Progression prediction in idiopathic inflammatory myopathy-associated interstitial lung disease: a combination of initial high attenuation areas and anti-melanoma differentiation-associated gene 5-positive

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Abstract Objective

To analyse quantitative lung densitometry and clinical baseline data of individuals with idiopathic inflammatory myopathy (IIM) and identify risk factors capable of predicting the progression of interstitial lung disease (ILD).

Methods

We utilised quantitative lung densitometry and clinical baseline data as explanatory variables. Univariate and multivariate Cox regression analyses were employed to pinpoint effective risk factors for predicting ILD progression in IIM patients.

Results

The findings from the Cox univariate regression analysis indicate that elevated carcinoembryonic antigen levels (HR=1.036, 95% CI 1.004-1.069) are connected to an elevated risk of ILD progression in patients with IIM (P=0.027), while PO₂ (HR=0.980, 95% CI 0.962-0.997), forced vital capacity (HR=0.551, 95% CI 0.320-0.946) are protective factors for ILD progression in patients with IIM (p=0.025, p=0.031, respectively), anti-EJ positivity (HR=0.399, 95% CI 0.175-0.912) and anti-Ro52 positivity (HR=0.437, 95% CI 0.199-0.960) are risk factors for ILD progression in patients with IIM (p=0.029, p=0.039, respectively). Furthermore, the results of Cox multivariate regression analysis reveal that high attenuation areas (HAA) (>465.745 cm³) (HR=5.007, 95% CI 0.041-0.396) are autonomous prognostic risk factors for ILD progression in individuals with IIM (p=0.002, p<0.001, respectively).

Conclusion

Among IIM patients, those who are anti-MDA5-positive, and exhibit HAA (>465.745cm³) are more likely to experience ILD progression.

Key words

idiopathic inflammatory myopathy, interstitial lung disease, high attenuation areas, anti-melanoma differentiation-associated gene 5

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Received on August 31, 2023; accepted in revised form on October 9, 2023.

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Idiopathic inflammatory myopathies (IIMs) constitute a diverse group of connective tissue disorders characterised by differing degrees of acquired skeletal muscle inflammation. This group includes conditions such as polymyositis (PM), dermatomyositis (DM), and clinically amyopathic dermatomyositis (CADM) (1, 2). IIM can involve multiple systems and organs throughout the body, influencing the patient outcomes, with interstitial lung disease (ILD) emerging as a significant comorbidity. A meta-analysis conducted by Sun in 2021 revealed that among individuals globally diagnosed with PM/ DM, the incidence of ILD is reported to reach as high as 41%, with an even more pronounced occurrence in Asian populations (3).

Some individuals with IIM-ILD may develop rapidly progressive ILD within one to two months of symptom onset, and this condition often proves resistant to immunosuppressive treatment, presenting considerable challenges in clinical management (4, 5). Notably, a study has revealed that within a few months after diagnosis of IIM, ILD accounted for the mortality of 71% of CADM patients and 60% of individuals with primary DM (6). Although ILD findings may manifest before, during, or after the diagnosis of IIM, the clinical course remains variable. Therefore, early assessment of the unknown risk of IIM-ILD progression is essential to optimise patient management.

High-resolution computed tomography (HRCT) plays a crucial and integral role in the identification and description of alterations in the lung's interstitial space (7, 8). However, the inconsistencies among various observers and even within the same observer make HRCT lack reliable and quantitative evaluation for ILD diagnosis (9). To address this challenge and enhance the accuracy of predicting IIM-ILD, a more objective and dependable approach to assessment is required.

Quantitative lung densitometry is a method that employs computer algorithms to provide a quantitative assessment of lung tissue density. It utilises HRCT images to evaluate lung diseases based on density values, usually represented in Hounsfield units (HU). This approach allows for the quantification of density changes in lung tissue, thereby assisting doctors in detecting and diagnosing lung diseases, particularly ILD. Through quantitative lung densitometry, clinicians can achieve more precise assessments of lung lesions, providing improved guidance for clinical treatment (10-13).

Currently, there is a lack of relevant research on quantitative lung densitometry in forecasting the progression of ILD in individuals with IIM. Hence, this study strives to investigate predictive markers for ILD progression among patients with IIM, leveraging baseline quantitative lung densitometry alongside pertinent clinical data.

Methods

Study subjects

and diagnostic criteria

Between May 2018 and December 2022, we gathered comprehensive data from a cohort of 126 patients diagnosed with IIM by utilising the electronic medical records system of the Second Affiliated Hospital of Anhui Medical University. All enrolled patients had complete medical records, chest HRCT images acquired at the time of initial diagnosis, and follow-up records (with the final follow-up time point set as March 30, 2023). The time duration commencing from the initial diagnosis of IIM and extending until either ILD progression, or the conclusion of the final follow-up period was designated as the "survival time". ILD progression was identified when the follow-up period showed the presence of two or more of the following conditions: worsening respiratory symptoms (e.g. dyspnoea upon exertion), decreased lung parenchymal opacity observed on chest HRCT, a decline of more than 10% in forced vital capacity (FVC), or a drop of more than 10 mmHg in arterial oxygen tension (PaO_2).

This study obtained endorsement from the Ethics Committee of the Second Affiliated Hospital of Anhui Medical University and strictly adhered to ethical standards. All participants provided written informed consent.

Competing interests: none declared.



Fig. 1. Density plot.

Quantitative lung densitometry analysis

To ensure accuracy, all chest HRCT scans were carefully examined for potential complications, including pleural effusions, oedema, haemorrhages, and bacterial or viral infections. Any scan with such complications was excluded from the final analysis. Fortunately, no examination was deemed ineligible in this series. Importantly, all CT examinations were performed without the use of contrast enhancement.

In this study, we conducted pulmonary parenchymal analysis using the Infer-Read CT Lung software provided by InferVision (Fig. 1). The software automatically segmented the lung images and analysed the following parameters: total lung volume (TLV; -1024 HU to -250 HU), normal lung density (NLD; -950 HU to -700 HU), high attenuation areas (HAA; -700 HU to -250 HU), and mean lung attenuation (MLA). Normal lung index (NLI) and HAA% represent the proportions of TLV occupied by NLD and HAA, respectively (Fig. 2). These methods were based on literature reports (14, 15).

Data collection

The patients' clinical baseline data were extracted from their initial electronic medical records and follow-up visits (Table I).





Fig. 2. Quantitative lung densitometry variables.

Statistical analysis

We conducted the statistical analysis using SPSS 23.0 software. Quantitative data were expressed as $\bar{x} \pm s$, while categorical data were displayed as frequency and percentage. To compare the advanced ILD group with the nonadvanced ILD group, we employed the t-test or Mann-Whitney U-test for continuous data, and the χ^2 test or Fisher's exact test for categorical data.

To identify meaningful predictors of ILD progression, we employed receiver operating characteristic (ROC) analysis along with the Youden's index to determine the optimal cut-off values and groupings. COX univariate and multivariate regression analyses were utilized to identify independent predictors of ILD progression in patients with IIM. A *p*-value of<0.05 was considered significant.

Results

Among the 126 IIM patients, 35 were males, average age was 53.1 ± 14.6 years, ranging from 13 to 88 years. Based on diagnostic criteria, 93 (73.81%), 26 (20.63%), and 7 (5.56%) cases were classified under DM, PM, and CADM groups, respectively. The patients were monitored for an average of 23.5 months, with a range of 0.5 to 56 months.

Comparison of ILD progressors and non-ILD progressors

Table I provides the variables' comparison between the ILD progressors and non-ILD progressors. Out of 126 patients, 37 (29.4%) experienced ILD deterioration, the median age was 58.5 years. It is worth noting that variables such as male sex, height, weight, body mass index (BMI), smoking status, IIM diagnosis, procalcitonin (PCT), interleukin (IL)-6, Krebs von den Lungen-6 (KL-6), rheumatoid factors (RF), creatine kinase (CK), aspartate aminotransferase (AST), alanine aminotransferase (ALT), triglyceride (TG), anti-cyclic citrullinated peptide antibody (anti-CCP), positron emission tomography-computed tomography (PET-CT) score, PCO₂, total lung capacity (TLC), diffusing lung capacity (DLCO), DLCO% predicted, and forced expiratory volume in one second (FEV1) did not exhibit statistically significant differences between ILD progressors and non-ILD progressors. However, older age, ILD, elevated Creactive protein (CRP), erythrocyte sedimentation rate (ESR), peak ferritin and carcino-embryonic antigen (CEA), higher A-aDO₂, higher FEV1/FVC ratio, and complications such as CMV infection and EB infection were associated with ILD progressors. Moreover, progressors had lower FVC and PO2 values than non-progressors. With respect to the utilisation of myositisspecific antibodies (MSAs) and Muscle-specific autoantibodies (MAAs), anti-glycyl tRNA synthetase (anti-EJ), anti-melanoma differentiation-associated gene 5 (anti-MDA5), and Anti-Ro52 were more significant for ILD progressors, while Anti-histidyl-tRNA synthetase (anti-Jo-1) and Anti-transcription intermediary factor-gamma

Table I. Baseline characteristics of patients with IIM.

Variables	ILD progressors (n=37)		Non-ILD progressors (n=89)		р
	Value	Available data per outcome	Value	Available data per outcome	
Demographics					
Age at onset, years	58.49±12.18	37 (100%)	50.82±15.01	89 (100%)	0.007
Male	14 (37.84%)	37 (100%)	21 (23.60%)	89 (100%)	0.104
ILD	37 (100%)	37 (100%)	63 (70.79%)	89 (100%)	<0.001
height, cm	159.85±8.13	34 (91.9%)	160.82±8.38	88 (98.9%)	0.566
weight, kg	54.50 (51.00, 62.00)	34 (91.9%)	58.00 (50.00,64.00)	88 (98.9%)	0.459
BMI, kg/m ²	22.00 ± 2.64	34 (91.9%)	22.41 ± 3.01	88 (98.9%)	0.482
CMN	9(24.32%)	37(100%) 21(82.78%)	12 (13.48%) 1 (1.02%)	89 (100%) 52 (58 42%)	0.112
FR	(3.25%) 6 (19.35%)	31 (83.78%) 31 (83.78%)	1 (1.92%) 7 (14.29%)	32 (38.43%) 49 (55.06%)	0.009
PIP	3 (100%)	3 (8.11%)	3 (100%)	3 (3.37%)	0.24
Diagnosis	(),				
PM	6 (16.22%)	37 (100%)	20 (22.47%)	89 (100%)	0.429
DM	29 (78.38%)	37 (100%)	64 (71.91%)	89 (100%)	0.452
CADM	2 (5.41%)	37 (100%)	5 (5.62%)	89 (100%)	0.664
Laboratory parameters	10.50 (0.85.25.60)	27 (100% 2.20)	1.00 (10.50)	00 (100%)	0.001
CRP, mg/dL PCT_ng/ml	12.50 (2.85,35.60) 0.06 (0.03.0,11)	37 (100%, 3.20) 26 (70.3%, 0.06)	1.20 (10.50)	89 (100%)	0.001
II_6_pg/ml	(0.00, (0.03, 0.11))	20 (70.3%,0.00) 27 (73.0% 18.30)	6.25 (69.10)	41 (461%)	0.594
ESR mm/h	29.00 (14.75.48.25)	36 (97.3% 18.50)	7.00 (33.25)	86 (96.6%)	0.008
KL-6, U/mL	887.00 (479.00,1635.00)	27 (73.0%,624.00)	364.00 (1167.00)	39 (43.8%)	0.094
RF (IU/ml)	16.25 (10.55,336.45)	6 (16.2%,24.55)	15.25 (34.80)	12 (13.5%)	0.820
Ferritin, ng/mL	756.25 (275.75,1432.00)	36 (97.3%,220.00)	115.00 (448.00)	77 (86.5%)	<0.001
CK, U/L	388.00 (73.50,993.00)	37 (100%,345.00)	67.00 (1584.00)	89 (100%)	0.533
AST, U/L	60.00 (33.50,147.00)	37 (100%,54.00)	31.50 (118.50)	89 (100%)	0.541
ALI, U/L	54.00 (30.50,84.00)	37 (100%,49.00)	28.00 (85.50)	89 (100%)	0.750
IG (mmol/L)	1.52 (1.14, 2.03) 3.52 (2.08, 6.00)	34 (91.9%) 25 (04.6%)	1.36) (1.00,1.99) 1.67) 0.04.2.50)	87 (97.8%) 78 (87.6%)	0.237
anti-CCP (RU/ml)	9.20 (3.43,18,93)	24 (64.9%)	5.55) 2.78 11 18)	56 (62.9%)	0.250
PO., L mmHg	73.40 (63.10.84.93)	32 (86.5%)	86.80) 75.70.102.80)	51 (57.3%)	0.004
PCO ₂	35.12±5.35	30 (81.1%)	36.64±5.80	51 (57.3%)	0.246
A-aDO,	36.25 (25.78,48.73)	30 (81.1%)	23.35) 7.63,31.43)	50) 56.2%)	0.004
PET-CT score	19.50 (8.50,30.75)	28 (75.7%)	28.00) 7.50,40.00)	57) 64.0%)	0.203
TIC	3.50(2.833.92)	8 (21.6%)	4.09 (3.22.4.86)	32(360%)	0 197
FVC%predicted %	69.76 (56.38.80.89)	33 (89.2%)	77.08 (69.23.94.59)	52 (50.070)	0.011
DLCO%predicted, %	46.50 (39.62,85.16)	9 (24.3%)	61.47 (42.46,76.28)	32 (36.0%)	0.297
FVC	2.07±0.70	33 (89.2%)	2.39±0.72	60 (67.4%)	0.045
DLCO	3.59 (3.04,6.50)	9 (24.3%)	5.19 (3.33,6.36)	32 (36.0%)	0.410
FEV1	1.71 (1.39,2.18)	32 (86.5%)	1.87 (1.52,2.39)	59 (66.3%)	0.125
FEV1/FVC	86.06±7.72	32 (86.5%)	82.97±6.58	59 (66.3%)	0.047
MSA					
Anti-SRP	0 (0%)	36 (97.3%)	4 (4.65%)	86 (96.6%)	0.242
Anti-Jo-I	5 (13.89%)	36 (97.3%)	26(30.23%)	86 (96.6%)	0.045
Anti PL-/	0 (10.07%) 3 (8.33%)	36 (97.3%)	3(3.81%)	86 (96.6%)	0.003
Anti-EI	7 (19.44%)	36 (97.3%)	4 (465%)	86 (96.6%)	0.015
Anti-OJ	1 (2.78%)	36 (97.3%)	2(2.33%)	86 (96.6%)	0.653
Anti-SAE1	0 (0%)	36 (97.3%)	1 (1.16%)	86 (96.6%)	0.705
Anti-SAE2	0 (0%)	36 (97.3%)	1 (1.16%)	86 (96.6%)	0.705
Anti-Mi-2	2 (5.56%)	36 (97.3%)	4 (4.65%)	86 (96.6%)	0.574
Anti-TIFy	2 (5.56%)	36 (97.3%)	17 (19.77%)	86 (96.6%)	0.038
Anti-MDA5	15 (41.67%)	36 (97.3%)	(12.79%)	86 (96.6%)	0.001
Anti-NXP2 Anti Ku	1 (2.78%)	36 (97.3%) 26 (07.2%)	2(2.33%)	86 (96.6%) 86 (06.6%)	0.053
ΜΔΔ	0 (0%)	50 (97.5%)	4 (4.05%)	80 (90.0%)	0.242
Anti-PMSc175	2 (5 56%)	36 (97.3%)	3 (3.49%)	86 (96.6%)	0 463
Anti-PMSc1100	3 (8.33%)	36 (97.3%)	1 (1.16%)	86 (96.6%)	0.077
Anti-Ro52	28 (77.78%)	36 (97.3%)	47 (54.65%)	86 (96.6%)	0.013
Quantitative Lung Densit	ometry				
ŤLV	2513.64 (1941.35,3332.00)	37 (100%)	2904.57 (2173.37,3723.04)	89 (100%)	0.109
NLD	1489.03 (1074.66,2185.62)	37 (100%)	1929.13 (1397.56,2513.66)	89 (100%)	0.018
NLI, %	59.43±8.78	37 (100%)	65.86±10.27	89 (100%)	0.001
HAA	645.85±177.27	37 (100%)	513.35 ± 173.00	89 (100%)	<0.001
HAA, %	25.36 (20.13,34.60)	3/(100%)	17.49 (11.07,28.63) 750.00 (805.24,606.62)	89 (100%) 80 (100%)	0.002
Type of ILD	- 099.03(-742.43,-047.63	57 (100%)	-139.90 (-003.24,-090.03)	69 (100%)	<0.001
NSIP	27 (72.97%)	37 (100%)	49 (55.06%)	89 (100%)	0.061
OP	2 (5.41%)	37 (100%)	5 (5.62%)	89 (100%)	0.664
NSIP/OP	3 (8.11%)	37 (100%)	4 (4.49%)	89 (100%)	0.336
UIP	3 (8.11%)	37 (100%)	6 (6.74%)	89 (100%)	0.524
AIP	2 (5.41%)	37 (100%)	0 (0%)	89 (100%)	0.085

Results are presented as mean \pm standard deviation, median (interquartile range), and frequency (percentage). T-test or Mann-Whitney U-test for continuous data, and the χ^2 test or Fisher's exact test for categorical data.

or Fisher's exact test for categorical data. ILD: interstitial lung; disease BMI: body mass index; CMV: cytomegalovirus; EB: Epstein-Barr virus; PJP: pneumocystis Jiroveci pneumonia; PM: polymyositis; DM: dermatomyositis; CADM: clinically amyopathic dermatomyositis; CRP: C-reactive protein; PCT: procalcitonin; IL-6: interleukin-6; ESR: erythrocyte sedimentation rate; KL-6: Krebs von den Lungen-6; RF: rheumatoid factor; CK: creatine kinase; AST: aspartate aminotransferase; ALT: alanine aminotransferase; TG: triglyceride; CEA: carcinoembryonic antigen; CCP: cyclic citrullinated peptide; PET-CT: positron emission tomography-computed Timography; TLC: total lung capacity; FVC: forced vital capacity; DLCO: diffusing lung capacity; FEV1: forced expiratory volume in one second; MSA: myositis-specific antibody; MAA: myositis-associated autoantibody; TLV: total lung volume; NLD: normal lung density; NLI: normal lung index; HAA: high attenuation areas; MLA: mean lung attenuation; NSIP: non-specific interstitial pneumonia; OP: organising pneumonia; UIP: usual interstitial pneumonia; AIP: acute interstitial pneumonia. (anti-TIF1 γ) were more significant for non-ILD progressors.

Quantitative lung densitometry findings

Except for TLV, all quantitative lung densitometry variables significantly differed between the groups, especially HAA and MLA. The HAA was significantly higher in progressors (645.9cm³±177.3cm³) than in nonprogressors $(513.4 \text{ cm}^3 \pm 173.00 \text{ cm}^3),$ (p < 0.001). Similarly, the MLA was higher in progressors [-699.0HU IQR: (-742.5HU - -647.6HU)] than in nonprogressors [-759.9HU IQR: (-805.2HU -.696.6], (p<0.001). Moreover, progressors had a higher HAA% [25.4% IQR: (20.1%-34.6%)] than non-progressors [17.5% IOR: (11.1%-28.6%)], (p=0.002).The NLD [1489.0cm³ $(1074.7 \text{ cm}^3 - 2185.6 \text{ cm}^3)$ IOR: and NLI 59.4% (8.8%) of progressors were lower than those of non-progressors [1929.1cm3 IQR: (1397.6cm3 -2513.7 cm³)] (*p*=0.018) and 65.9% (10.3%) (p=0.001), respectively.

Predictors of ILD progression in IIM patients

As potential risk factors of ILD progression in individuals with IIM, we selected 15 factors for analysis, including ILD, CRP, CEA, PO₂, A-aDO₂, FVC, FEV1/FVC, NLD, NLI, HAA, HAA%, MLA, Anti-Jo-1, Anti-EJ, Anti-MDA5, Anti-Ro52, and CMV/EB infection. Patients were divided into two groups using optimal cut-off values for NLD, NLI, HAA, HAA%, and MLA, respectively. Each index underwent COX univariate analysis, which identified several significant factors, including ILD, CEA, PO2, HAA (>465.745 cm³), HAA% (>20.835%), MLA (> -744.29 HU), anti-EJ, Anti-MDA5, and Anti-Ro52.Among these factors, HAA (>465.745 cm³), HAA% (>20.835%), and MLA (>-744.29 HU) are all quantitative lung densitometry analysis indices. A correlation analysis revealed significant associations among these three indices. Consequently, they were subjected to multivariate analysis along with other relevant single-factor analysis indices. The multivariate analysis results revealed that HAA (>465.745

Variables	Univariate analysis		Multivariate analysis	
	HR (95% CI)	<i>p</i> -value	HR (95% CI)	<i>p</i> -value
ILD	0.032 (0.001~0.819)	0.037		
CRP, mg/dL	1.005 (0.998~1.012)	0.173		
CEA, ng/ml	1.036 (1.004~1.069)	0.027		
PO ₂	0.980 (0.962~0.997)	0.025		
A-aDO ₂	1.005 (0.997~1.013)	0.236		
FVC	0.551 (0.320~0.946)	0.031		
FEV1/FVC	1.050 (0.997~1.105)	0.064		
Anti-Jo-1	2.559 (0.994~6.588)	0.052		
Anti-EJ	0.399 (0.175~0.912)	0.029		
Anti-MDA5	0.239 (0.122~0.468)	<0.001	0.127 (0.041~0.396)	<0.001
Anti-Ro52	0.437 (0.199~0.960)	0.039		
CMV	0.776 (0.106~5.696)	0.803		
EB	0.631 (0.259v1.541)	0.312		
HAA (>465.745cm ³)	5.007 (1.773~14.144)	0.002	4.758 (1.009~22.440)	0.049

Univariate and multivariate Cox regression were used to identify predictors of disease outcomes.



Fig. 3. The AUC for HAA (>465.745 cm³), anti-MDA5 positivity, and the combined diagnosis using both.

cm³) and positive Anti-MDA5 status emerged as independent risk factors associated with ILD progression in IIM individuals (Table II). Intriguingly, the risk of ILD progression was prominently amplified, showing a remarkable 7.874-fold increase in patients who exhibited positive anti-MDA5 results compared to their negative counterparts. HAA (>465.745cm³) were positively correlated with ILD progression. The combination of HAA (>465.745 cm³) and anti-MDA5 positivity was employed for diagnosing ILD progression in IIM patients, and the ROC curve analysis yielded an AUC of 0.745. This AUC value was higher than that of either HAA (>465.745 cm³) or Anti-MDA5 positivity alone (Fig. 3).

Discussion

In recent years, the utilisation of MSAs as autoantibodies exclusively found in IIM individuals has garnered significant attention due to their association with specific clinical symptoms, disease progression, and treatment response (16, 17). These antibodies have become pivotal in the clinical diagnosis and prognostic assessment of IIM, particularly with regard to anti-ARS and anti-MDA5, which have been identified as playing a critical role in IIM-ILD.

Remarkably, MDA5 has emerged as a pivotal prognostic marker, particularly concerning the onset of RP-ILD in individuals grappling with DM-ILD (18-20). In our study, we reaffirmed the predictive efficacy of anti-MDA5 positivity. The outcomes of the multivariate Cox regression analysis clearly revealed that patients with a positive anti-MDA5 status were significantly more inclined to experience the progression of ILD (p<0.001). However, it is essential to acknowledge that despite the robust predictive potential of anti-MDA5 positivity, a significant portion of patients with this marker may still undergo a stable trajectory of ILD over an extended duration. To avoid excessive clinical intervention due to overemphasis on anti-MDA5, this study integrated a comprehensive analysis of multiple baseline clinical data to enhance diagnostic efficiency and maximise clinical benefits by enabling joint prediction.

After conducting a retrospective analysis of initial features of 126 individuals with IIM and their relationship to ILD progression, the study identified HAA (>465.745cm³) and anti-MDA5 positivity as independent prognostic risk factors for ILD progression in IIM patients (p=0.049, p=0.001, respectively). The combination of HAA (>465.745 cm³) and anti-MDA5 positivity significantly enhances the diagnostic efficiency for ILD progression in individuals with IIM.

This study represents a pioneering investigation into the use of quantitative lung densitometry and clinical data for predicting ILD progression in patients with IIM. The advantage of quantitative lung densitometry over CT evaluation conducted by radiologists lies in its quantitative nature and operatorindependence, making it easily applicable to standard clinical CT scans. The study incorporated six quantitative CT indexes (QIs), and in the final multiplefactor COX regression analysis, HAA (>465.745cm³) emerged as an independent prognostic risk factor of ILD progression. The study revealed that IIM patients with a HAA (>465.745 cm³) had a 4.758 times higher risk of ILD progression compared to those with a HAA ($\leq 465.745 \text{ cm}^3$) (p=0.049). HAA, a crucial parameter in quantitative lung densitometry, serves as a quantitative measure of abnormal lung density distribution, aiding in the diagnosis and monitoring of pulmonary conditions. In this study, HAA encompasses lung regions with attenuation densities exceeding -700HU, which, in the chest CT scans of our study cohort, primarily correspond to ground glass opacities, linear high-density opacities, and thickening of interlobular septa. From a histopathological perspective, HAA is regarded as a consequence of subclinical alveolar epithelial injury, pulmonary inflammation, extracellular matrix remodelling, and interstitial fibrosis, making it a potential surrogate density marker for subclinical interstitial lung disease (12).

Elevated HAA levels have shown significant associations with increased serum levels of MMP-7 and IL-6, alongside diminished FVC, reduced exercise capacity, and heightened ILD-related mortality (21). These research findings underscore a robust correlation between elevated inflammatory markers, clinical events linked to ILD progression, and higher HAA levels. Prior investigations have indicated that HAA could serve as a valuable marker for detecting interstitial lung abnormalities (ILA), carrying substantial implications for the timely identification and effective management of RA-ILD (22). Furthermore, some studies have suggested that even modest elevations in HAA are linked to a heightened risk of clinical ILD events, further emphasising the clinical relevance of HAA (23).

This study breaks new ground by introducing quantitative lung densitometry analysis as an indicator for forecasting ILD advancement in individuals with IIM. Importantly, it conclusively establishes that HAA (>465.745cm³) stands as an independent predictor of ILD progression. HAA is considered indicative of latent lung injury, subclinical pulmonary inflammation, and possible initial extracellular matrix remodelling (21). Given that HAA may reflect early pathological changes in ILD, potentially representing structural alveolar alterations preceding clinically relevant ILD, the observation of elevated HAA levels in this study may effectively signal the likelihood of ILD progression in IIM patients. Consequently, it is believed that HAA can serve as an indicator of disease progression in ILA or ILD, holding significant clinical implications.

The utilisation of quantitative lung densitometry analysis allows radiologists to automatically obtain HAA values based on CT scans, facilitating the evaluation of lesions that may not be easily quantifiable by visual examination and aiding in the prediction of disease progression. Combining these research findings, it was observed that IIM patients with HAA (>465.745cm³) are more susceptible to experiencing ILD progression. In addition to the findings related to quantitative lung densitometry, our analysis of clinical data also revealed important associations with ILD progression in patients with IIM. Notably, our observations highlighted a significant association between advanced age and an increased likelihood of ILD progression in individuals with IIM (p=0.007). Moreover, individuals with CMV or EB virus infections demonstrated a heightened risk of ILD progression as well (p=0.009, p=0.007, respectively).

IIM-ILD can be classified into different types based on CT presentations, including non-specific interstitial pneumonia (NSIP), organising pneumonia (OP), NSIP/OP overlap, usual interstitial pneumonia (UIP), and acute interstitial pneumonia (AIP). It is generally acknowledged that the first three ILD patterns are relatively more common in IIM-ILD patients, which aligns with the distribution observed in our cases. The majority of our cases indeed conform to the NSIP pattern.

Prior research has suggested a potential association between disease progression and the OP pattern (24), while other studies have not found a significant correlation between ILD patterns and disease progression (25). In our study, we conducted a comparative analysis of different ILD patterns and disease progression and did not identify any significant correlations.

Furthermore, a comparison between patient groups, in line with the outcomes of the univariate Cox regression analysis, highlighted that those with pre-existing ILD at the time of initial diagnosis showed an elevated susceptibility to undergo ILD progression over the course of the follow-up (p=0.037). These comparisons of clinical data offer valuable insights into potential risk factors linked to ILD progression in IIM patients, further underscoring the significance of a comprehensive assessment in forecasting disease outcomes.

Our study encompassed a range of laboratory examination parameters, including routine biochemical markers, inflammatory factors, and autoantibody indicators. These indicators have consistently demonstrated diverse degrees of relevance with the diagnosis, treatment, and prognosis of IIM or ILD in both earlier studies and clinical practice (25-29). Consequently, we included these factors in our investigation to explore their potential correlation with ILD progression in individuals with IIM.

Prior investigations have highlighted that heightened CEA levels are frequently detected in individuals with CADM and could potentially serve as an indicator of RP-ILD, a condition associated with an adverse prognosis (30). Similarly, research has shown that elevated CEA levels are autonomously associated with an increased risk of RP-ILD in patients diagnosed with anti-synthetase syndrome (ASS) (31). Therefore, we included CEA as a predictive parameter in our study. The outcome of the Cox univariate regression analysis indicated that CEA (HR=1.036, 95% CI 1.004-1.069) emerged as a notable contributor to the progression of ILD (p=0.027) in individuals with IIM. These findings indicate that elevated CEA levels suggest an elevated risk of ILD progression in individuals with IIM, underscoring the importance of early detection and intervention.

Regarding arterial blood gas analysis for ILD patients, we focused on whether the main manifestation was hypoxemia with a decrease in arterial partial pressure of carbon dioxide and an increased alveolar-arterial oxygen pressure difference. Thus, we included these three indicators to explore their predictive ability in ILD progression in individuals with IIM. The findings from the Cox univariate analysis indicated that an increase in PO₂ was a protective factor for ILD progression (p=0.025), which is consistent with our clinical practice. Furthermore, pulmonary function tests (PFTs) play a vital role in examining potential IIM-ILD patients, primarily used to detect the presence of restrictive ventilation impairment and decreased diffusion function, aiding in disease screening and progression evaluation. Our study's COX univariate analysis findings suggested that FVC serves as a safeguard against ILD progression (p=0.031). These results provide valuable insights into the potential factors influencing ILD progression in IIM patients and underscore the significance of monitoring these indicators in clinical practice.

Among patients with IIM-ILD, anti-ARS antibodies were the most frequently identified. Numerous independent studies have consistently shown that anti-ARS-positive patients are markedly more susceptible to developing ILD than their negative counterparts. Moreover, this ILD is known to typically follow a chronic clinical course (32-34).

Regarding anti-EJ, although it belongs to the anti-ARS category, it is relatively uncommon, and there is limited research on its link to ILD progression. Li et al. conducted a study in 2019 and found that ILD was more frequent among patients with anti-EJ antibodies (OR 14.202, 95% CI 1.696-118.902), and the OR value was higher than that for other anti-ARS antibodies. However, that study did not specifically investigate the prediction of ILD progression (28). Our study aimed to investigate the link between different anti-ARS antibodies and predicting ILD progression. The outcomes of the univariate Cox regression analysis in our investigation unveiled that anti-EJ stood out as a noteworthy risk factor for ILD progression (p=0.029).

MAAs can be detected in various connective tissue diseases, although their specificity for IIM is relatively low (35). Among these, Anti-Ro52 is the most prevalent MAA and is frequently coexistent with MSAs (36). In our study, the findings from the univariate Cox regression analysis highlighted Anti-Ro52 as a notable risk factor for ILD progression (p=0.039). Additionally, prior studies have shown that patients positive for anti-MDA5 or anti-Jo1 antibodies often have concurrent positivity for anti-Ro52 antibodies. In anti-Ro52-positive patients, particularly when also positive for anti-MDA5 antibodies, the risk of RP-ILD development increases, leading to poorer prognostic outcomes (37, 38).

This study aimed to differentiate baseline clinical and imaging characteristics between IIM patients with and without ILD progression and predict ILD progression in IIM patients. However, several potential limitations should be acknowledged.

Firstly, we did not exclude the influence of drug treatments for the disease, which may have affected the accuracy of our predictions. Medications and their dosages could potentially impact disease progression and outcomes, and not accounting for these factors might have introduced some confounding effects.

Secondly, our study did not stratify ILD progression into different risk levels for more in-depth prediction. Such stratification could offer more precise and tailored clinical guidance based on individual patient risk profiles.

Thirdly, the CT scans were conducted using different models of CT machines, which were not standardised. This variability may have resulted in differences in HU values and could have affected the accuracy of our predictions.

Finally, it has been suggested that there may be a correlation between COV-ID-19 and certain forms of IIMs. The widespread dissemination of the novel coronavirus in recent years may have implications for research on this condition (39).

Despite these limitations, our study has significant implications. Unlike previous observational studies that mainly focused on patients already diagnosed with IIM-ILD, which may have reflected late-stage disease and the impact of ILD rather than its underlying causes, our study included IIM patients with different ILD statuses, allowing for predictions of progression from non-existing to existing ILD, as well as from existing to worsened ILD. This approach provided a more objective and accurate prediction of ILD progression in the overall IIM population.

Moreover, our study utilised quantitative lung densitometry and clinical baseline data for prediction, making the process more objective and less reliant on subjective interpretations. This approach enhances the robustness of our findings and contributes to a more accurate assessment of ILD progression in individuals with IIM.

Conclusion

In conclusion, among IIM patients, those who are anti-MDA5-positive and exhibit HAA (>465.745cm³) are more likely to experience ILD progression. Importantly, the combined predictive efficacy of these two factors is even higher, suggesting their potential utility as valuable indicators for identifying individuals at risk of ILD progression in this patient population.

References

- LUNDBERG IE, DE VISSER M, WERTH VP: Classification of myositis. Nat Rev Rheumatol 2018; 14(5): 269-78. https://doi.org/10.1038/nrrheum.2018.41
- 2. DALAKAS MC, HOHLFELD R: Polymyositis and dermatomyositis. *Lancet* 2003; 362(9388): 971-82. https:// doi.org/10.1016/S0140-6736(03)14368-1
- SUN KY, FAN Y, WANG YX, ZHONG YJ, WANG GF: Prevalence of interstitial lung disease in polymyositis and dermatomyositis: A metaanalysis from 2000 to 2020. *Semin Arthritis Rheum* 2021; 51(1): 175-91. https:// doi.org/10.1016/j.semarthrit.2020.11.009
- KAMEDA H, NAGASAWA H, OGAWA H et al.: Combination therapy with corticosteroids, cyclosporin A, and intravenous pulse cyclophosphamide for acute/subacute interstitial pneumonia in patients with dermatomyositis. J Rheumatol 2005; 32(9): 1719-26.
- BETTERIDGE Z, MCHUGH N: Myositisspecific autoantibodies: an important tool to support diagnosis of myositis. J Intern Med 2016; 280(1): 8-23. https://doi.org/10.1111/joim.12451

6. YAMASAKI Y, YAMADA H, OHKUBO M et al.: Longterm survival and associated risk factors in patients with adult-onset idiopathic inflammatory myopathies and amyopathic dermatomyositis: experience in a single institute in Japan. J Rheumatol 2011; 38(8): 1636-43. https://doi.org/10.3899/jrheum.101002

 ZOU J, GUO Q, CHI J, WU H, BAO C: HRCT score and serum ferritin level are factors associated to the 1-year mortality of acute interstitial lung disease in clinically amyopathic dermatomyositis patients. *Clin Rheumatol* 2015; 34(4): 707-14.

https://doi.org/10.1007/s10067-015-2866-5

- ZUO Y, YE L, LIU M et al.: Clinical significance of radiological patterns of HRCT and their association with macrophage activation in dermatomyositis. *Rheumatology* (Oxford) 2020; 59(10): 2829-37. https:// doi.org/10.1093/rheumatology/keaa034
- COLLINS CD, WELLS AU, HANSELL DM et al.: Observer variation in pattern type and extent of disease in fibrosing alveolitis on thin section computed tomography and chest radiography. Clin Radiol 1994; 49(4): 236-40. https:// doi.org/10.1016/s0009-9260(05)81847-1
- XU Y, VAN BEEK EJ, HWANJO Y, GUO J, MCLENNAN G, HOFFMAN EA: Computeraided classification of interstitial lung diseases via MDCT: 3D adaptive multiple feature method (3D AMFM). Acad Radiol 2006; 13(8): 969-78.

https://doi.org/10.1016/j.acra.2006.04.017 11. CAMICIOTTOLI G. ORLANDI I. BARTOLUCCI

- M *et al.*: Lung CT densitometry in systemic sclerosis: correlation with lung function, exercise testing, and quality of life. *Chest* 2007; 131(3): 672-681. https://doi.org/10.1378/chest.06-1401
- LEDERER DJ, ENRIGHT PL, KAWUT SM et al.: Cigarette smoking is associated with subclinical parenchymal lung disease: the Multi-Ethnic Study of Atherosclerosis (MESA)lung study. Am J Respir Crit Care Med 2009; 180(5): 407-14. https:// doi.org/10.1164/rccm.200812-1966OC
- HOFFMAN EA, JIANG R, BAUMHAUER H et al.: Reproducibility and validity of lung density measures from cardiac CT Scans -The Multi-Ethnic Study of Atherosclerosis (MESA) Lung Study. Acad Radiol 2009; 16(6): 689-99.
- https://doi.org/10.1016/j.acra.2008.12.024 14. HOCHHEGGER B, PELAEZ A, MACHUCA T *et al.*: CT imaging findings in lung transplant recipients with COVID-19. *Eur Radiol* 2023; 33(3): 2089-95.
- https://doi.org/10.1007/s00330-022-09148-w 15. BARROS MC, HOCHHEGGER B, ALTMAYER S
- et al.: The normal lung index from quantitative computed tomography for the evaluation of obstructive and restrictive lung disease. J Thorac Imaging 2022; 37(4): 246-52. https:// doi.org/10.1097/rti.000000000000629
- 16. MARIAMPILLAI K, GRANGER B, AMELIN D et al.: Development of a new classification system for idiopathic inflammatory myopathies based on clinical manifestations and myositis-specific autoantibodies. JAMA Neurol 2018; 75(12): 1528-37. https:// doi.org/10.1001/jamaneurol.2018.2598
- BETTERIDGE Z, TANSLEY S, SHADDICK G et al.: Frequency, mutual exclusivity and clinical associations of myositis autoantibodies in a combined European cohort of idiopathic inflammatory myopathy patients. J Autoimmun 2019; 101: 48-55. https://doi.org/10.1016/j.jaut.2019.04.001
- MOGHADAM-KIA S, ODDIS CV, SATO S, KUWANA M, AGGARWAL R: Anti-melanoma differentiation-associated gene 5 is associated with rapidly progressive lung disease

and poor survival in US patients with AMY-OPATHIC and myopathic dermatomyositis. *Arthritis Care Res* (Hoboken) 2016; 68(5): 689-94. https://doi.org/10.1002/acr.22728

- 19. CHEN Z, CAO M, PLANA MN *et al.*: Utility of anti-melanoma differentiation-associated gene 5 antibody measurement in identifying patients with dermatomyositis and a high risk for developing rapidly progressive interstitial lung disease: a review of the literature and a meta-analysis. *Arthritis Care Res* (Hoboken) 2013; 65(8): 1316-24. https://doi.org/10.1002/acr.21985
- 20. KOGA T, FUJIKAWA K, HORAI Y *et al.*: The diagnostic utility of anti-melanoma differentiation-associated gene 5 antibody testing for predicting the prognosis of Japanese patients with DM. *Rheumatology* (Oxford) 2012; 51(7): 1278-84.
- https://doi.org/10.1093/rheumatology/ker518 21. PODOLANCZUK AJ, OELSNER EC, BARR RG *et al.*: High attenuation areas on chest computed tomography in community-dwelling adults: the MESA study. *Eur Respir J* 2016; 48(5): 1442-52. https:// doi.org/10.1183/13993003.00129-2016
- ALEVIZOS MK, DANOFF SK, PAPPAS DA et al.: Assessing predictors of rheumatoid arthritis-associated interstitial lung disease using quantitative lung densitometry. *Rheumatology* (Oxford) 2022; 61(7): 2792-804. https:// doi.org/10.1093/rheumatology/keab828
- 23. PODOLANCZUK AJ, OELSNER EC, BARR RG et al.: High-attenuation areas on chest computed tomography and clinical respiratory outcomes in community-dwelling adults. Am J Respir Crit Care Med 2017; 196(11): 1434-42.
- https://doi.org/10.1164/rccm.201703-0555OC 24. SAMBATARO D, SAMBATARO G, PIGNATARO F et al.: Patients with interstitial lung disease secondary to autoimmune diseases: how to recognize them? *Diagnostics* (Basel) 2020; 10(4): 208.

https://doi.org/10.3390/diagnostics10040208

25. SAMBATARO G, SAMBATARO D, SPICUZZA L *et al.*: Progression and prognosis of interstitial pneumonia with autoimmune features: a longitudinal, prospective, multi-centre study. *Clin Exp Rheumatol* 2023; 41(5): 1140-8. https://

doi.org/10.55563/clinexprheumatol/lycdca

- 26. KELLY CA, SARAVANAN V, NISAR M et al.: Rheumatoid arthritis-related interstitial lung disease: associations, prognostic factors and physiological and radiological characteristicsa large multicentre UK study. *Rheumatology* (Oxford) 2014; 53(9): 1676-82. https:// doi.org/10.1093/rheumatology/keu165
- 27. ZHENG M, LOU A, ZHANG H, ZHU S, YANG M, LAI W: Serum KL-6, CA19-9, CA125 and CEA are diagnostic biomarkers for rheumatoid arthritis-associated interstitial lung disease in the Chinese population. *Rheumatol Ther* 2021; 8(1): 517-27. https://doi.org/10.1007/s40744-021-00288-x
- 28. LI S, GE Y, YANG H et al.: The spectrum and clinical significance of myositis-specific autoantibodies in Chinese patients with idiopathic inflammatory myopathies. Clin Rheumatol 2019; 38(8): 2171-9. https://doi.org/10.1007/s10067-019-04503-7

29. JABLONSKI R, BHORADE S, STREK ME, DE-MATTE J: Recognition and management of myositis-associated rapidly progressive interstitial lung disease. *Chest* 2020; 158(1): 252-63.

https://doi.org/10.1016/j.chest.2020.01.033

- 30. ZHU D, QIAO J, TANG S et al.: Elevated carcinoembryonic antigen predicts rapidly progressive interstitial lung disease in clinically amyopathic dermatomyositis. *Rheumatology* (Oxford) 2021; 60(8): 3896-903. https:// doi.org/10.1093/rheumatology/keaa819
- 31. ZUO Y, YE L, CHEN F et al.: Different multivariable risk factors for rapid progressive interstitial lung disease in anti-MDA5 positive dermatomyositis and anti-synthetase syndrome. Front Immunol 2022; 13: 845988. https://doi.org/10.3389/fimmu.2022.845988
- 32. FUKAMATSU H, HIRAI Y, MIYAKE T et al.: Clinical manifestations of skin, lung and muscle diseases in dermatomyositis positive for anti-aminoacyl tRNA synthetase an-

tibodies. J Dermatol 2019; 46(10): 886-97. https://doi.org/10.1111/1346-8138.15049

- 33. YAMASAKI Y, YAMADA H, NOZAKI T et al.: Unusually high frequency of autoantibodies to PL-7 associated with milder muscle disease in Japanese patients with polymyositis/ dermatomyositis. Arthritis Rheum 2006; 54(6): 2004-9.
- https://doi.org/10.1002/art.21883
- 34. SATO S, MURAKAMI A, KUWAJIMA A et al.: Clinical utility of an enzyme-linked immunosorbent assay for detecting anti-melanoma differentiation-associated gene 5 autoantibodies. PLoS One 2016; 11(4): e0154285. https:// https://
 - doi.org/10.1371/journal.pone.0154285
- BETTERIDGE Z, MCHUGH N: Myositisspecific autoantibodies: an important tool to support diagnosis of myositis. *J Intern Med* 2016; 280(1): 8-23.
- https://doi.org/10.1111/joim.12451 36. SCHULTE-PELKUM J, FRITZLER M, MAHLER

M: Latest update on the Ro/SS-A autoantibody system. *Autoimmun Rev* 2009; 8(7): 632-7.

https://doi.org/10.1016/j.autrev.2009.02.010 37. GUI X, SHENYUN S, DING H *et al.*: Anti-Ro52 antibodies are associated with the prognosis of adult idiopathic inflammatory myopathyassociated interstitial lung disease. *Rheumatology* (Oxford) 2022; 61(11): 4570-8. https://

doi.org/10.1093/rheumatology/keac090

- DOURADO E, BOTTAZZI F, CARDELLI C et al.: Idiopathic inflammatory myopathies: one year in review 2022. Clin Exp Rheumatol 2023; 41(2): 199-213. https:// doi.org/10.55563/clinexprheumatol/jof6qn
- CAVAGNA L, FERRO F, ZANFRAMUNDO G, LA ROCCA G, PUXEDDU I: Idiopathic inflammatory myopathies and COVID-19: an intriguing liaison? *Clin Exp Rheumatol* 2023; 41(2): 217-20. https://

doi.org/10.55563/clinexprheumatol/njaff0