Seven clinical conundrums in the treatment of ANCA-associated vasculitis

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ABSTRACT

Granulomatosis with polyangiitis and microscopic polyangiitis are two autoimmune diseases characterised by necrotising small-vessel vasculitis and presence of antineutrophil cytoplasm autoantibodies (ANCA). Current immunosuppressive regimes that combine cyclophosphamide and glucocorticoids have dramatically improved the outcome for these patients. However, these treatments are associated with toxic effects and do not lead to permanent remission in the majority of cases. Newer approaches have been sought during the last 15 years, with improvement in medication protocols and inclusion of novel therapies. This review develops on seven clinical conundrums of evidence-based therapeutic strategies for ANCA-vasculitis, posed as questions on aspects such as the role of established drugs in both remission induction and maintenance: glucocorticoids (and its duration), oral cyclophosphamide, methotrexate, TNF-a blockers, plasma exchange, mycophenolate mofetil, plus one related to newer developments in treatment with agents blocking the complement system and the possible role of sequential or combined therapies, mainly directed against B cells.

Introduction

Granulomatosis with polyangiitis (GPA, Wegener's) and microscopic polyangiitis (MPA) are small-vessel systemic vasculitis associated with antineutrophil cytoplasmic autoantibodies (ANCA) (1). Prior to the introduction of current standard treatment, GPA had a mortality rate of 80% within the first year (2). Forty years after first use of cyclophosphamide (CYC) as effective therapy in ANCA-associated vasculitis (AASV), survival is close to 80% at 5 years, but morbidity and drug-related toxic effects are still frequent (3-5). Although treatment options have increased in the last decade, with other immunosuppressives, biological drugs and plasma exchange (PLEX) (6), several questions remain unanswered. We have developed on some based on available evidence.

A MEDLINE search for English-language articles published between January 1970 and November 2012 was done. Terms included ANCA-associated vasculitis, granulomatosis with polyangiitis, Wegener granulomatosis, microscopic polyangiitis, renal limited vasculitis and necrotising glomerulonephritis plus combinations with treatment, therapy, refractory, cyclophosphamide, methotrexate, TNF-a blockers, etanercept, infliximab, adalimumab, certolizumab, golimumab, plasma exchange, plasmapheresis, mycophenolate mofetil, rituximab, intravenous immunoglobulin, and complement system. We identified and sought relevant articles from the references listed in retrieved articles. Late-breaking communications from international meetings were reviewed. Both authors reviewed wholly relevant articles.

Conundrum 1: Which is the best glucocorticoid regime and how long should it be used?

Although glucocorticoids (GC) are the cornerstone of the treatment, the optimal initial dose, tapering scheme and duration for remission-induction and maintenance phases are not well established.

Initial dose

It is common to start the with prednisone (PDN) or prednisonole at 1 mg/ kg/day (maximum 60-80 mg/d) (6). In addition, in severe disease, pulsed

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methylprednisolone is used (7 to 15 mg/kg, max 500-1000 mg qd for 3 consecutive days) (6, 7). Although common practice (7-12), these schemes are empirical, unsupported on evidencebased studies. Thus, best initial PDN (or equivalents) doses are debatable.

GC exert their anti-inflammatory and immunosupressive actions through genomic and non-genomic pathways (13, 14). Recent investigations reveal that the genomic pathway (responsible of inhibition of many pro-inflammatory molecules, but also toxic effects) is fully activating at PDN doses between 30 and 100 mg qd (15-18). This suggests that, at least theoretically, lower initial doses of PDN (0.5 mg/kg/d) could be as effective as higher to manage inflammatory disorders. There is evidence that schemes with reduced PDN or lower methylprednisolone doses, are non-inferior to standard high GC regimes in the treatment of lupus nephritis (19-21). This possibility, applied to the AASV merits formal evaluation. The PEXIVAS study (22) (ClinicalTrials.gov Identifier: NCT00987389) has introduced one low-dose GC arm (0.5 mg/kg) to evaluate if the reduced cumulative dosing regimen is as effective as standard doses for patients with severe AASV.

Tapering protocol

PDN tapering schedules are heterogeneous (7-12). Most protocols reach a dose of 20 mg qd at month two (12, 23), and no lower than 15 mg qd for the first 3 months of therapy (6). Decreasing GC protocols must be appropriate to reach disease control but to prevent the development of adrenal insufficiency. In this regard, it has been estimated that patients who receive ≥20 mg of PDN/ day for more than 3 weeks will suffer hypothalamic-pituitary-adrenal (HPA)axis inhibition (24); thus, PDN should be gradually tapered. However, some patients are unable to tolerate PDN doses below 10-15 mg/day without relapsing or experiencing smoldering activity (25-27), while others on GC physiologic doses (7.5-10 mg/d) cannot tolerate complete withdrawal. This is particularly true for GPA patients with granulomatous disease (28, 29). Accordingly, individual clinical criteria are necessary to achieve a balance between the minimum dose required to control activity and that predisposing to adverse effects.

Treatment duration

Total duration of GC therapy is controversial. It ranges from 6 to 18 months or more to control persistent systemic symptoms (7, 26, 30). The former is particularly possible when vasculitic manifestations, like glomerulonephritis or pulmonary haemorrhage due to lung capillaritis are present, but could take longer in those with predominant granulomatous manifestations. A recent guideline endorsed by EULAR recommends a maintenance dose of 10 mg qd or less during remission, but does not specify for how long (6).

Two recent studies (31, 32) evaluating the effects of duration of GC therapy in AASV are contradictory. In the first (31), -a metaanalysis of 983 patients-, a three-fold higher relapse rate took place in patients with GC discontinued within the first 12 months, compared with patients in which it was done after, or not at all (43% vs. 14%, respectively). These results contrast with those reported by McGregor et al. (32), who observed that length of GC therapy (shorter or longer than 6 months) had no effect on relapse number, progression to end-stage renal disease (ESRD) or death. Importantly, those treated beyond 6 months had significantly greater incidence of infections and new-onset diabetes mellitus.

In the absence of clear evidence to limit the PDN to ≤ 6 months, our opinion is to use it at low dose for long term in patients at high risk of relapse (upper respiratory tract disease, lung involvement, positive PR3-ANCA (33, 34)), in order to maintain control and reduce the accumulation of disease-related scarring. Also, it seems that some patients with persistant disease, especially with eye and retro-orbital involvement, and/ or rhinosinusal disease, are not able to wholly withdraw low-dose GC.

Conundrum 2:

Who is a likely candidate for remission induction with methotrexate?

Treatment with long-term, high-dose CYC is associated with well-known

toxicity at multiple levels that influence long-term morbidity and mortality of patients (5, 35). In an effort to reduce adverse effects (AE), methotrexate (MTX) has been used instead of CYC for remission induction (8, 12, 23, 27, 36-39). However, there are several questions about its use.

Dose

Added to GC, oral or parenteral formulations have been used starting at 15 mg/week (0.25–0.3 mg/kg) with increases of 2.5 mg weekly up to 20-25 mg/week between the first and third months (8, 23, 27, 36-39). Once remission is achieved, treatment should be continued for at least one year, probably 18-24 months, then tapered by 2.5 mg/week until stopped (8, 23, 30, 37).

Target population

MTX is recommended as an alternative in patients with AASV with early systemic disease (6). However, there is lack of an uniform definition for such (8, 12, 23, 27, 36-39), being currently described as non-organ or life threatening disease with a serum creatinine (Cr) level <1.35 mg/dL (40).

From published literature (8, 12, 23, 27, 36-39), MTX should be used in patients with recent GPA with or without constitutional symptoms and limited pulmonary involvement (focal pulmonary infiltrates or nodules with room air PO2 >70 mm Hg or pulse oximetry >92%), mild eye inflammatory disease (conjunctivitis, episcleritis, dacriocystitis), ENT manifestations, skin disease without extensive areas of gangrene, or musculoskeletal symptoms. MTX is not advised in cases of severe or lifethreatening disease including acute renal failure, pulmonary haemorrhage associated with bilateral infiltrates, gastrointestinal or central nervous system involvement, progressive neuropathy, severe eye manifestations or heart failure due to pericarditis or myocarditis. In addition, chronic liver disease, alcoholism, pregnancy, pre-existing blood dyscrasias and age >75 years are current restrictions for MTX administration. It should be stressed that combination of co-trimoxazole - used previously for maintenance of remission in GPA patients with limited disease (41) – and MTX is contraindicated as this may result in pancytopenia. Of interest, MTX has been related to inflammatory and/or fibrotic lung disease in 1-5% of rheumatoid arthritis patients (42). Although pre-existing lung disease is not considered a contraindication to MTX administration, it has been identified as risk factor for pulmonary toxicity (43). This is particularly true in MPA patients with associated interstitial lung disease (44, 45).

General efficacy and

management of renal disease

Remission has been achieved in 60%-92% of patients treated with MTX and GC (8, 12, 36, 37, 46). In the only randomised clinical trial comparing CYC and MTX for remission induction, (NO-RAM study), MTX was non-inferior to CYC (8). In 100 patients (90% GPA), remission was achieved in 90% of those treated with MTX and in 93.5% treated with CYC. MTX was less effective in patients with extense disease and those with pulmonary involvement, requiring longer time to achieve remission. In addition, more MTX-treated patients relapsed (69.5% vs. 46.5%, similar to other reports, 36%-52% (25, 36, 37, 47)) and required higher cumulative doses of PDN (8.8 gr for MTX vs. 6.2 gr for CYC). No difference in infections was documented. Two drawbacks were the slow escalating schedule to reach target dose, which besides was 20-25 mg/week. If tolerated, it could have probably been of 30 mg/week.

NORAM's long-term follow-up was recently published. MTX-treated patients had shorter sustained remission than those who received CYC (48), cumulative relapse-free survival tended to be higher in CYC, and exposure to GC and other immunossupressive drugs was longer in MTX patients. However, no differences in cumulative survival (MTX 88% vs. CYC 90%), development of end-stage renal disease (ESRD), cancer or serious infections occurred. There is debate about efficacy and safety

of MTX in patients with renal involvement. MTX is mainly kidney-eliminated, and toxic potential is enhanced when renal clearance is impaired (49). In all referenced studies (8, 12, 23, 27, 36-39), patients with severe renal disease (Cr increase of >25% above baseline or Cr >2.5 mg/dL, red blood cell casts and proteinuria >1 gr qd) were excluded. However, good results have been reported in patients with mild renal disease (23, 48). In Langford et al. series (23), all patients (n=21) with active glomerulonephritis (mean Cr level of 1.4 mg/dL) achieved remission and 90% patients remained with stable or improved Cr levels at the end of follow-up (median of 6 years). It must be stressed that 60-70% of relapses in patients with maintenance therapy with MTX involve the kidneys (25, 47). However, progression to ESRD is rarely observed (2-5%) (12, 25, 47, 48), and patients with active renal manifestations without severe disease could be MTX-treated. We agree with this notion, but would achieve a maximal MTX dose (25-30 mg/week) faster than the NORAM trial did. They did it in 12 weeks, while we advocate doing it by 6-8 weeks. Also, some issues like dividing the dose on the same day or on 2 days, or giving it subcutaneously, which may, as in rheumatoid arthritis patients, increase absoption and bioavailability, could result favourable. However, in these diseases this has not been specifically proven. Regarding other manifestations, MTX seems adequate for predominant granulomatous disease.

Conundrum 3: Is TNF-α inhibition out of sight?

Tumour necrosis factor- α (TNF- α) has been implicated in the pathogenesis of the AASV (50-54). *In vitro*, it enhances the ability of neutrophils to degranulate in presence of ANCA (54). In addition, high expression of TNF- α is detected at sites of vasculitis injury (52) and in the granulomas of patients with GPA (53). In the latter, increased circulating levels of TNF- α and its soluble receptor are present in active disase (50, 51).

Initial promising results with TNF- α inhibition (55, 56) led to the WGET trial, a randomised, placebo-controlled study. Results showed etanercept's inefficacy for remission maintenance, and a high rate of increased solid organ tumours (12, 57).

Although no randomised trial has established the efficacy of infliximab in AASV, information derived from small case series (58-60) or open label cohorts (61, 62) allows to speculate on its benefits. In five studies comprising 72 patients (GCA=53, MPA=19) (58-62), it induced remission in refractory or relapsing patients, mostly those who had received one or more immunosuppressives including CYC, MTX, azathioprine (AZA) or mycophenolate mofetil (MMF). Between 6 to 26 weeks, it induced remission or response in 85-90% of patients. In contrast, in a recently randomised study comparing the long term efficacy of RTX (n=8) and infliximab (n=9), only 33% of refractory GPA patients reached remission at 12 months (63). Also, it seems effective for controlling severe clinical manifestations like rapidly progressive glomerulonephritis, neuropathy, central nervous system manifestations (pachymeningitis and pituitary involvement), gastrointestinal vasculitis, severe ocular disease and refractory ENT and lung symptoms, and exhibits a GC-sparing effect. However, relapses are frequent with the use of infliximab (20-60%) even while still on therapy or after discontinuation. Some relapses have been successfully retreated with reintroduction of the biological. Regarding AE, infections are high (20%) with one fatality in 72 reported cases, while in relation to malignancy, one previously CYC-treated patient developed lymphoma.

Clinical differences in efficacy and side effects of etanercept and other anti TNF- α agents exist, in particular in granulomatous disorders such as Crohn's disease (64, 65). Variations in bioavailability and stability of union to the TNF- α molecule reside in the structure of different TNF-inhibitors. Etanercept is a fusion protein composed of two extracellular p75 TNF receptor domains (66), while infliximab is a chimeric monoclonal antibody (67). In contrast to etanercept, infliximab binds to circulating and membrane-bound TNF- α and induces apoptosis in cells expressing this molecule (64, 65, 68). In addition, infliximab has higher affinity and avidity, resulting in longer bind-

ing to TNF- α (61). Lack of favourable response observed in the WGET trial with etanercept was probably due to these differences but also because the dose used may not have been sufficient to achieve a therapeutic effect. Available information supports infliximab use as second line therapy for refractory or uncontrolled relapsing AASV patients. Concurrent use of CYC and anti-TNF- α is not advised, and patients that have received both medications should be followed closely.

There is one series of 14 patients with severe AASV (GPA=9, MPA=5) treated with adalimumab (69), in where remission was achieved in 78.5% of patients within the first 14 weeks of treatment, allowing concomitant GC reduction. Infections were reported in 3 patients (21%). Further studies are needed to gain insight about the potential place of adalimumab or infliximab for AASV.

Conundrum 4:

Plasma exchange in alveolar haemorrhage: as effective as in renal disease?

In patients with AASV and severe kidney disease, PLEX has reduced ESRD development, although it did not improve overall survival (70-76).

Diffuse alveolar haemorrhage (DAH) is present in 10-45% of patients with GPA and 10–30% with MPA (5, 38, 77-79). In the acute setting, 30% of DAH patients died, and in those who survived the initial episode, life expectancy at 1 and 5 years was reduced to 82% and 68%, respectively (79). Evidence for PLEX addition in AASV-DAH is scarce, heterogeneous and composed of case reports (80-85) and small retrospective series (86-89).

Most published case reports (80-83) include MPA, MPO-ANCA positive patients with respiratory failure. In them, combination of intense immunosuppression with CYC, IV-methylprednisolone and PLEX was associated with clinical and radiological improvement. However, populations were heterogeneous and the PLEX protocols used were diverse or not detailed. In the largest series (n=20 patients) (89), DAH resolved in 100% cases when treated with IV-methylprednisolone for 3 days and/or IV-CYC in combination with daily full-volume PLEX (average of 6.4 exchanges (range 4-9)). These results contrast with other two case series (87, 88), where mortality was as high as 36-42%, particularly in the first month. Based on the available information, it is not possible to unequivocally recommed PLEX as useful in DAH. In our limited experience (unpublished), we have had good results, similar to those of the MEPEX trial, in where some patients also had DAH, with resolution or improvement of this complication. The results of the PEXIVAS trial (22) will probably clarify if PLEX is useful for DAH. Five-hundred patients with DAH and/or severe renal disease will be randomised to adjunctive PLEX or no PLEX besides standard or reduced GC treatment.

Conundrum 5: What is the current role of oral cyclophosphamide?

The current use of oral CYC is under review (4,90) due to proved efficacy of pulsed IV-CYC (91, 92) and rituximab (RTX) (93-95).

In contrast to data regarding IV-CYC or RTX, there is long-term experience regarding oral CYC in AASV patients (4, 90), with remission or response in 77%-94% of cases, sustained remission in 43%-58% of cases (5, 7, 8, 33, 96), and 5-year survival between 70% to 90% (3, 97). Nonetheless, relapses are frequent in CYC-treated patients (29%-50%) and permanent disease morbidity is observed in 86% (5, 7, 8, 33, 96). The major disadvantage of oral CYC is high frequency of toxic effects, seen in 80% of individuals (8). Adverse effects (AE) associated with CYC are: amenorrhea (57%), cystitis (43%), leukopenia (35%), hair loss (17%), bladder cancer (2.8%), myelodysplasia (2%) and haematological malignancies (1 %) (5). Remarkably, 8% to 46% of GPA or MPA patients experience serious infections, with 20% mortality (5). Efforts to reduce the AE of high cumulative doses of CYC include dosage adjustment or IV administration, but although the latter and RTX are equally effective for inducing remission in patients with generalised AASV (91-94, 98, 99) they are not superior to oral CYC, remaining an option. Indeed, some patients who did not improve with IV-CYC, did with daily administration (100)

When compared to oral CYC, IV-CYC and RTX have a similar incidence of severe AE, except for leukopenia (91-94, 98, 99). Of note, infections rate has not decreased with RTX and IV-CYC (94, 101, 102). This probably reflects the effect of prolonged administration of high oral CYC doses used in older cohorts (3-5 mg/kg/d for severe disease) (5) in comparison to adjusted doses used in the most recent studies (1.5–2 mg/kg/d) (103).

Patients which seem to benefit most from oral CYC include those with generalised disease without predictors of relapse, i.e. PR3-ANCA+, upper respiratory tract, lung disease (33, 34, 96, 104), cardiac involvement or granulomatous predominant disorder (105), patients with active disease despite RTX, or those intolerant to it, and those with DAH severe enough to require mechanical ventilation or with rapidly progressive glomerulonephritis and serum Cr levels >4mg/dL. Additionally, oral CYC could also be an option for individuals in countries where the economic burden of biologics is very high. Patients preference may contribute to the regime choice once advantages and disadvantages are explained.

For those patients receiving oral CYC, we recommend an initial dose of 1.5–2 mg/kg qd (maximum 150 mg/day) for a minimum of 3, and a maximum of 6 months in non-refractory cases (92, 106, 107). Excellent previous reviews (92, 106, 107) have described dose adjustments, monitoring guidelines, and preventive measures regarding opportunistic infections, urinary and gonadal toxicity.

Conundrum 6: Is MMF effective for either induction or maintenance therapy?

MMF selectively inhibits T and B cell proliferation by suppressing guanine synthesis and blocking DNA structure (108). It is extensively used in organ rejection prevention following renal, cardiac or hepatic transplants (109, 110). In addition, MMF has been explored with good results in other autoimmune disorders, paticularly in lupus nephritis (111-113).

In AASV, MMF has been used for both remission induction (114-119) and maintenance therapy (117, 120, 121). Experience as induction therapy includes almost 100 patients reported in small case series (114, 116-119) and one single centre non-blinded clinical trial (115). In these studies, patients with GPA and MPA were treated with MMF instead of CYC because of intolerance or refractory/relapsing disease. Remarkably, half of these patients had mild to moderate active renal disease. Overall, complete remission was attained in 60%-82% of these refractory/ relapsing cases (114-117) with daily doses of 1.5-3 g. Progression to ESRD was uncommon and, in approximately 60% of patients with kidney involvement, renal function remained stable or improved (114, 116-119).

Direct comparison between MMF and CYC for remission induction has been performed in a small single-centre trial (n=35, most patients MPO-ANCA+) (115). Remission rate at 6 months was higher with MMF than CYC (77.8% vs. 47.1%), with renal function improvement. In contrast, results of an international, multi-centre, controlled trial (MYCYC) (122), -which was ultimately underporwered-, patients were randomised to MMF (2-3 g/day) or 6-10 pulses of IV CYC (15mg/kg). In the preliminary results, recently published in abstract form, remission at 6 months with MMF was 66% vs. 69% with CYC, but confidence intervals were wide and surprisingly, overall expected remission rates were inferior to estimated; also, MMF did not demostrate significant glucocorticoid-sparing effect, and both drugs were associated with similar rates of adverse effects.

Regarding maintenance therapy, case series (117, 120, 123, 124) summing approximately 70 patients reported stabilisation of renal function and decreased activity indices in the majority of patients. However, relapses are common, presented in 10%–76% of cases (117, 120, 123, 124). More importantly, the

results of an open-label, randomised multi-center trial (IMPROVE) (121), showed that MMF was less effective than AZA for the prevention of relapses in AASV. In 156 patients with newly diagnosed GPA/MPA, MMF (2 g/day) was compared to AZA (2 mg/kg/day) for maintenance after induction of remission with CYC. At median followup of 39 months, relapses were significantly more frequent among those in the MMF limb compared with the AZA arm (55% vs. 38%). Also, in the referenced studies (114, 115, 117, 121), MMF related AE were commonly observed (22%-71%), mostly infections and/or gastrointestinal intolerance. Based on the available information, it remains to elucidate if MMF could be considered as a first line therapy for remission induction, and in maintenance AZA and MTX seem better. However, it can be considered as reasonable option for patients with non-life threatening relapsing or refractory disease.

Conundrum 7: Which recent advances seem promising?

Through this review we have discussed on unsolved dilemmas in treatment of AASV. There is clearly a need for newer therapies with better efficacy and fewer adverse effects. In the last 2 years, with better understanding of AASV pathogenesis and efforts of collaborative groups around the world, new strategies to achieve and maintain remission have emerged as potential options (125). Belimumab, abatacept or gusperimus could be new future alternatives, provided extended information and precise situations in which their role is endorsed develop. However, in the final part of this paper we will review two exciting options with known drugs: 1) combination of RTX with high-dose intravenous immunoglobulin (IVIG) and 2) inhibition of the complement system.

1) The hypothetic role of combined or sequential schemes of B-cell depletion therapy and IVIG

Two randomised clinical trials, *i.e.* RAVE (94) and RITUXVAS (93) and several case series and cohort studies

(126-141) (pooled $n = \sim 250$ patients), demonstrated that RTX combined with GC is not inferior as standard therapy with CYC for remission induction of patients with newly severe AASV and, probably more effective in refractory/relapsing cases. On the other hand, small case series (142-146) have showed that administration of highdose IVIG (total dose 2 g/kg) in cases with uncontrolled and persistent ANCA disease resulted in improvement of disease activity in 50%-80% of patients. However, in these last studies low rate of complete remission was attained (145) and benefits of IVIG were limited to the first 3 months after infusion (144, 146).

Combination schemes using RTX and IVIG have already been tried with good results in a case of an acquired inhibitor of coagulation factor VIII (147), for renal transplant recipients (148, 149) and in patients with pemphigus vulgaris. (150, 151). Importantly, a favourable safety profile was reported in all (152). RTX and IVIG share some mechanisms of action which can act synergistically, such as B-cell apoptosis induction, antibody-dependent cellular cytotoxicity, modulation of several pro-inflammatory cytokines and improvement of the suppressive functions of Treg (152-154). In addition, some of their immune-modulatory effects can be complementary. For example, IVIG may act rapidly reducing the immune activity by interacting with Fc-receptors of inflammatory cells (155, 156) and by the presence of anti-idiotypic antibodies that could theoretically neutralise ANCA (157-159). These highspeed, short-lived effect of IVIG can be followed by the more prolonged and durable actions of RTX (153). As experience with combination schemes is virtually inexistent in AASV, it remains to determine the optimal dose for each drug, suitability of concomitant or sequential administration and, importantly, the safety profile. However, we think such an option deserves formal evaluation.

2) Inhibition of the complement system in light of newer discoveries Recent data suggest a role for the the

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complement system (CS) in the pathogenesis of AASV (reviewed in (160-162)). Briefly, based on mice models of anti-MPO-induced glomerulonephritis (163, 164), it has been demostrated that the development of renal disease and vascular inflammation depend on complement activation (165, 166), specifically, the alternative pathway (166). Of note, C5a and its receptor C5aR appear to be fundamental for induction of damage in these models (164, 165). In addition, in vitro studies have shown that ANCA-stimulated neutrophils release factors (probably properdin) that cause complement activation with generation of C3a (166) and C5a (164), initiating the cascade that culminates in inflammation and damage of vessel walls. In humans there is evidence to suggest that the alternative pathway is involved in vasculitic renal damage. Complement factors C3c, C3d and membrane attack complex (MAC) have been detected in renal specimens of patients with ANCA-associated glomerulonephritis (167, 168). Interestingly, demostration of glomerular deposition of C3c was associated with higher levels of proteinuria and poorer renal function (167). Finally, a recent published study have disclosed elevated plasma and urinary concentrations of C5a in 24 patients with active MPO ANCA-vasculitis with glomerular disease (169). Based on these data, blocking the activity of the complement system appears interesting, particularly C5 inhibition. For this purpose, eculizumab, a monoclonal antibody (mAb) that binds to C5, blocking its activation seems a logical choice. In fact, a clinical trial targeting complement activation in AASV with eculizumab is currently recruiting patients (ClinicalTrials. gov Identifier: NCT01275287). In this study, patients with active ANCA vasculitis and renal disease will be randomised to receive conventional therapy in addition to the biologic therapy or placebo. Other inhibitory molecules of the C5-C5aR axis that could be candidates for AASV treatment include pexelizumab (mAb targeted C5), neutrazumab (antibody blocking the C5aR), and molecules like ARC905 and CCX168 (170).

Conclusion

Current treatment of AASV is the result of multiple contributions and refinements made in the last 40 years. Major progress has been made in patient survival. However, we face now chronic diseases that can severely impair the quality of life. Therefore, finding new and better treatments is a priority for those dedicated to the study of systemic vasculitides.

References

- JENNETTE JC, FALK RJ, BACON PA et al.: 2012 Revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides. Arthritis Rheum 2013; 65: 1-11.
- WALTON EW: Giant-cell granuloma of the respiratory tract (Wegener's granulomatosis). Br Med J 1958; 2: 265-70.
- FLOSSMANN O, BERDEN A, DE GROOT K et al.: Long-term patient survival in ANCAassociated vasculitis. Ann Rheum Dis 2011; 70: 488-94.
- FAUCI AS, WOLFF SM: Wegener's granulomatosis: studies in eighteen patients and a review of the literature. *Medicine* (Baltimore) 1973; 52: 535-61.
- HOFFMAN GS, KERR GS, LEAVITT RY et al.: Wegener granulomatosis: an analysis of 158 patients. Ann Intern Med 1992; 116: 488-98.
- MUKHTYAR C, GUILLEVIN L, CID MC et al.: EULAR recommendations for the management of primary small and medium vessel vasculitis. Ann Rheum Dis 2009; 68: 310-7.
- GUILLEVIN L, CORDIER JF, LHOTE F et al.: A prospective, multicenter, randomized trial comparing steroids and pulse cyclophosphamide versus steroids and oral cyclophosphamide in the treatment of generalized Wegener's granulomatosis. *Arthritis Rheum* 1997; 40: 2187-98.
- DE GROOT K, RASMUSSEN N, BACON PA et al.: Randomized trial of cyclophosphamide versus methotrexate for induction of remission in early systemic antineutrophil cytoplasmic antibody-associated vasculitis. *Arthritis Rheum* 2005; 52: 2461-9.
- JAYNE D, RASMUSSEN N, ANDRASSY K et al.: A randomized trial of maintenance therapy for vasculitis associated with antineutrophil cytoplasmic autoantibodies. N Engl J Med 2003; 349: 36-44.
- JAYNE DR, RASMUSSEN N: Treatment of antineutrophil cytoplasm autoantibody-associated systemic vasculitis: initiatives of the European Community Systemic Vasculitis Clinical Trials Study Group. *Mayo Clin Proc* 1997; 72: 737-47.
- KOLDINGSNES W, NOSSENT JC: Baseline features and initial treatment as predictors of remission and relapse in Wegener's granulomatosis. J Rheumatol 2003; 30: 80-8.
- 12. WEGENER'S GRANULOMATOSIS ETANERCEPT TRIAL (WGET) RESEARCH GROUP: Etanercept plus standard therapy for Wegener's granulomatosis. *N Engl J Med* 2005; 352: 351-61.

- KLEIMAN A, TUCKERMANN JP: Glucocorticoid receptor action in beneficial and side effects of steroid therapy: lessons from conditional knockout mice. *Mol Cell Endocrinol* 2007; 275: 98-108.
- STAHN C, BUTTGEREIT F: Genomic and nongenomic effects of glucocorticoids. Nat Clin Pract Rheumatol 2008; 4: 525-33.
- 15. BUTTGEREIT F, DA SILVA JA, BOERS M et al.: Standardised nomenclature for glucocorticoid dosages and glucocorticoid treatment regimens: current questions and tentative answers in rheumatology. Ann Rheum Dis 2002; 61: 718-22.
- LIPWORTH BJ: Therapeutic implications of non-genomic glucocorticoid activity. *Lancet* 2000; 356: 87-9.
- RUIZ-IRASTORZA G, DANZA A, KHAMASH-TA M: Glucocorticoid use and abuse in SLE. *Rheumatology* (Oxford) 2012; 51: 1145-53.
- STAHN C, LOWENBERG M, HOMMES DW, BUTTGEREIT F: Molecular mechanisms of glucocorticoid action and selective glucocorticoid receptor agonists. *Mol Cell Endocrinol* 2007; 275: 71-8.
- BADSHA H, EDWARDS CJ: Intravenous pulses of methylprednisolone for systemic lupus erythematosus. *Semin Arthritis Rheum* 2003; 32: 370-7.
- 20. ILLEI GG, AUSTIN HA, CRANE M et al.: Combination therapy with pulse cyclophosphamide plus pulse methylprednisolone improves long-term renal outcome without adding toxicity in patients with lupus nephritis. Ann Intern Med 2001; 135: 248-57.
- KONG KO, BADSHA H, LIAN TY, EDWARDS CJ, CHNG HH: Low-dose pulse methylprednisolone is an effective therapy for severe SLE flares. *Lupus* 2004; 13: 212-3.
- 22. WALSH M, MERKEL P, CASIAN A, JAYNE D: PEXIVAS: design of an RCT of PLEX and GC dosing in the treatment of severe ANCA-AAV. *Clin Exp Immunol* 2011; 164: 67.
- 23. LANGFORD CA, TALAR-WILLIAMS C, SNEL-LER MC: Use of methotrexate and glucocorticoids in the treatment of Wegener's granulomatosis. Long-term renal outcome in patients with glomerulonephritis. *Arthritis Rheum* 2000; 43: 1836-40.
- 24. MEIKLE AW, TYLER FH: Potency and duration of action of glucocorticoids. Effects of hydrocortisone, prednisone and dexamethasone on human pituitary-adrenal function. *Am J Med* 1977; 63: 200-7.
- 25. LANGFORD CA, TALAR-WILLIAMS C, BAR-RON KS, SNELLER MC: Use of a cyclophosphamide-induction methotrexate-maintenance regimen for the treatment of Wegener's granulomatosis: extended follow-up and rate of relapse. *Am J Med* 2003; 114: 463-9.
- 26. DE GROOT K, REINHOLD-KELLER E, TAT-SIS E *et al.*: Therapy for the maintenance of remission in sixty-five patients with generalized Wegener's granulomatosis. Methotrexate versus trimethoprim/sulfamethoxazole. *Arthritis Rheum* 1996; 39: 2052-61.
- HOFFMAN GS, LEAVITT RY, KERR GS, FAUCI AS: The treatment of Wegener's granulomatosis with glucocorticoids and methotrexate. *Arthritis Rheum* 1992; 35: 1322-9.

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- STONE JH: Limited versus severe Wegener's granulomatosis: baseline data on patients in the Wegener's granulomatosis etanercept trial. Arthritis Rheum 2003; 48: 2299-309.
- 29. HOLLE JU, GROSS WL, HOLL-ULRICH K et al.: Prospective long-term follow-up of patients with localised Wegener's granulomatosis: does it occur as persistent disease stage? Ann Rheum Dis 2010; 69: 1934-9.
- PAGNOUX C, MAHR A, HAMIDOU MA et al.: Azathioprine or methotrexate maintenance for ANCA-associated vasculitis. N Engl J Med 2008; 359: 2790-803.
- 31. WALSH M, MERKEL PA, MAHR A, JAYNE D: Effects of duration of glucocorticoid therapy on relapse rate in antineutrophil cytoplasmic antibody-associated vasculitis: A meta-analysis. Arthritis Care Res (Hoboken) 2010; 62: 1166-73.
- 32. MCGREGOR JG, HOGAN SL, HU Y, JENN-ETTE CE, FALK RJ, NACHMAN PH: Glucocorticoids and relapse and infection rates in anti-neutrophil cytoplasmic antibody disease. *Clin J Am Soc Nephrol* 2012; 7: 240-7.
- 33. NACHMAN PH, HOGAN SL, JENNETTE JC, FALK RJ: Treatment response and relapse in antineutrophil cytoplasmic autoantibody-associated microscopic polyangiitis and glomerulonephritis. J Am Soc Nephrol 1996; 7: 33-9.
- WALSH M, FLOSSMANN O, BERDEN A et al.: Risk factors for relapse of antineutrophil cytoplasmic antibody-associated vasculitis. *Arthritis Rheum* 2012; 64: 542-8.
- 35. TALAR-WILLIAMS C, HIJAZI YM, WALTHER MM *et al.*: Cyclophosphamide-induced cystitis and bladder cancer in patients with Wegener granulomatosis. *Ann Intern Med* 1996; 124: 477-84.
- 36. SNELLER MC, HOFFMAN GS, TALAR-WIL-LIAMS C, KERR GS, HALLAHAN CW, FAUCI AS: An analysis of forty-two Wegener's granulomatosis patients treated with methotrexate and prednisone. *Arthritis Rheum* 1995; 38: 608-13.
- VILLA-FORTE A, CLARK TM, GOMES M et al.: Substitution of methotrexate for cyclophosphamide in Wegener granulomatosis: a 12-year single-practice experience. *Medicine* (Baltimore) 2007; 86: 269-77.
- 38. REINHOLD-KELLER E, BEUGE N, LATZA U et al.: An interdisciplinary approach to the care of patients with Wegener's granulomatosis: long-term outcome in 155 patients. Arthritis Rheum 2000; 43: 1021-32.
- 39. STONE JH, TUN W, HELLMAN DB: Treatment of non-life threatening Wegener's granulomatosis with methotrexate and daily prednisone as the initial therapy of choice. J Rheumatol 1999; 26: 1134-9.
- 40. HELLMICH B, FLOSSMANN O, GROSS WL et al.: EULAR recommendations for conducting clinical studies and/or clinical trials in systemic vasculitis: focus on anti-neutrophil cytoplasm antibody-associated vasculitis. Ann Rheum Dis 2007; 66: 605-17.
- 41. STEGEMAN CA, TERVAERT JW, DE JONG PE, KALLENBERG CG: Trimethoprim-sulfamethoxazole (co-trimoxazole) for the prevention of relapses of Wegener's granulomatosis. Dutch Co-Trimoxazole Wegener Study Group. N Engl J Med 1996; 335: 16-20.

- 42. SALLIOT C, VAN DER HEIJDE D: Long-term safety of methotrexate monotherapy in patients with rheumatoid arthritis: a systematic literature research. *Ann Rheum Dis* 2009; 68: 1100-4.
- 43. GOLDEN MR, KATZ RS, BALK RA, GOLDEN HE: The relationship of preexisting lung disease to the development of methotrexate pneumonitis in patients with rheumatoid arthritis. J Rheumatol 1995; 22: 1043-7.
- 44. TZELEPIS GE, KOKOSI M, TZIOUFAS A et al.: Prevalence and outcome of pulmonary fibrosis in microscopic polyangiitis. Eur Respir J 2010; 36: 116-21.
- 45. ARULKUMARAN N, PERISELNERIS N, GASKIN G et al.: Interstitial lung disease and ANCA-associated vasculitis: a retrospective observational cohort study. *Rheumatology* (Oxford) 2011; 50: 2035-43.
- 46. DE GROOT K, MUHLER M, REINHOLD-KEL-LER E, PAULSEN J, GROSS WL: Induction of remission in Wegener's granulomatosis with low dose methotrexate. *J Rheumatol* 1998; 25: 492-5.
- 47. REINHOLD-KELLER E, FINK CO, HERLYN K, GROSS WL, DE GROOT K: High rate of renal relapse in 71 patients with Wegener's granulomatosis under maintenance of remission with low-dose methotrexate. *Arthritis Rheum* 2002; 47: 326-32.
- 48. FAURSCHOU M, WESTMAN K, RASMUSSEN N et al.: Long-term outcome of a clinical trial comparing methotrexate to cyclophosphamide for remission induction of early systemic ANCA-associated vasculitis. Arthritis Rheum 2012.
- 49. FURST DE: Practical clinical pharmacology and drug interactions of low-dose methotrexate therapy in rheumatoid arthritis. *Br J Rheumatol* 1995; 34 (Suppl. 2): 20-5.
- NASSONOV EL, SAMSONOV MY, TILZ GP et al.: Serum concentrations of neopterin, soluble interleukin 2 receptor, and soluble tumor necrosis factor receptor in Wegener's granulomatosis. J Rheumatol 1997; 24: 666-70.
- JONASDOTTIR O, PETERSEN J, BENDTZEN K: Tumour necrosis factor-alpha (TNF), lymphotoxin and TNF receptor levels in serum from patients with Wegener's granulomatosis. *APMIS* 2001; 109: 781-6.
- NORONHA IL, KRUGER C, ANDRASSY K, RITZ E, WALDHERR R: In situ production of TNF-alpha, IL-1 beta and IL-2R in ANCApositive glomerulonephritis. *Kidney Int* 1993; 43: 682-92.
- 53. KINDLER V, SAPPINO AP, GRAU GE, PIGUET PF, VASSALLI P: The inducing role of tumor necrosis factor in the development of bactericidal granulomas during BCG infection. *Cell* 1989; 56: 731-40.
- 54. FALK RJ, TERRELL RS, CHARLES LA, JEN-NETTE JC: Anti-neutrophil cytoplasmic autoantibodies induce neutrophils to degranulate and produce oxygen radicals in vitro. *Proc Natl Acad Sci U S A* 1990; 87; 4115-9.
- 55. STONE JH, UHLFELDER ML, HELLMANN DB, CROOK S, BEDOCS NM, HOFFMAN GS: Etanercept combined with conventional treatment in Wegener's granulomatosis: a six-month open-label trial to evaluate safety. Arthritis Rheum 2001; 44: 1149-54.

- LANGFORD C, TALAR-WILLIAMS C, BAR-RON K, MCCABE K, SNELLER M: Phase I/II trial of etanercept in Wegener's granulomatosis (WG): safety and preliminary experience [abstract]. Arthritis and Rheum 2000; 43: S163.
- 57. SILVA F, SEO P, SCHROEDER DR *et al.*: Solid malignancies among etanercept-treated patients with granulomatosis with polyangiitis (Wegener's): long-term followup of a multicenter longitudinal cohort. *Arthritis Rheum* 2011; 63: 2495-503.
- LAMPRECHT P, VOSWINKEL J, LILIENTHAL T et al.: Effectiveness of TNF-alpha blockade with infliximab in refractory Wegener's granulomatosis. *Rheumatology (Oxford)* 2002; 41: 1303-7.
- 59. BOOTH A, HARPER L, HAMMAD T et al.: Prospective study of TNFalpha blockade with infliximab in anti-neutrophil cytoplasmic antibody-associated systemic vasculitis. J Am Soc Nephrol 2004; 15: 717-21.
- BARTOLUCCI P, RAMANOELINA J, COHEN P et al.: Efficacy of the anti-TNF-alpha antibody infliximab against refractory systemic vasculitides: an open pilot study on 10 patients. *Rheumatology* (Oxford) 2002; 41: 1126-32.
- JOSSELIN L, MAHR A, COHEN P et al.: Infliximab efficacy and safety against refractory systemic necrotising vasculitides: longterm follow-up of 15 patients. Ann Rheum Dis 2008; 67: 1343-6.
- MORGAN MD, DRAYSON MT, SAVAGE CO, HARPER L: Addition of infliximab to standard therapy for ANCA-associated vasculitis. *Nephron Clin Pract* 2011; 117: c89-97.
- 63. DE MENTHON M, COHEN P, PAGNOUX C et al.: Infliximab or rituximab for refractory Wegener's granulomatosis: long-term follow up. A prospective randomised multicentre study on 17 patients. Clin Exp Rheumatol 2011; 29: S63-71.
- 64. LIN J, ZIRING D, DESAI S *et al.*: TNFalpha blockade in human diseases: an overview of efficacy and safety. *Clin Immunol* 2008; 126: 13-30.
- WONG M, ZIRING D, KORIN Y et al.: TNFalpha blockade in human diseases: mechanisms and future directions. *Clin Immunol* 2008; 126: 121-36.
- 66. MORELAND LW, BAUMGARTNER SW, SCHIFF MH *et al.*: Treatment of rheumatoid arthritis with a recombinant human tumor necrosis factor receptor (p75)-Fc fusion protein. *N Engl J Med* 1997; 337: 141-7.
- 67. LIPSKY PE, VAN DER HEIJDE DM, ST CLAIR EW et al.: Infliximab and methotrexate in the treatment of rheumatoid arthritis. Anti-Tumor Necrosis Factor Trial in Rheumatoid Arthritis with Concomitant Therapy Study Group. N Engl J Med 2000; 343: 1594-602.
- 68. VAN DEN BRANDE JM, BRAAT H, VAN DEN BRINK GR et al.: Infliximab but not etanercept induces apoptosis in lamina propria T-lymphocytes from patients with Crohn's disease. Gastroenterology 2003; 124: 1774-85.
- 69. LAURINO S, CHAUDHRY A, BOOTH A, CONTE G, JAYNE D: Prospective study of TNFalpha blockade with adalimumab in ANCA-associated systemic vasculitis with

REVIEW

renal involvement. *Nephrol Dial Transplant* 2010; 25: 3307-14.

- 70. COLE E, CATTRAN D, MAGIL A *et al.*: A prospective randomized trial of plasma exchange as additive therapy in idiopathic crescentic glomerulonephritis. The Canadian Apheresis Study Group. *Am J Kidney Dis* 1992; 20: 261-9.
- 71. GUILLEVIN L, LHOTE F, COHEN P et al.: Corticosteroids plus pulse cyclophosphamide and plasma exchanges versus corticosteroids plus pulse cyclophosphamide alone in the treatment of polyarteritis nodosa and Churg-Strauss syndrome patients with factors predicting poor prognosis. A prospective, randomized trial in sixty-two patients. *Arthritis Rheum* 1995; 38: 1638-45.
- PUSEY CD, REES AJ, EVANS DJ, PETERS DK, LOCKWOOD CM: Plasma exchange in focal necrotizing glomerulonephritis without anti-GBM antibodies. *Kidney Int* 1991; 40: 757-63.
- 73. WALSH M, CATAPANO F, SZPIRT W et al.: Plasma exchange for renal vasculitis and idiopathic rapidly progressive glomerulonephritis: a meta-analysis. Am J Kidney Dis 2011: 57: 566-74.
- 74. ZAUNER I, BACH D, BRAUN N et al.: Predictive value of initial histology and effect of plasmapheresis on long-term prognosis of rapidly progressive glomerulonephritis. Am J Kidney Dis 2002; 39: 28-35.
- 75. JAYNE DR, GASKIN G, RASMUSSEN N et al.: Randomized trial of plasma exchange or high-dosage methylprednisolone as adjunctive therapy for severe renal vasculitis. J Am Soc Nephrol 2007; 18: 2180-8.
- GLASSOCK RJ: Intensive plasma exchange in crescentic glomerulonephritis: help or no help? Am J Kidney Dis 1992; 20: 270-5.
- 77. ANDERSON G, COLES ET, CRANE M et al.: Wegener's granuloma. A series of 265 British cases seen between 1975 and 1985. A report by a sub-committee of the British Thoracic Society Research Committee. Q J Med 1992; 83: 427-38.
- CORDIER JF, VALEYRE D, GUILLEVIN L, LOIRE R, BRECHOT JM: Pulmonary Wegener's granulomatosis. A clinical and imaging study of 77 cases. *Chest* 1990; 97: 906-12.
- 79. GUILLEVIN L, DURAND-GASSELIN B, CEVALLOS R *et al.*: Microscopic polyangiitis: clinical and laboratory findings in eighty-five patients. *Arthritis Rheum* 1999; 42: 421-30.
- BOSCH X, FONT J, MIRAPEIX E, REVERT L, INGELMO M, URBANO-MARQUEZ A: Antimyeloperoxidase autoantibody-associated necrotizing alveolar capillaritis. *Am Rev Respir Dis* 1992; 146: 1326-9.
- MACKAY J: Rapidly progressive pulmonary haemorrhage in a case of microscopic polyangiitis. *BMJ Case Rep* 2011; 2011.
- 82. WANG CC, SHIANG JC, TSAI MK et al.: Prompt plasmapheresis successfully rescue pulmonary-renal syndrome caused by ANCA-negative microscopic polyangiitis. *Clin Rheumatol* 2009; 28: 1457-60.
- 83. AZUMA M, SASAKI S, MOCHIDUKI Y et al.: [A case of ANCA-associated vasculitis with diffuse alveolar hemorrhage successfully treated by plasmapheresis]. Nihon Kokyuki

Gakkai Zasshi 2007; 45: 21-5.

- 84. SUGIMOTO T, DEJI N, KUME S et al.: Pulmonary-renal syndrome, diffuse pulmonary hemorrhage and glomerulonephritis, associated with Wegener's granulomatosis effectively treated with early plasma exchange therapy. *Intern Med* 2007; 46: 49-53.
- 85. NGUYEN T, MARTIN MK, INDRIKOVS AJ: Plasmapheresis for diffuse alveolar hemorrhage in a patient with Wegener's granulomatosis: case report and review of the literature. J Clin Apher 2005; 20: 230-4.
- 86. PONS-ESTEL GJ, SALERNI GE, SERRANO RM et al.: Therapeutic plasma exchange for the management of refractory systemic autoimmune diseases: report of 31 cases and review of the literature. Autoimmun Rev 2011; 10: 679-84.
- 87. SASAKI S, MOCHIZUKI Y, NAKAHARA Y et al.: [Seven cases of diffuse alveolar hemorrhage with acute respiratory failure treated by plasma exchange]. Nihon Kokyuki Gakkai Zasshi 2010; 48: 10-6.
- GALLAGHER H, KWAN JT, JAYNE DR: Pulmonary renal syndrome: a 4-year, singlecenter experience. *Am J Kidney Dis* 2002; 39: 42-7.
- 89. KLEMMER PJ, CHALERMSKULRAT W, REIF MS, HOGAN SL, HENKE DC, FALK RJ: Plasmapheresis therapy for diffuse alveolar hemorrhage in patients with small-vessel vasculitis. *Am J Kidney Dis* 2003; 42: 1149-53.
- 90. FAUCI AS, HAYNES BF, KATZ P, WOLFF SM: Wegener's granulomatosis: prospective clinical and therapeutic experience with 85 patients for 21 years. *Ann Intern Med* 1983; 98: 76-85.
- DE GROOT K, ADU D, SAVAGE CO: The value of pulse cyclophosphamide in ANCA-associated vasculitis: meta-analysis and critical review. *Nephrol Dial Transplant* 2001; 16: 2018-27.
- 92. DE GROOT K, HARPER L, JAYNE D *et al.*: Pulse versus daily oral cyclophosphamide for induction of remission in antineutrophil cytoplasmic antibody-associated vasculitis: a randomized trial. *Ann Intern Med* 2009; 150: 670-80.
- 93. JONES RB, TERVAERT JW, HAUSER T et al.: Rituximab versus cyclophosphamide in ANCA-associated renal vasculitis. N Engl J Med 2010; 363: 211-20.
- 94. STONE JH, MERKEL PA, SPIERA R et al.: Rituximab versus cyclophosphamide for ANCA-associated vasculitis. N Engl J Med 2010; 363: 221-32.
- ALBAMA, FLORES-SUAREZ LF: [Rituximab for the treatment of ANCA associated vasculitis: the future today?]. *Reumatol Clin* 2011; 7 Suppl 3: S41-6.
- 96. HOGAN SL, FALK RJ, CHIN H et al.: Predictors of relapse and treatment resistance in antineutrophil cytoplasmic antibody-associated small-vessel vasculitis. Ann Intern Med 2005; 143: 621-31.
- 97. MUKHTYAR C, FLOSSMANN O, HELLMICH B et al.: Outcomes from studies of antineutrophil cytoplasm antibody associated vasculitis: a systematic review by the European League Against Rheumatism systemic vasculitis task force. Ann Rheum Dis 2008; 67: 1004-10.

- 98. GAYRAUD M, GUILLEVIN L, COHEN P et al.: Treatment of good-prognosis polyarteritis nodosa and Churg-Strauss syndrome: comparison of steroids and oral or pulse cyclophosphamide in 25 patients. French Cooperative Study Group for Vasculitides. Br J Rheumatol 1997; 36: 1290-7.
- 99. GAYRAUD M, GUILLEVIN L, LE TOUMELIN P et al.: Long-term followup of polyarteritis nodosa, microscopic polyangiitis, and Churg-Strauss syndrome: analysis of four prospective trials including 278 patients. Arthritis Rheum 2001; 44: 666-75.
- 100. SEROR R, PAGNOUX C, RUIVARD M et al.: Treatment strategies and outcome of induction-refractory Wegener's granulomatosis or microscopic polyangiitis: analysis of 32 patients with first-line induction-refractory disease in the WEGENT trial. Ann Rheum Dis 2010; 69: 2125-30.
- 101. HARPER L, MORGAN MD, WALSH M et al.: Pulse versus daily oral cyclophosphamide for induction of remission in ANCA-associated vasculitis: long-term follow-up. Ann Rheum Dis 2012; 71: 955-60.
- 102. SPECKS U, STONE JH: Long-term effi cacy and safety results of the RAVE trial. *Clin EXp Immunol* 2011; 164: 65.
- 103. HOLLE JU, GROSS WL, LATZA U *et al.*: Improved outcome in 445 patients with Wegener's granulomatosis in a German vasculitis center over four decades. *Arthritis Rheum* 2011; 63: 257-66.
- 104. PAGNOUX C, HOGAN SL, CHIN H et al.: Predictors of treatment resistance and relapse in antineutrophil cytoplasmic antibody-associated small-vessel vasculitis: comparison of two independent cohorts. Arthritis Rheum 2008; 58: 2908-18.
- 105. PIERROT-DESEILLIGNY DESPUJOL C, POU-CHOT J, PAGNOUX C, COSTE J, GUILLEVIN L: Predictors at diagnosis of a first Wegener's granulomatosis relapse after obtaining complete remission. *Rheumatology* (Oxford) 2010; 49: 2181-90.
- 106. LANGFORD CA: Cyclophosphamide as induction therapy for Wegener's granulomatosis and microscopic polyangiitis. *Clin Exp Immunol* 2011; 164 (Suppl. 1): 31-4.
- 107. LANGFORD CA, TALAR-WILLIAMS C, BAR-RON KS, SNELLER MC: A staged approach to the treatment of Wegener's granulomatosis: induction of remission with glucocorticoids and daily cyclophosphamide switching to methotrexate for remission maintenance. *Arthritis Rheum* 1999; 42: 2666-73.
- 108. ALLISON AC: Mechanisms of action of mycophenolate mofetil. *Lupus* 2005; 14 (Suppl. 1): s2-8.
- 109. SOLLINGER HW: Mycophenolate mofetil for the prevention of acute rejection in primary cadaveric renal allograft recipients. U.S. Renal Transplant Mycophenolate Mofetil Study Group. *Transplantation* 1995; 60: 225-32.
- 110. LANG P, PARDON A, AUDARD V: Long-term benefit of mycophenolate mofetil in renal transplantation. *Transplantation* 2005; 79: S47-8.
- 111. APPEL GB, CONTRERAS G, DOOLEY MA et al.: Mycophenolate mofetil versus cyclophosphamide for induction treatment of

lupus nephritis. J Am Soc Nephrol 2009; 20: 1103-12.

- 112. GINZLER EM, DOOLEY MA, ARANOW C et al.: Mycophenolate mofetil or intravenous cyclophosphamide for lupus nephritis. N Engl J Med 2005; 353: 2219-28.
- 113. KAMANAMOOL N, MCEVOY M, ATTIA J, INGSATHIT A, NGAMJANYAPORN P, THAK-KINSTIAN A: Efficacy and adverse events of mycophenolate mofetil versus cyclophosphamide for induction therapy of lupus nephritis: systematic review and meta-analysis. *Medicine* (Baltimore) 2010; 89: 227-35.
- 114. STASSEN PM, TERVAERT JW, STEGEMAN CA: Induction of remission in active antineutrophil cytoplasmic antibody-associated vasculitis with mycophenolate mofetil in patients who cannot be treated with cyclophosphamide. Ann Rheum Dis 2007; 66: 798-802.
- 115. HU W, LIU C, XIE H, CHEN H, LIU Z, LI L: Mycophenolate mofetil versus cyclophosphamide for inducing remission of ANCA vasculitis with moderate renal involvement. *Nephrol Dial Transplant* 2008; 23: 1307-12.
- 116. JOY MS, HOGAN SL, JENNETTE JC, FALK RJ, NACHMAN PH: A pilot study using mycophenolate mofetil in relapsing or resistant ANCA small vessel vasculitis. *Nephrol Dial Transplant* 2005; 20: 2725-32.
- 117. KOUKOULAKI M, JAYNE DR: Mycophenolate mofetil in anti-neutrophil cytoplasm antibodies-associated systemic vasculitis. *Nephron Clin Pract* 2006; 102: c100-7.
- 118. PESAVENTO T, FALKENHAIN M, ROVIN B, HEBERT L: Mycophenolate mofetil therapy (MMF) in anti-neutrophil cytoplasmic antibody (ANCA) vascultis [abstract]. J Am Soc Nephrol 1999; 10: 114A.
- 119. HAIDINGER M, NEUMANN I, GRUTZMACH-ER H, BAYER P, MEISL F: Mycophenolate mofetil (MMF) treatment of ANCA-assocaited small-vessel vasculitis: a pharmacokinetically controlled study. [abstract]. Clin Exp Immunol 2000; 120 (Suppl. 1): 72.
- 120. IATROU C, ZERBALA S, REVELA I et al.: Mycophenolate mofetil as maintenance therapy in patients with vasculitis and renal involvement. *Clin Nephrol* 2009; 72: 31-7.
- 121. HIEMSTRA TF, WALSH M, MAHR A et al.: Mycophenolate mofetil vs azathioprine for remission maintenance in antineutrophil cytoplasmic antibody-associated vasculitis: a randomized controlled trial. JAMA 2010; 304: 2381-8.
- 122. JONES R, WALSH M: A Randomised Trial of Mycophenolate Mofetil versus Cyclophosphamide for Remission Induction in ANCA-Associated Vasculitis: MYCYC [Abstract]. J Am Soc Nephrol 2012; 23.
- 123. LANGFORD CA, TALAR-WILLIAMS C, SNELLER MC: Mycophenolate mofetil for remission maintenance in the treatment of Wegener's granulomatosis. *Arthritis Rheum* 2004; 51: 278-83.
- 124. NOWACK R, GOBEL U, KLOOKER P, HERGES-ELL O, ANDRASSY K, VAN DER WOUDE FJ: Mycophenolate mofetil for maintenance therapy of Wegener's granulomatosis and microscopic polyangiitis: a pilot study in 11

patients with renal involvement. J Am Soc Nephrol 1999; 10: 1965-71.

- 125. TALARICO R, BALDINI C, DELLA ROSSA A et al.: Large- and small-vessel vasculitis: a critical digest of the 2010-2011 literature. *Clin Exp Rheumatol* 2012; 30: S130-8.
- 126. ARIES PM, HELLMICH B, VOSWINKEL J et al.: Lack of efficacy of rituximab in Wegener's granulomatosis with refractory granulomatous manifestations. Ann Rheum Dis 2006; 65: 853-8.
- 127. BRIHAYE B, AOUBA A, PAGNOUX C, VIG-NAUX O, LE HELLO C, GUILLEVIN L: Rituximab reversed cardiac involvement of Wegener's granulomatosis: magnetic resonance imaging assessment. *Presse Med* 2008; 37: 412-5.
- 128. ERIKSSON P: Nine patients with anti-neutrophil cytoplasmic antibody-positive vasculitis successfully treated with rituximab. *J Intern Med* 2005; 257: 540-8.
- 129. HENES JC, FRITZ J, KOCH S *et al.*: Rituximab for treatment-resistant extensive Wegener's granulomatosis--additive effects of a maintenance treatment with leflunomide. *Clin Rheumatol* 2007; 26: 1711-5.
- 130. JONES RB, FERRARO AJ, CHAUDHRY AN *et al.*: A multicenter survey of rituximab therapy for refractory antineutrophil cytoplasmic antibody-associated vasculitis. *Arthritis Rheum* 2009; 60: 2156-68.
- 131. KEOGH KA, WYLAM ME, STONE JH, SPECKS U: Induction of remission by B lymphocyte depletion in eleven patients with refractory antineutrophil cytoplasmic antibody-associated vasculitis. *Arthritis Rheum* 2005; 52: 262-8.
- 132. KEOGH KA, YTTERBERG SR, FERVENZA FC, CARLSON KA, SCHROEDER DR, SPECKS U: Rituximab for refractory Wegener's granulomatosis: report of a prospective, open-label pilot trial. Am J Respir Crit Care Med 2006; 173: 180-7.
- 133. LOVRIC S, ERDBRUEGGER U, KUMPERS P et al.: Rituximab as rescue therapy in antineutrophil cytoplasmic antibody-associated vasculitis: a single-centre experience with 15 patients. Nephrol Dial Transplant 2009; 24: 179-85.
- 134. MANSFIELD N, HAMOUR S, MARIE-HABIB A *et al.*: Prolonged disease-free remission following rituximab and low-dose cyclophosphamide therapy for renal ANCA-associated vasculitis. *Nephrol Dial Transplant* 2011.
- 135. OMDAL R, WILDHAGEN K, HANSEN T, GUNNARSSON R, KRISTOFFERSEN G: Anti-CD20 therapy of treatment-resistant Wegener's granulomatosis: favourable but temporary response. *Scand J Rheumatol* 2005; 34: 229-32.
- 136. ROCCATELLO D, BALDOVINO S, ROSSI D et al.: Long-term effects of anti-CD20 monoclonal antibody treatment of cryoglobulinaemic glomerulonephritis. Nephrol Dial Transplant 2004; 19: 3054-61.
- 137. SANCHEZ-CANO D, CALLEJAS-RUBIO JL, ORTEGO-CENTENO N: Effect of rituximab on refractory Wegener granulomatosis with predominant granulomatous disease. *J Clin Rheumatol* 2008; 14: 92-3.
- 138. SEO P, SPECKS U, KEOGH KA: Efficacy of

rituximab in limited Wegener's granulomatosis with refractory granulomatous manifestations. *J Rheumatol* 2008; 35: 2017-23.

- 139. SMITH KG, JONES RB, BURNS SM, JAYNE DR: Long-term comparison of rituximab treatment for refractory systemic lupus erythematosus and vasculitis: Remission, relapse, and re-treatment. *Arthritis Rheum* 2006; 54: 2970-82.
- 140. STASI R, STIPA E, DEL POETA G, AMADORI S, NEWLAND AC, PROVAN D: Long-term observation of patients with anti-neutrophil cytoplasmic antibody-associated vasculitis treated with rituximab. *Rheumatology* (Oxford) 2006; 45: 1432-6.
- 141. TAMURA N, MATSUDAIRA R, HIRASHIMA M *et al.*: Two cases of refractory Wegener's granulomatosis successfully treated with rituximab. *Intern Med* 2007; 46: 409-14.
- 142. FORTIN PM, TEJANI AM, BASSETT K, MUSINI VM: Intravenous immunoglobulin as adjuvant therapy for Wegener's granulomatosis. *Cochrane Database Syst Rev* 2009; CD007057.
- 143. JAYNE DR, CHAPEL H, ADU D et al.: Intravenous immunoglobulin for ANCA-associated systemic vasculitis with persistent disease activity. QJM 2000; 93: 433-9.
- 144. JAYNE DR, LOCKWOOD CM: Intravenous immunoglobulin as sole therapy for systemic vasculitis. *Br J Rheumatol* 1996; 35: 1150-3.
- 145. MARTINEZ V, COHEN P, PAGNOUX C et al.: Intravenous immunoglobulins for relapses of systemic vasculitides associated with antineutrophil cytoplasmic autoantibodies: results of a multicenter, prospective, openlabel study of twenty-two patients. *Arthritis Rheum* 2008; 58: 308-17.
- 146. RICHTER C, SCHNABEL A, CSERNOK E, DE GROOT K, REINHOLD-KELLER E, GROSS WL: Treatment of anti-neutrophil cytoplasmic antibody (ANCA)-associated systemic vasculitis with high-dose intravenous immunoglobulin. *Clin Exp Immunol* 1995; 101: 2-7.
- 147. MUZAFFAR J, KATRAGADDA L, HAIDER S, JAVEDA, ANAISSIE E, USMANI S: Rituximab and intravenous immunoglobulin (IVIG) for the management of acquired factor VIII inhibitor in multiple myeloma: case report and review of literature. *Int J Hematol* 2012; 95: 102-6.
- 148. VO AA, LUKOVSKY M, TOYODA M et al.: Rituximab and intravenous immune globulin for desensitization during renal transplantation. N Engl J Med 2008; 359: 242-51.
- 149. HONG YA, KIM HG, CHOI SR *et al.*: Effectiveness of rituximab and intravenous immunoglobulin therapy in renal transplant recipients with chronic active antibody-mediated rejection. *Transplant Proc* 2012; 44: 182-4.
- 150. FELDMAN RJ, CHRISTEN WG, AHMED AR: Comparison of immunological parameters in patients with pemphigus vulgaris following rituximab and IVIG therapy. Br J Dermatol 2012; 166: 511-7.
- 151. AHMED AR, SPIGELMAN Z, CAVACINI LA, POSNER MR: Treatment of pemphigus vulgaris with rituximab and intravenous im-

mune globulin. N Engl J Med 2006; 355: 1772-9.

- 152. BAYRY J, NEGI VS, KAVERI SV: Intravenous immunoglobulin therapy in rheumatic diseases. *Nat Rev Rheumatol* 2011; 7: 349-59.
- 153. KESSEL A, ROSNER I, TOUBI E: Rituximab: beyond simple B cell depletion. *Clin Rev Allergy Immunol* 2008; 34: 74-9.
- 154. KESSEL A, AMMURI H, PERI R et al.: Intravenous immunoglobulin therapy affects T regulatory cells by increasing their suppressive function. J Immunol 2007; 179: 5571-5.
- 155. SAMUELSSON A, TOWERS TL, RAVETCH JV: Anti-inflammatory activity of IVIG mediated through the inhibitory Fc receptor. *Science* 2001; 291: 484-6.
- 156. TEELING JL, JANSEN-HENDRIKS T, KUI-JPERS TW *et al.*: Therapeutic efficacy of intravenous immunoglobulin preparations depends on the immunoglobulin G dimers: studies in experimental immune thrombocytopenia. *Blood* 2001; 98: 1095-9.
- 157. JAYNE DR, DAVIES MJ, FOX CJ, BLACK CM, LOCKWOOD CM: Treatment of systemic vasculitis with pooled intravenous immunoglobulin. *Lancet* 1991; 337: 1137-9.
- 158. DIETRICH G, KAVERI SV, KAZATCHKINE MD: Modulation of autoimmunity by intravenous immune globulin through interac-

tion with the function of the immune/idiotypic network. *Clin Immunol Immunopathol* 1992; 62: S73-81.

- 159. KAVERI SV, DIETRICH G, HUREZ V, KAZAT-CHKINE MD: Intravenous immunoglobulins (IVIg) in the treatment of autoimmune diseases. *Clin Exp Immunol* 1991; 86: 192-8.
- 160. CHEN M, DAHA MR, KALLENBERG CG: The complement system in systemic autoimmune disease. J Autoimmun 2010; 34: J276-86.
- 161. FLORES-SUAREZ LF: [The complement system in the pathogenesis of antineutrophil cytoplasm antibodies-associated vasculitis]. *Reumatol Clin* 2011; 7 Suppl 3: S18-21.
- 162. VAN TIMMEREN MM, CHEN M, HEERINGA P: Review article: Pathogenic role of complement activation in anti-neutrophil cytoplasmic auto-antibody-associated vasculitis. *Nephrology* (Carlton) 2009; 14: 16-25.
- 163. XIAO H, HEERINGA P, HU P et al.: Antineutrophil cytoplasmic autoantibodies specific for myeloperoxidase cause glomerulonephritis and vasculitis in mice. J Clin Invest 2002; 110: 955-63.
- 164. SCHREIBER A, XIAO H, JENNETTE JC, SCHNEIDER W, LUFT FC, KETTRITZ R: C5a receptor mediates neutrophil activation and ANCA-induced glomerulonephritis.

J Am Soc Nephrol 2009; 20: 289-98.

- 165. HUUGEN D, VAN ESCH A, XIAO H et al.: Inhibition of complement factor C5 protects against anti-myeloperoxidase antibody-mediated glomerulonephritis in mice. *Kidney Int* 2007; 71: 646-54.
- 166. XIAO H, SCHREIBER A, HEERINGA P, FALK RJ, JENNETTE JC: Alternative complement pathway in the pathogenesis of disease mediated by anti-neutrophil cytoplasmic autoantibodies. *Am J Pathol* 2007; 170: 52-64.
- 167. CHEN M, XING GQ, YU F, LIU G, ZHAO MH: Complement deposition in renal histopathology of patients with ANCA-associated pauci-immune glomerulonephritis. *Nephrol Dial Transplant* 2009; 24: 1247-52.
- 168. XING GQ, CHEN M, LIU G et al.: Complement activation is involved in renal damage in human antineutrophil cytoplasmic autoantibody associated pauci-immune vasculitis. J Clin Immunol 2009; 29: 282-91.
- 169. YUAN J, GOU SJ, HUANG J, HAO J, CHEN M, ZHAO MH: C5a and its receptors in human anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis. *Arthritis Res Ther* 2012; 14: R140.
- 170. WOODRUFF TM, NANDAKUMAR KS, TED-ESCO F: Inhibiting the C5-C5a receptor axis. *Mol Immunol* 2011; 48: 1631-42.