Interstitial lung disease in microscopic polyangiitis and granulomatosis with polyangiitis: demographic, clinical, serological and radiological features of an Italian cohort from the Italian Society for Rheumatology

A. Manfredi¹, G. Cassone^{1,2}, R. Izzo³, G. Dallagiacoma⁴, M. Felicetti^{5,6}, A. Cariddi⁷, A. Berti⁶,
A. Giollo^{5,8}, C. Nannini⁹, S. Bettio¹⁰, S. Monti¹¹, E. Conticini¹², M. Govoni¹³, L. Quartuccio¹⁴,
L.M. Argolini¹⁵, S. Kaleci¹⁶, G. Emmi¹⁷, C. Dejaco^{4,18}, R. Padoan⁵, L. Dagna⁷, M. Rossini⁸,
F. Cantini⁹, C. Montecucco¹¹, B. Frediani¹², S. De Vita¹⁴, R. Caporali¹⁵, F. Muratore¹⁹,
M. Sebastiani¹, C. Salvarani^{1,19}, from the Italian Study Group of the Italian Society for Rheumatology (SIR)

¹Rheumatology Unit, Azienda Ospedaliera Policlinico di Modena, University of Modena and Reggio Emilia, Modena; ²Clinical and Experimental Medicine, PhD Programme, University of Modena and Reggio Emilia, Modena; ³Rheumatology Unit, Department of Clinical Internal, Anaesthesiologic and Cardiovascular Sciences, Sapienza University of Rome; ⁴Department of Rheumatology, Hospital of Bruneck (ASAA-SABES); ⁵Operative Unit of Rheumatology, Department of Medicine DIMED, University of Padova; ⁶Rheumatology Unit, Santa Chiara Hospital, Trento; ⁷Unit of Immunology, Rheumatology, Allergy and Rare Diseases (UnIRAR), IRCCS San Raffaele Hospital & Vita-Salute San Raffaele University, Milano; ⁸Rheumatology Section, Department of Medicine, University of Verona;⁹Division of Rheumatology, Prato Hospital, Prato; ¹⁰General Medicine Unit 1, Regional Centre for Rare Immunological and Rheumatological Diseases, Ca' Foncello, University Hospital of Treviso; ¹¹Department of Rheumatology, IRCCS Policlinico S. Matteo Fondazione, University of Pavia; ¹²Rheumatology Unit, Department of Medicine, Surgery and Neurosciences, University of Siena; ¹³Rheumatology Unit, S. Anna Hospital (AZOU) Ferrara, Department of Medical Sciences, University of Ferrara; ¹⁴Rheumatology Clinic, Department of Medicine, University of Udine, ASU FC, Udine; ¹⁵Division of Rheumatology, ASST Gaetano Pini, Milan; ¹⁶Department of Surgical, Medical, Dental and Morphological Sciences related to Transplant, Oncology and Regenerative Medicine, Modena; ¹⁷Department of Experimental and Clinical Medicine, University of Florence, Italy; ¹⁸Department of Rheumatology, Medical University Graz, Austria; ¹⁹Rheumatology Unit, IRCCS Arcispedale Santa Maria Nuova, Azienda Unità Sanitaria Locale-IRCCS di Reggio Emilia, Reggio Emilia, Italy.

Abstract

Objective

Interstitial lung disease (ILD) has been described as a possible pulmonary involvement in antineutrophil cytoplasmic antibodies (ANCA)-associated vasculitides (AAV), mainly granulomatosis with polyangiitis (GPA) and microscopic polyangiitis (MPA). The aim of this cross-sectional Italian national study was to describe demographic, clinical and serological profile of ILD related to MPA and GPA and investigate possible correlations between radiologic patterns of ILD and vasculitis features.

Methods

We enrolled 95 consecutive patients with AAV-ILD, 56 affected by MPA (58.9%) and 39 by GPA (41.1%).

Results

NSIP was the most frequently detected ILD pattern, observed in c-ANCA patients in 60.9% of cases, followed by UIP pattern mainly observed in p-ANCA patients (47.7%, p=0.03). ILD represented the first clinical manifestation, preceding vasculitis diagnosis in 22.1% of cases and, globally, ILD was already detectable at AAV diagnosis in 66.3% of patients. The diagnosis of ILD preceded that of AAV in 85.7% of p-ANCA positive-patients, while only one patient with c-ANCA developed ILD before AAV (p=0.039). Multivariate analysis confirmed the correlation of UIP pattern with p-ANCA-positivity and a diagnosis of ILD before AAV, also when adjusted for age and sex.

Conclusion

Our study confirms that UIP is a frequent pattern of lung disease in AAVILD patients. Our results also suggest that ILD can represent an early complication of AAV but also occur in the course of the disease, suggesting the need of a careful evaluation by both pulmonologist and rheumatologist to achieve an early diagnosis. Further prospective studies are needed to define ILD prevalence and evolution in AAV patients.

Key words

interstitial lung disease, ANCA-associated vasculitides, granulomatosis with polyangiitis, microscopic polyangiitis, usual interstitial pneumonia

Andreina Manfredi, MD, PhD Giulia Cassone, MD Raffaella Izzo, MD Gloria Dallagiacoma, MD Mara Felicetti, MD Adriana Cariddi, MD Alvise Berti, MD Alessandro Giollo, MD Carlotta Nannini, MD Silvano Bettio, MD Sara Monti, MD Edoardo Conticini, MD Marcello Govoni, Prof Luca Quartuccio, Prof Lorenza M. Argolini, MD Shaniko Kaleci Giacomo Emmi, MD Christian Dejaco, Prof Roberto Padoan, MD Lorenzo Dagna, Prof Maurizio Rossini, Prof Fabrizio Cantini, MD Carlomaurizio Montecucco, Prof Bruno Frediani, Prof Salvatore De Vita, Prof Roberto Caporali, Prof Francesco Muratore, MD Marco Sebastiani, Prof Carlo Salvarani, Prof Please address correspondence to: Andreina Manfredi, U.O. di Reumatologia, Azienda Policlinico of Modena, Università di Modena e Reggio Emilia, Via del Pozzo 71, 41121 Modena, Italy. E-mail: andreina.manfredi@gmail.com Received on April 13, 2022; accepted in revised form on July 25, 2022. © Copyright CLINICAL AND

EXPERIMENTAL RHEUMATOLOGY 2023.

Competing interests: page 827.

Introduction

Antineutrophil cytoplasmic antibodies (ANCA)-associated vasculitides (AAV) are a heterogeneous group of vasculitis affecting small vessels. They are represented by granulomatosis with polyangiitis (GPA, formerly Wegener's granulomatosis), microscopic polyangiitis (MPA), together with renal-limited vasculitis, and eosinophilic granulomatosis with polyangiitis (EGPA, formerly Churg Strauss disease) (1, 2).

Myeloperoxidase (MPO) and proteinase-3 (PR3) represent the target of ANCA (MPO-ANCA and PR3-ANCA respectively). The former is more frequent in MPA and EGPA patients, the latter can be found in about 80% of patients with GPA. However, these ANCA specificities are not pathognomonic for different AAV and the negativity of ANCA does not exclude a diagnosis of AAV. Diagnosis is generally defined in presence of a combination of clinical and serological findings and/or histological evidence of necrotising pauciimmune small-vessel vasculitis (1, 3-6). Different types of pulmonary involvement have been described in AAV patients; however, the association with interstitial lung disease (ILD) has been also described, mainly in MPA and GPA patients (7, 8). ILD is a rare condition that is well defined in its idiopathic forms, such as idiopathic pulmonary fibrosis (IPF), but also recognised as a frequent complication occurring in immune-mediated rheumatic diseases, such as systemic sclerosis, rheumatoid arthritis, idiopathic inflammatory myopathies, and Sjögren's syndrome (9-11). Our knowledge about ILD in course of AAV is mainly derived from case series, showing that ILD frequently occurs in elderly patients with detectable circulating MPO-ANCA, with usual interstitial pneumonia (UIP) being the most frequent pattern at high-resolution computed tomography (HRCT). Interestingly, in a percentage of patients, the diagnosis of ILD preceded that of AAV (8).

The aim of our cross-sectional multicentric Italian national study was to describe the demographic, clinical and serological profile of ILD secondary to MPA and GPA and investigate possible correlations between HRCT patterns and vasculitis features.

Patients and methods

Diagnosis of AAV

In this national multicentre cross-sectional study, we enrolled all consecutive patients with a diagnosis of ILD associated to confirmed MPA or GPA attending the Rheumatology Units of 14 Italian Centres for a 6-month period (from July 1, 2019 to December 31, 2019).

Diagnosis of GPA or MPA was made on the basis of current classification criteria (12-15), or by expert rheumatologists according to clinical presentation, laboratory and imaging finding (16). Classification of enrolled AAV patients was revised after the publication of new 2022 American College of Rheumatology/European Alliance of Associations for Rheumatology Classification Criteria for MPA and GPA (17, 18).

The serum ANCA positivity was tested on ethanol-fixed neutrophils by indirect immuno-fluorescence (IIF), and/or with a standard commercial enzymelinked immunosorbent assays (ELISA) to detect specific antibodies directed against PR3 or MPO, according to the practice of each immunology laboratory (19).

In particular, all patients underwent IIF, and all patients but 3 underwent ELISA (2 patients were negative at IIF and 1 patient was p-ANCA positive at IIF).

ILD assessment

The diagnosis of ILD was made by means of chest HRCT and multidisciplinary discussion involving at least rheumatologist, pulmonologist and radiologist. Other possible causes of ILD, different from AAV, were excluded. The last available HRCT at the moment of enrolment was considered to determine the radiological pattern. For each centre, HRCT scans were reassessed by an expert chest radiologist who interpreted the radiologic pattern of ILD according to the Fleischner Society White Paper statement on the diagnosis of idiopathic pulmonary fibrosis (IPF) (20). The pattern of disease was recorded as definite, probable UIP or indeterminate for UIP. If a pattern indeterminate for UIP was noted, it was further classified as nonspecific interstitial pneumonia (NSIP), organising pneumonia (OP) or other patterns, that can also be observed in combination (20-22).

The last available HRCT and pulmonary function tests (PFTs), if available, were recorded.

PFTs were performed in all centres according to standard methods (23, 24). The results of PFTs were expressed as percentage of the predicted value of each parameter and corrected for age, gender and height. Pulmonary function was evaluated by means of forced vital capacity (FVC). Single-breath diffusing capacity of the lung for carbon monoxide (DLCO-SB) was used to assess gas transfer.

Vasculitis assessment

For all patients, demographic data, clinical and serological characteristics of vasculitis, time of diagnosis of MPA or GPA and ILD, as well as the presence of pulmonary symptoms such as dry cough and dyspnoea were recorded at the time of vasculitis or ILD diagnosis. Past and current treatments for each patient were also collected. All data were recorded on a standardised case-report form and entered into a computerised database.

Disease activity was determined using the Birmingham Vasculitis Activity Score (BVAS), version 3 (25).

Ethics

The study was approved by the local Institutional Review Board "Comitato Etico Area Vasta Nord" (approval number: AOU0011234/20).

Statistical analysis

Statistical analysis was performed using STATA® software v. 14 (StataCorp. 2015. Stata Statistical Software: Release 14. College Station, TX: StataCorp LP.). Descriptive statistics were presented for baseline demographic clinical characteristics for the entire group, as well as for the groups of patients. Continuous variables were presented as the number of patients (N), median and interquartile range (IQR), and compared between subgroups using non-

Table I. Demographic, clinical and serological features of patients

		n	%
Total number	95		
GPA/MPA	39/56	41/59	
Ever smokers	28	29.5	
Male/Female	ratio	52/43	54.7/45.3
pANCA	65	68.4	
cANCA	23	24.2	
Anti-MPO	52	54.7	
Anti-PR3	21	22.1	
ILD pattern			
F	UIP	36	37.9
	NSIP	49	51.6
	OP	10	10.5
ILD occurrent	ce.		
	Concurrent	42	44.2
	Preceding	21	22.1
	Following	32	33.7
II D accurran	Tonowing	52	55.1
	Until 6 months*	63	66.3
	More than 6 months*	32	33.7
Elizate all in the		52	55.1
First clinical i	nanifestation of vasculitis (more results possible)	0	0.5
	Skin vasculitis	9	9.5
	Renai involvement	34	35.8
	Neurological involvement	15	15.8
	ENT involvement	18	18.9
	Lung involvement	60	63.2
	Other	25	26.3
Clinical mani	festations of vasculitis		
	Skin vasculitis	20	21.1
	Renal involvement	49	51.6
	Neurological involvement	37	40.6
	ENT involvement	36	37.9
	Arthritis	29	30.5
Lung involve	ment		
Build in our of	Asthma	4	4.2
	Alveolar haemorrhage	13	13.7
	Lung cavitation	11	11.6
	Lung infiltrates	35	36.8
	Serositis	13	13.7
	ARDS	13	14 7
Cough	111120	35	36.8
Dysphoea		55	57.9
_ / °P11004			***
		Median	IQR
Median age y	ears (IQR)	69	62,75
Median diseas	se duration months (IQR)	31	8,74
Median interv	al vasculitis-ILD onset months (IQR)	0	-12, 16
B-VAS diagno	osis	14	8,20
B-VAS enrolr	nent	1	0,5
FVC % (IQR)	1	85	70,101
DLCO % (IQ	R)	57	44,77
Treatments		n	%
Cyclophospha	mide	48	50.5
Rituximab		44	46.3
Mycophenola	te mofetil	24	25.3
Azathioprine		38	40
Methotrexate		18	18.9
Glucocorticoi	ds	93	97.9

*since the diagnosis of AAV.

GPA: granulomatosis with polyangiitis; MPA: microscopic polyangiitis; ANCA: antineutrophil cytoplasmic antibodies; MPO: myeloperoxidase; PR3: proteinase-3; ILD: interstitial lung disease; UIP: usual interstitial pneumonia; NSIP: nonspecific interstitial pneumonia; OP: organising pneumonia; ENT: ear, nose and throat ; ARDS: Acute Respiratory Distress Syndrome; B-VAS: Birmingham Vasculitis Activity Score; FVC: forced vital capacity; DLCO: diffusing capacity of the lung for carbon monoxide.

parametric Mann-Whitney test; while categorical variables were presented as frequency (n, percentage [%]) and compared using Pearson's chi-squared test or Fisher's exact test as appropriate. The association between these parameters and UIP versus other patterns was assessed using a logistic regression model with stepwise forward selection. In the first step, the intercept-only model was fitted and individual score statistics for the potential variables were evaluated. A significance level of p < 0.05 was used to allow a variable into the model. In stepwise selection, an attempt was made to remove any insignificant variables from the model before adding a significant variable to the model. The Hosmer and Lemeshow test was used to evaluate "goodness of fit" in the selection model. Data from the univariate and multivariate logistic regression analyses were expressed as odds ratio (OR) and 95% confidence interval (CI). A p-value <.05 was considered statistically significant.

Results

Characteristics of AAV-ILD patients A total of 95 patients were enrolled, 56 affected by MPA (58.9%) and 39 by GPA (41.1%). Fifty-two patients were males (54.7%) and 43 females (45.3%), a median age of 69 years (62, 75), and the median disease duration of AAV was of 31 months (8, 74). P-ANCA positivity was detected in 65 patients (68.4%), c-ANCA in 23 patients (24.2%). MPO-ANCA were detected in 52 patients (54.7%), while PR3-ANCA were observed in 21 patients (22.1%). Among HRCT patterns, a UIP, NSIP, or other patterns, were observed in 36, 49 and 10 patients (37.9, 51.6, 10.5% respectively).

Details about demographic, serological and clinical features of AAV-ILD patients are summarised in Table I.

Comparison between

p-ANCA and c-ANCA patients Our population enclosed 65 patients with p-ANCA (31 females and 34 males respectively) and 23 patients with c-ANCA positivity (7 females and 16 males respectively), see Table II for more details. Table II. Demographic, clinical and serological features of patients with pANCA and cANCA.

	pANCA n (%)		cANCA n (%)		р
Total number	65	(68.4)	23	(24.2)	
Male/Female ratio	34/31		16/7		0.13
ILD pattern					
UIP	31	(47.7)	4	(17.4)	
NSIP	29	(44.6)	14	(60.9)	0.03
Other	5	(7.7)	5	(21.7)	
ILD occurrence					
Following	18	(27.7)	10	(47.8)	
Concurrent	29	(44.6)	11	(47.8)	0.13
Preceding	18	(27.7)	1	(4.3)	
Median age years (IQR)	69.5	(63, 75)	66	(59.5, 74.5)	0.07
Median disease duration months (IQR)	25	(8,71)	27.5	(4, 133)	0.46
Median interval vasculitis-ILD onset months (IQR)	-2.5	(-14.5, 2.5)	60	(0, 113)	<0.01
B-VAS diagnosis (IQR)	16	(10, 21)	17	(7,21)	0.42
B-VAS enrolment (IQR)	0	(0,5)	2	(0, 5)	0.47
FVC % (IQR)	86	(70.5, 101.5)	81	(72.5, 103,5)	0.65
DLCO % (IQR)	56.5	(46, 74)	66	(53.5, 88.5)	0.18

ILD: interstitial lung disease; UIP: usual interstitial pneumonia; NSIP: nonspecific interstitial pneumonia; OP: organising pneumonia; B-VAS: Birmingham Vasculitis Activity Score; FVC: forced vital capacity; DLCO: diffusing capacity of the lung for carbon monoxide.

Case	Diagnosis	Clinical picture at diagnosis
1. Female, 66	GPA	lung nodules, peripheral neuropathy, cardiac involvement, ENT in- volvement*
2. Male, 75	GPA	lung nodules and cavitations, glomerulonephritis**, purpura, chronic sinusitis, ENT involvement*
3. Female, 59	GPA	lung nodules and infiltrates, ENT involvement, arthritis, constitutional symptoms
4. Female, 59	GPA	lung involvement with infiltrates and respiratory failure (ARDS), pe- ripheral neuropathy, CNS involvement, chronic sinusitis, ENT involve- ment*
5. Female, 57	GPA	middle ear involvement**, mononeuritis multiplex, arthritis, CNS involvement
6. Female, 53	GPA	multiple mononeuritis**, arthritis, central nervous system involvement
7. Male, 73	MPA	skin vasculitis**, transient lung infiltrates, peripheral neuropathy, con- stitutional symptoms

*Ear, nose, throat, including otitis media, bloody nasal discharge and crusting. **confirmed by biopsy.

GPA: granulomatosis with polyangiitis; MPA: microscopic polyangiitis; ENT: ear, nose and throat; ARDS: Acute Respiratory Distress Syndrome.

ANCA were negative in 7 patients. No differences were observed with regard to median age at diagnosis and disease duration of vasculitis, while a short disease duration between AAV and ILD diagnosis was observed in p-ANCA patients (p<0.01). UIP pattern was more frequent in p-ANCA than in c-ANCA patients (47.7%), while NSIP pattern was detected in 60.9% of c-ANCA pa-

tients, with a statistically significant difference (p=0.03).

Six out of seven (85.7%) ANCA negative patients showed an NSIP pattern, and ILD diagnosis followed AAV diagnosis in 3/7 cases (42.8%). General features of ANCA negative patients are summarised in Table III.

All 7 patients had a diagnosis of ANCA negative vasculitis according to the

opinion of a rheumatologist expert in vasculitis and satisfied the new ACR/ EULAR AAV classification criteria (17, 18). Furthermore, the diagnosis of vasculitis was confirmed by biopsy in 4 patients and histology was compatible with 2012 Revised International Chapel Hill Consensus Conference Nomenclature (CHCC) for MPA or GPA (14). All 7 patients had long-term follow-up and other diagnoses were excluded.

Comparison according to temporal correlation between ILD and AAV diagnosis

ILD was diagnosed before AAV in 21 patients (22.1%), the diagnoses of ILD and AAV were concomitant in 42 patients (44.2%), while in 32 patients (33.7%) ILD followed the diagnosis of AAV.

P-ANCA positivity was observed in 18/21 patients (85.7%) in whom the diagnosis of ILD preceded that of AAV, while c-ANCA were detected only in 1 case. Evaluating ANCA specificities, we observed that PR3-ANCA were detected in patients with ILD preceding AAV only in one case (p=0.039).

Patients in whom ILD was detected before the diagnosis of AAV revealed mainly a UIP pattern (41.7%) while NSIP pattern occurred less frequently (10.2%). Regarding ILD occurrence in relation to the time of AAV diagnosis, the differences were statistically significant (p=0.003) (Table IV).

Comparison according to HRCT pattern of UIP or NSIP

NSIP pattern was detected in 49 patients (51.6%), 25 females and 24 males (51% and 49% respectively), while UIP pattern was detected in 36 patients (37.9%), 13 females and 23 males (36.1% and 63.9%, respectively). UIP pattern was observed more frequently in patients with a diagnosis of MPA (27/56, 48.2%) while NSIP pattern was more frequent in GPA patients (30/39, 76.9%) (p=0.018). P-ANCA were more frequently observed in patients with UIP pattern compared to those with NSIP pattern (86.1% ver*sus* 59.2%, *p*<0.001), furthermore ever smoking was found to be significantly associated with UIP pattern (51.6%

Table IV. Demographic, clinical and serological features of patients according to ILD onset.

	preceding n (%)	concurrent n (%)	following n (%)	р
Total number	21 (22.1)	42 (44.2)	32 (33.7)	
GPA/MPA	4/17	19/23	16/16	0.062
Male/Female ratio	14/7	20/22	18/14	0.351
pANCA	18 (85.7)	29 (69)	18 (56.3)	
cANCA	1 (4.8)	11 (26.2)	11 (34.4)	0.13
ANCA-negative	2 (9.5)	2 (4.8)	3 (9.4)	
Anti-MPO	13 (61.9)	25 (59.5)	14 (48.3)	0.548
Anti-PR3	1 (4.8)	14 (33.3)	6 (20.7)	0.037
ILD pattern				
ÛIP	15 (71.4)	10 (23.8)	11 (34.4)	
NSIP	5 (23.8)	25 (59.5)	19 (59.4)	0.011
Other	1 (4.8)	7 (16.7)	2 (6.2)	
Median age years (IQR)	70 (66, 74)	65 (62, 75)	69 (55, 75)	0.074
Median disease duration months (IQR)	32 (5.5, 70.5)	12 (6,56)	52 (24, 137)	0.025
Median interval vasculitis-ILD onset months (IQR)	-22 (-65, -12)	0 (-3,0)	47 (15, 118)	<0.001
B-VAS diagnosis (IQR)	13 (7, 17)	15.5 (11,21)	15 (9,23)	0.25
B-VAS enrolment (IQR)	1 (0,3)	0 (0,4)	2(0,8)	0.1
FVC % (IQR)	75 (64, 101)	98 (78, 112)	78 (69, 93)	0.076
DLCO % (IQR)	47 (34, 71)	64 (51, 89)	61 (52, 74)	0.173

GPA: granulomatosis with polyangiitis; MPA: microscopic polyangiitis; ANCA: antineutrophil cytoplasmic antibodies; MPO: myeloperoxidase; PR3: proteinase-3; ILD: interstitial lung disease; UIP: usual interstitial pneumonia; NSIP: nonspecific interstitial pneumonia; B-VAS: Birmingham Vasculitis Activity Score; FVC: forced vital capacity; DLCO: diffusing capacity of the lung for carbon monoxide.

Table V. Demographic, clinical	and serological fe	eatures of patients	according to the radio-
logical pattern.			

	UIP	Ν	ISIP	р
Total number	36 (37.9) 49	(51.6)	
GPA/MPA	9/27	30/	/29	0.018
Male/Female ratio	23/13	24	/25	0.172
Ever smokers	16 (51.6) 10	(25)	0.021
pANCA	31 (86.1) 29	(59.2)	< 0.001
cANCA	4 (11.1) 14	(28.6)	0.06
Anti-MPO	24 (66.7) 24	(51.1)	0.154
Anti-PR3	6 (16.7) 11	(23.4)	0.451
ILD occurrence				
Preceding	15 (41.7) 6	(10.2)	
Concurrent	10 (27.8) 25	(51)	0.003
Following	11 (30.6) 19	(38.8)	
ILD occurrence				
Until 6 months	25 (69.4) 38	(64.4)	0.66
More than 6 months	11 (30.6) 21	(35.6)	
Median age years (IQR)	69.5 (63,	75) 69	(62, 75)	0.21
Median disease duration months (IQR)	23 (6,6)	3) 52.5	(11,97)	0.83
Median interval vasculitis-ILD onset months (IQR)	-3 (-24,	13) 0	(-5,96)	0.393
B-VAS diagnosis	14.5 (10.5	, 21.5) 16	(8, 20)	0.59
B-VAS enrolment	1 (0,9)	1	(0,3)	0.48
FVC % (IQR)	86 (68, 1	101) 83.5	(69, 100)	0.921
DLCO % (IQR)	60 (44,	77) 56.5	(46, 73)	0.882

GPA: granulomatosis with polyangiitis; MPA: microscopic polyangiitis; ANCA: antineutrophil cytoplasmic antibodies; MPO: myeloperoxidase; PR3: proteinase-3; ILD: interstitial lung disease; B-VAS: Birmingham Vasculitis Activity Score; FVC: forced vital capacity; DLCO: diffusing capacity of the lung for carbon monoxide.

versus 25%, p=0.02), but no correlation was detected between smoking and ANCA pattern (p=0.79). No significant differences were observed in re-

gard to MPO-ANCA and PR3-ANCA, median age at AAV diagnosis, vasculitis duration and interval between AAV and ILD diagnoses (Table V).

Parameter	Standard error	OD	95% CI	р
ANCA (p-ANCA)	3.13	4.4	1.10-17.73	0.04
Smoke	1.34	2.13	0.62-7.32	0.23
Occurrence before AAV (preceding)	5.23	7.6	1.97-29.31	< 0.01
Male sex	1.14	1.79	0.51-6.22	0.36
Age	0.02	1.01	0.96-1.06	0.76

 Table VI. Multivariate analysis: factors associated to UIP pattern in AAV-ILD patients.

OR: odds ratio; CI: confidence interval; ANCA: antineutrophil cytoplasmic antibodies; AAV: ANCA-associated vasculitis.

Clinical and organ involvement, namely skin vasculitis, arthritis, renal, neurological and ENT involvement didn't correlate with HRCT pattern of ILD, neither with ANCA serology or time of ILD appearance (data not showed).

Multivariate analysis

Univariate analysis was performed to evaluate possible correlations between HRCT UIP pattern and AAV clinicalserological features, showing a significant association with MPA, smoke, p-ANCA and occurrence of ILD before AAV diagnosis. On the basis of the these results a multivariate regression model was also performed.

Therefore, the model included type of ANCA, smoking, and timing of ILD occurrence in comparison to vasculitis diagnosis. The analysis confirmed the correlation of UIP pattern with p-ANCA (OR 4.40; 95%CI: 1.10–17.72) and ILD diagnosed before AAV appearance (OR 7.60; 95%CI 1.97–29.31), also when adjusted for age and sex (Table VI).

Discussion

In this paper, we investigated for the first time in a cross-sectional study in an Italian population, the serological, clinical and radiological features of ILD associated to MPA and GPA patients. Our results showed that NSIP was the HRCT ILD pattern most frequently detected in AAV patients, and the diagnosis of ILD was predominantly made before or concurrently with that of vasculitis. Moreover, UIP pattern was more frequently observed in p-ANCA positive patients.

During the last few years, an increasing number of publications reported the association between ILD and AAV. In 2014, Comarmond *et al.* described a population of 49 patients with ILD associated with AAV. The diagnosis of ILD preceded the diagnosis of vasculitis in 45% of patients, was concurrent in 43% and followed the diagnosis of AAV in 12% of patients. In 88% of cases, ANCA were MPO. In regard to radiologic pattern, UIP was the most frequently detected (43%), while other patterns were: atypical UIP (14%), fibrotic NSIP (7%), combined pulmonary fibrosis and emphysema (CPFE) (21.4%), NSIP (9.5%), and finally indeterminate (neither UIP nor NSIP although reticulation was present) in the remaining patients (4.8%) (26).

Recently, a larger AAV-ILD cohort was described by Maillet et al. including 62 AAV-ILD patients, namely 53 MPA and 9 GPA. Also, in this case, ILD was diagnosed mostly before (52%) or simultaneously (39%) compared to the vasculitis diagnosis. The majority of patients (63%) showed a UIP pattern, while only 24 patients have a NSIP pattern (35%) (27). These authors also compared AAV-ILD to AAV patients without ILD observing that the former had a lower prevalence of systemic manifestations. Finally, patients with UIP pattern showed a shorter survival at Cox regression analysis, suggesting that AAV-ILD cases had less active vasculitis but a higher risk of death because of UIP, which progressed independently of vasculitis (27).

To better investigate features of AAV-ILD we compared different subgroups, according to antibody profile, HRCT pattern of ILD, and chronological relationship with AAV. To our knowledge, the population described in our paper represents the largest in literature, and we confirmed in this population some results already shown by previous studies. We observed that ILD represented the first clinical manifestation, preceding the vasculitis diagnosis, in 22.1% of cases and that globally ILD was already detectable at AAV diagnosis in 66.3% of patients. The new classification criteria for MPA highlights the importance of ILD for diagnosis of MPA, but, when vasculitis is absent, diagnosis of AAV remains challenging. In a multicentre Italian-Spanish study, only 9/58 ILDpatients with anti-MPO antibodies developed an MPA in a 39-month period, while 4 patients developed other rheumatic diseases. This result confirms that anti-MPO antibodies are necessary, but not sufficient, to make a diagnosis of MPA in ILD patients.

Other authors reported the presence of ANCA in patients with IPF, with a prevalence up to 35% in the Japanese population (28-30). However, the occurrence of vasculitis, mainly MPA, has been described in a minority of these cases and only 7-23% of ANCA-positive subjects with a diagnosis of IPF subsequently developed a clinically overt vasculitis, mainly in patients with anti-MPO antibodies (28, 29). Our data confirm the association between p-AN-CA and the development of vasculitis in patients with ILD, because only one c-ANCA patient developed ILD before the occurrence of AAV.

In 2015, IPAF classification criteria were proposed, in order to define patients with interstitial pneumonia and some clinical, serological or morphological characteristics suggestive of rheumatic disease, but not sufficient to make a definite diagnosis. One limitation of these classification criteria was the lack of ANCA antibodies in the serological domain. Therefore, patients with ANCA and ILD could not be classified as IPAF in any case (31).

Until now, only a few studies have evaluated the features of ILD in c-ANCA positive and ANCA-negative patients with MPA and GPA (32-37). Interestingly, although limited by the low number of patients, we observed that c-ANCA positive and ANCA negative patients (30 patients, 31.6% of population) showed similar features being prevalently characterised by NSIP pattern (20/30 patients, 66.7%) and preceding the diagnosis of vasculitis only in a minority of patients (3/30, 10%).

This subgroup of patients could also explain the high prevalence of NSIP HRCT pattern recorded in our study if compared with other studies. In fact, according to the currently published studies (27, 38), our data confirm that UIP is the prevalent pattern observed in p-ANCA positive patients (47.7%), but we also observed that NSIP is predominant in c-ANCA positive and ANCA negative cases.

These results suggest that we can probably distinguish two different clinical phenotypes. In the first, ILD usually precedes vasculitis, patients are more frequently p-ANCA positive and have a predominant UIP pattern at HRCT. A number of these patients are initially classified as IPF, and therefore they need a careful assessment by the pulmonologist of clinical manifestations and laboratory findings during their follow-up for an early and appropriate management and treatment of vasculitis. In addition, this subgroup of patients has a poor prognosis more related to ILD than to vasculitis, and, in analogy to IPF, anti-fibrotic agents might be useful (8, 39, 40). This clinical subgroup supports the biological hypothesis of a fibrosing effect of AN-CA-MPO (41).

On the other hand, in the second phenotype, NSIP pattern is frequently detected concomitantly with diagnosis of AAV or in the course of the disease; these patients are more frequently c-ANCA positive or ANCA-negative suggesting the need for a regular assessment also for ILD in patients with AAV, particularly in those with GPA.

Our study has many limitations, but also some strengths. First, since data at disease onset and diagnosis were retrospectively collected, incomplete data sets may have influenced the findings. Other limitations include possible referral bias of cases; patients in this cohort may represent the those with more severe ILD, while asymptomatic or mildly symptomatic patients may have been less frequently identified and included in the study.

Finally, the lack of a central reading of the HRCTs could represent a bias in HRCT pattern definition, but on the other side all the participant centres had thoracic radiologists with high expertise in parenchymal lung diseases.

A strength of this study is the large number of consecutive cases which should provide a more reliable perspective of the spectrum of ILD in MPA and GPA than the previous smaller series. Furthermore, the patients were uniformly evaluated with a standardised data collection.

Further prospective studies with larger and homogeneous populations are needed to better define the ILD prevalence and evolution in AAV, its clinical phenotype and possibly the better therapeutic approach.

Acknowledgments

We thank the Italian Society for Rheumatology - Società Italiana di Reumatologia (SIR) - and in particular the Vasculitis Study Group.

Competing interests

A. Berti has served on advisory boards and has received lecturing fees from GSK, all unrealted to the present publication. A. Giollo has received speaker and consultancy fees from Galapagos, Novartis and Eli Lilly. C. Dejaco has received research grants from Abbvie, honoraria for consultancies and/or speaker's bureau from Abbvie, Pfizer, Lilly, Galapagos, Roche, Sanofi, Novartis and Janssen. R. Caporali has received consulting fees from Abbvie, BMS, Amgen, Celltrion, Lilly, Fresenius-Kabi, Galapagos, Janssen, MSD, Novartis, Pfizer and UCB.

The other authors have declared no competing interests.

Contributors

F. Schiavon, E. Marasco, A. Gattamelata, R. Bortolotti, D. Malandrino, B. Maranini.

References

- NAKAZAWA D, MASUDA S, TOMARU U, ISHIZU A: Pathogenesis and therapeutic interventions for ANCA-associated vasculitis. *Nat Rev Rheumatol* 2019; 15: 91-101. https://doi.org/10.1038/s41584-018-0145-y
- FERRO F, QUARTUCCIO L, MONTI S et al.: One year in review 2021: systemic vasculitis. *Clin Exp Rheumatol* 2021; 39 (Suppl. 129): S3-12. https://
- doi.org/10.55563/clinexprheumatol/v1tpfo 3. KALLENBERG CGM: Key advances in the

clinical approach to ANCA-associated vasculitis. *Nat Rev Rheumatol* 2014; 10: 484-93. https://doi.org/10.1038/nrrheum.2014.104

4. CORNEC D, GALL EC LE, FERVENZA FC, SPECKS U: ANCA-associated vasculitis-clinical utility of using ANCA specificity to classify patients. *Nat Rev Rheumatol* 2016; 12: 570-9.

https://doi.org/10.1038/nrrheum.2016.123

- LIONAKI S, BLYTH ER, HOGAN SL et al.: Classification of antineutrophil cytoplasmic autoantibody vasculitides: The role of antineutrophil cytoplasmic autoantibody specificity for myeloperoxidase or proteinase 3 in disease recognition and prognosis. Arthritis Rheum 2012; 64: 3452-62. https://doi.org/10.1002/art.34562
- 6. WÓJCIK K, BIEDROŇ G, WAWRZYCKA-AD-AMCZYK K *et al.*: Subphenotypes of ANCAassociated vasculitis identified by latent class analysis. *Clin Exp Rheumatol* 2021; 39 (Suppl. 129): S62-8. https://
- doi.org/10.55563/clinexprheumatol/d01o72
 7. ALBA MA, FLORES-SUÁREZ LF, HENDERSON AG et al.: Interstital lung disease in ANCA vasculitis. Autoimmun Rev 2017; 16: 722-9. https://doi.org/10.1016/j.autrev.2017.05.008
- SEBASTIANI M, MANFREDI A, VACCHI C et al.: Epidemiology and management of interstitial lung disease in ANCA-associated vasculitis. *Clin Exp Rheumatol* 2020; 38 (Suppl. 124): S221-31.
- WALLIS A, SPINKS K: The diagnosis and management of interstitial lung diseases. *BMJ* 2015; 350: h2072. https://doi.org/10.1136/bmj.h2072
- COTTIN V, HIRANI NA, HOTCHKIN DL et al.: Presentation, diagnosis and clinical course of the spectrum of progressive-fibrosing interstitial lung diseases. *Eur Respir Rev* 2018; 27: 180076.

https://doi.org/10.1183/16000617.0076-2018 11. MIRA-AVENDANO I, ABRIL A, BURGER CD

- 11. MIRA-AVENDANO I, ABRIL A, BURGER CD et al.: Interstitial lung disease and other pulmonary manifestations in connective tissue diseases. *Mayo Clin Proc* 2019; 94: 309-25. https://
- doi.org/10.1016/j.mayocp.2018.09.002
- 12. WATTS R, LANE S, HANSLIK T *et al.*: Development and validation of a consensus methodology for the classification of the ANCA-associated vasculitides and polyarteritis no-dosa for epidemiological studies. *Ann Rheum Dis* 2007; 66: 222-7.
 - https://doi.org/10.1136/ard.2006.054593
- LEAVITT RY, FAUCI AS, BLOCH DA *et al.*: The American College of Rheumatology 1990 criteria for the classification of wegener's granulomatosis. *Arthritis Rheum* 1990; 33: 1101-7.
 - https://doi.org/10.1002/art.1780330807
- 14. JENNETTE JC, FALK RJ, BACON PA et al.: 2012 Revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides. Arthritis Rheum 2013; 65: 1-11. https://doi.org/10.1002/art.37715
- 15. MAHR A, KATSAHIAN S, VARET H et al.: Revisiting the classification of clinical phenotypes of anti-neutrophil cytoplasmic antibody-associated vasculitis: A cluster analysis. Ann Rheum Dis 2013; 72: 1003-10. https:// doi.org/10.1136/annrheumdis-2012-201750

- 16. CORRAL-GUDINO L, GONZÁLEZ-VÁZQUEZ E, CALERO-PANIAGUA I et al.: The complexity of classifying ANCA-associated smallvessel vasculitis in actual clinical practice: data from a multicenter retrospective survey. *Rheumatol Int* 2020; 40: 303-11. https://doi.org/10.1007/s00296-019-04406-5
- SUPPIAH R, ROBSON JC, GRAYSON PC et al.: 2022 American College of Rheumatology/European Alliance of Associations for Rheumatology classification criteria for microscopic polyangiitis. Ann Rheum Dis 2022; 81: 321-6. https://
- doi.org/10.1136/annrheumdis-2021-221796
 18. ROBSON JC, GRAYSON PC, PONTE C et al.: 2022 American College of Rheumatology/ European Alliance of Associations for Rheumatology classification criteria for granulomatosis with polyangiitis. Ann Rheum Dis 2022; 81: 315-20. https://
- doi.org/10.1136/annrheumdis-2021-221795 19. BOSSUYT X, COHEN TERVAERT J, ARIMURA
- Y *et al.*: Position paper: Revised 2017 international consensus on testing of ANCAs in granulomatosis with polyangiitis and microscopic polyangiitis. *Nat Rev Rheumatol* 2017; 13: 683-92.
- https://doi.org/10.1038/nrrheum.2017.140
 20. LYNCH DA, SVERZELLATI N, TRAVIS WD et al.: Diagnostic criteria for idiopathic pulmonary fibrosis: a Fleischner Society White Paper. Lancet Respir Med 2018; 6: 138-53. htt-
- ps://doi.org/10.1016/s2213-2600(17)30433-2
 21. TANAKA N, KIM JS, NEWELL JD *et al.*: Rheumatoid arthritis-related lung diseases: CT findings. *Radiology* 2004; 232: 81-91. https://doi.org/10.1148/radiol.2321030174
- 22. TRAVIS WD, COSTABEL U, HANSELL DM et al.: An Official American Thoracic Society/European Respiratory Society Statement: Update of the International Multidisciplinary Classification of the Idiopathic Interstitial Pneumonias. Am J Respir Crit Care Med 2013; 188: 733-48.
- https://doi.org/10.1164/rccm.201308-1483st
 KARIMI-SHAH BA, CHOWDHURY BA: Forced vital capacity in idiopathic pulmonary fibrosis FDA review of Pirfenidone and Nintedanib. N Engl J Med 2015; 372: 1189-91.

https://doi.org/10.1056/nejmp1500526

- 24. GRAHAM BL, STEENBRUGGEN I, BARJAK-TAREVIC IZ *et al.*: Standardization of Spirometry 2019 Update. An Official American Thoracic Society and European Respiratory Society Technical Statement. *Am J Respir Crit Care Med* 2019; 200: e70-88. https://doi.org/10.1164/rccm.201908-1590st
- 25. MUKHTYAR C, LEE R, BROWN D et al.: Modification and validation of the Birmingham vasculitis activity score (version 3). Ann Rheum Dis 2009; 68: 1827-32. https://doi.org/10.1136/ard.2008.101279
- 26. COMARMOND C, CRESTANI B, TAZI A et al.: Pulmonary fibrosis in antineutrophil cytoplasmic antibodies (ANCA)-associated vasculitis. *Medicine* (Baltimore) 2014; 93: 340-9. https://

doi.org/10.1097/md.000000000000217

- MAILLET T, GOLETTO T, BELTRAMO G et al.: Usual interstitial pneumonia in ANCA-associated vasculitis: A poor prognostic factor. J Autoimmun 2020; 106: 102338. https://doi.org/10.1016/j.jaut.2019.102338
- ANDO M, MIYAZAKI E, ISHII T et al.: Incidence of myeloperoxidase anti-neutrophil cytoplasmic antibody positivity and microscopic polyangitis in the course of idiopathic pulmonary fibrosis. *Respir Med* 2013; 107: 608-15.
- https://doi.org/10.1016/j.rmed.2013.01.006
 29. KANG BH, PARK JK, ROH JH et al.: Clinical significance of serum autoantibodies in idiopathic interstitial pneumonia. J Korean Med Sci 2013; 28: 731.
 - https://doi.org/10.3346/jkms.2013.28.5.731
- 30. SEBASTIANI M, LUPPI F, SAMBATARO G et al.: Interstitial lung disease and anti-myeloperoxidase antibodies: not a simple association. J Clin Med 2021; 10: 2548. https://doi.org/10.3390/jcm10122548
- 31. FISCHER A, ANTONIOU KM, BROWN KK et al.: An official European Respiratory Society/American Thoracic Society research statement: Interstitial pneumonia with autoimmune features. Eur Respir J 2015; 46: 976-87. https:// doi.org/10.1183/13993003.00150-2015
- 32. KATSUMATA Y, KAWAGUCHI Y, YAMANAKA H: Interstitial lung disease with ANCA-associated vasculitis. *Clin Med Insights Circ*

Respir Pulm Med 2015; 9 (Suppl. 1): 51-6. https://doi.org/10.4137/ccrpm.s23314

- 33. HIRAYAMA K, KOBAYASHI M, USUI J et al.: Pulmonary involvements of anti-neutrophil cytoplasmic autoantibody-associated renal vasculitis in Japan. Nephrol Dial Transpl 2015; 30 (Suppl. 1): i83-93. https://doi.org/10.1093/ndt/gfu385
- 34. CHEN M, YU F, ZHANG Y, ZHAO MH: Antineutrophil cytoplasmic autoantibodyassociated vasculitis in older patients. *Medicine* (Baltimore) 2008; 87: 203-9. https:// doi.org/10.1097/md.0b013e31817c744b
- 35. FOULON G, DELAVAL P, VALEYRE D et al.: ANCA-associated lung fibrosis: Analysis of 17 patients. *Respir Med* 2008; 102: 1392-8. https://doi.org/10.1016/j.rmed.2008.04.023
- 36. NOZU T, KONDO M, SUZUKI K, TAMAOKI J, NAGAI A: A comparison of the clinical features of ANCA-positive and ANCA-negative idiopathic pulmonary fibrosis patients. *Respiration* 2009; 77: 407-15. https://doi.org/10.1159/000183754
- HOZUMI H, ENOMOTO N, OYAMA Y *et al.*: Clinical implication of proteinase-3-antineutrophil cytoplasmic antibody in patients with idiopathic interstitial pneumonias. *Lung* 2016; 194: 235-42.
 - https://doi.org/10.1007/s00408-016-9851-x
- 38. MOHAMMAD AJ, MORTENSEN KH, BABAR J et al.: Pulmonary involvement in antineutrophil cytoplasmic antibodies (ANCA)associated vasculitis: the influence of ANCA subtype. J Rheumatol 2017; 44: 1458-67. https://doi.org/10.3899/jrheum.161224
- 39. SGALLA G, COMES A, RICHELDI L: An updated safety review of the drug treatments for idiopathic pulmonary fibrosis. *Expert Opin Drug Saf* 2021; 20: 1035-48. https:// doi.org/10.1080/14740338.2021.1921143
- 40. COLLINS BF, LUPPI F: Diagnosis and management of fibrotic interstitial lung diseases. *Clin Chest Med* 2021; 42: 321-35. https://doi.org/10.1016/j.ccm.2021.03.008
- 41. POPAT RJ, HAKKI S, THAKKER A *et al.*: Anti-myeloperoxidase antibodies attenuate the monocyte response to LPS and shape macrophage development. *JCI Insight* 2017; 2: e87379.

https://doi.org/10.1172/jci.insight.87379