### The role of asymmetric dimethylarginine alone and in combination with N-terminal pro-B-type natriuretic peptide as a screening biomarker for systemic sclerosis-related pulmonary arterial hypertension: a case control study

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*E-mail: m.nikpour@unimelb.edu.au Received on April 14, 2015; accepted in* 

revised form on January 8, 2016. Clin Exp Rheumatol 2016; 34 (Suppl. 100): S129-S136.

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**Key words:** systemic sclerosis, pulmonary arterial hypertension, biomarker, screening, asymmetric dimethylarginine (ADMA)

For funding and competing interests see page S-134.

#### ABSTRACT

**Objective.** Asymmetric dimethylarginine (ADMA) is a novel biomarker of endothelial cell dysfunction. In this proof of concept study, we sought to evaluate the role of ADMA as a screening biomarker for incident systemic sclerosis-related pulmonary arterial hypertension (SSc-PAH).

Methods. ADMA levels were measured using high performance liquid chromatography in 15 consecutive treatmentnaive patients with newly-diagnosed SSc-PAH and compared with 30 SSccontrols without PAH. Logistic regression models were used to evaluate the independent association of ADMA with PAH. The optimal cut-point of ADMA for SSc-PAH screening was determined. NT-proBNP levels were previously measured in the same patients and the optimal cut-point of NT-proBNP of  $\geq$ 210ng/mL was coupled with the optimal cut-point of ADMA to create a screening model that combined the two biomarkers.

Results. The PAH group had significantly higher mean ADMA levels than the control group (0.76±0.14 µM versus 0.59±0.07 µM; p<0.0001). ADMA levels remained significantly associated with PAH after the adjustment for specific disease characteristics, cardiovascular risk factors and other SSc-related vascular complications (all p < 0.01). An ADMA level  $\geq 0.7 \ \mu M$  had a sensitivity of 86.7%, specificity of 90.0% and AUC of 0.86 for diagnosing PAH. A screening model that combined an NT-proBNP ≥210ng/mL and/or ADMA  $\geq 0.7$  ng/mL resulted in a sensitivity of 100% and specificity of 90% for the detection of SSc-PAH.

**Conclusion.** In this small study, use of ADMA in combination with NT-proB-

NP produced excellent sensitivity and specificity for the non-invasive identification of SSc-PAH. The role of ADMA as a screening biomarker for SSc-PAH merits further evaluation.

#### Introduction

Systemic sclerosis (SSc) is a heterogeneous, multisystem connective tissue disease characterised by autoantibodies, vasculopathy and fibrosis affecting the skin and a number of internal organs. Systemic sclerosis related pulmonary arterial hypertension (SSc-PAH) is a major cause of morbidity in this disease, conferring a significantly increased risk of death and accounting for approximately 30% of SSc related mortality (1-3). In this context, screening for PAH has emerged as an important consideration in the optimal management of patients with SSc (4). The rationale behind screening for SSc-PAH is based on the early identification of an aggressive but treatable complication in an at risk population, allowing prompt initiation of therapies that appear to offer significant clinical benefit. A recent French study showed higher 3-, 5- and 8-yr survival rates in patients identified by screening, compared with patients diagnosed during routine care, where symptoms and/ or signs directed further investigation (81%, 73% and 64% vs. 31%, 25% and 17%, respectively) (5).

Right heart catheterisation (RHC) is necessary for making a definitive diagnosis of SSc-PAH but is not feasible for general screening as it is invasive. Instead, current international guidelines recommend regular trans-thoracic echocardiography (TTE), with or without diffusing capacity for carbon monoxide (DLCO), for screening in order to

help identify high-risk individuals who should undergo RHC (6-8). However, current screening tests perform better when PAH is more advanced (9) as the high capacitance physiology of the pulmonary microcirculation means that it is only after >50% of the pulmonary vasculature is obstructed that the mean pulmonary artery pressure rises (10). Therefore, interest is turning to alternative strategies such as incorporating blood biomarkers reflecting the pathophysiology, pathogenesis or genetics of the condition, to assist in the risk stratification of SSc patients leading to the earlier detection of PAH.

In case control and cohort studies, we have previously evaluated the role of serum N-terminal pro-B-type natriuretic peptide (NT-proBNP), released as a result of myocardial stress, in screening for SSc-PAH and found a level  $\geq$  210ng/mL to have optimal properties for diagnosing PAH, with a sensitivity of 93.3%, specificity of 100% and AUC of 0.94 (11, 12).

Asymmetrical dimethylarginine (ADMA, or NG-NG-Dimethyl-L-arginine) is also a potential biomarker of interest in pulmonary hypertension (13). It is the principal endogenous inhibitor of nitric oxide synthase (NOS) and results in impaired NO production (see Figure 1) (14, 15). ADMA also directly affects endothelial cell gene expression including bone morphogenetic protein (BMP), which has been associated with idiopathic PAH (16). Monomethylarginine (MMA) and symmetrical dimethyarginine (SDMA) are released alongside ADMA in the process of arginine methylation, but only ADMA and MMA are thought to directly inhibit NOS. In SSc, Dimitroualis et al. reported significantly higher mean ADMA levels in SSc patients with a systolic pulmonary artery pressure on transthoracic echocardiography  $(sPAP_{TTE}) \ge 40 \text{ mmHg compared with}$ SSc patients with a sPAP<sub>TTE</sub> <40mmHg; however, the lack of RHC in this study was a major limitation (17). Elevated ADMA levels have also been reported in other types of pulmonary hypertension including idiopathic PAH (iPAH), chronic thromboembolic pulmonary (CTEPH), congenital hypertension

heart disease related PH and PAH secondary to sickle cell disease, suggesting that ADMA may be a biomarker of endothelial cell dysfunction in pulmonary hypertension (18-21).

The goal of this study was to evaluate the relationship between serum ADMA levels in SSc patients with and without PAH confirmed by RHC, and to determine the clinical utility of ADMA alone and in combination with NT-proBNP as a screening biomarker for incident SSc-PAH.

#### Patients and methods

Study design and population In this case-control study, sera of SSc patients with and without PAH were collected, analysed and compared. All patients fulfilled ACR criteria for SSc (22). All clinical parameters, TTE and pulmonary function tests (PFTs) including DLCO were assessed within three months of the serum collection and prospectively recorded in the Australian Scleroderma Cohort Study (ASCS) database as previously described (23). The ASCS is approved by the human research ethics committees of the 13 participating Australian centres, and patients provide written informed consent at recruitment.

Group 1 comprised patients with SSc-PAH and included 15 consecutive patients with a new diagnosis of RHC confirmed PAH based on a mean pulmonary artery pressure (mPAP)  $\geq$ 25mmHg and pulmonary capillary wedge pressure (PCWP)  $\leq$ 15mmHg. These patients had no more than minor changes of interstitial lung disease (ILD) on high-resolution CT lung (HRCT).

Group 2 (n=30) were SSc-controls who had no evidence of cardiopulmonary complications, based on sPAP<sub>TTE</sub> <30mmHg, normal myocardial function on TTE, DLCO corrected for haemoglobin (DLCO<sub>corr</sub>) >70% predicted, (forced expiratory volume in one second/forced vital capacity (FEV1/FVC) % predicted >0.7, no ILD on HRCT (and in those without an HRCT, FVC ≥80% predicted), and World Health Organisation Functional Class (WHO-FC) I or II.

Exclusion criteria for both groups included the presence of abnormal left ventricular systolic or diastolic function for age measured at TTE, abnormal left atrial size, an unrecordable tricuspid regurgitant Doppler signal, and estimated glomerular filtration rate (eGFR) <30ml/min.

#### Cardiac and pulmonary assessments

Left ventricular systolic and diastolic function was determined by 2-dimensional TTE performed within three months of collection of serum. The sPAP<sub>TTE</sub> was measured at rest, based on peak velocity of the tricuspid regurgitant jet and estimation of right atrial pressure of 5-10 mmHg based on the diameter and respiratory variation of the inferior vena cava. TTE was performed only at tertiary centres for SSc assessment. Pulmonary involvement was assessed by PFT and/or HRCT within 3 months of serum collection. HRCT were reported as no, mild, moderate or severe ILD by a radiologist. All DLCO<sub>corr</sub> (ml/mmHg/min) values are reported as % predicted values, corrected for haemoglobin. All FEV1 (litres), FVC (litres) and FVC/DLCO<sub>corr</sub> values are reported as percentage predicted for sex, race and height (24).

#### Serum samples

All sera were collected from patients within three months of their annual clinical assessment and cardiopulmonary investigations. All PAH patients (Group 1) had serum collected for ADMA measurement at the time of their RHC and prior to the commencement of pulmonary vasodilator therapy. Blood samples were collected at rest into tubes containing EDTA. Samples were centrifuged and stored at -80°C until used.

#### Measurement of ADMA, SDMA and L-arginine levels

ADMA, SDMA and L-arginine levels were determined by High Performance Liquid Chromatography (HPLC) with Solid Phase Extraction (SPE) at the Cardiology Unit, The Queen Elizabeth Hospital, University of Adelaide, using an established, published method (25). HPLC with SPE is one of the most accessible methods for ADMA determination, and very high accuracy can be



Fig. 1. NO metabolism and the effects of ADMA

NOS converts L-Arginine to NO and L-citrulline. ADMA is the principal inhibitor of all NOS isoforms. ADMA (along with SDMA and MMA) is produced as a result of degradation of methylated proteins, and cleared predominantly by the action of dimethylarginine dimethylaminohydrolase (DDAH). ADMA competes with arginine and blocks NO production by acting as a false substrate for NOS. ADMA also influences endothelial cell gene expression, in particular BMP signalling. NOS: nitric oxide synthase; NO: nitric oxide; ADMA: asymmetric dimethylarginine; SDMA: symmetrical dimethylarginine; MMA:monomethylarginine; BMP:bone morphogenetic protein.

achieved with standardised, validated techniques (26). Briefly, ADMA and SDMA were extracted from serum by strong cation exchange (SCX), which was followed by fluorescent derivatisation. The resultant highly stable fluorescent derivatives were then processed by HPLC, which achieved an excellent separation of the arginine metabolites from internal standard and endogenous serum components. The interassay variability for ADMA/SDMA was 6%, and 8% for arginine (n=17 sets). The % recovery in spiked samples for ADMA/ SDMA was 90%, and 76% for arginine. The limit of detection for ADMA/ SDMA is 0.1 µM.

#### Statistical analysis

In this case-control study design, ADMA, SDMA and L-arginine levels in patients with RHC-proven SSc-PAH and SSc-controls were compared using the student t-test after appropriate transformations were performed to satisfy the assumptions of normality and homogeneity of variance. Logistic regression models were used to explore the independent association of ADMA with PAH, after the adjustment for each covariate in individual models. Important correlations of ADMA and SDMA were assessed using Spearman's rank correlation coefficient (rho). Receiver operator characteristic (ROC) curve analysis was used to determine the optimal cut-points that maximised desired test properties, namely sensitivity and negative predictive value. Based on ROC curve analysis, combination biomarker models were tested using contingency tables. All statistical analyses were performed using STATA 12.1 (Statacorp, College Station, TX, USA).

#### Results

## Characteristics of the patient population

ADMA levels were evaluated in 45 SSc patients, consisting of 15 treatmentnaïve SSc patients with PAH (group 1; SSc-PAH) and 30 SSc patients with no PAH (group 2; SSc-controls). Demographic, clinical and investigative characteristics of these patients are summarised in Tables I and II.

#### Comparison of ADMA levels: SSc-PAH vs. SSc-controls

ADMA levels were compared between groups (Fig. 2). The PAH group had significantly higher mean ADMA levels than the control group ( $0.76\pm0.14$  $\mu$ M vs. 0.59 $\pm0.07$   $\mu$ M; p<0.0001). Serum SDMA levels were also shown to be significantly higher in SSc-PAH ( $0.79\pm0.26$   $\mu$ M vs. 0.46 $\pm0.07$ ; p<0.0001) (Fig. 3). L-Arginine levels were significantly lower in SSc-PAH versus SSc-controls (97.28 $\pm27.40$  vs. 117.45 $\pm26.70\mu$ M, p=0.017). A significantly lower L-arginine to ADMA ratio was noted in SSc-PAH (p<0.0001).

# Adjusting ADMA levels for the influence of covariates

Individual logistic regression models were used to evaluate the independent association of ADMA with PAH, after the adjustment for each of the covariates in turn. Accordingly, due to a limited sample size, each model contained only two variables, ADMA level and the covariate. Covariates included clinical characteristics, co-morbidities, and vascular complications (see Table III). As older age and longer disease duration have been associated with PAH, and were available in the study cohort, the effect of these variables was adjusted for using logistic regression. ADMA was associated with PAH (OR =1.14, 95%CI: 1.03-1.25, p=0.008) after adjustment for age (OR =1.12, 95%CI: 1.01-1.24, p=0.031). Similarly, ADMA was associated with PAH (OR =1.16, 95%CI: 1.05-1.27, p=0.003) after adjustment for disease duration (OR 1.09, 95%CI: 0.98-1.20, v=0.104). ADMA levels remained significantly associated with PAH after adjustment for cardiovascular risk factors (including hypertension, hypercholesterolaemia, diabetes, current or previous smoking), vascular complications (including Raynaud's phenomenon and digital ulcers) as well as calcium channel blocker usage (see Table III). ADMA levels were also significantly associated with PAH after adjustment for disease subtype, modified Rodnan skin score and body mass index.

The independent associations of SDMA with PAH, after adjustment for covariates were similarly significant (Table IV).

# Correlations of ADMA with RHC parameters in SSc-PAH

The correlations of ADMA levels with RHC parameters were evaluated in SSc-PAH patients. There was no significant correlation of ADMA level with mPAP (rho =0.12, p=0.661), PVR (rho =0.30, p=0.341) or mRAP (rho =0.10, p=0.739). Similarly, SDMA did not correlate with mPAP (rho =-0.23, p=0.41), PVR (rho =0.13, p=0.68) or mRAP (rho =-0.004, p=0.99).

#### ROC curve analysis

As can be seen in Table V, an ADMA level  $\geq 0.694 \ \mu\text{M}$  for PAH *versus* controls, had a sensitivity of 86.7% (95%CI: 58.4-97.7%), specificity of 90.0% (95%CI: 72.3-97.4%), with an AUC of 0.86 (95%CI: 0.7-1.0) for diagnosing PAH. On the other hand, an SDMA level  $\geq 0.621 \mu\text{M}$  had a sensitivity of 80.0% and specificity of 100%, with an AUC of 88% for diagnosing PAH (95%CI: 74-100%).

#### Screening model and properties

The optimal biomarker cut-points derived from ROC curve analysis were used to form screening models for SSc-PAH versus SSc-controls (see Table V). The goal of developing these composite biomarker models was to maximise the sensitivity (*i.e.* eliminate false negatives), as would be required of a screening model, whilst maintaining specificity. As can be seen in Table V, the screening model that combined an NT-proBNP ≥210 ng/mL (as determined in our previous study (11)) and/ or ADMA ≥0.7 ng/mL resulted in a sensitivity of 100%, which was higher than either biomarker in isolation; the specificity of this model was 90%.

#### Discussion

The results of this study suggest that serum ADMA levels may serve as an important screening biomarker for SSc-PAH. Additionally, a composite biomarker screening algorithm, using NTproBNP in combination with ADMA, may achieve an excellent sensitivity (and also specificity) for the identification of SSc-PAH.

Serum ADMA levels, measured by HPLC, were shown to be significantly

Table I. Comparison of clinical characteristics between groups.

Characteristics	Group 1 SSc-PAH (mean ± SD)	Group 2 SSc-controls (mean ± SD)	<i>p</i> -value	
Number (n)	15	30	N/A	
Age at onset (y)	$44.5 \pm 12.9$	$40.6 \pm 13.2$	0.355	
Age at study (y)	$62.1 \pm 10.9$	$48.7 \pm 10.1$	< 0.0001	
Dis. duration (y)	$19.2 \pm 12.4$	$7.8 \pm 7.2$	0.0011	
Female, n (%)	12 (80)	30 (100)	0.011	
Male, n (%)	3 (20)	0		
Limited (n)	13	23	0.429	
Diffuse (n)	2	7		
ANA, n	14	29	0.160	
Anti-Scl70, n	1	5	0.333	
Anti-cent, n	7	16	0.592	
MRSS	$10.4 \pm 12.1$	$7.6 \pm 7.5$	0.432	
WHO FC				
-1	0	25	< 0.0001	
-2	2	5		
-3	11	0		
-4	2	0		

Dis. duration: disease duration; PAH: pulmonary arterial hypertension; ILD: interstitial lung disease; ANA: Anti-nuclear antibody; Anti-Scl70: anti- topoisomerase-1 antibody; Anti-cent: anti-centromere antibody; MRSS: modified Rodnan skin score; WHO FC: World Health Organisation functional class.

Table II.	Compariso	on of invest	stigation	parameters	between	groups
	-		0			0

Investigations	Group 1 PAH	Group 2 Controls	<i>p</i> -value	
TTE parameters				
TR vel (m/s)	$3.8 \pm 0.7$	$2.20 \pm 0.2$	< 0.0001	
sPAP (mmHg)	$65.8 \pm 27.3$	$26.3 \pm 2.6$	< 0.0001	
RHC results				
mPAP(mmHg)	$40.2 \pm 12.5$	N/A	N/A	
mRAP (mmHg)	$10.1 \pm 3.1$	N/A	N/A	
PVR (wood units)	$6.2 \pm 3.4$	N/A	N/A	
PFT				
FVC (% pred)	$75.5 \pm 24.3$	$102.8 \pm 13.4$	< 0.0005	
DLCO	$45.6 \pm 11.7$	$86.8 \pm 13.0$	< 0.0001	
FVC/DLCO <sub>corr</sub>	$1.76 \pm 0.38$	$1.20 \pm 0.20$	<0.0001	
6MWD (m)	$337 \pm 100$	$520 \pm 48.6$	< 0.0001	
C-reactive protein	$8.6 \pm 9.7$	$5.9 \pm 10.0$	0.379	
ESR	$24.7 \pm 13.3$	$12.4 \pm 14.5$	0.0094	

TRV: tricuspid regurgitant velocity; sPAP: systolic pulmonary artery pressure; mPAP: mean pulmonary artery pressure; mRAP: mean right atrial pressure; PVR: pulmonary vascular resistance; FVC: forced vital capacity (% predicted); DLCO: diffusion capacity of lung for carbon monoxide (% predicted); 6MWD: six minute walk distance; m: metres; Erythrocyte Sedimentation Rate (ESR); PAH: pulmonary arterial hypertension.

higher in SSc-PAH than SSc-controls, even after adjusting for a number of the systemic vascular and fibrotic complications that characterise SSc. This is in line with the work of Kielstein *et al.* and Pulamsetti *et al.*, who showed higher levels of ADMA in iPAH patients compared to healthy controls (18, 19). Our study overcomes some of the limitations of the previous study in SSc patients which reported higher ADMA levels in SSc patients with an sPAP<sub>TTE</sub>  $\geq$ 40mmHg compared with those with a sPAP<sub>TTE</sub> < 40mmHg but did not confirm the presence or absence of PAH with RHC. This concern was further highlighted by the unexpectedly similar DLCO % predicted among cases and controls in that study (61±21 vs. 65±22, *p*=NS), which is unusual given that SSc-PAH is usually associated with a markedly decreased DLCO compared to SSc controls (17). In the wider literature, 'normal'ADMA levels









in healthy subjects have been reported to lie within a narrow range of 0.4 - 0.6 $\mu$ M, with relatively small elevations of ADMA associated with adverse outcomes such as acute coronary events and mortality in patients with coronary artery disease, organ failure and haemodialysis (26-30). In this study, the mean ADMA concentration of  $0.76\pm0.14 \mu$ M in SSc-PAH was clearly outside the published normal range and significantly above the ADMA levels seen in SSc patients without PAH. The ROC curve analysis also gave an optimal ADMA cutpoint of  $\geq 0.7 \ \mu M$  for the identification of SSc-PAH. Taken together, these results suggest that elevated ADMA levels may be associated with SSc-PAH.

ADMA levels were not found to correlate with specific RHC haemodynamics (mPAP, mRAP or PVR). This is not surprising when we consider that measured serum levels of ADMA are very much below the 10µM that would be required to directly affect NO production (15). Instead, it seems more likely that ADMA may act as a 'surrogate' marker of intracellular changes in methylarginine metabolism, with small changes in serum ADMA reflecting greater changes in intracellular ADMA concentrations (31). ADMA is also known to directly influence endothelial cell gene expression, including BMP signalling, which has been associated with iPAH (16). In a population of 57 iPAH patients, Kielstein et al. showed a mild but significant correlation of ADMA with mRAP (r =0.39, p<0.0003), with both ADMA and mRAP independently predicting survival (18). Kielstein et al. directly sampled mixed venous blood from the pulmonary arterial vascular bed at RHC, and this may have enabled the more accurate correlation of ADMA levels Alternately, it may reflect inherent differences in SSc-PAH vs. iPAH, or simply the greater sample size of that study. Nonetheless, an abnormal ADMA level presents an attractive non-invasive biomarker for SSc-PAH, and further studies are required.

In this study, serum measurement of ADMA with a cut-point of 0.7 µM achieved a very good sensitivity and specificity for detection of PAH (86.7% and 90.0%, respectively). However, a high sensitivity is crucial in screening for SSc-PAH, and in order to improve this further, ADMA was combined with NT-proBNP. NT-proBNP is a 76 amino-acid polypeptide that is released by cardiac myocytes in response increased ventricular wall stress, as typically occurs with volume overload and ventricular contractile dysfunction (32). NTproBNP is a well-studied candidate biomarker for SSc-PAH, demonstrating utility in the screening, diagnosis, and prognosis of SSc-PAH (23, 33, 34). One of factors impeding the wider use of NT-proBNP in the screening and diagnosis of SSc-PAH has been the lack of sufficiently high sensitivity as a 'stand-alone' test (35, 36). In order to improve the sensitivity of these biomarkers, ADMA and NT-proBNP were combined. This 'biomarker-based' model achieved an

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Covariate	OR of ADMA for PAH	95% CI	<i>p</i> -value	
Age (y)	1.14	1.03, 1.25	0.008	
Disease duration (y)	1.16	1.05, 1.27	0.003	
Disease subtype	1.18	1.07, 1.30	0.001	
Hypertension	1.16	1.06, 1.28	0.002	
High Cholesterol	1.15	1.04, 1.26	0.004	
Smoking status	1.17	1.06, 1.29	0.002	
MRSS	1.17	1.06, 1.29	0.001	
BMI	1.15	1.04, 1.26	0.006	
Raynaud's disease	1.18	1.07, 1.29	0.001	
Digital ulceration	1.19	1.07, 1.32	0.001	
Calcium channel blocker us	le 1.19	1.07-1.31	0.001	

\*OR are reported per 0.01µM increase in ADMA. In each logistic regression model, there were only two independent variables: ADMA and each of the covariates in turn.

ADMA: asymmetric dimethylarginine; PAH: pulmonary arterial hypertension; OR: odds ratio; CI: confidence interval; y: years; MRSS: modified Rodnan skin score; BMI: body mass index.

Table IV. Independent associations of SDMA with PAH, after adjustment for covariates.

Covariate	OR of SDMA for PAH	95% CI	<i>p</i> -value	
Age (y)	1.12	1.01, 1.25	0.025	
Disease duration (y)	1.16	1.04, 1.29	0.008	
Disease subtype	1.17	1.06, 1.30	0.001	
Hypertension	1.17	1.05, 1.31	0.005	
High cholesterol	1.16	1.05, 1.29	0.005	
Smoking status	1.15	1.05, 1.28	0.004	
MRSS	1.17	1.06, 1.29	0.002	
BMI	1.16	1.04, 1.29	0.007	
Raynaud's disease	1.17	1.06, 1.29	0.002	
Digital ulceration	1.19	1.06, 1.32	0.002	
Calcium channel blocker u	se 1.18	1.06, 1.31	0.004	

\*OR are reported per  $0.01\mu$ M increase in SDMA. In each logistic regression model, there were only two independent variables: SDMA and each of the covariates in turn.

SDMA: symmetric dimethylarginine; PAH: pulmonary arterial hypertension; OR: odds ratio; CI: confidence interval; y: years; MRSS: modified Rodnan skin score; BMI: body mass index.

excellent sensitivity and specificity (100% and 90%, respectively) for the detection of SSc-PAH. Thus, it may be possible to screen for SSc-PAH using a non-invasive combination biomarker model using NT-proBNP coupled with ADMA. This finding clearly needs to be validated in a larger prospective cohort of SSc-PAH.

Interestingly, raised SDMA levels were found in SSc-PAH. While the role of SDMA in endothelial dysfunction is uncertain, it has been hypothesised that SDMA may indirectly inhibit NO synthesis by interfering with arginine uptake through the inhibition of the human cationic amino acid transporter hCAT-2B (37). Raised SDMA levels have also been previously reported in iPAH (19). While SDMA performed well in this study as a stand-alone biomarker the sensitivity of SDMA for the detection of PAH was less than that of ADMA; furthermore, SDMA did not improve the performance of NTproBNP. More studies are required to confirm these findings, and the possible role SDMA may have as a biomarker for PAH.

This study has a number of strengths. Firstly, PAH was defined according to internationally accepted RHC criteria. Secondly, ADMA levels were assayed in newly diagnosed SSc-PAH patients, before the commencement of advanced pulmonary vasodilator therapies. Thirdly, accurate and validated techniques of ADMA measurement were employed using HPLC. Lastly, clinically relevant ADMA cut-points were determined in isolation, and in combination with NTproBNP, which may enable clinicians to non-invasively identify patients who should undergo further testing for PAH. This study has some important limitations. The study was small and had an observational case-control design. Patients with significant left ventricular and renal dysfunction were excluded, as both of these conditions have been associated with raised ADMA levels. Therefore, larger studies inclusive of these population subsets would provide further information about the strengths and limitations of ADMA in identifying patients with SSc-PAH. Furthermore, prospective, sequential sampling of ADMA levels, particularly in patients developing PAH, would help better understand the utility of ADMA levels in at risk individuals, as well as providing valuable information about the stability of ADMA over time and the response to therapy.

In summary, this study demonstrates the association of increased ADMA levels with SSc-PAH and the potential utility of ADMA in the screening and diagnosis of SSc-PAH. This has important clinical implications as noninvasive, biomarker-based screening models may enable the convenient and accurate identification of SSc patients who require further diagnostic evaluation for the presence of PAH.

#### Acknowledgements

We wish to thank the following people for assistance with sample preparation and data collection: Alison Batty, Kerry Cooper, Kathleen Elford, Dot Fowler, Barbara Gemmell, Trish Lewis, Helen Marsden, Leah McWilliams and Bev Wilson. We wish to thank the patients who participated in this study and St Vincent's Hospital Melbourne IT department for their expert advice and continued support of the Australian Scleroderma Cohort Study and Database.

#### Funding

V. Thakkar was funded by a National Health and Medical Research Council of Australia Scholarship (APP1038612) and The Australian Scleroderma Interest Group Clinical Research Fellowship. M. Nikpour holds a National Health and Medical Research Council of Australia Early Career Fellowship (APP1071735) and is supported by a David Bickart Clinician Research Award from the University of Mel-

#### Table V. ROC analysis for PAH compared to controls.

Variable	Optimal cutpoint	Sensitivity (95% CI)	Specificity (95% CI)	+LR	-LR	AUC
ADMA (µM)	≥0.7	86.7% (58.4%, 97.7%)	90.0% (72.3%-97.4%)	8.7 (2.9-25.8)	0.15 (0.04-0.54)	0.86 (0.70, 1.0)
SDMA (µM)	≥0.621	73.3% (44.8%, 91.1%)	100.0% (85.6%, 99.7%)	-	0.27 (0.12-0.62)	0.88 (0.74-1.0)
ADMA ≥0.7 &/or SDMA ≥0.621	-	86.7% (58.4, 97.7%)	90% (72.3%, 97.4%)	8.7 (2.9-25.8)	0.15 (0.04, 0.54)	-
SDMA ≥0.621 &/or NTproBNP ≥210	-	93.3% (66.0%, 99.7%)	100% (85.9%, 99.7%)	-	0.07 (0.01, 0.44)	-
ADMA ≥0.7 &/or NTproBNP ≥210	-	100% (74.7%, 99.4%)	90% (72.3%, 97.4%)	10.0 (3.4-29.3)	-	-

ADMA: asymmetric dimethylarginine; SDMA: symmetrical dimethylarginine (SDMA); NT-proBNP: N-terminal pro-B-type natriuretic peptide; CI: confidence interval; LR: likelihood ratio; AUC: area under the curve.

bourne Faculty of Medicine, Dentistry and Health Sciences. Other sources of funding include Scleroderma Australia, Arthritis Australia, St. Vincent's Research Endowment Foundation, Pfizer (including a Pfizer CardioVascular Lipid research grant), Actelion Australia, Bayer, CSL Biotherapies and Glaxo-SmithKline Australia and BMS.

#### **Competing interests**

V. Thakkar has received honoraria from Actelion Australia and an unrestricted educational grant on behalf of the Department of Rheumatology, Liverpool Hospital. E. Gabbay has received honoraria for speaking at the Actelion Excellence Programme Annual Scientific Meeting, and research support. S.M. Proudman has received honoraria and/or research grants from Actelion Pharmaceuticals Australia and Bayer. M. Nikpour has received honoraria for presentations at meetings from Actelion Pharmaceuticals and grants/research support from Actelion, Pfizer and BMS. All the other authors have delared no cometing interests.

#### References

- STEEN VD, MEDSGER TA: Changes in causes of death in systemic sclerosis, 1972-2002. *Ann Rheum Dis* 2007; 66: 940-4.
- TYNDALL AJ, BANNERT B, VONK M et al.: Causes and risk factors for death in systemic sclerosis: a study from the EULAR Scleroderma Trials and Research (EUSTAR) database. Ann Rheum Dis 2010; 69: 1809-15.
- HACHULLA E, CARPENTIER P, GRESSIN V et al.: Risk factors for death and the 3-year survival of patients with systemic sclerosis: the French ItinerAIR-Sclerodermie study. *Rheu*matology 2008; 48: 304-8.

- KHANNA D, TAN M, FURST DE et al.: Recognition of pulmonary hypertension in the rheumatology community: lessons from a quality enhancement research initiative. Clin Exp Rheumatol 2014; 32 (Suppl. 86): S21-7.
- HUMBERT M, YAICI A, DE GROOTE P, et al.: Screening for pulmonary arterial hypertension in patients with systemic sclerosis: Clinical characteristics at diagnosis and long-term survival. Arthritis Rheum 2011; 63: 3522-30.
- Consensus statement on the management of pulmonary hypertension in clinical practice in the UK and Ireland. *Heart* 2008; 94 (Suppl. 1): i1-41.
- GALIE N, HOEPER MM, HUMBERT M et al.: Guidelines for the diagnosis and treatment of pulmonary hypertension: The Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS), endorsed by the International Society of Heart and Lung Transplantation (ISHLT). Eur Heart J 2009; 30: 2493-537.
- MCLAUGHLIN V, HUMBERT M, COGHLAN G, NASH P, STEEN V: Pulmonary arterial hypertension: the most devastating vascular complication of systemic sclerosis. *Rheumatol*ogy (Oxford) 2009; 48 (Suppl. 3): iii25-31.
- MUKERJEE D, ST GEORGE D, KNIGHT C et al.: Echocardiography and pulmonary function as screening tests for pulmonary arterial hypertension in systemic sclerosis. *Rheuma*tology (Oxford) 2004; 43: 461-6.
- PEACOCK A: Prevention and early diagnosis of pulmonary hypertension. In: Pulmonary Vascular Pathology: a Clinical Update. DEMEDTS M, DELCROIX M, VERHAEGE R, VERLEDEN GM (Eds.) vol. 27. Sheffield: European Respiratory Society Monograph; 2003: 227-42.
- 11. THAKKAR V, STEVENS WM, PRIOR D et al.: N-terminal pro-brain natriuretic peptide in a novel screening algorithm for pulmonary arterial hypertension in systemic sclerosis: a case-control study. Arthritis Res Ther 2012; 14: R143.
- 12. THAKKAR V, STEVENS W, PRIOR D et al.: The inclusion of N-terminal pro-brain natriuretic peptide in a sensitive screening strategy for systemic sclerosis-related pulmonary ar-

terial hypertension: a cohort study. *Arthritis Res Ther* 2013; 15: R193.

- 13. NINABER MK, HAMERSMA WB, SCHOUFFO-ER AA et al.: Detection of pulmonary vasculopathy by novel analysis of oxygen uptake in patients with systemic sclerosis: association with pulmonary arterial hypertension. Clin Exp Rheumatol 2014; 32 (Suppl. 86): S60-7.
- 14. ZAKRZEWICZ D, EICKELBERG O: From arginine methylation to ADMA: a novel mechanism with therapeutic potential in chronic lung diseases. *BMC Pulm Med* 2009; 9: 5.
- DWEIK RA: The lung in the balance: arginine, methylated arginines, and nitric oxide. *Am J Physiol Lung Cell Mol Physiol* 2007; 292: L15-17.
- 16. SMITH CL, ANTHONY S, HUBANK M, LEIPER JM, VALLANCE P: Effects of ADMA upon gene expression: an insight into the pathophysiological significance of raised plasma ADMA. PLoS Med 2005; 2: e264.
- DIMITROULAS T, GIANNAKOULAS G, SFET-SIOS T *et al.*: Asymmetrical dimethylarginine in systemic sclerosis-related pulmonary arterial hypertension. *Rheumatology* (Oxford) 2008; 47: 1682-5.
- KIELSTEIN JT, BODE-BOGER SM, HESSE G et al.: Asymmetrical dimethylarginine in idiopathic pulmonary arterial hypertension. Arterioscler Thromb Vasc Biol 2005; 25: 1414-8.
- PULLAMSETTI S, KISS L, GHOFRANI HA et al.: Increased levels and reduced catabolism of asymmetric and symmetric dimethylarginines in pulmonary hypertension. Faseb J 2005; 19: 1175-7.
- 20. SKORO-SAJER N, MITTERMAYER F, PANZEN-BOECK A *et al.*: Asymmetric Dimethylarginine Is Increased in Chronic Thromboembolic Pulmonary Hypertension. *Am J Resp Crit Care Med* 2007; 176: 1154-60.
- 21. LANDBURG PP, TEERLINK T, VAN BEERS EJ et al.: Association of asymmetric dimethylarginine with sickle cell disease-related pulmonary hypertension. *Haematologica* 2008; 93: 1410-2.
- 22. VAN DEN HOOGEN F, KHANNA D, FRANSEN J et al.: 2013 classification criteria for systemic sclerosis: an American college of rheumatology/European league against rheumatism collaborative initiative. Ann Rheum Dis 2013; 72: 1747-55.

- 23. THAKKAR V, STEVENS WM, PRIOR D *et al.*: N-terminal pro-brain natriuretic peptide in a novel screening algorithm for pulmonary arterial hypertension in systemic sclerosis: a case-control study. *Arthritis Res Ther* 2012; 14: R143.
- 24. MACINTYRE N, CRAPO RO, VIEGI G et al.: Standardisation of the single-breath determination of carbon monoxide uptake in the lung. Eur Respir J 2005; 26: 720-35.
- 25. HERESZTYN T, WORTHLEY MI, HOROWITZ JD: Determination of l-arginine and NG, NG and NG, NG' -dimethyl-L-arginine in plasma by liquid chromatography as AccQ-Fluor fluorescent derivatives. J Chromatogr B Analyt Technol Biomed Life Sci 2004; 805: 325-9.
- 26. HOROWITZ JD, HERESZTYN T: An overview of plasma concentrations of asymmetric dimethylarginine (ADMA) in health and disease and in clinical studies: methodological considerations. J Chromatogr B Analyt Technol Biomed Life Sci 2007; 851: 42-50.
- 27. VALLANCE P, LEONE A, CALVER A, COLLIER J, MONCADA S: Accumulation of an endogenous inhibitor of nitric oxide synthesis in

chronic renal failure. *Lancet* 1992; 339: 572-5.

- VALKONEN VP, PAIVA H, SALONEN JT *et al.*: Risk of acute coronary events and serum concentration of asymmetrical dimethylarginine. *Lancet* 2001; 358: 2127-8.
- 29. SCHNABEL R, BLANKENBERG S, LUBOS E *et al.*: Asymmetric dimethylarginine and the risk of cardiovascular events and death in patients with coronary artery disease: results from the AtheroGene Study. *Circ Res* 2005; 97: e53-9.
- 30. NIJVELDT RJ, TEERLINK T, VAN DER HOVEN B *et al.*: Asymmetrical dimethylarginine (ADMA) in critically ill patients: high plasma ADMA concentration is an independent risk factor of ICU mortality. *Clin Nutr* 2003; 22: 23-30.
- ZAKRZEWICZ D, EICKELBERG O: From arginine methylation to ADMA: A novel mechanism with therapeutic potential in chronic lung diseases. *BMC Pulm Med* 2009; 9: 5.
- 32. BRAUNWALD E: Biomarkers in heart failure. *N Engl J Med* 2008; 358: 2148-59.
- 33. DIMITROULAS T, GIANNAKOULAS G,

KARVOUNIS H, GATZOULIS MA, SETTAS L: Natriuretic peptides in systemic sclerosisrelated pulmonary arterial hypertension. *Sem Arthritis Rheum* 2010; 39: 278-84.

- 34. THAKKAR V, STEVENS W, PRIOR D et al.: The inclusion of N-terminal pro-brain natriuretic peptide in a sensitive screening strategy for systemic sclerosis-related pulmonary arterial hypertension: a cohort study. Arthritis Res Ther 2013; 15: R193.
- 35. MUKERJEE D, YAP LB, HOLMES AM *et al.*: Significance of plasma N-terminal pro-brain natriuretic peptide in patients with systemic sclerosis-related pulmonary arterial hypertension. *Respir Med* 2003; 97: 1230-6.
- 36. WILLIAMS MH, HANDLER CE, AKRAM R et al.: Role of N-terminal brain natriuretic peptide (N-TproBNP) in scleroderma-associated pulmonary arterial hypertension. Eur Heart J 2006; 27: 1485-94.
- 37. CLOSS EI, BASHA FZ, HABERMEIER A, FOR-STERMANN U: Interference of L-arginine analogues with L-arginine transport mediated by the y+ carrier hCAT-2B. *Nitric Oxide* 1997; 1: 65-73.