

Angiogenesis in vasculitides

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This study was supported by the Associazione Italiana per la Ricerca sul Cancro (AIRC, National and Regional Funds) Milan, Ministry for Education, the Universities and Research (Project CARSO n. 72/2; FIRB 2001 and PRIN 2005), Rome, and Fondazione Italiana per la Lotta al Neuroblastoma, Genoa, Italy.

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Received on June 18, 2007; accepted in revised form on October 2, 2007.

Clin Exp Rheumatol 2008; 26; 476-483.

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Key words: Angiogenesis, inflammation, vasculitides.

ABSTRACT

Vasculitides, including Wegener's granulomatosis, Takayasu's arteritis, giant cell arteritis, Kawasaki disease, Behçet disease, thromboangiitis obliterans and erythema elevatum diutinum, are inflammatory diseases of blood vessel wall characterized by myointimal proliferation, fibrosis and thrombus formation leading to stenosis or occlusion of the vascular lumen, and finally to tissue ischemia. In these diseases the hypoxic environment subsequent to stenosis or occlusion of the vascular lumen is a potent signal for the generation of new blood vessels. Angiogenesis may be a compensatory response to ischemia and to the increased metabolic activity and may be also a further inflammatory stimulus because endothelial cells of newly-formed vessels express adhesion molecules and produce colony-stimulating factors and chemokines for leukocytes.

Angiogenesis

Angiogenesis is the formation of newly-formed capillaries from pre-existing vessels. It is characterized by a well-programmed cascade of events which contains a number of distinct steps (1). Angiogenic factors activate endothelial cells, inducing them to produce, in turn, proteolytic enzymes such as matrix metalloproteinases and plasminogen activators, which are responsible of the degradation of the basement membrane and of the perivascular extracellular matrix. The proliferation and migration of endothelial cells into the perivascular area and the subsequent lumenation of these "primary sprouts" forming "capillary loops" and the synthesis of new basement membrane, lead finally to new vessel formation. Endothelial cells of the "primary sprouts" proliferate and migrate to generate secondary and further generations of sprouts. Angiogenesis is regulated by several

angiogenic and anti-angiogenic factors (2) (Table I). Vascular endothelial growth factor (VEGF) family members, fibroblast growth factor (FGF) family members, platelet-derived growth factor (PDGF), transforming growth factor alpha and beta (TGF- α and β), tumor necrosis factor alpha (TNF- α), interleukins (IL), chemokines, angiogenin, and angiopoietins have a positive regulatory activity. On the contrary, angiostatin, endostatin, and thrombospondin have a negative regulatory activity (3-5). The net balance between these positive and negative factors, with a prevalence of positive regulators, or a downregulation of the expression of negative regulators, is responsible for inducing and regulating the angiogenic process (2) (Fig. 1).

Angiogenesis in chronic inflammation

Angiogenesis plays a key role in the pathogenic events leading to chronic inflammation. Chronic inflammatory lesions occurring in several diseases, such as rheumatoid arthritis, Crohn's disease, ulcerative colitis, primary biliary cirrhosis, and vasculitides (6-10) are characterized by cellular infiltrates and newly-formed vessels involved in favouring inflammatory cells recruitment and providing a compensatory response to ischemia and to the increased metabolic activity (11, 12).

Angiogenic factors induce endothelial cells to express adhesion molecules, cytokines and chemokines, which may have additive stimulatory effects on chronic inflammation. VEGF promotes the migration of inflammatory cells, such as monocytes and lymphocytes, into the extracellular matrix via inducing vascular permeability and endothelial cells expression of adhesion molecules, such as VCAM-1 and ICAM-1 (13, 14). FGF-1 and FGF-2 promote endothelial cell production of

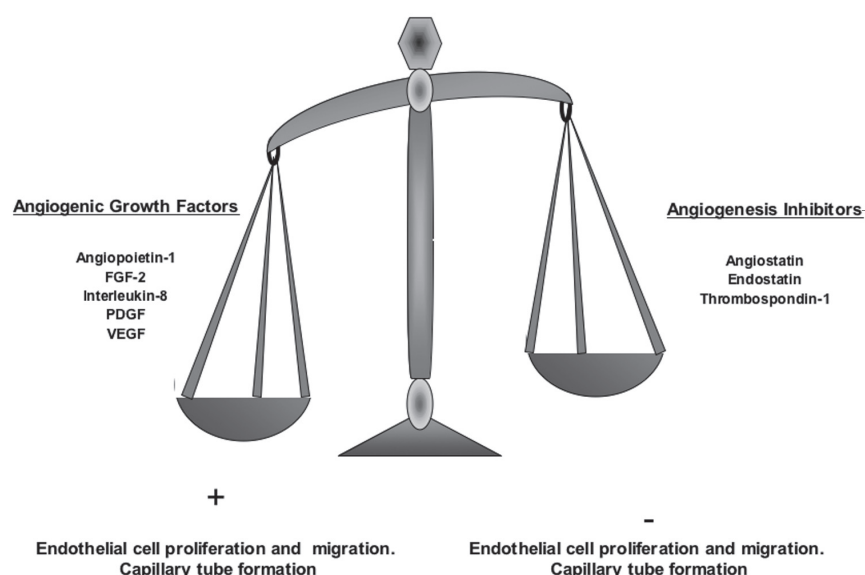
Competing interests: none declared.

Table I. Main angiogenic and anti-angiogenic factors that regulate angiogenesis.

Angiogenic factors*	Angiogenic activity			
	<i>In vitro</i> assays			<i>In vivo</i> assays
	Proliferation	Migration	Capillary tube formation	
Vascular endothelial growth factor (VEGF)	S	S	S	S
Fibroblast growth factor-2 (FGF-2)	S	S	S	S
Platelet derived growth factor (PDGF)	N	S	=	S
Transforming growth factors beta (TGF- β)	I	N	S	S
Angiopoietin-1	N	S	S	S
Anti-angiogenic factors*				
Thrombospondin-1	I	I	I	I
Angiostatin	I	I	I	I
Endostatin	I	I	I	I

*All the angiogenic and the anti-angiogenic factors may be considered as pleiotropic growth factors, which recognize several producing and effector cells, such as endothelial cells, pericytes, inflammatory cells and tumor cells.

I: inhibition; N: no effect; S: stimulation; =: no findings available.

**Fig. 1.** Angiogenesis results from the balance between pro- and anti-angiogenic factors.

plasminogen activator and collagenase which allow the migration of inflammatory cells via degradation of the extracellular matrix (15). Other angiogenic factors involved in inflammatory cells recruitment are chemokines containing the ELR motif (glutamyl-leucyl-arginyl sequence), such as CXC chemokines (16, 17).

Most angiogenic factors, such as TNF- α , IL-1, IL-6, IL-8, and IL-18 are also inflammatory factors involved in increasing the production of other inflammatory cytokines and cell adhesion molecules, and in enhancing matrix metallo-proteinases and/or cyclooxygenase activity (18).

Vasculitides

Vasculitides are a heterogeneous group of disorders including giant cell arteritis, Takayasu's arteritis, Cogan's syndrome, Behçet disease, polyarteritis nodosa, thromboangiitis obliterans, Kawasaki disease, primary angitis of the central nervous system, Goodpasture's disease, cutaneous leukocytoclastic angitis, Henoch-Schönlein purpura, hypocomplementemic urticarial vasculitis, essential cryoglobulinemia, erythema elevatum diutinum, Wegener's granulomatosis, microscopic polyangiitis, Churg-Strauss syndrome, renal-limited vasculitis and secondary forms of vasculitis (19).

At the present time, the Chapell Hill Consensus Conference Nomenclature of primary vasculitides (Table II) provide a useful guide to clinician and pathologist for evaluating a patient with a idiopathic form of vasculitis. This classification is based on the predominant size of vessels affected and describes the main clinico-pathologic features of the various clearly defined types of systemic vasculitis.

Vasculitides are also classified on the basis of their histopathologic features into neutrophilic or leukocytoclastic vasculitis, lymphocytic vasculitis, and granulomatous vasculitis (20). Leukocytoclastic vasculitis is an inflammation of small-vessel characterized by segmental angiocentric neutrophilic inflammation, endothelial cell damage and fibrinoid necrosis. These histopathologic features may be found in cryoglobulinemia and rarely in multiple myeloma (21). Lymphocytic vasculitis, characterized by lymphocytic recruitment of inflammatory cells and cytotoxic reactions, is the histopathologic pattern of lymphocytic endovascularitis, lymphocytic lichenoid vasculitis and granulomatous vasculitis (22). Granulomatous vasculitis is a granulomatous inflammation with extensive necrosis and a variegated cellular infiltrate. Wegener's granulomatosis is an example of this histopathologic pattern (23).

These disorders are characterized by inflammation of blood vessel wall. The final result is myointimal proliferation, fibrosis and thrombus formation leading to stenosis or occlusion of the vascular lumen, and finally to tissue ischemia (24). Even if aetiology of vasculitides is still unknown, infections, inflammatory disorders, autoimmune disorders and drugs are involved as triggers for disease activity in vasculitides (25-27).

Because the etiologies of most from of vasculitis remain unknown, the most valid basis for classifying the vasculitides is the size of the predominant blood vessels involved. Large vessel vasculitis involves the aorta and its major branches; medium vessel vasculitis involves vessels large enough to be observed in gross pathological specimens or visualized by angiography; small

Table II. Names and definitions of vasculitides adopted by the Chapel Hill Consensus Conference.

Giant cell arteritis

Granulomatosis arteritis of the aorta and its major branches, with a predilection for the extracranial branches of the carotid artery. Often involves the temporal artery.

Takayasu's arteritis

Granulomatosis arteritis of the aorta and its major branches.

Polyarteritis nodosa

Necrotizing inflammation of the medium-sized or small arteries.

Kawasaki's disease

Arteritis involving large, medium sized and small arteries. Coronary arteries are often involved.

Wegener's granulomatosis

Necrotizing vasculitis affecting small to medium-sized vessels.

Churg-Strauss syndrome

Necrotizing vasculitis affecting small to medium-sized vessels.

Microscopic polyangitis

Necrotizing vasculitis affecting small vessels.

Henoch-Schönlein purpura

Vasculitis affecting small vessels.

Essential cryoglobulinemia vasculitis

Vasculitis affecting small vessels

Cutaneous leukocytoclastic vasculitis

Isolated cutaneous leukocytoclastic angitis without systemic vasculitis.

vessel vasculitis involves capillaries, post-capillary venules and arterioles.

Several studies have demonstrated that angiogenesis is involved in the pathogenesis of vasculitides, such as Wegener's granulomatosis, Takayasu's arteritis, giant cell arteritis, Kawasaki disease, Behçet disease, thromboangiitis obliterans and erythema elevatum diutinum. The angiogenic response is more intense in small vessel vasculitis, as compared to medium- and large-vessels vasculitis, because angiogenesis generally involves capillary and post-capillary venules. The hypoxic environment subsequent to stenosis or occlusion of the vascular lumen is a potent signal for the generation of new blood vessels.

A dual role of angiogenesis in vasculitides has been proposed (9). On the one hand, angiogenesis may be a compensatory response to ischemia and to the increased metabolic activity above all in strong acute phase of disease. On the

other hand, endothelial cells of newly-formed vessels express adhesion molecules and produce colony-stimulating factors and chemokines for leukocytes (28-32). In this way, circulating leukocytes arrive to sites of inflammation where constitute a further inflammatory stimulus (32, 33). Antineutrophil cytoplasmic antibodies (ANCA) potentiate neutrophil and monocyte mediated endothelial cell activation (34).

Angiogenesis in giant cell arteritis

Giant cell arteritis is a vasculitis that mainly affects extracranial medium-sized and large arteries, aorta and its principal branches (35, 36). The aorta typically demonstrates aortic root dilatation, with medial dissection in occasional cases of aortic rupture. The intima is wrinkled demonstrating a tree-back appearance. Histologically, the inflammatory infiltrate consists of lymphocytes, plasma cells and histiocytes. There is disruption of the internal elastic lamina with fragmentation and a giant cell reaction. The disruption may be accompanied by a significant degree of necrosis. Vessel occlusion occurring in giant cell arteritis causes ischemia. Cid *et al.* (9) have hypothesized that angiogenesis has an essential role in reducing ischemic tissue damage above all in giant cell arteritis target organs (37), such as small arteries supplying the optic nerve, and potentiates inflammation via expression of adhesion molecules and production of colony-stimulating factors and chemokines for leukocytes (28-32). Angiogenesis has been associated with severe inflammation in giant cell arteritis (32). On the other hand, severe inflammation, manifested by very high levels of erythrocyte sedimentation rate greater than 100 mm/1st hour or chronic anemia, has been associated with protection against visual ischemic manifestations or other severe ischemic manifestations of this vasculitis (38-40).

VEGF, FGF-2, TGF- β , PDGF, TNF- α , monocyte chemoattractant protein-1 (MCP-1) and IL-8 have been observed in giant cell arteritis lesions (41, 42). Moreover, high levels of VEGF, FGF-2 and soluble ICAM-1 have been found in sera of giant cell arteritis patients

(43). TNF- α and IL-6 levels are higher in giant cell arteritis acute phase (44), when haptoglobin, a carrier for free hemoglobin with angiogenic properties, is highly produced (45).

Constitutive (PECAM-1, ICAM-1, ICAM-2, and P-selectin) and inducible (E-selectin and VCAM-1) endothelial cell adhesion molecules for leukocytes are over-expressed by endothelial cells of adventitial microvessels and neovessels in giant cell arteritis vascular lesions. Leukocytes interactions with these ligands are responsible for the formation of inflammatory infiltrates in giant cell arteritis lesions (32). In adventitia, CD4⁺ T cells produce interferon- γ (IFN- γ). IFN- γ is correlated with multinucleated giant cells (MGCs) formation and intimal hyperplasia (46). MGCs are specialized fused cells derived by macrophages that accumulate in media-intima of arterial wall where are involved in VEGF production. VEGF and IFN- γ , in addition to tissue hypoxia subsequent to arterial wall thickening and luminal stenosis, are probably responsible of the angiogenesis in giant cell arteritis lesions (47). Blindness, stroke, and jaw claudication are often seen in giant cell arteritis patients with high levels of IFN- γ in their lesions (46). On the contrary, systemic manifestations in giant cell arteritis patients are well correlated with low levels of IFN- γ (47). Interestingly, a microsatellite dinucleotide (CA) repeat polymorphism in the first intron of the IFN- γ gene was associated with some differences between biopsy-proven giant cell arteritis, with or without visual ischemic manifestations. In this regard, an association between a high IFN- γ producer allele with giant cell arteritis patients who experienced visual ischemic events, and an inverse correlation with individuals carrying a low IFN- γ producer allele was found (48). Also, a functional VEGF-634 G $\rightarrow$ C gene polymorphism was found to be associated with severe ischemic complications in giant cell arteritis patients from Northwestern Spain (49). The low VEGF producer VEGF-634 G allele of this genetic polymorphism was significantly overexpressed in giant cell arteritis

patients with ischemic complications and additionally, a higher risk of developing severe ischemic complications was observed for VEGF-634 GG homozygous individuals (49).

MGCs are also involved in elastic membranes degradation via matrix metalloproteinase (MMP)-2 production (50-52). Moreover, MGCs produce PDGF-AA and PDGF-BB, both involved in intimal proliferation (53).

Giant cell arteritis is characterized by the release, in the systemic acute-phase, of proinflammatory cytokines, such as IL-1, TNF- α , and IL-6 (54), which may influence vascular responses such as vessel occlusion or regeneration, involved in the pathogenesis of giant cell arteritis inflammatory lesions (28, 55, 56). IL-6-induced angiogenesis is responsible of inflammation in giant cell arteritis and patients with elevated IL-6 levels needed higher corticosteroid dosage to limit their inflammatory activity (44, 57, 58).

Decorin, a chondroitin/dermatan sulfate proteoglycan of the extracellular matrix (59), has been found in giant cell arteritis lesions (60). Even if no factors responsible for the induction of decorin production have been still found, it is known that endothelial cells of newly-formed vessels in giant cell arteritis temporal artery may produce decorin. Decorin probably interacts with type I collagen (61, 62) and fibronectin (63, 64), two extracellular matrix proteins, and may induce endothelial cells proliferation by modifying the structural organization of these molecules (65, 66). It was found that when cultured endothelial cells spontaneously change their morphology from a polygonal shape to a sprouting phenotype they concomitantly initiate decorin synthesis and deposition, indicating that decorin is associated with angiogenesis (67). Decorin, a ligand for the epidermal growth factor receptor (EGF-R) (68), that is involved in endothelial cells proliferation, inhibits apoptosis of endothelial cells (69), induces collagenase expression (70) and phospholipase A2 activity (71), both involved in angiogenesis (72-74), inhibits TGF-beta (75, 76) and induces FGF-2 activity (77). Nelimarkka *et al.*

(78) have demonstrated that the intimal thickening and angiogenesis are coupled in giant cell arteritis and that the capillary neovessels within the inflamed temporal artery wall contain decorin.

Angiogenesis in Kawasaki disease

Kawasaki disease is a vasculitis of young childhood, characterized by aneurysmal dilatation and acute thrombosis of coronary arteries (79-83). Kawasaki disease lesions are characterized by thinning of vascular media, inflammation and destruction of the extracellular matrix in the internal elastic lamina and in the trilaminar structure of the vascular wall (79, 80, 83, 84). Fibrosis, granulation and angiogenesis have been seen in these lesions (79, 80, 83-85). Increased VEGF levels have been found in Kawasaki disease and TGF- β 1 has been demonstrated to upregulate VEGF expression in acute phase of disease (86). Moreover, endothelial cells of newly-formed blood vessels in myointima and adventitia of Kawasaki disease coronary artery aneurysms produce high levels of MMP-2 (84). Moreover, Gavin *et al.* (84) have demonstrated that in Kawasaki disease coronary aneurysms, MMP-9 was also expressed, but its expression is not confined to aneurysmal arteries. Systemic arterial expression of MMP-9 in acute Kawasaki disease, even in absence of inflammatory changes in the vessel, suggests induction by a circulating factor, or by an infectious agent with tropism for arterial tissue. Moreover, the significantly elevated MMP-9 levels during acute phase of Kawasaki disease may reflect vascular remodeling or an inflammatory response to a microbial agent, suggesting a pathophysiological role for MMP-9 in coronary aneurysm formation (87).

Miura *et al.* (88) have demonstrated that endothelial cells of newly-formed blood vessels in coronary artery aneurysm in Kawasaki disease express E-selectin, involved in leukocytes adhesion on endothelial cells, and VCAM-1, important in leukocytes adhesion and extravasation into tissues. On the contrary, luminal endothelial cells of coronary arteries without aneurysms

do not express E-selectin and VCAM-1, such as endothelial cells of newly-formed blood vessels in polyarteritis nodosa and giant cell arteritis (29, 32). This particular feature may be due to the vasculitic process on luminal endothelial cells.

Angiogenesis in Behçet's disease

Behçet's disease is characterized by increased VEGF expression in oral aphthous lesions and in the ocular inflammation (89, 90) and by increased interleukin-8 levels in synovial fluids (91). Moreover, increased levels of VEGF and MCP-1 were detected in sera of Behçet disease patients (92).

Recombinant human interferon alpha-2a is effective in ocular Behçet's disease, leading to significant improvement of vision and complete remission of ocular vasculitis in the majority of patients (93). The mode of action of interferon may be explained by an anti-angiogenic effect, as it has been demonstrated since 1980, when interferon alpha was shown to inhibit endothelial cell migration in a dose-dependent manner (94).

Angiogenesis in thromboangiitis obliterans

Angiogenesis plays a crucial role in the inflammatory process of thromboangiitis obliterans. Vascular lesions are characterized by increased levels of TNF- α and ICAM-1, VCAM-1 and E-selectin are expressed on the endothelial cells of thromboangiitis obliterans newly-formed vessels contributing to leukocyte adhesion (95).

Angiogenesis in leukocytoclastic vasculitis

Erythema elevatum diutinum, a variation of leukocytoclastic vasculitis, is characterized by cutaneous lesions over the shins, buttocks and extensor surfaces of the knees, elbows, and finger joints (96). Macroscopically, erythema elevatum diutinum lesions in the acute inflammatory phase are raised erythematous plaques, resolving in fibrous nodules with storiform or concentric fibrosis (97). Microscopically, three stages are described: a first stage characterized by capillary proliferation and fibrinoid degeneration of vessel wall; a

second stage characterized by fibrosis, dermal aggregates of neutrophils and areas of granulation tissue; a third stage characterized by fibrosis of the dermis with small foci of persistent leukocytoclastic vasculitis.

Dermal dendrocytes (DD), present in the adventitial and reticular dermis, have phagocytic and antigen presenting function and may modulate the regulatory mechanisms of deposition of extracellular macromolecules in the dermis. Moreover, DD are involved in the tissue repair mechanisms, fibrogenesis and angiogenesis (98, 99). Pacheco *et al.* (100) have demonstrated that DD may play a role in inflammatory skin diseases and found that DD in erythema elevatum diutinum and leukocytoclastic vasculitis are increased in number, hypertrophic and show stellate morphology. Even if erythema elevatum diutinum was characterized by a higher number of newly-formed blood vessels than leukocytoclastic vasculitis, no statistically significant difference in the number of DD has been observed between these two pathological conditions (100).

Angiogenesis in lymphocytic vasculitis

Lymphocytic vasculitis can be secondary to treatment of rheumatoid arthritis and Crohn's disease with infliximab, etanercept, or adalimumab (26, 27). These drugs rarely cause autoimmune disorders, such as vasculitis and lupus, probably due to low TNF- α levels (101-106), which are unable to suppress autoreactive B and T cells responsible in turn of autoimmune reactions (107). Srivastava *et al.* (27) found an increase of VEGF and chemokine levels, especially RANTES, in serum of a patient with T-cell lymphocytic vasculitis secondary to treatment with etanercept, which worsened significantly with switch to infliximab. Moreover, angiogenic mediators, including TNF- α , IL-1 β , IL-15, IL-18, were found in the vasculitic lesions of the patient.

Further evidences of angiogenesis in vasculitides: haptoglobin

Haptoglobin, a carrier for free hemoglobin, has angiogenic properties and

is probably involved in endothelial cell differentiation and proliferation in systemic vasculitis, including Wegener's granulomatosis, Takayasu's arteritis and giant cell arteritis. Cid *et al.* (45) found a potent angiogenic activity in association with high levels of haptoglobin in sera from patients affected by Wegener's granulomatosis, Takayasu's arteritis and giant cell arteritis, using both *in vitro* and *in vivo* angiogenesis models. Haptoglobin levels are particularly high in Wegener's granulomatosis patients in the period of clinically active disease characterized by intense angiogenic activity.

Concluding remarks

In vasculitides, angiogenesis may be a compensatory response to ischemia and to the increased metabolic activity and may be also a further inflammatory stimulus because endothelial cells of newly-formed vessels express adhesion molecules and produce colony-stimulating factors and chemokines for leukocytes (28-33). Even if further studies are needed to elucidate the pathogenic mechanism of vasculitides, the emergency of angiogenesis as a key player in the pathogenic events leading to Wegener's granulomatosis, Takayasu's arteritis, giant cell arteritis, Kawasaki disease, Behçet's disease, thromboangiitis obliterans and erythema elevatum diutinum vasculitides, may provide a basis for a rational approach to the development of an anti-angiogenic therapy.

It is well established that the angiogenic phenotype results from the imbalance between positive and negative regulator factors, so that the contribution of each angiogenic factor may play a different role in the definition of the angiogenic phenotype in vasculitides, when increased production of angiogenic stimuli and/or reduced production of angiogenic inhibitors may lead to abnormal neovascularization.

The development of a clinical trial requires the identification and characterization of the physiological targets involved in angio-stimulatory and angio-inhibitory activities.

Angiogenesis inhibitors are now being approved and introduced into medical practice throughout the world. Much

research has been concentrated on the role of angiogenesis in cancer, and inhibition of angiogenesis is a major area of therapeutic development for the treatment of this disease. It is conceivable that this therapeutical approach might involve also the treatment of chronic inflammatory diseases, such as vasculitides.

References

1. RISAU W: Mechanisms of angiogenesis. *Nature* 1997; 386: 671-4.
2. PEPPER MS: Manipulating angiogenesis: from basic science to the bedside. *Arter Thromb Vasc Biol* 1997; 17: 605-19.
3. DAMERON KM, VOLPERT OV, TAINSKY MA, BOUCK N: Control of angiogenesis in fibroblasts by p53 regulation of thrombospondin-1. *Science* 1994; 265: 1582-4.
4. O'REILLY MS, HOLMGREN L, SHING Y *et al.*: Angiostatin: a novel angiogenesis inhibitor that mediates the suppression of metastases by a Lewis lung carcinoma. *Cell* 1994; 79: 315-28.
5. O'REILLY MS, BOEHM T, SHING Y *et al.*: Endostatin: an endogenous inhibitor of angiogenesis and tumor growth. *Cell* 1997; 88: 277-85.
6. FIOCCCHI C: Inflammatory bowel disease etiology and pathogenesis. *Gastroenterology* 1998; 115: 182-205.
7. BOUSVAROS A, LEICHTNER A, ZURAKOWSKI D *et al.*: Elevated serum vascular endothelial growth factor in children and young adults with Crohn's disease. *Dig Dis Sci* 1999; 44: 424-30.
8. LEE SS, JOO YS, KIM WU *et al.*: Vascular endothelial growth factor levels in the serum and synovial fluid of patients with rheumatoid arthritis. *Clin Exp Rheumatol* 2001; 19: 321-4.
9. CID MC, HERNÁNDEZ-RODRÍGUEZ J, ESTEBAN MJ *et al.*: Tissue and Serum angiogenic activity is associated with low prevalence of ischemic complications in patients with giant-cell arteritis. *Circulation* 2002; 106: 1664.
10. MEDINA J, SANZ-CAMENO P, GARCÍA-BUEY L, MARTÍN-VÍLCHEZ S, LÓPEZ-CABRERA M, MORENO-OTERO R: Evidence of angiogenesis in primary biliary cirrhosis: an immunohistochemical descriptive study. *J Hepatol* 2005; 42: 124-31.
11. BALLARA SC, MIOTLA JM, PALEOLOG EM: New vessels, new approaches: angiogenesis as a therapeutic target in musculoskeletal disorders. *Int J Exp Pathol* 1999; 80: 235-50.
12. BIAN XW, CHEN JH, JIANG XF, BAI JS, WANG QL, ZHANG X: Angiogenesis as an immunopharmacologic target in inflammation and cancer. *Int Immunopharmacol* 2004; 4: 1537-47.
13. CLAUS M, GERLACH M, GERLACH H *et al.*: Vascular permeability factor: a tumor-derived polypeptide that induces endothelial cell and monocyte procoagulant activity, and promotes monocyte migration.

- J Exp Med* 1990; 172: 1535-45.
14. MELDER RJ, KOENIG GC, WITWER BP, SAFABAKHSH N, MUNN LL, JAIN RK: During angiogenesis, vascular endothelial growth factor and basic fibroblast growth factor regulate natural killer cell adhesion to tumor endothelium. *Nat Med* 1996; 2: 992-7.
 15. FOLKMAN J, KLAGSBRUN M, SASSE J, WADZINSKI M, INGBER D, VLODAVSKY I: A heparin-binding angiogenic protein – basic fibroblast growth factor – is stored within basement membrane. *Am J Pathol* 1988; 130: 393-400.
 16. BUCKLEY CD, AMFT N, BRADFIELD PF *et al.*: Persistent induction of the chemokine receptor CXCR4 by TGF- β 1 on synovial T cells contributes to their accumulation within the rheumatoid synovium. *J Immunol* 2000; 165: 3423-9.
 17. BUCKLEY CD, PILLING D, LORD JM, AKBAR AN, SCHEEL-TOELLNER D, SALMON M: Fibroblasts regulate the switch from acute resolving to chronic persistent inflammation. *Trends Immunol* 2001; 22: 199-204.
 18. MARUOTTI N, CANTATORE FP, CRIVELLATO E, VACCA A, RIBATTI D: Macrophages in rheumatoid arthritis. *Histol Histopathol* 2007; 22: 581-6.
 19. SALEH A, STONE JH: Classification and diagnostic criteria in systemic vasculitis. *Best Pract Res Clin Rheumatol* 2005; 19: 209-21.
 20. COPEMAN PWM, RYAN TJ: The problems of classification of cutaneous angitis with reference to histopathology and pathogenesis. *Br J Dermatol* 1970; 82: 2-14.
 21. CEM AR M, SOYSAL T, HATEMI G, SALIHOGLU A, YAZICI H, ULKU B: Successful management of cryoglobulinemia-induced leukocytoclastic vasculitis with thalidomide in a patient with multiple myeloma. *Ann Hematol* 2005; 84: 609-13.
 22. KOSSARD S: Defining lymphocytic vasculitis. *Australas J Dermatol* 2000; 41: 149-55.
 23. HAMMAR SP: Granulomatous vasculitis. *Semin Respir Infect* 1995; 10: 107-20.
 24. LUQMARI RA, ROBINSON H: Introduction to, and classification of the systemic vasculitides. *Best Pract Res Clin Rheumatol* 2001; 15: 187-202.
 25. NIKKARI S, RELMAN A: Molecular approaches for identification of infectious agents in Wegener's granulomatosis. *Curr Opin Rheumatol* 1999; 11: 11-6.
 26. SRIVASTAVA MD: Immunomodulatory effects of etanercept (TNFR:Fc) and its use in a patient with Crohn's disease. *Res Commun Mol Pathol Pharmacol* 2001; 109: 125-41.
 27. SRIVASTAVA MD, ALEXANDER F, TUTHILL RJ: Immunology of cutaneous vasculitis associated with both etanercept and infliximab. *Scand J Immunol* 2005; 61: 329-36.
 28. MANTOVANI A, BUSSOLINO F, DEJANA E: Cytokine regulation of endothelial cell function. *FASEB J* 1992; 6: 2591-9.
 29. COLL-VINENT B, CEBRIAN M, CID MC *et al.*: Dynamic pattern of endothelial cell adhesion molecule expression in muscle and perineural vessels from patients with classic polyarteritis nodosa. *Arthritis Rheum* 1998; 41: 435-44.
 30. SZEKANECZ A, SZEGEDI G, KOCH AE: Angiogenesis in rheumatoid arthritis: pathogenic and clinical significance. *J Invest Med* 1998; 46: 27-41.
 31. CARMELIET P, JAIN RK: Angiogenesis in cancer and other diseases. *Nature* 2000; 407: 249-57.
 32. CID MC, CEBRIAN M, FONT C *et al.*: Cell adhesion molecules in the development of inflammatory infiltrates in giant cell arteritis: inflammation-induced angiogenesis as the preferential site of leukocyte/endothelial cell interactions. *Arthritis Rheum* 2000; 43: 184-94.
 33. CID MC, ESPARZA J, JUAN M *et al.*: Signaling through CD50 (ICAM-3) stimulates T lymphocyte binding to human umbilical vein endothelial cells and extracellular matrix proteins via an increase in β 1 and β 2 integrin function. *Eur J Immunol* 1994; 24: 1377-82.
 34. CID MC, SEGARRA M, GARCIA-MARTINEZ A, HERNANDEZ-RODRIGUEZ J: Endothelial cells, antineutrophil cytoplasmic antibodies, and cytokines in the pathogenesis of systemic vasculitis. *Curr Rheumatol Rep* 2004; 6: 184-94.
 35. NORDBORG E, NORDBORG C, MALMVALL BE, ANDERSSON R, BENGTSSON BA: Giant cell arteritis. *Rheum Dis Clin North Am* 1995; 21: 1013-26.
 36. HUNTER GG: Giant cell arteritis and polymyalgia rheumatica. *Med Clin North Am* 1997; 81: 195-219.
 37. ESTEBAN MJ, FONT C, HERNANDEZ-RODRIGUEZ J *et al.*: Small-vessel vasculitis surrounding a spared temporal artery: clinical and pathologic findings in a series of twenty-eight patients. *Arthritis Rheum* 2001; 44: 1387-95.
 38. SALVARANI C, CIMINO L, MACCHIONI P *et al.*: Risk factors for visual loss in an Italian population-based cohort of patients with giant cell arteritis. *Arthritis Rheum* 2005; 53: 293-7.
 39. GONZALES-GAY MA, GARCIA-PORRUA C, LLORCA J *et al.*: Visual manifestations of giant cell arteritis. Trends and clinical spectrum in 161 patients. *Medicine (Baltimore)* 2000; 79: 283-92.
 40. GONZALES-GAY MA, BARROS S, LOPEZ-DIAZ MJ, GARCIA-PORRUA C, SANCHEZ-ANDRADE A, LLORCA J: Giant cell arteritis: disease patterns of clinical presentation in a series of 240 patients. *Medicine (Baltimore)* 2005; 84: 269-76.
 41. WEYAND CM, HICOK KC, HUNTER GG, GORONZY JJ: Tissue cytokine patterns in patients with polymyalgia rheumatica and giant-cell arteritis. *Ann Intern Med* 1994; 121: 484-91.
 42. WEYAND CM, GORONZY JJ: Arterial wall injury in giant-cell arteritis. *Arthritis Rheum* 1999; 42: 844-53.
 43. COLL-VINENT B, VILARDELL C, FONT C *et al.*: Circulating soluble adhesion molecules in patients with giant-cell arteritis: correlation between soluble intercellular adhesion molecule-1 (SCAM-1) and disease activity. *Ann Rheum Dis* 1999; 58: 189-92.
 44. HERNANDEZ-RODRIGUEZ J, GARCIA-MARTINEZ A, CASADEMONT J *et al.*: A strong initial systemic inflammatory response is associated with higher corticosteroid requirements and longer duration of therapy in patients with giant-cell arteritis. *Arthritis Rheum* 2002; 47: 29-35.
 45. CID MC, GRANT DS, HOFFMAN GS, AUERBACH R, FAUCI AS, KLEINMAN HK: Identification of haptoglobin as an angiogenic factor in sera from patients with systemic vasculitis. *J Clin Invest* 1993; 91: 977-85.
 46. WEYAND CM, TETZLAFF N, BJORNSSON J, BRACK A, YOUNGE B, GORONZY JJ: Disease patterns and tissue cytokine profiles in giant cell arteritis. *Arthritis Rheum* 1997; 40: 19-26.
 47. KAISER M, YOUNGE B, BJORNSSON J, GORONZY JJ, WEYAND CM: Formation of new vasa vasorum in vasculitis. *Am J Pathol* 1999; 155: 765-74.
 48. GONZALES-GAY MA, HAJEER AH, DABABNEH A *et al.*: Interferon-gamma gene microsatellite polymorphisms in patients with biopsy-proven giant cell arteritis and isolated polymyalgia rheumatica. *Clin Exp Rheumatol* 2004; 22 (Suppl. 36): S18-20.
 49. RUEDA B, LOPEZ-NEVOT MA, LOPEZ-DIAZ MJ, GARCIA-PORRUA C, MARTIN J, GONZALES-GAY MA: A functional variant of vascular endothelial growth factor is associated with severe ischemic complications in giant cell arteritis. *J Rheumatol* 2005; 32: 1737-41.
 50. LIE JT: Histopathologic specificity of systemic vasculitis. *Rheum Dis Clin North Am* 1995; 21: 883-909.
 51. WEYAND CM, WAGNER AD, BJORNSSON J, GORONZY JJ: Correlation of the topographical arrangement and the functional pattern of tissue-infiltrating macrophages in giant cell arteritis. *J Clin Invest* 1996; 98: 1642-9.
 52. NIKKARI S, HOYHTYAM, ISOLAJ, NIKKARI T: Macrophages contain 92-kd gelatinase (MMP-9) at the site of degenerated internal elastic lamina in temporal arteritis. *Am J Pathol* 1996; 149: 1427-33.
 53. KAISER M, WEYAND CM, BJORNSSON J, GORONZY JJ: Platelet-derived growth factor, intimal hyperplasia, and ischemic complications in giant cell arteritis. *Arthritis Rheum* 1998; 41: 623-33.
 54. BAUMANN H, GAULDIE J: The acute phase response. *Immunol Today* 1994; 15: 74-80.
 55. ROSSI V, BREVIARIO F, GHEZZI P, DEJANA E, MANTOVANI A: Prostacyclin synthesis induced by in vascular cells by interleukin-1. *Science* 1985; 225: 174-6.
 56. MANTOVANI A, GARLANDA C, INTRONA M, VECCHI A: Regulation of endothelial cell function by pro- and anti-inflammatory cytokines. *Transplant Proc* 1998; 30: 4239-43.
 57. HERNANDEZ-RODRIGUEZ J, SEGARRA M, VILARDELL C *et al.*: Elevated production of interleukin-6 is associated with a lower incidence of disease-related ischemic events in patients with giant-cell arteritis. *Circulation* 2003; 107: 2428-34.
 58. HERNANDEZ-RODRIGUEZ J, SEGARRA M, VILARDELL C *et al.*: Tissue production of proinflammatory cytokines (IL-1 β , TNF- α , and IL-6) correlates with the intensity of the systemic inflammatory response and

- with corticosteroid requirements in giant-cell arteritis. *Rheumatology* 2004; 43: 294-301.
59. IOZZO RV: The family of the small leucine-rich proteoglycans: key regulators of matrix assembly and cellular growth. *Crit Rev Biochem Mol Biol* 1997; 32: 141-74.
 60. NELIMARKKA L, SALMINEN H, KUOPIO T *et al.*: Decorin is produced by capillary endothelial cells in inflammation-associated angiogenesis. *Am J Pathol* 2001; 158: 345-53.
 61. SCHÖNHERR E, WITSCH-PREHM P, HARRACH B, ROBENEK H, RAUTERBERG J, KRESSE H: Interaction of biglycan with type I collagen. *J Biol Chem* 1995; 270: 2776-83.
 62. SVENSSON L, HEINEGARD D, OLDBERG A: Decorin-binding sites for collagen type I are mainly located in leucine-rich repeats 4-5. *J Biol Chem* 1995; 270: 20712-6.
 63. LEWANDOWSKA K, CHOI HU, ROSENBERG LC, ZARDI L, CULP LA: Fibronectin-mediated adhesion of fibroblasts: inhibition by dermatan sulfate proteoglycan and evidence for a cryptic glycosaminoglycan-binding domain. *J Cell Biol* 1987; 105: 1443-54.
 64. SCHMIDT G, ROBENEK H, HARRACH B *et al.*: Interaction of small dermatan sulfate proteoglycan from fibroblasts with fibronectin. *J Cell Biol* 1987; 104: 1683-91.
 65. JACKSON CJ, JENKINS KL: Type I collagen fibrils promote rapid vascular tube formation upon contact with the apical side of cultured endothelium. *Exp Cell Res* 1991; 192: 319-23.
 66. VERNON RB, LARA SL, DRAKE CJ *et al.*: Organized type I collagen influences endothelial patterns during "spontaneous angiogenesis in vitro": planar cultures as models of vascular development. *In Vitro Cell Dev Biol Anim* 1995; 31: 120-31.
 67. JARVELAIMEN HT, IRUELA-ARISPE ML, KINSELLA MG, SANDELL LJ, SAGE EH, WIGHT TN: Expression of decorin by sprouting bovine aortic endothelial cells exhibiting angiogenesis *in vitro*. *Exp Cell Res* 1992; 203: 395-401.
 68. IOZZO RV, MOSCATELLO DK, MCQUILLAN DJ, EICHSTETTER I: Decorin is a biological ligand for the epidermal growth factor receptor. *J Biol Chem* 1999; 274: 4489-92.
 69. SCHÖNHERR E, O'CONNELL BC, SCHITTNY J *et al.*: Paracrine or virus-mediated induction of decorin expression by endothelial cells contributes to tube formation and prevention of apoptosis in collagen lattices. *Eur J Cell Biol* 1999; 78: 44-55.
 70. HUTTENLOCHER A, WERB Z, TREMBLE P, HUHTALA P, ROSENBERG L, DAMSKY CH: Decorin regulates collagenase gene expression in fibroblasts adhering to vitronectin. *Matrix Biol* 1996; 15: 239-50.
 71. SARTIPY P, JOHANSEN B, GASVIK K, HURT CAMEJO E: Molecular basis for the association of group IIA phospholipase A(2) and decorin in human atherosclerotic lesions. *Circ Res* 2000; 86: 707-14.
 72. JACKSON JR, BOLOGNESE B, MANGAR CA, HUBBARD WC, MARSHALL LA, WINKLER JD: The role of platelet activating factor and other lipid mediators in inflammatory angiogenesis. *Biochim Biophys Acta* 1998; 1392: 145-52.
 73. MACH F, SCHONBECK U, FABUNMI RP *et al.*: T lymphocytes induce endothelial cell matrix metalloproteinase expression by a CD40L-dependent mechanism: implications for tubule formation. *Am J Pathol* 1999; 154: 229-38.
 74. O'CONNOR DS, SCHECHNER JS, ADIDA C *et al.*: Control of apoptosis during angiogenesis by survivin expression in endothelial cells. *Am J Pathol* 2000; 156: 393-8.
 75. YAMAGUCHI Y, MANN DM, RUOSLAHTI E: Negative regulation of transforming growth factor-beta by the proteoglycan decorin. *Nature* 1990; 346: 281-4.
 76. HILDEBRAND A, ROMARIS M, RASMUSSEN LM *et al.*: Interaction of the small interstitial proteoglycans biglycan, decorin and fibromodulin with transforming growth factor beta. *Biochem J* 1994; 302: 527-34.
 77. PENC SF, POMAHAC B, WINKLER T *et al.*: Dermatan sulfate released after injury is a potent promoter of fibroblast growth factor-2 function. *J Biol Chem* 1998; 273: 28116-21.
 78. NELIMARKKA L, SALMINEN H, KUOPIO T *et al.*: Decorin is produced by capillary endothelial cells in inflammation-associated angiogenesis. *Am J Pathol* 2001; 158: 345-53.
 79. FUJIWARA H, HAMASHIMA Y: Pathology of the heart in Kawasaki disease. *Pediatrics* 1978; 61: 100-7.
 80. AMANO S, HAZAMA F, HAMASHIMA Y: Pathology of Kawasaki disease: pathology and morphogenesis of the vascular changes. *Jpn Circ J* 1979; 43: 633-43.
 81. KATO H, INOUE O, AKAGI T: Kawasaki disease: cardiac problems and management. *Pediatr Rev* 1988; 9: 209-17.
 82. ROWLEY AH, SHULMAN ST: Kawasaki syndrome. *Pediatr Clin North Am* 1999; 46: 313-29.
 83. NAOE S, TAKAHASHI K, MASUDA H, TANAKA N: Kawasaki disease with particular emphasis on arterial lesions. *Acta Pathol Jpn* 1991; 41: 785-97.
 84. GAVIN PJ, CRAWFORD SE, SHULMAN ST, GARCIA FL, ROWLEY AH: Systemic arterial expression of matrix metalloproteinases 2 and 9 in acute Kawasaki disease. *Arterioscler Thromb Vasc Biol* 2003; 23: 576-81.
 85. MITCHELL RN, COTRAN RS: Repair: cell regeneration, fibrosis, and wound healing. In KUMAR V, COTRAN RS, ROBBINS SL (Eds.) *Basic Pathology*. 6th ed. Philadelphia: WB Saunders, 1997: 47-59.
 86. TERAJI M, YASUKAWA K, NARUMOTO S, TATENO S, OANA S, KOHNO Y: Vascular endothelial growth factor in acute Kawasaki disease. *Am J Cardiol* 1999; 83: 337-9.
 87. CHUA PK, MELISH ME, YU Q, YANAGIHARA R, YAMAMOTO KS, NERURKAR VR: Elevated levels of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 during the acute phase of Kawasaki disease. *Clin Diagn Lab Immunol* 2003; 10: 308-314.
 88. MIURA M, GARCIA FL, CRAWFORD SE, ROWLEY AH: Cell adhesion molecule expression in coronary artery aneurysms in acute Kawasaki disease. *Pediatr Infect Dis J* 2004; 23: 931-6.
 89. NAME EJ, HAN SW, KIM SU *et al.*: Association of vascular endothelial growth factor gene polymorphisms with Behçet disease in a Korean population. *Hum Immunol* 2005; 66: 1068-73.
 90. YACIN B, ARDA N, TEZEL GG, ERMAN M, ALLI N: Expressions of vascular endothelial growth factor and CD34 in oral aphthous lesions of Behçet's disease. *Anal Quant Cytol Histol* 2006; 28: 303-6.
 91. ERDEM H, PAY S, SERDAR M *et al.*: Different ELR (+) angiogenic CXCL chemokine profiles in synovial fluid of patients with BD, familial Mediterranean fever, rheumatoid arthritis, and osteoarthritis. *Rheumatol Int* 2005; 26: 162-7.
 92. BOZOGLU E, DINC A, ERDEM H, PAY S, SIMSEK I, KOCAR IH: Vascular endothelial growth factor and monocyte chemoattractant protein-1 in BD patients with venous thrombosis. *Clin Exp Rheumatol* 2005; 23: S42-8.
 93. KÖTTER I, ZIERHUT M, ECKSTEIN AK *et al.*: Human recombinant interferon alfa-2a for the treatment of Behçet's disease with sight threatening posterior or panuveitis. *Br J Ophthalmol* 2003; 87: 423-31.
 94. BROUTY-BOYE D, ZETTER BR: Inhibition of cell motility by interferon. *Science* 1980; 208: 516-8.
 95. HALACHEVA K, GULUBOVA MV, MANOLOVA I, PETKOV D: Expression of ICAM-1, VCAM-1, E-selectin and TNF-alpha on the endothelium of femoral and iliac arteries in thromboangiitis obliterans. *Acta Histochem* 2002; 104: 177-84.
 96. KATZ SI: Inflammatory and neoplastic disorders of the dermis: Erythema elevatum diutinum. In FREEDBERG IM, EISEN AZ, WOLF K *et al.* (Eds.): *Fitzpatrick's Dermatology in General Medicine*, 5th ed. New York, McGraw-Hill 1993; 1: 1124.
 97. LE BOITPE, BENEDICTYEN TSB, WINTROUB B: The evolution of lesion in erythema elevatum diutinum. *Am J Dermatopathol* 1986; 8: 392.
 98. NICKOLOFF BJ, GRIFFITHS CEM: The spindle-shaped cells in cutaneous Kaposi's sarcoma. Histologic simulators include factor XIIIa dermal dendrocytes. *Am J Pathol* 1989; 135: 793-800.
 99. PENNEYS SP: Factor XIII expression in the skin: observations and a hypothesis. *J Am Acad Dermatol* 1990; 22: 484-8.
 100. PACHECO LS, SOTTO MN: Factor XIIIa+ dermal dendrocytes in erythema elevatum diutinum and ordinary cutaneous leukocytoclastic vasculitis lesions. *J Cutan Pathol* 2000; 27: 136-40.
 101. GALARIA NA, WERTH VP, SCHUMACHER HR: Leukocytoclastic vasculitis due to etanercept. *J Rheumatol* 2000; 27: 2041-4.
 102. MCCAIN ME, QUINET RJ, DAVIS WE: Etanercept and infliximab associated with cutaneous vasculitis. *Rheumatology* 2002; 41: 116-7.
 103. LIVERMORE PA, MURRAY KJ: Anti-tumor necrosis factor therapy associated with cutaneous vasculitis. *Rheumatology* 2002; 41: 1450-2.

104. SHAKOOR N, MICHALSKA M, HARRIS C, BLOCK J: Drug-induced systemic lupus erythematosus associated with etanercept therapy. *Lancet* 2003; 359: 579-80.
105. JARRETT SJ, CUNNANE G, CONAGHAN PG *et al.*: Antitumor necrosis factor-alpha therapy-induced vasculitis: case series. *J Rheumatol* 2003; 30: 2287-91.
106. MCILWAIN L, CARTER JD, BIN-SAGHEERS, VASEY FB, NORD J: Hypersensitivity vasculitis with leukocytoclastic vasculitis secondary to infliximab. *J Clin Gastroenterol* 2003; 36: 411-3.
107. SEO SJ, FIELDS ML, BUCKLER JL *et al.*: The impact of T helper and T regulatory cells on the regulation of anti-double stranded DNA B cells. *Immunity* 2002; 16: 535-46.