
Two types of myeloperoxidase-antineutrophil cytoplasmic autoantibodies with a high affinity and a low affinity in small vessel vasculitis

M. Yoshida, M. Sasaki, I. Nakabayashi, M. Akashi, T. Tomiyasu,
N. Yoshikawa, T. Kojima, N. Ohno¹, M. Yamada²

Renal Unit of Internal Medicine,
Hachioji Medical Center, Tokyo Medical
University, Tokyo; ¹Laboratory for
Immunopharmacology of Microbial
Products, School of Pharmacy, Tokyo
University of Pharmacy, Tokyo;
²International Graduate School of Arts
and Sciences, Yokohama City University,
Yokohama, Japan.

Masaharu Yoshida, MD, PhD
Mariko Sasaki, MA
Iwao Nakabayashi, MD, PhD
Masakazu Akashi, MD
Tomohiro Tomiyasu, MD
Noriko Yoshikawa, MD, PhD
Tadasu Kojima, MD
Naohito Ohno, PhD
Michiyuki Yamada, PhD

This study was funded by a grant from
the Research Committee of Intractable
Vasculitis and Multicenter Trial of
Treatment in ANCA Associated Vasculitis
of the Ministry of Labor and Welfare of
Japan.

Please address correspondence to:
Dr Masaharu Yoshida, MD, PhD,
Dept. of Renal Unit of Internal Medicine,
Hachioji Medical Center of Tokyo Medical
University, 1163 Tate-Machi,
Hachioji City, Tokyo, Japan.

E-mail: myoshida@tokyo-med.ac.jp

Received on June 6, 2008; accepted in
revised form on January 7, 2009.

Clin Exp Rheumatol 2009; 27 (Suppl. 52):
S28-S32.

© Copyright CLINICAL AND
EXPERIMENTAL RHEUMATOLOGY 2009.

Key words: Myeloperoxidase (MPO)-
antineutrophil cytoplasmic antibody
(ANCA), affinity of MPO-ANCA,
small vessel vasculitis, vasculitis
activity.

Competing interests: none declared.

ABSTRACT

Objective. Myeloperoxidase (MPO)
-anti-neutrophil cytoplasmic autoanti-
bodies (ANCAs) are detected at a high
rate in microscopic polyangiitis and re-
nal-limited vasculitis. MPO-ANCA tit-
ers are not always reflected in the dis-
ease activity. We studied the titer and
affinity of MPO-ANCA in sera from pa-
tients in relation to vasculitis activity.

Methods. Blood samples were collect-
ed from 27 newly diagnosed or relapsed
patients with MPO-ANCA-associated
vasculitides. The MPO-ANCA titer was
determined by a direct enzyme-linked
immunosorbent assay (ELISA) using
homogeneously purified human MPO
of leukocytes. The MPO-ANCA affini-
ty was expressed as IC₅₀ that was
determined by a competitive inhibition
method using the ELISA.

Results. The MPO-ANCA affinity of 27
sera from 27 patients could be classi-
fied into a high-affinity type (14 sera)
and a low-affinity type (13 sera). The
mean values for IC₅₀ in the two types
were 0.15±0.06 µg/ml and 0.54±0.15
µg/ml, and the difference was statisti-
cally significant ($p < 0.0000000684$).
Between the two groups of patients
divided by the affinity, there were dif-
ferences in the Birmingham Vasculitis
Activity Score (BVAS): and in C-reactive
protein (CRP): ($p < 0.00093$ and
 $p < 0.00129$, respectively). However, the
difference in titer was not statistically
significant ($p < 0.0265$). The affinity re-
mained steady from the disease onset
to remission or relapse.

Conclusions. The affinity of MPO-
ANCA from patients with MPO-ANCA-
associated vasculitides were largely
distinguished into a high and a low af-
finity, irrespective of the level of MPO-
ANCA titers, and may be helpful for as-
sessment of vasculitis activity affecting
mainly the kidney and the lung.

Introduction

Proteinase 3 (PR 3) - anti-neutrophil
cytoplasmic autoantibodies (ANCAs)
are frequently detected in Wegener's
granulomatosis, while myeloperoxi-
dase (MPO)-ANCAs are detected at a
high rate in microscopic polyangiitis
and renal-limited form of vasculitis
(RLV). These diseases are referred to as
ANCA-associated vasculitides (1). The
level of disease activity of these ANCA-
associated vasculitides and the ANCA
titer have been found to generally fluctu-
ate in close correlation (1). There are
patients with high MPO-ANCA titers
but almost no disease activity, as well as
patients with low or negative titers but
a high level of vasculitis activity (2-4).
However, the acute 3 or 4 fold increase
in PR-3 and MPO-ANCA titer is used
as an important index when monitoring
disease activity (5, 6). MPO-ANCA-as-
sociated vasculitides are more common
in Asian countries than in the USA or
Europe where 80% of ANCA associ-
ated vasculitis is PR-3 ANCA (7). In
Japan, MPO-ANCAs have been report-
ed to account for 90% of patients with
ANCA-associated vasculitides (8-10).
The relationships between the MPO-
ANCA titer and the affinity in patients
with small vessel vasculitis are not
comprehensively understood (11, 12).
Here we report the two types of MPO-
ANCA having a high affinity and a low
affinity in small vessel vasculitis.

Materials and methods

Between January 2005 and August
2008, blood samples were collected
from 27 patients with ANCA-associ-
ated vasculitides who could be peri-
odically followed up at Tokyo Medical
University Hachioji Medical Center.
These patients were either newly di-
agnosed or relapsed cases of ANCA-
associated vasculitides, and each had

given written informed consent to participation in the clinical study. The titer and affinity of the serum MPO-ANCA were determined prior to the start of therapy or at the time of relapse. The MPO-ANCA titer was determined by a standard procedure of a direct enzyme-linked immunosorbent assay (ELISA) (13) using a homogeneously purified human MPOIII of peripheral blood leukocytes (50 μ l of 1 μ g/ml) for coating solid phase surface (14). IC₅₀ was taken as an MPO-ANCA affinity. IC₅₀ was the concentration of MPO which produces 50% inhibition of MPO-ANCA binding at the dose response curve in the liquid phase of ELISA. Briefly, the MPO (1 μ g/ml) (14) was immobilized on the surface of microtiter wells at 50 μ l/well, and then 1% skimmed milk-phosphate buffered saline – 0.05% Tween 20 was added to each well as a blocking agent. Each of the serum samples was first suitably diluted to adjust its OD by the direct ELISA to about 1.0 (measured at 405 nm), and then each diluted serum sample (25 μ l/well) and serially diluted MPO (0-100 μ g/ml; 25 μ l/well) were simultaneously added to the MPO-coated plates and incubated for 1h at 37°C. Then, the bound IgG was detected using the system consisting of anti-human IgG alkaline phosphatase conjugate (1 h 37°C) and colour development at 405 nm with substrate (0.5 h 37°C). The competitive inhibition rate (%) was calculated by defining the inhibition at the highest MPO concentration (50 μ g/ml) as 100%. An approximation curve was drawn based on the assay results, and then the liquid-phase MPO concentration causing 50% competitive inhibition (IC₅₀) of MPO-ANCA binding to the solid-phase MPO was calculated from the curve.

Each assay was performed in duplicate and all serum samples were assayed at least 2-3 times.

Results

Most of antigen specificity for MPO-ANCAs was confirmed on the basis of 100% inhibition by adding an excess antigen of 50 μ g/ml to the liquid phase (Fig. 1). In few MPO-ANCAs, 5-10% MPO-uninhibitable binding activity was observed. This amount

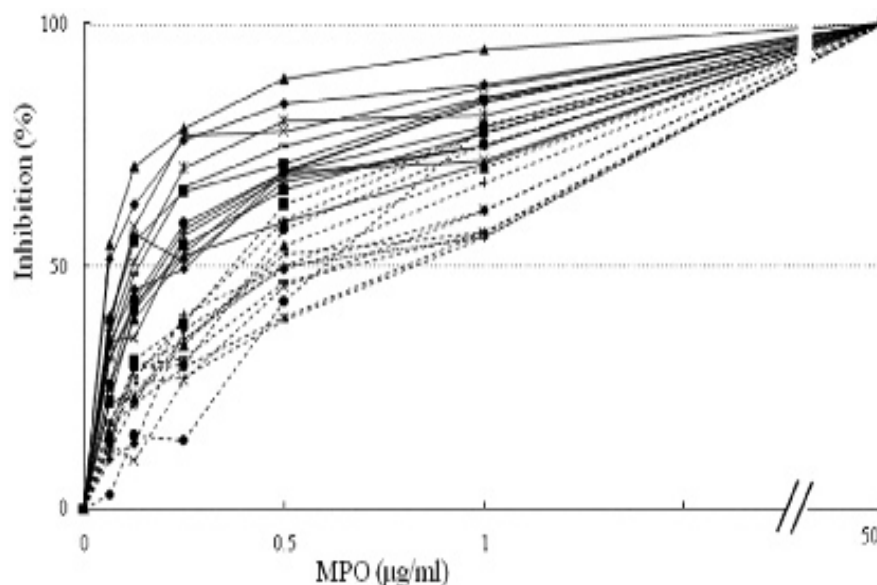
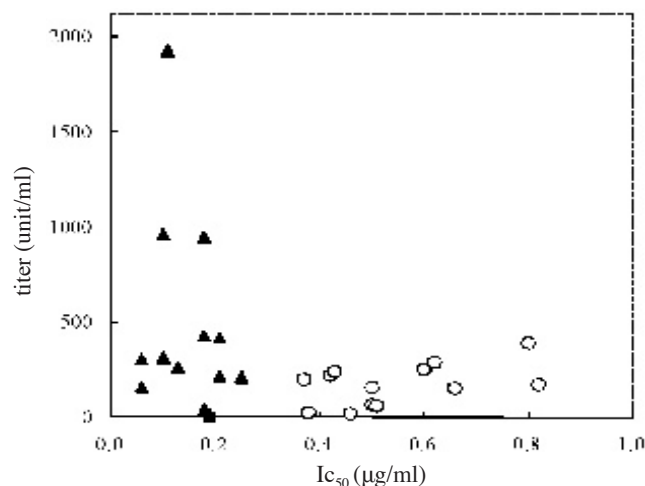


Fig. 1. MPO-inhibition of MPO-ANCA in ELISA.

The MPO-ANCA affinity (IC₅₀) of the 27 patients with MPO-ANCA associated vasculitides could be classified into a high-affinity group (solid line 14 patients) and a low-affinity group (dotted line 13 patients). Each point was assayed in duplicate.

Fig. 2. Correlation of MPO-ANCA titer and affinity (IC₅₀). solid triangle (\blacktriangle), high affinity group (n=14) open circle (\circ), low affinity group (n=13)



was excluded in 100% inhibition because the more excess antigen did not increase inhibition. Figure 1 showed that IC₅₀ of sera of the 27 patients with MPO-ANCA-associated vasculitides could be classified into a high-affinity type (14 patients) ranging from 0.06 to 0.25 μ g/ml and a low-affinity type (13 patients) ranging from 0.37 to 0.82 μ g/ml. Figure 2 showed that correlation of MPO-ANCA titer and affinity (IC₅₀) of the 27 patients with MPO-ANCA associated vasculitides. As shown in Table I, the mean values \pm standard deviation for IC₅₀ in the high affinity type were 0.15 \pm 0.06 μ g/ml and those in the low affinity type were 0.54 \pm 0.15

μ g/ml, and the difference between the two types was statistically significant ($p < 0.0000000684$). The Birmingham Vasculitis Activity Score (BVAS) (15) and C-reactive protein (CRP), which are clinical indicators of vasculitis activity (16, 17), were 18 \pm 6 and 10.8 \pm 7.7, respectively, in the high-affinity patient group (14 patients) and 11 \pm 8 and 2.8 \pm 4.1, respectively, in the low-affinity patient group (13 patients). There were statistically significant differences in BVAS and CRP scores between the two groups (BVAS: $p < 0.000933$; CRP: $p < 0.00129$). Mean MPO-ANCA titer in the high affinity group was 465 \pm 507 unit/ml (ranging from 13 to 1926 unit/

Table I. MPO-ANCA affinity and clinical data.

Patient	Age	Gender	BVAS	Renal symptoms RPGN/CRF	Respiratory symptoms ARDS/CIP	CRP (mg/dl)	MPO-ANCA titers (ELISA unit/ml)	MPO-ANCA affinity IC50 (µg/ml)	Treatment
Group 1. High affinity (n=14)									
1	79	F	15	+/-	-/-	4.60	316	0.10	CS
2	58	M	22	+/-	-/-	9.00	261	0.13	CS, IS
3	72	M	22	+/-	-/-	13.00	210	0.25	CS, IS, PE, HD
4	79	F	9	-/-	+/-	10.00	38	0.18	CS
5	56	F	9	-/-	+/-	7.00	307	0.06	CS
6	78	M	22	+/-	-/-	31.30	965	0.10	CS, IS, PE,
7	81	M	22	+/-	-/-	14.60	1926	0.11	CS, IS, PE,
8	88	F	15	+/-	-/-	3.50	947	0.18	CS, IS, PE,
9	65	M	15	-/-	+/-	20.00	13	0.19	CS, IS
10	89	F	22	+/-	+/-	8.10	429	0.18	CS, IS, PE, HD
11	74	M	9	+/-	-/-	1.60	418	0.21	CS, IS
12	81	F	21	-/+	+/-	15.12	158	0.06	CS
13	77	F	20	-/+	-/-	7.50	306	0.10	CS, IS
14	63	F	26	-/+	-/-	6.42	213	0.21	CS, IS
mean	74	M6:F8	● 18±6	8(57%)/3(21%)	5(36%)/0(0%)	■ 10.8±7.7	▲ 465 (±507)	◆ 0.15 (±0.06)	
Group 2. Low affinity (n=13)									
15	77	M	12	-/+	-/+	0.35	156	0.50	CS
16	68	M	3	-/+	-/+	0.10	19	0.46	not done
17	65	F	3	-/+	-/-	0.10	150	0.66	CS
18	56	M	8	-/+	-/-	0.10	254	0.60	CS
19	63	F	15	-/+	-/+	0.50	388	0.80	CS
20	61	M	12	+/-	-/-	0.10	222	0.42	CS
21	82	F	9	-/+	-/-	6.00	64	0.50	CS
22	76	F	14	+/-	-/-	8.00	171	0.82	CS, IS
23	63	F	4	-/+	-/-	1.30	22	0.38	not done
24	61	M	15	-/+	-/+	7.00	59	0.51	CS, IS
25	67	M	12	-/+	-/+	0.10	199	0.37	CS
26	63	M	19	-/+	-/+	12.00	241	0.43	CS, IS
27	77	F	12	+/-	-/+	0.10	288	0.62	CS
mean	68	M7:F6	● 11±8	3(23%)/10(77%)	0(0%)/7(54%)	■ 2.8±4.1	▲ 172 (±110)	◆ 0.54 (±0.15)	

CS: corticosteroid; prednisolone; PE: plasma exchange; IS: immunosuppressant (cyclophosphamide, azathiopurine, mizoribine etc); HD: hemodialysis; RPGN: rapidly progressive renal failure; CRF: chronic renal failure; ARDS: acute respiratory distress syndrome (acute interstitial pneumonitis/lung bleeding); CIP: chronic interstitial pneumonitis.

Differences between Group 1 and Group 2: ● $p < 0.000933$; ■ $p < 0.00129$; ▲ $p < 0.0265$; ◆ $p < 0.0000000684$ (Welch T-test).

BVAS, CRP, MPO-ANCA at the time of first diagnosis or relapse.

All cases from 9 to 18 months long term observation of MPO-ANCA titer and affinity.

ml) and that in the low affinity group 172±110 unit/ml (ranging from 22 to 388 unit/ml). The criterion of the difference in titer between the two groups for statistical significance was the 0.03 level ($p < 0.0265$). The high affinity group (n=14) included mainly cases with severe vasculitis disease activity (*i.e.* rapidly progressive glomerulonephritis (RPGN) 57% and/or acute respiratory distress syndrome (ARDS) 36%), while the low affinity group (n=13) included mainly cases with mild vasculitis activity (*i.e.* chronic renal failure (CRF) 77% and/or chronic interstitial pneumonitis (CIP 54%)). In addition, time-course determinations of

the ANCA titer and IC₅₀ were performed using serum samples from 10 of 27 patients. Figure 3 shows the representation time course in groups 1 and 2. In patient 6 of group 1, the MPO-ANCA titer decreased from 2800 (units/ml) to 250 (units/ml) during the 12-month period from the early stage of RPGN after onset until remission was achieved (serum creatinine from 14 mg/dl to 1.5 mg/dl and CRP from 31 mg/dl to 0.3 mg/dl), but IC₅₀ was maintained at approximately 0.15 µg/ml during the same period. In patient 19 of group 2 of CRF (serum creatinine from 2.8 mg/dl to 2.1 mg/dl and CRP from 2.8 mg/dl to 2.0 mg/dl) and CIP, the MPO-ANCA titer

decreased from 820 (units/ml) to 300 (units/ml) during an 10-month period with a relapse, but IC₅₀ remained steady at 0.5 and 0.6 µg/ml, respectively, with no statistically significant change during the time-course (Fig. 3).

Discussion

We set out to determine whether MPO-ANCA activity could be a potential biomarker of the activity of ANCA-associated vasculitides. This study indicated the occurrence of the two types of MPO-ANCA in vasculitis: one with a high affinity (IC₅₀=0.15±0.06 µg MPO/ml) and one with low affinity (IC₅₀=0.54±0.15 µg MPO/ml). High

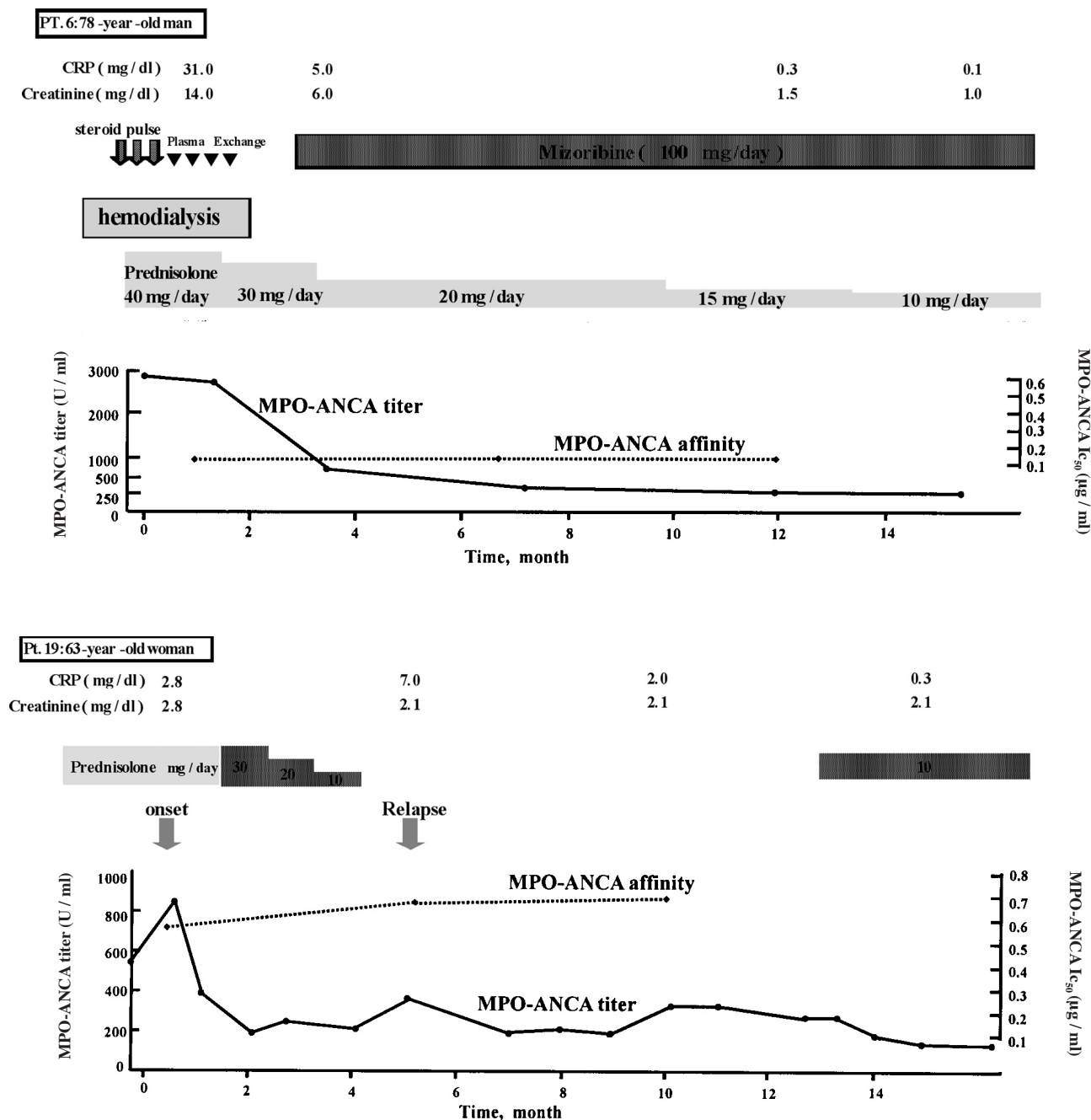


Fig. 3. Clinical course of titer and IC₅₀ of MPO-ANCA in patients with renal limited form of vasculitis. Upper panel: patient 6 (Group 1) with rapidly progressive glomerulonephritis; Lower panel: patient 19 (Group 2) with chronic renal failure.

affinity MPO-ANCAs were found in patients with mean BVAS of 18 ± 6 with severe vasculitis disease activity (16, 17) (*i.e.* RPGN and/or ARDS) and low affinity MPO-ANCAs in patients with mean BVAS of 11 ± 5 with mild vasculitis disease activity (16, 17) (*i.e.* CRF and CIP). The distinctive difference in affinity between the two types suggests that the number of antibody clones produced during disease

was limited. In addition, this notion is also supported by the observation in which no change in affinity of MPO-ANCA was longitudinally found from disease onset to remission or to the time of relapse. Native MPO is an N-glycosylated heme protein, which is a form of homodimer each consisting of a light chain and a heavy chain (18). Epitope mapping for MPO-ANCAs has indicated that epitopes are restricted to

mostly either an N-terminal 131 amino acid residues region or a C-terminal 228 amino acid residues region in the heavy chain, depending on the use of a panel of recombinant polypeptides or a series of recombinant human-mouse chimeric forms in a whole molecule (19, 20), respectively. These findings are also consistent with the notion that the antibody producing clones in each individual patient are limited. However,

whether these epitopes are associated with the affinities given for native MPO in this work remains to be clarified. Importantly, the finding that MPO ANCA with high or low affinity are found in patients with vasculitides will aid in mapping pathogenic epitope(s). As shown in this work, no correlation between MPO-ANCA titer and affinity was found as similarly as reported by others (11). In the longitudinal study the affinity remained unchanged during the follow-up. On the contrary, the two other groups reported that high apparent affinity constants aK of MPO-ANCAs in systemic vasculitis and propylthiouracil-induced vasculitis decrease to very low values within the short period of follow-up (11, 12). Discrepancies in affinity among these studies remain unsolved. The impurity of used MPO preparations and the presence of non-specific IgG binding to MPO-ELISA wells in some serum samples may affect affinity determination.

The present report is the first regarding the contribution of MPO-ANCA affinity in ANCA-associated vasculitides. Our results suggest that the affinity may be a useful index for assessment of the activity of vasculitis in the early period after disease onset and at the time of relapse. They also suggest that the chronological determinations of both titer and affinity of MPO-ANCA from the disease onset to the time of remission or relapse may be helpful for the assessment of the disease activity.

Conclusions

The affinity of MPO-ANCAs from patients with MPO-ANCA associated

vasculitides were largely distinguished into a high and low affinity, irrespective of the level of MPO-ANCA titers.

Acknowledgments

The authors are indebted to Associate Professor Breugelmanns of the International Medical Communications Center of Tokyo Medical University for his review of the manuscript.

References

1. KALLENBERG CGM: Antineutrophil cytoplasmic autoantibody-associated small-vessel vasculitis. *Curr Opin Rheumatol* 2007; 19: 17-24.
2. PETTERSSON E, HEIGL Z: Antineutrophil cytoplasmic antibody (cANCA and pANCA) titers in relation to disease activity in patients with necrotizing vasculitis: a longitudinal study. *Clin Nephrol* 1992; 37: 219-28.
3. SINICO RA, RADICE A, CORACE C *et al.*: Value of a new automated fluorescence immunoassay (EliA) for PR3 and MPO-ANCA in monitoring disease activity in ANCA-associated systemic vasculitis. *Ann NY Acad Sci* 2005; 1050: 185-92.
4. DE'OLIVIERA J, GASKIN G, DASH A *et al.*: Relationship between disease activity and anti-neutrophil cytoplasmic antibody concentration in long-term management of systemic vasculitis. *Am J Kidney Diseases* 1995; 25: 380-9.
5. HAN WK, CHOI HK, ROTH RM *et al.*: Serial ANCA titers: Useful tool for prevention of relapses in ANCA-associated vasculitis. *Kid Int* 2003; 63: 1079-85.
6. LURATI-RUIZ F, SPERTINI F: Predictive value of antineutrophil cytoplasmic antibodies in small-vessel vasculitis. *J Rheumatol.* 2005; 32: 2167-72.
7. CHEN M, YU F, ZHANG Y *et al.*: Characteristics of Chinese patients with Wegener's granulomatosis with anti-myeloperoxidase autoantibodies. *Kidney Int* 2005; 68: 2225-9.
8. YOSHIDA M, IWAHORI T, NAKABAYASHI I *et al.*: *In vitro* production of myeloperoxidase anti-neutrophil cytoplasmic antibody and establishment of Th1-type T cell lines from peripheral blood lymphocytes of patients. *Clin Exp Rheumatol* 2005; 23: 227-30.
9. OZAKI S: ANCA-associated vasculitis: diagnostic and therapeutic strategy. *Allergology Int* 2007; 56: 87-96.
10. FUJIMOTO S, UEZONO S, HISANAGA S *et al.*: Incidence of ANCA-associated primary renal vasculitis in the Miyazaki prefecture: the first population-based, retrospective, epidemiologic survey in Japan. *CJASN* 2006; 1: 1016-22.
11. KOKOLINA E, NOEL LH, NUSBAUM P *et al.*: Isotype and affinity of anti-myeloperoxidase autoantibodies in systemic vasculitis. *Kidney Int* 1994; 46: 177-84.
12. GAO Y, CHEN M, YE H *et al.*: Follow-up or avidity and titre of anti-myeloperoxidase antibodies in sera from patients with propylthiouracil-induced vasculitis. *Clin Endocrinol* 2007; 66: 543-7.
13. SAVIGE J, DAVIES D, FALK RJ *et al.*: Anti-neutrophil cytoplasmic antibodies and associated diseases: A review of the clinical and laboratory features. *Kidney Int* 2000; 57: 846-62.
14. SUZUKI K, YAMADA M, AKASHI K *et al.*: Similarity of kinetics of three types of myeloperoxidase from human leukocytes and four types from HL-60 cells. *Arch Biochem Biophys* 1986; 245: 167-73.
15. LUQMARI RA, BACON PA, MOOTS RJ *et al.*: Birmingham Vasculitis Activity Score (BVAS) in systemic necrotizing vasculitis. *QJM* 1994; 87: 671-8.
16. MUKHTYAR C, FLOSSMANN O, LUQMARI RA: Clinical and biological assessment in systemic necrotizing vasculitides. *Clin Exp Rheumatol* 2006; 24: S92-9.
17. MUKHTYAR C, LUQMARI R: Disease specific quality indicators, guidelines and outcome measures in vasculitis. *Clin Exp Rheumatol* 2007; 25: S120-9.
18. HANSSON M, OLSSON I, NAUSEEF WM: Biosynthesis processing and sorting of human myeloperoxidase. *Arch Biochem Biophys* 2006; 445: 214-24.
19. SUZUKI K, KOBAYASHI S, YAMAZAKI K *et al.*: Analysis of risk epitopes of anti-neutrophil antibody MPO-ANCA in vasculitis in Japanese population. *Microbiol Immunol* 2007; 51: 1215-20.
20. ERDBRUGGER U, HELLMARK T, BUNCH DO *et al.*: Mapping of myeloperoxidase epitopes recognized by MPO-ANCA using human-mouse MPO chimeras. *Kidney Int* 2006; 69: 1799-805.