses against self-antigens, leading to damage of tissues or organs. Even though single autoimmune diseases are rare, as a group they affect up to 6% of the industrialised general population (1) and are among the leading causes of morbidity and mortality in middle aged women (2, 3).

Importantly, around 78% of those with an autoimmune disease are women (4). The magnitude of female predilection correlates with disease’s prevalence since common diseases show a higher female skewing. For example, autoimmune thyroiditis, the most common organ specific autoimmune disorder, as well as systemic lupus erythematosus (SLE) and Sjögren’s syndrome (SS), the prototypical systemic autoimmune disorders, share the most striking female sex biases. Earlier epidemiologic studies supported a female to male ratio in SLE and SS of 9-11:1, but recent data shows the gender ratio in SS to be even higher at 20:1 (5). Therefore, SS constitutes the most female predominant systemic autoimmune disease, and it presents the best research model to investigate and elucidate the implicated mechanisms favouring a women bias (Table I).

Growing scientific evidence shows that the adult females mount stronger innate and adaptive immunological responses, reflected by a lower risk for serious infections and a better vaccine antibody response, although the pathophysiology has not been thoroughly worked out (6-11). The ongoing worldwide severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pandemic, showing a case mortality ratio of 1,7 males versus 1 female, is another pertinent example (12). In this line, when it comes to immune deregulation, the precise pathogenetic processes that govern autoimmune disorders’ significant sexual dimorphism and predilection are still undetermined, concealing a crucial step both in understanding the fundamental underlying mechanisms of autoimmunity and tailoring a true precision medicine therapeutic approach. It is commonly believed that inherent differences between genders

Competing interests: none declared.

**Table I.** Female to male ratio between systemic and organ specific autoimmune diseases.

Autoimmune disease	Female: Male ratio	Reference
<i>Systemic autoimmune disease</i>		
Sjögren’s syndrome	20:1	[5]
Systemic lupus erythematosus	9:1	[110]
Systemic sclerosis	3:1	[111]
Rheumatoid arthritis	2:1	[112]
<i>Vasculitides</i>		
Giant cell arteritis	3:1	[113]
ANCA vasculitis	1:1	[114]
Takayasu’s arteritis	11:1	[115]
Inflammatory myopathies	2:1	[116]
<i>Seronegative spondyloarthropathies</i>		
Ankylosing spondylitis	1:3	[117]
Psoriatic arthritis	1:1.5	[117]
Reactive arthritis	1:1.2	[117]
<i>Organ specific systemic disease</i>		
Hashimoto thyroiditis	10:1	
Grave’s disease	4:1	[118]
Multiple sclerosis	3:1	[119]
Vitiligo	1:1	[120]
Addison’s disease	1:1.7	[121]
Diabetes mellitus type I	1:1.8	[122]

might contribute to this phenomenon. For decades, the unique female hormonal milieu was considered the sole contributing factor, though offering only partial interpretation, since female predilection exists before puberty or after menopause, and disease course remains mostly unaffected in patients on oral contraceptives or estrogen replacement therapy (13). Recently, the inherent genetic imbalance caused by the unequal dosages of genes on the sex chromosomes has been implicated as well. Females carry two X chromosomes and males one X and one Y, a clamant but until recently overlooked discrepancy between genders. Supportive evidence for the contribution of the X chromosome in the pathogenesis of autoimmune diseases emerges from chromosomal disorders such as the Turner syndrome (X, O), where patients have a very low risk of developing SLE, and the Klinefelter syndrome (XXY), where patients have a 14-fold risk of developing SLE or SS, almost equal to that of euploid women (XX) (14-16). Epigenetic changes related to X chromosome inactivation and skewing as well as maternal microchimerism, a maternal-embryo trafficking of circulating cells during embryogenesis, are also under investigation as con-

tributing factors to the phenomenon. The notion that the second, rich in immune related genes, X chromosome in women, instead of the much smaller but sex determining Y chromosome in males is accountable, partly, for the female predilection in autoimmunity is compelling and emphasizes once more the fate-shaping power of genetics and epigenetics. In the present review we highlight the role of the X chromosome as an important sex biased determinant, the X factor, using Sjögren’s syndrome as a systemic autoimmune disease model.

**Dissecting the X and Y chromosomes**

Both X and Y chromosomes, even though vastly different in structure, size, function, and number of genes, originate from the same pair of homologous chromosomes, known as autosomes (17). Approximately 200 million years ago a male determining mutation occurred in one of the autosomal genes triggering various stepwise processes that hindered recombination, thus allowing the 2 sex chromosomes to evolve independently (18, 19). Despite evolutionary divergence, 2 regions at both termini of X and Y chromosome, called pseudo-autosomal

regions (PAR 1 and PAR 2) containing at least 29 genes, retained homology and the ability to recombine during meiosis (20, 21). The X chromosome encodes around 1100 annotated genes, with few if any implicated in sex determination, compared to the Y chromosome that contains 78 male specific genes crucial for germ cell differentiation, fertility, and masculinization (20, 22). Evidenced by the high number of X related genes implicated in the development of immune cells (Bruton tyrosine kinase, IL2RG), in the innate (TLR 7, TLR 8, IRAK) and adaptive (CD40L, FOXP3) immune responses, is the instrumental role X chromosome imparts in regulating immune homeostasis and maintaining tolerance (23). In this line, mutations in specific genes of the X chromosome may result in immunodeficiency syndromes sharing autoimmune manifestations, including the hyper IgM syndrome, the immunodysregulation polyendocrinopathy enteropathy X-linked (IPEX), and the X-linked recessive severe combined immunodeficiency (XSCID) syndromes caused by mutations in the CD40 ligand, the forkhead box P3 (FOXP3) and the IL-2 receptor genes respectively, all residing in different parts of the X chromosome (24-27). Another recently discovered novel X chromosome-based mutation is that of SAM and SH3 domain-containing 3 (SASH3), which is postulated to orchestrate signal transductions downstream pathways in lymphocytes that, when dysfunctional, result in a clinical phenotype that combines susceptibility to various infections and autoimmune cytopenias (28). Similarly, a limited number of reports have linked specific Y located genes with genome wide regulatory properties, such as the Sly and Rbmy genes charged with immune modulation and conferring susceptibility to autoimmune diseases (29). The X chromosome accounts for more than 5% of the total DNA in each cell, approximately 3 times more than the Y chromosome, an imbalance even more apparent in the gene apparatus, since the former contains more than 100 times the number of transcriptionally active genes leading to an obvious ge-

netic inequity between females (XX) and males (XY). To relieve those imbalances, in female mammals, early in embryogenesis, either paternal or maternal X chromosome gets epigenetically inactivated randomly by a process called X chromosome inactivation (XCI), forming a heterochromatin nucleolar satellite in contact with the nucleus membrane, called Barr body (30-32). Failure of X chromosome silencing leads to lethality due to the overabundance of X gene related products that interfere with normal cell function (33, 34). In each embryonic cell the assortment of X chromosome rendered inactive is random and irreversible, resulting in a mosaic pattern of expression in cells originating from different germ layers. However, the supposedly random delegation process is not always perfect, and skewed patterns in which more than 75% of cells preferentially inactivate the maternal or paternal X chromosome have been observed. Cross-sectional studies have supported a possible influence of aging on the degree of skewing, though without confirmation by more recent longitudinal studies (35). Interestingly, patients with autoimmune diseases have been related to extreme skewing (>90%), despite several conflicting reports (36). A proposed pathogenetic explanation connecting chromosome skewing with the ignition of the autoimmunity cascade, implicates the X linked self-antigens derived from the residual cell population after non-random skewing. In this case, these self-autoantigens behave as cryptoantigens that may escape central thymic T cell tolerance, resulting in autoantigen presentation and generation of pathologic autoreactive T cell clones later in life (37-40). It is known, however, that in females harbouring structural alterations on one X chromosome, a preferential expression of the other normal chromosome occurs. The ability of the cell to deliberately silence the harmful chromosome, as it happens in X-linked agammaglobulinemia and Wiskott-Aldrich syndrome, is an evolutionary phenomenon, clearly offering a survival advantage in women carriers. A complete explanation for skewing

when both chromosomes are structurally sound remains elusive (41-43). Except for the extraordinary epigenetic transcriptional silencing mechanism of skewed X-chromosome inactivation, a post transcriptional crucial regulatory role has been also assigned to the X chromosome, since it affords the second largest number of genes among all chromosomes transcribing non-coding RNA molecules called (miRNAs). A number of these genes have been associated with a tendency to develop autoimmune diseases, uncloning an additional epigenetic immune modulating role of the X chromosome (44, 45).

The most intriguing property of XCI process, however, is that it is an incomplete process, lacking a telomere-to-telomere chromosomal wide approach, opening the pandoras box of genetic material imbalance between genders. Evaluation of sex gene expression in humans has revealed that 15 to 23% of X resident genes escape inactivation and are expressed seamlessly from both the active and the inactive chromosome, creating at least for some antigens an active protein abundance in females. Pervasive heterogeneity exists in the loci of the escape genes in the X chromosome [except for the telomeric regions belonging to the pseudo-autosomal regions (PAR 1 and 2), which by definition are not susceptible to inactivation], between female individuals, and even among cells from different tissues of the same individual, since complete biallelic expression in only one tissue appears to be quite rare, at around 6% (46, 47). Despite the emergence of novel biotechnologies currently available, it is predominantly the use of single cell RNA seq analysis that has enabled scientists to identify repeatedly established escaping genes, but a consensus on the complete list of those genes which evade silencing has not been reached, as several reports are not concordant (31, 48, 49). To be enlisted, a gene requires a biallelic read in a single cell or a tissue, provided that XCI is completely skewed. Genes that escape epigenomic silencing might explain phenotypic variability among females heterozygous for X-linked conditions, gene de-

letions that occasionally lead to disease because of dosage sensitivity (theory proposed by Miyake *et al.* referring to Kabuki syndrome) as well as various clinical manifestations in patients with X-chromosome aneuploidy (50). So, the towering question of why autoimmunity privileges females might be answered by the immune related genes escaping inactivation, a fact that sets those genes at the centre of scientific inquiry. Below we will briefly elaborate on the X chromosome immune genes that have been shown to escape silencing and display an established or putative role in SS pathogenesis, building in that way a probable association with the striking SS female predilection. A selection of genes that play a significant role in various cell processes and/or in the immune system operation that have been shown to escape XCI silencing but have been meagerly or not at all studied in SS are shown in Table II.

#### **Sjögren’s syndrome related XCI escapees’ profile**

##### *Toll-like receptor 7 (TLR 7)*

Toll like receptors (TLRs) are type I transmembrane proteins in charge of recognising evolutionary conserved molecular patterns from microbes or cell damage debris, initiating an immune response. Among the pattern recognition receptors (PRRs), phylogenetic analysis has shown that TLRs are the most ancient class (especially TLR3), bearing the broadest recognition repertoire (51, 52). In humans, 10 TLR subtypes have been identified, classified into 2 categories according to their location: i) located in the plasma membrane (TLR1, TLR2, TLR4, TLR5, TLR6) and ii) intracellular TLRs (TLR3, TLR7, TLR8, TLR9, TLR 10) (53). TLR 7 has the ability to sense single stranded RNA (ssRNA) and upon ligation, to produce many proinflammatory cytokines (54, 55). Among the downstream inflammatory pathways, TLR 7 induces the production of type I interferons mainly by plasmacytoid dendritic cells (pDCs) via the interferon regulatory factor 7 (IRF7) transcription factor. In SS patients, type I interferon (IFN)-inducible genes, have

**Table II.** A selection of immune related genes that escape XCI silencing and have not been studied extensively in Sjögren’s syndrome.

GENE NOMECLATURE	FUNCTION
Lysosome-associated membrane protein 2 (LAMP 2)	Crucial role in chaperone-mediated autophagy
DEAD-Box Helicase 3 X-Linked (DDX3X)	RNA helicase mediating INF signaling
Methyl-CpG Binding Protein 2 (MECP2)	Epigenetically regulates methylation sensitive genes, important role in immune homeostasis
Cluster of differentiation 99 (CD99)	Important transmembrane protein for leukocyte trafficking and T cell adhesion
Interleukin 13 Receptor Subunit Alpha 1 (IL13RA1)	Important for type II inflammatory responses
Ubiquitin Specific Peptidase 27 X-Linked (USP27X)	Deubiquitinating enzyme related to increased cell apoptosis
Ubiquitin Specific Peptidase 9 X-Linked (USP9X)	Deubiquitylating enzyme with important role in neurodevelopment
Ornithine carbamoyltransferase (OTC)	Located in liver mitochondria and functions as part of the urea cycle
Ribosomal Protein S6 Kinase A3 (RPS6KA3)	A serine/threonine kinase involved in the MAPK pathway
Lysine-specific demethylase 6A (KDM6a)	Functions as histone demethylase
The eukaryotic translation initiation factor 2 complex (EIF2S3)	A part of the protein synthesis cascade

shown a remarkable overexpression, at both the inflamed salivary glands and peripheral blood mononuclear cells (PBMCs), suggesting a pivotal role in the pathogenesis of the disease. Among the IFN-induced genes, a special attention should be given to B-cell activating factor (BAFF) which serves as a biomarker and therapeutic target in SS, and can be produced by salivary gland epithelial cells (SGECs) after ssRNA stimulation (56, 57). In accordance, TLR 7 has been found upregulated in blood samples, PMBCs and cultured salivary gland epithelial cells (SGECs) of patients with SS compared to gender matched controls (58-60). In addition, a recent study by Davies et al showed that SS patients’ PBMC TLR7 stimulation significantly alters the downstream molecular phosphorylation profile compared to healthy controls (61). Another mechanism through which TLR 7 activation might contribute to Sjögren’s pathogenesis is via age-associated B cells (ABCs). ABCs is a newly discovered B cell subset that expresses the transcription factor T-bet and accumulates with normal aging. Interestingly ABCs also display a premature expansion in various autoimmune diseases, including SS, having the capacity to produce harmful autoantibodies (62-64). It has been also shown that TLR 7 is the driving force behind the proliferation and inflamed tissue accumulation of autoanti-

body secreting ABCs involved in lupus manifestations, seen in an autoimmune prone mice model (65).

Evidently, TLR 7 is involved in the pathogenesis of SS, but whether a biallelic expression plays a role in the female sex bias observed in autoimmune diseases remains unknown. Souyris *et al.* showed that B lymphocytes, monocytes, and interferon producing plasmacytoid dendritic cells, not only in females but also in males with Klinefelter syndrome, express elevated levels of TLR7 and exhibit a propensity for class switch toward IgG producing plasma cells in contrast to the behaviour of immune cells from euploid males (XY) (66). Moreover, a recent study focusing on pDCs confirmed that TLR7 escapes inactivation in this cell line as well, resulting in higher mRNA transcription of IFN $\alpha$  and IFN $\beta$  that is further augmented substantially by IFN $\alpha$  exposure (67), creating a vicious cycle. Interestingly, a gain of function polymorphism mutation in TLR 7 gene predisposes male individuals to develop SLE (65).

*CD40 ligand*

CD 40 ligand (CD40L), also known as CD154, is a transmembrane protein type II located primarily on T helper cells. Its primary target is CD40 on the surface of B cells. When a CD40-CD40L interaction occurs, it acts as a second co-stimulatory signal, vital for

B cell activation, isotype class switching and germinal centre formation (68, 69). Further studies have shown that CD40 and its ligand is also expressed in non-lymphoid cell types including monocytes, basophils, eosinophils, dendritic cells, megakaryocytes, platelets, fibroblasts, smooth muscle, and endothelial cells, mediating both humoral and cellular immunity (70, 71). SS is a disease characterised by the presence of germinal centre-like regions in the lip biopsy of more than 20% of patients. Even though being ectopic (outside of the secondary lymphoid organs), these structures are fully functional and equipped with all the required molecular tools, such as CD40L and activation-induced cytidine deaminase (AID), to activate B cells and produce large quantities of (auto) antibodies (72-74). The presence of ectopic germinal centre in the lip biopsy has been associated with an increased risk for future lymphoma development, a higher focus score, more systemic manifestations, and enriched autoantibody profile (75, 76). It is important that this knowledge should be guiding clinicians with their patients’ follow up plans (77). As expected, reports dating from the early 2000, showed increased expression of CD40L in lip biopsies of 17 SS patients and their cultured SGECs compared to healthy donors (78, 79), indicating a putative role of T and B cell interactions in the patho-



genesis of the disease (80). In addition, T cells carrying CD40L might mediate epithelial gland dysfunction through interaction with the CD40 molecule expressed on epithelial cells membrane (81). The first therapeutic initiative to block the CD40 pathway in lupus patients ended in early discontinuation of the trial, because of an increase in thromboembolic events caused by the cross-linking of anti-CD40L antibodies to platelet CD40L and Fc gamma Receptor IIa on adjacent platelets (82). However, newer therapeutic anti-CD40 agents conferring different amino acid sequence in the Fc region preventing platelet aggregation have completed phase II studies, with promising preliminary results (83, 84).

Given the aforementioned crucial role of CD40 in the pathogenesis of Sjogren’s syndrome, the fact that a number of women’s T cells have biallelic CD40L expression may provide a mechanistic explanation for the sexual divergence (85). Supporting evidence comes from murine models in which CD40 overexpression resulted in high titers of autoantibodies (86) and from human subjects with CD40L gene duplication who develop autoimmune manifestations and lymphopenia (87). Finally, a recent study revealed that stimulated PBMCs from normal women and males with Klinefelter syndrome harboured higher expression of CD40L on T cells, compared to euploid males and females with Turner syndrome (88).

#### *C-X-C motif chemokine receptor 3*

The only chemokine receptor gene encoded by the X chromosome is C-X-C motif chemokine receptor 3 (CXCR3), a crucial Th1 response regulator (89). Its expression is ubiquitous among all immune cell lineages, but its role has been especially studied in activated T cells, serving as a receptor for interferon (IFN)- $\gamma$  inducible chemokines CXCL9, CXCL10, and CXCL11. All these chemokines are not constitutively expressed unless induced by an inflammatory stimulus and mainly regulate immune cell migration, differentiation, and activation (90). Given its immune cell expression, selective ligation,

and prominent function, CXCR3 has been implicated in autoimmune disease pathogenesis (91). Especially in the salivary and lacrimal glands of SS patients, CXCR3 and its ligands have been shown to be upregulated (92, 93). In addition, CXCR3 mRNA levels in CD4<sup>+</sup> T cells of female donors have shown increased levels compared to males, despite the fact that this difference abated at the protein level, suggesting additional post-transcriptional regulation (94). Initial studies on XCI escape genes failed to recognize CXCR3 as an escapee. A recent study, however, exploiting more sensitivity single-cell RNA FISH analysis exhibited biallelic expression patterns for CXCR3 a finding that was confirmed by *in vivo* fluorescence in a specially designed dual reporter mouse model (85, 95). Conclusively, CXCR3 seems to play a role in the pathogenetic mechanisms of SS among other autoimmune diseases and its biallelic expression and ample protein production if confirmed in future studies, could become a potential contributor to the observed preponderance of the female sex.

#### *Interleukin-1 receptor associated kinase*

Interleukin-1 receptor associated kinase (IRAK-1) contributes to a promiscuous innate immunity proinflammatory signal transduction, relaying signals from Toll/IL-1 receptor (TIR) family (96). Upon ligation, these membrane receptors dimerize and recruit MYD88 (except for TLR 3), an adaptor protein that interacts with IRAK-4 and then phosphorylates IRAK-1 and 2 (97). Upon its activation, IRAK-1 phosphorylates TRAF-6 in a proline-serine-threonine-rich domain and activates several downstream pathways, including NF- $\kappa$ B, leading to various gene expression of cytokines (IL-1, IL-6, IL-12, TNF- $\alpha$ ), chemokines and proliferative factors (G-CSF, M-CSF). Understanding the role of IRAK-1 in SS awaits further studies, as there is only a single report investigating its expression in SS patients’ PBMCs, demonstrating decreased expression compared to healthy donors (98). In lupus, however, human and murine studies

have established IRAK-1 as a prominent protein involved in its pathogenesis. Indeed, IRAK-1 deficient SLE mice models eschew all lupus-associated manifestations with concurrent reversal of any abnormal dendritic cell function, while in humans, polymorphisms of IRAK-1 gene have been correlated with increased lupus incidence and increased IRAK-1 levels in CD4<sup>+</sup> T cells from SLE patients (99-101). Clearly, further studies investigating its role in the pathophysiology of SS via biallelic expression of specific immune cell population or salivary gland epithelial cells, are the next core steps to uncloak a potential link that might prove fruitful in the elucidation of the remarkable sex disparity among SS patients.

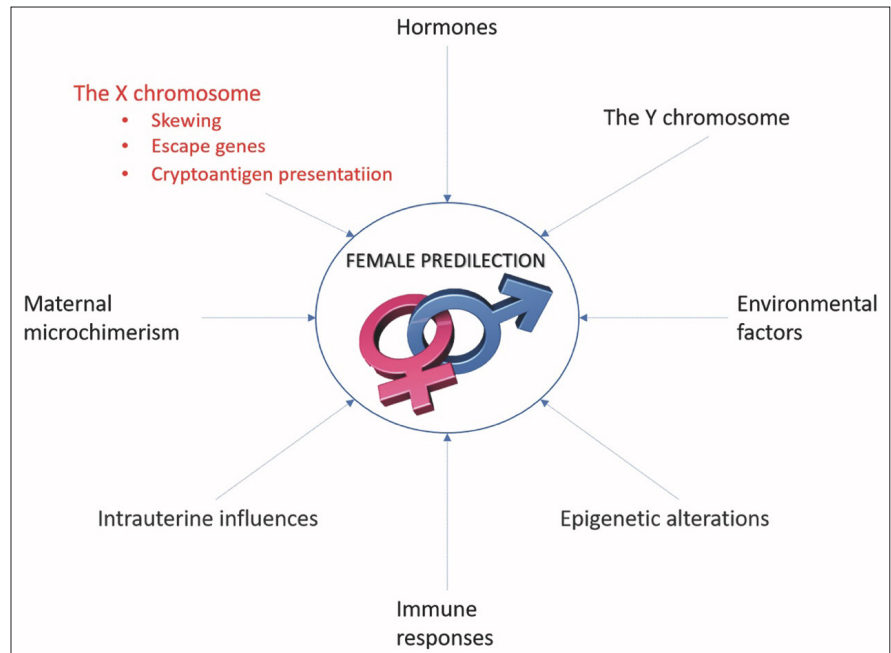
#### *Chromosome X open reading frame 21 (CXorf21)*

Until fairly recently, the function of the protein encoded by CXorf21 was a mystery, even though GWAS studies had already identified CXorf21 among the SLE and SS susceptibility genes and its levels had been shown to parallel the disease activity (102-104). In 2020, Heinz *et al.* renamed the protein as “TLR adaptor interacting with SLC15A4 on the lysosome” (TASL) and highlighted its key function as a gatekeeper in the recruitment and activation of the transcription factor IRF5 after endosomal TLR 7, 8 and 9 ligation (105, 106), resembling innate immune adaptor proteins STING, MAVS and TRIF. CXorf21 gene may variably escape X chromosome inactivation and its levels are increased in monocytes, B cells and lymphoblastoid cell lines of females compared to males (107). On this, functional studies on SS and SLE patients revealed sexual dimorphism in their lysosomal cell pH (lower pH in female compared to males’ cells), possibly disrupting normal endolysosomal antigen processing owing to higher levels of CXorf21. Thus, CXorf21 gene combined with the aforementioned TLR 7 is a duo of closely related inactivation evading genes, since their product proteins are constantly collaborating in innate immune signaling (108). Both genes display a female

gender predominant protein expression and have a proven crucial role in the pathogenesis of autoimmune diseases and as such, they hold the promise that their further study might shed light to our understanding of the factors and processes underpinning the observed sex bias among autoimmune patients including SS. (107).

### Conclusion

Autoimmune diseases favour women in a striking and enigmatic manner. To date, many contributing factors have been incriminated in this phenomenon, creating a largely unexplored pathogenetic tapestry that mediates women’s predilection for autoimmunity. Components of the tapestry that define gender bias in autoimmune diseases encompass the hormonal milieu and sex chromosomes on the one hand and innate and acquired immunity on the other, through a dynamic and complex interplay (Fig. 1). For many years, hormones took a central role when scientists attempted to clarify nature’s gender discrimination, with either inconclusive or even conflicting results. To fill this knowledge gap, the scientific community turned to nature’s own imperfection regarding the homeostatic process of balancing the genetic material between the sexes. Indeed, a failure to completely silence the second, rich in immune related genes, X chromosome may engender an environment that renders women particularly vulnerable to autoimmune disorders. The multilevel regulation of gene expression also involves sex chromosomes (epigenetic, transcriptional, post-transcriptional regulation), highlighting the difficulty to identify those key genes and their effector protein products, driving the female predilection in autoimmunity. Apart from the well-established loci with immune functions, there are also loci with still ill-defined functions which may indirectly interfere with both innate and acquired immunity. In addition, the underlying immune deregulation may involve the effector, memory, or regulatory component of acquired immunity. Thus, an overly complex network of interactions among the key contributors may eventually define the female predilection in autoimmunity.



**Fig. 1.** Female predilection in autoimmunity may be the end result of complex interactions between several confounding factors, with the X chromosome’s unique features constituting a key contributor.

Availability of a suitable disease model in humans that might allow promising lines of research to be further explored in depth, would be a considerable advance and perhaps act as the holy grail in our quest to illuminate sex bias in autoimmunity. Such a model would encompass a) a robust female predominance, b) a high prevalence, c) a wide age distribution with disease onset before puberty as well as after menopause and d) a practical histologic access to the autoimmune disorder’s main target organ. Sjögren’s syndrome, exhibiting a female predominance more pronounced than in any other systemic autoimmune disease, a wide age gamut and an easily obtainable target tissue specimen by the safe and highly informative minor salivary gland biopsy, is indubitably the disease candidate of choice (109). So far, studies in SS are limited in number and mainly involve sporadic reports of biallelic expression of certain XCI escape genes in the peripheral blood, as described above. No studies have been performed in cultured salivary gland epithelial cells or immune cells infiltrating the salivary glands. Such studies could shed light on the different protein apparatus and its subsequent downstream effects be-

tween genders and prove in a mechanistic order whether X chromosome is the true “X factor” underlying sexual divergence in autoimmunity.

### References

- MORONI L, BIANCHI I, LLEO A: Geoepidemiology, gender and autoimmune disease. *Autoimmun Rev* 2012; 11: A386-92.
- WALSH SJ, RAU LM: Autoimmune diseases: a leading cause of death among young and middle-aged women in the United States. *Am J Public Health* 2000; 90: 1463-6.
- AL MAINI M, ADELOWO F, AL SALEH J *et al.*: The global challenges and opportunities in the practice of rheumatology: white paper by the World Forum on Rheumatic and Musculoskeletal Diseases. *Clin Rheumatol* 2015; 34: 819-29.
- JACOBSON DL, GANGE SJ, ROSE NR, GRAHAM NM: Epidemiology and estimated population burden of selected autoimmune diseases in the United States. *Clin Immunol Immunopathol* 1997; 84: 223-43.
- CHATZIS L, PEZOULAS VC, FERRO F *et al.*: Sjögren’s Syndrome: The Clinical Spectrum of Male Patients. *J Clin Med* 2020; 9: 2620.
- AMADORI A, ZAMARCHI R, DE SILVESTRO G *et al.*: Genetic control of the CD4/CD8 T-cell ratio in humans. *Nat Med* 1995; 1: 1279-83.
- NALBANDIAN G, KOVATS S: Understanding sex biases in immunity: effects of estrogen on the differentiation and function of antigen-presenting cells. *Immunol Res* 2005; 31: 91-106.
- MANUEL RSJ, LIANG Y: Sexual dimorphism in immunometabolism and autoimmunity: Impact on personalized medicine. *Autoimmun Rev* 2021; 20: 102775.

9. STERLING TR, VLAHOV D, ASTEMBORSKI J, HOOVER DR, MARGOLICK JB, QUINN TC: Initial plasma HIV-1 RNA levels and progression to AIDS in women and men. *N Engl J Med* 2001; 344: 720-5.
10. HERTZ D, SCHNEIDER B: Sex differences in tuberculosis. *Semin Immunopathol* 2019; 41: 225-37.
11. FLANAGAN KL, FINK AL, PLEBANSKI M, KLEIN SL: Sex and Gender Differences in the Outcomes of Vaccination over the Life Course. *Annu Rev Cell Dev Biol* 2017; 33: 577-99.
12. SCULLY EP, HAVERFIELD J, URSIN RL, TANNENBAUM C, KLEIN SL: Considering how biological sex impacts immune responses and COVID-19 outcomes. *Nat Rev Immunol* 2020; 20: 442-7.
13. HOLROYD CR, EDWARDS CJ: The effects of hormone replacement therapy on autoimmune disease: rheumatoid arthritis and systemic lupus erythematosus. *Climacteric* 2009; 12: 378-86.
14. JØRGENSEN KT, ROSTGAARD K, BACHE I *et al.*: Autoimmune diseases in women with Turner’s syndrome. *Arthritis Rheum* 2010; 62: 658-66.
15. HARRIS VM, SHARMA R, CAVETT J *et al.*: Klinefelter’s syndrome (47,XXY) is in excess among men with Sjögren’s syndrome. *Clin Immunol* 2016; 168: 25-9.
16. SEMINOG OO, SEMINOG AB, YEATES D, GOLDACRE MJ: Associations between Klinefelter’s syndrome and autoimmune diseases: English national record linkage studies. *Autoimmunity* 2015; 48: 125-8.
17. BACHTROG D, MANK JE, PEICHEL CL *et al.*: Sex determination: why so many ways of doing it? *PLoS Biol* 2014; 12: e1001899.
18. BACHTROG D: Y-chromosome evolution: emerging insights into processes of Y-chromosome degeneration. *Nature Rev Genet* 2013; 14: 113-24.
19. ABBOTT JK, NORDÉN AK, HANSSON B: Sex chromosome evolution: historical insights and future perspectives. *Proc Biol Sci* 2017; 284: 20162806.
20. ROSS MT, GRAFHAM DV, COFFEY AJ *et al.*: The DNA sequence of the human X chromosome. *Nature* 2005; 434: 325-37.
21. HELENA MANGS A, MORRIS BJ: The Human Pseudoautosomal Region (PAR): Origin, Function and Future. *Curr Genomics* 2007; 8: 129-36.
22. SKALETSKY H, KURODA-KAWAGUCHI T, MINX PJ *et al.*: The male-specific region of the human Y chromosome is a mosaic of discrete sequence classes. *Nature* 2003; 423: 825-37.
23. LAMBERT NC: Nonendocrine mechanisms of sex bias in rheumatic diseases. *Nature Rev Rheumatol* 2019; 15: 673-86.
24. YAZDANI R, FEKRVAND S, SHAHKARAMI S *et al.*: The hyper IgM syndromes: Epidemiology, pathogenesis, clinical manifestations, diagnosis and management. *Clin Immunol* 2019; 198: 19-30.
25. LU L, BARBI J, PAN F: The regulation of immune tolerance by FOXP3. *Nat Rev Immunol* 2017; 17: 703-17.
26. VAN DE VEN AA, WARNATZ K: The autoimmune conundrum in common variable immunodeficiency disorders. *Curr Opin Allergy Clin Immunol* 2015; 15: 514-24.
27. POWELL BR, BUIST NR, STENZEL P: An X-linked syndrome of diarrhea, polyendocrinopathy, and fatal infection in infancy. *J Pediatr* 1982; 100: 731-7.
28. DELMONTE OM, BERGERSON JRE, KAWAIT *et al.*: SASH3 variants cause a novel form of X-linked combined immunodeficiency with immune dysregulation. *Blood* 2021; Apr 19.
29. CASE LK, WALL EH, DRAGON JA *et al.*: The Y chromosome as a regulatory element shaping immune cell transcriptomes and susceptibility to autoimmune disease. *Genome Res* 2013; 23: 1474-85.
30. LYON MF: Gene action in the X-chromosome of the mouse (*Mus musculus* L.). *Nature* 1961; 190: 372-3.
31. FANG H, DISTECHE CM, BERLETCHE JB: X Inactivation and Escape: Epigenetic and Structural Features. *Front Cell Dev Biol* 2019; 7: 219.
32. GALUPA R, HEARD E: X-Chromosome Inactivation: A Crossroads Between Chromosome Architecture and Gene Regulation. *Annu Rev Genet* 2018; 52: 535-66.
33. BORENSZTEIN M, SYX L, ANCELIN K *et al.*: Xist-dependent imprinted X inactivation and the early developmental consequences of its failure. *Nat Struct Mol Biol* 2017; 24: 226-33.
34. MARAHRENS Y, PANNING B, DAUSMAN J, STRAUSS W, JAENISCH R: Xist-deficient mice are defective in dosage compensation but not spermatogenesis. *Genes Dev* 1997; 11: 156-66.
35. MENGEL-FROM J, LINDAHL-JACOBSEN R, NYGAARD M *et al.*: Skewness of X-chromosome inactivation increases with age and varies across birth cohorts in elderly Danish women. *Sci Rep* 2021; 11: 4326.
36. GREGERSEN PK, CHITNIS S, MONTEIRO J, SALMON J: Increased X-inactivation skewing in SLE? *Immunol Today* 1999; 20: 152; author reply 153.
37. UZ E, MUSTAFA C, TOPALOGLU R *et al.*: Increased frequency of extremely skewed X chromosome inactivation in juvenile idiopathic arthritis. *Arthritis Rheum* 2009; 60: 3410-2.
38. OZCELIK T, UZ E, AKYERLI CB *et al.*: Evidence from autoimmune thyroiditis of skewed X-chromosome inactivation in female predisposition to autoimmunity. *Eur J Hum Genet* 2006; 14: 791-7.
39. OZBALKAN Z, BAGIŞLAR S, KIRAZ S *et al.*: Skewed X chromosome inactivation in blood cells of women with scleroderma. *Arthritis Rheum* 2005; 52: 1564-70.
40. STEWART JJ: The female X-inactivation mosaic in systemic lupus erythematosus. *Immunol Today* 1998; 19: 352-7.
41. PODOLSKA A, KOBELT A, FUCHS S *et al.*: Functional monosomy of 6q27-qter and functional disomy of Xpter-p22.11 due to X; 6 translocation with an atypical X-inactivation pattern. *Am J Med Genet A* 2017; 173: 1334-41.
42. FAVILLA BP, MELONI VA, PEREZ AB *et al.*: Spread of X-chromosome inactivation into autosomal regions in patients with unbalanced X-autosome translocations and its phenotypic effects. *Am J Med Genet A* 2021; 185: 2295-305.
43. ORSTAVIK KH: X chromosome inactivation in clinical practice. *Hum Genet* 2009; 126: 363-73.
44. KHALIFA O, PERS YM, FERREIRA R *et al.*: X-Linked miRNAs Associated with Gender Differences in Rheumatoid Arthritis. *Int J Mol Sci* 2016; 17: 1852.
45. PINHEIRO I, DEJAGER L, LIBERT C: X-chromosome-located microRNAs in immunity: might they explain male/female differences? The X chromosome-genomic context may affect X-located miRNAs and downstream signaling, thereby contributing to the enhanced immune response of females. *BioEssays* 2011; 33: 791-802.
46. TUKIAINEN T, VILLANI AC, YEN A *et al.*: Landscape of X chromosome inactivation across human tissues. *Nature* 2017; 550: 244-8.
47. GARIERI M, STAMOULIS G, BLANC X *et al.*: Extensive cellular heterogeneity of X inactivation revealed by single-cell allele-specific expression in human fibroblasts. *Proc Natl Acad Sci USA* 2018; 115: 13015-20.
48. WAINER KATSIR K, LINIAL M: Human genes escaping X-inactivation revealed by single cell expression data. *BMC Genomics* 2019; 20: 201.
49. ZHANG Y, CASTILLO-MORALES A, JIANG M *et al.*: Genes that escape X-inactivation in humans have high intraspecific variability in expression, are associated with mental impairment but are not slow evolving. *Mol Biol Evol* 2013; 30: 2588-601.
50. MIYAKE N, MIZUNO S, OKAMOTO N *et al.*: KDM6A point mutations cause Kabuki syndrome. *Hum Mutat* 2013; 34: 108-10.
51. LIU G, ZHANG H, ZHAO C, ZHANG H: Evolutionary History of the Toll-Like Receptor Gene Family across Vertebrates. *Genome Biol Evol* 2020; 12: 3615-34.
52. JIMÉNEZ-DALMARONI MJ, GERSWHIN ME, ADAMOPOULOS IE: The critical role of toll-like receptors--From microbial recognition to autoimmunity: A comprehensive review. *Autoimmun Rev* 2016; 15: 1-8.
53. TAKEDA K, KAISHO T, AKIRA S: Toll-like receptors. *Ann Rev Immunol* 2003; 21: 335-76.
54. DIEBOLD SS, KAISHO T, HEMMI H, AKIRA S, REIS C, SOUSA E: Innate antiviral responses by means of TLR7-mediated recognition of single-stranded RNA. *Science* 2004; 303: 1529-31.
55. PETES C, ODOARDI N, GEE K: The Toll for Trafficking: Toll-Like Receptor 7 Delivery to the Endosome. *Front Immunol* 2017; 8: 1075.
56. VAKALOGLOU KM, MAVRAGANI CP: Activation of the type I interferon pathway in primary Sjögren’s syndrome: an update. *Curr Opin Rheumatol* 2011; 23: 459-64.
57. YAO Y, LIU Z, JALLAL B, SHEN N, RÖNNBLÖM L: Type I interferons in Sjögren’s syndrome. *Autoimmun Rev* 2013; 12: 558-66.
58. MARIA NI, STEENWIJK EC, IPMA AS *et al.*: Contrasting expression pattern of RNA-sensing receptors TLR7, RIG-I and MDA5 in interferon-positive and interferon-negative patients with primary Sjögren’s syn-



- drome. *Ann Rheum Dis* 2017; 76: 721-30.
59. ZHENG L, ZHANG Z, YU C, YANG C: Expression of Toll-like receptors 7, 8, and 9 in primary Sjögren’s syndrome. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2010; 109: 844-50.
  60. SHIMIZU T, NAKAMURA H, TAKATANI A *et al.*: Activation of Toll-like receptor 7 signaling in labial salivary glands of primary Sjögren’s syndrome patients. *Clin Exp Immunol* 2019; 196: 39-51.
  61. DAVIES R, SARKAR I, HAMMENFORS D *et al.*: Single Cell Based Phosphorylation Profiling Identifies Alterations in Toll-Like Receptor 7 and 9 Signaling in Patients With Primary Sjögren’s Syndrome. *Front Immunol* 2019; 10: 281.
  62. CANCRO MP: Age-Associated B Cells. *Annu Rev Immunol* 2020; 38: 315-40.
  63. RUBTSOVA K, RUBTSOV AV, CANCRO MP, MARRACK P: Age-Associated B Cells: A T-bet-Dependent Effector with Roles in Protective and Pathogenic Immunity. *J Immunol* 2015; 195: 1933-7.
  64. HAO Y, O’NEILL P, NARADIKIAN MS, SCHOLZ JL, CANCRO MP: A B-cell subset uniquely responsive to innate stimuli accumulates in aged mice. *Blood* 2011; 118: 1294-304.
  65. RUBTSOV AV, RUBTSOVA K, KAPPLER JW, MARRACK P: TLR7 drives accumulation of ABCs and autoantibody production in autoimmune-prone mice. *Immunol Res* 2013; 55: 210-6.
  66. SOUYRIS M, CENAC C, AZAR P *et al.*: TLR7 escapes X chromosome inactivation in immune cells. *Sci Immunol* 2018; 3: eaap8855.
  67. HAGEN SH, HENSELING F, HENNESEN J *et al.*: Heterogeneous Escape from X Chromosome Inactivation Results in Sex Differences in Type I IFN Responses at the Single Human pDC Level. *Cell Rep* 2020; 33: 108485.
  68. KAWABE T, NAKA T, YOSHIDA K *et al.*: The immune responses in CD40-deficient mice: impaired immunoglobulin class switching and germinal center formation. *Immunity* 1994; 1: 167-78.
  69. ARMITAGE RJ, FANSLAW WC, STROCKBINE L *et al.*: Molecular and biological characterization of a murine ligand for CD40. *Nature* 1992; 357: 80-2.
  70. KARNELL JL, ALBULESCU M, DRABIC S *et al.*: A CD40L-targeting protein reduces autoantibodies and improves disease activity in patients with autoimmunity. *Sci Transl Med* 2019; 11: eaar6584.
  71. INWALD DP, MCDOWALL A, PETERS MJ, CALLARD RE, KLEIN NJ: CD40 is constitutively expressed on platelets and provides a novel mechanism for platelet activation. *Circ Res* 2003; 92: 1041-8.
  72. BOMBARDIERI M, LEWIS M, PITZALIS C: Ectopic lymphoid neogenesis in rheumatic autoimmune diseases. *Nat Rev Rheumatol* 2017; 13: 141-54.
  73. BOMBARDIERI M, BARONE F, HUMBY F *et al.*: Activation-induced cytidine deaminase expression in follicular dendritic cell networks and interfollicular large B cells supports functionality of ectopic lymphoid neogenesis in autoimmune sialoadenitis and MALT lymphoma in Sjögren’s syndrome. *J Immunol* 2007; 179: 4929-38.
  74. BOMBARDIERI M, ARGYROPOULOU OD, FERRO F *et al.*: One year in review 2020: pathogenesis of primary Sjögren’s syndrome. *Clin Exp Rheumatol* 2020; 38 (Suppl. 126): S3-9.
  75. THEANDER E, VASAITIS L, BAECKLUND E *et al.*: Lymphoid organisation in labial salivary gland biopsies is a possible predictor for the development of malignant lymphoma in primary Sjögren’s syndrome. *Ann Rheum Dis* 2011; 70: 1363-8.
  76. SÈNE D, ISMAEL S, FORIEN M *et al.*: Ectopic Germinal Center-Like Structures in Minor Salivary Gland Biopsy Tissue Predict Lymphoma Occurrence in Patients With Primary Sjögren’s Syndrome. *Arthritis Rheumatol* 2018; 70: 1481-8.
  77. CHATZIS L, VLACHOYIANNPOULOS PG, TZIOUFAS AG, GOULES AV: New frontiers in precision medicine for Sjögren’s syndrome. *Expert Rev Clin Immunol* 2021; 17: 127-41.
  78. DIMITRIOU ID, KAPSOGEOGOU EK, MOUTSOPOULOS HM, MANOUSSAKIS MN: CD40 on salivary gland epithelial cells: high constitutive expression by cultured cells from Sjögren’s syndrome patients indicating their intrinsic activation. *Clin Exp Immunol* 2002; 127: 386-92.
  79. GOULES A, TZIOUFAS AG, MANOUSAKIS MN, KIROU KA, CROW MK, ROUTSIAS JG: Elevated levels of soluble CD40 ligand (sCD40L) in serum of patients with systemic autoimmune diseases. *J Autoimmun* 2006; 26: 165-71.
  80. OHLSSON M, SZODORAY P, LORO LL, JOHANNESEN AC, JONSSON R: CD40, CD154, Bax and Bcl-2 expression in Sjögren’s syndrome salivary glands: a putative anti-apoptotic role during its effector phases. *Scand J Immunol* 2002; 56: 561-71.
  81. PING L, OGAWA N, SUGAI S: Novel role of CD40 in Fas-dependent apoptosis of cultured salivary epithelial cells from patients with Sjögren’s syndrome. *Arthritis Rheum* 2005; 52: 573-81.
  82. BOUMPAS DT, FURIE R, MANZI S *et al.*: A short course of BG9588 (anti-CD40 ligand antibody) improves serologic activity and decreases hematuria in patients with proliferative lupus glomerulonephritis. *Arthritis Rheum* 2003; 48: 719-27.
  83. WIECZOREK G, BIGAUD M, PFISTER S *et al.*: Blockade of CD40-CD154 pathway interactions suppresses ectopic lymphoid structures and inhibits pathology in the NOD/ShiLJ mouse model of Sjögren’s syndrome. *Ann Rheum Dis* 2019; 78: 974-8.
  84. FISHER B, SZANTO A, NG W-F *et al.*: Assessment of the anti-CD40 antibody iscalimab in patients with primary Sjögren’s syndrome: a multicentre, randomised, double-blind, placebo-controlled, proof-of-concept study. *Lancet Rheumatol* 2020; 2.
  85. WANG J, SYRETT CM, KRAMER MC, BASU A, ATCHISON ML, ANGUERA MC: Unusual maintenance of X chromosome inactivation predisposes female lymphocytes for increased expression from the inactive X. *Proc Natl Acad Sci USA* 2016; 113: E2029-38.
  86. CLEGG CH, RULFFES JT, HAUGEN HS *et al.*: Thymus dysfunction and chronic inflammatory disease in gp39 transgenic mice. *Int Immunol* 1997; 9: 1111-22.
  87. LE COZ C, TROFA M, SYRETT CM *et al.*: CD40LG duplication-associated autoimmune disease is silenced by nonrandom X-chromosome inactivation. *J Allergy Clin Immunol* 2018; 141: 2308-11.e7.
  88. SARMIENTO L, SVENSSON J, BARCHETTA I, GIWERCMAN A, CILIO CM: Copy number of the X-linked genes TLR7 and CD40L influences innate and adaptive immune responses. *Scand J Immunol* 2019; 90: e12776.
  89. LOETSCHER M, LOETSCHER P, BRASS N, MEESE E, MOSER B: Lymphocyte-specific chemokine receptor CXCR3: regulation, chemokine binding and gene localization. *Eur J Immunol* 1998; 28: 3696-705.
  90. METZEMAEKERS M, VANHEULE V, JANSSENS R, STRUYF S, PROOST P: Overview of the Mechanisms that May Contribute to the Non-Redundant Activities of Interferon-Inducible CXCR3 Chemokine Receptor 3 Ligands. *Front Immunol* 2017; 8: 1970.
  91. LACOTTE S, BRUN S, MULLER S, DUMORTIER H: CXCR3, inflammation, and autoimmune diseases. *Ann NY Acad Sci* 2009; 1173: 310-7.
  92. OGAWA N, PING L, ZHENJUN L, TAKADA Y, SUGAI S: Involvement of the interferon-gamma-induced T cell-attracting chemokines, interferon-gamma-inducible 10-kd protein (CXCL10) and monokine induced by interferon-gamma (CXCL9), in the salivary gland lesions of patients with Sjögren’s syndrome. *Arthritis Rheum* 2002; 46: 2730-41.
  93. YOON KC, PARK CS, YOU IC *et al.*: Expression of CXCL9, -10, -11, and CXCR3 in the tear film and ocular surface of patients with dry eye syndrome. *Invest Ophthalmol Vis Sci* 2010; 51: 643-50.
  94. HEWAGAMA A, GORELIK G, PATEL D *et al.*: Overexpression of X-linked genes in T cells from women with lupus. *J Autoimmun* 2013; 41: 60-71.
  95. OGHUMU S, VARIKUTI S, STOCK JC *et al.*: Cutting Edge: CXCR3 Escapes X Chromosome Inactivation in T Cells during Infection: Potential Implications for Sex Differences in Immune Responses. *J Immunol* 2019; 203: 789-94.
  96. MARTIN MU, WESCHE H: Summary and comparison of the signaling mechanisms of the Toll/interleukin-1 receptor family. *Biochim Biophys Acta* 2002; 1592: 265-80.
  97. GOTTIPATI S, RAO NL, FUNG-LEUNG WP: IRAK1: a critical signaling mediator of innate immunity. *Cell Signal* 2008; 20: 269-76.
  98. ZILAH E, TARR T, PAPP G, GRIGER Z, SIPKA S, ZEHER M: Increased microRNA-146a/b, TRAF6 gene and decreased IRAK1 gene expressions in the peripheral mononuclear cells of patients with Sjögren’s syndrome. *Immunol Lett* 2012; 141: 165-8.
  99. JACOB CO, ZHU J, ARMSTRONG DL *et al.*: Identification of IRAK1 as a risk gene with critical role in the pathogenesis of systemic lupus erythematosus. *Proc Natl Acad Sci*



- USA 2009; 106: 6256-61.
100. KAUFMAN KM, ZHAO J, KELLY JA *et al.*: Fine mapping of Xq28: both MECP2 and IRAK1 contribute to risk for systemic lupus erythematosus in multiple ancestral groups. *Ann Rheum Dis* 2013; 72: 437-44.
  101. ZHAI Y, XU K, LENG RX *et al.*: Association of interleukin-1 receptor-associated kinase (IRAK1) gene polymorphisms (rs3027898, rs1059702) with systemic lupus erythematosus in a Chinese Han population. *Inflamm Res* 2013; 62: 555-60.
  102. BENTHAM J, MORRIS DL, GRAHAM DSC *et al.*: Genetic association analyses implicate aberrant regulation of innate and adaptive immunity genes in the pathogenesis of systemic lupus erythematosus. *Nat Genet* 2015; 47: 1457-64.
  103. MICELI-RICHARD C, WANG-RENAULT SF, BOUDAOU S *et al.*: Overlap between differentially methylated DNA regions in blood B lymphocytes and genetic at-risk loci in primary Sjögren’s syndrome. *Ann Rheum Dis* 2016; 75: 933-40.
  104. MACKAY M, OSWALD M, SANCHEZ-GUERRERO J *et al.*: Molecular signatures in systemic lupus erythematosus: distinction between disease flare and infection. *Lupus Sci Med* 2016; 3: e000159.
  105. HEINZ LX, LEE J, KAPOOR U *et al.*: TASL is the SLC15A4-associated adaptor for IRF5 activation by TLR7-9. *Nature* 2020; 581: 316-22.
  106. PISETSKY DS: The role of TASL in the pathogenesis of SLE: X marks the spot. *Ann Rheum Dis* 2021; 80: 6-7.
  107. HARRIS VM, HARLEY ITW, KURIEN BT, KOELSCH KA, SCOFIELD RH: Lysosomal pH Is Regulated in a Sex Dependent Manner in Immune Cells Expressing CXorf21. *Front Immunol* 2019; 10: 578.
  108. HARRIS VM, SCOFIELD RH, SIVILS KL: Genetics in Sjögren’s syndrome: where we are and where we go. *Clin Exp Rheumatol* 2019; 37 (Suppl. 118): S234-9.
  109. CHATZIS L, GOULES AV, PEZOULAS V *et al.*: A biomarker for lymphoma development in Sjögren’s syndrome: Salivary gland focus score. *J Autoimmun* 2021; 121: 102648.
  110. IZMIRLY PM, PARTON H, WANG L *et al.*: Prevalence of Systemic Lupus Erythematosus in the United States: Estimates From a Meta-Analysis of the Centers for Disease Control and Prevention National Lupus Registries. *Arthritis Rheumatol* (Hoboken), 2021; 73 :991-6.
  111. PEOPLES C, MEDSGER TA JR, LUCAS M, ROSARIO BL, FEGHALI-BOSTWICK CA: Gender differences in systemic sclerosis: relationship to clinical features, serologic status and outcomes. *J Scleroderma Relat Disord* 2016; 1: 177-240.
  112. MYASOEDOVA E, DAVIS J, MATTESON EL, CROWSON CS: Is the epidemiology of rheumatoid arthritis changing? Results from a population-based incidence study, 1985-2014. *Ann Rheum Dis* 2020; 79: 440-4.
  113. NORDBORG E, NORDBORG C: Giant cell arteritis: epidemiological clues to its pathogenesis and an update on its treatment. *Rheumatology* (Oxford) 2003; 42: 413-21.
  114. BERTIA, CORNEC D, CROWSON CS, SPECKS U, MATTESON EL: The Epidemiology of Antineutrophil Cytoplasmic Autoantibody-Associated Vasculitis in Olmsted County, Minnesota: A Twenty-Year US Population-Based Study. *Arthritis Rheumatol* (Hoboken) 2017; 69: 2338-50.
  115. ONEN F, AKKOC N: Epidemiology of Takayasu arteritis. *Presse Med* (France), 2017; 46: e197-e203.
  116. MEYER A, MEYER N, SCHAEFFER M, GOTTENBERG JE, GENY B, SIBILIA J: Incidence and prevalence of inflammatory myopathies: a systematic review. *Rheumatology* (Oxford) 2015; 54: 50-63.
  117. STOLWIJK C, BOONEN A, VAN TUBERGEN A, REVEILLE JD: Epidemiology of spondyloarthritis. *Rheum Dis Clin North Am* 2012; 38: 441-76.
  118. HUSSAIN YS, HOOKHAM JC, ALLAHABADIA A, BALASUBRAMANIAN SP: Epidemiology, management and outcomes of Graves’ disease-real life data. *Endocrine* 2017; 56: 568-78.
  119. HARBO HF, GOLD R, TINTORÉ M: Sex and gender issues in multiple sclerosis. *Ther Adv Neurol Disord* 2013; 6: 237-48.
  120. ALKHATEEB A, FAIN PR, THODY A, BENNETT DC, SPRITZ RA: Epidemiology of vitiligo and associated autoimmune diseases in Caucasian probands and their families. *Pigment Cell Res* 2003; 16: 208-14.
  121. BETTERLE C, PRESOTTO F, FURMANIAK J: Epidemiology, pathogenesis, and diagnosis of Addison’s disease in adults. *J Endocrinol Invest* 2019; 42: 1407-33.
  122. BLOHMÉ G, NYSTRÖM L, ARNQVIST HJ *et al.*: Male predominance of type 1 (insulin-dependent) diabetes mellitus in young adults: results from a 5-year prospective nationwide study of the 15-34-year age group in Sweden. *Diabetologia* 1992; 35: 56-62.